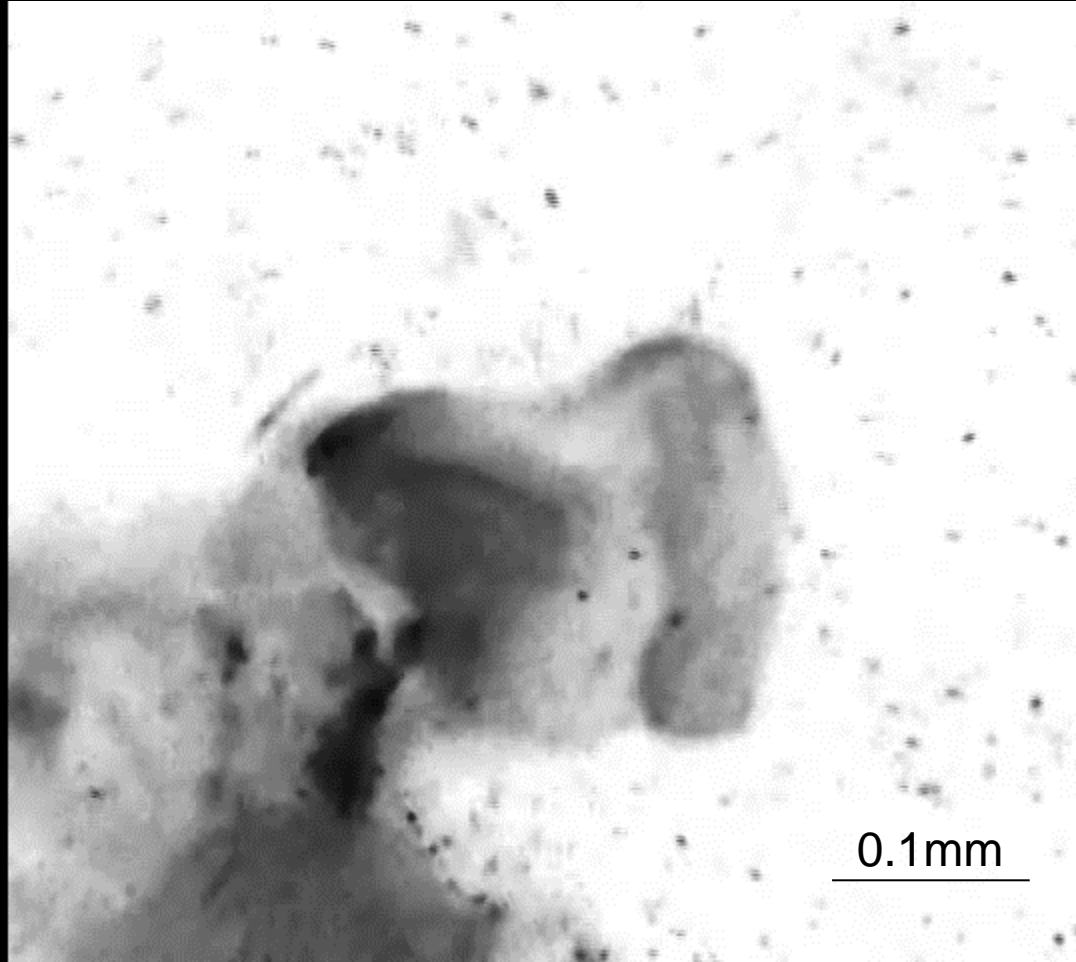


Cilia-driven flow fields for transport and selective capture of bacteria



Janna Nawroth, Ph.D.

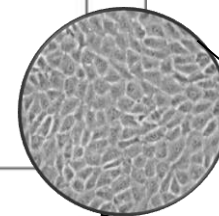
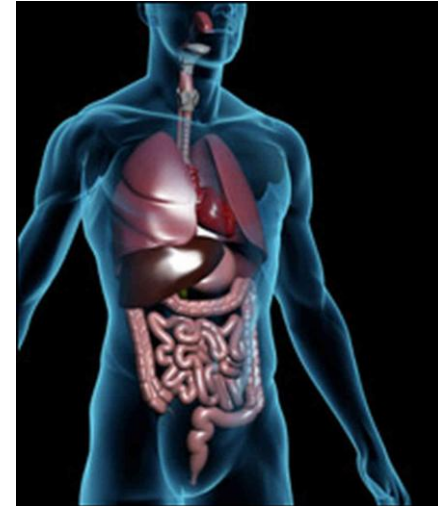
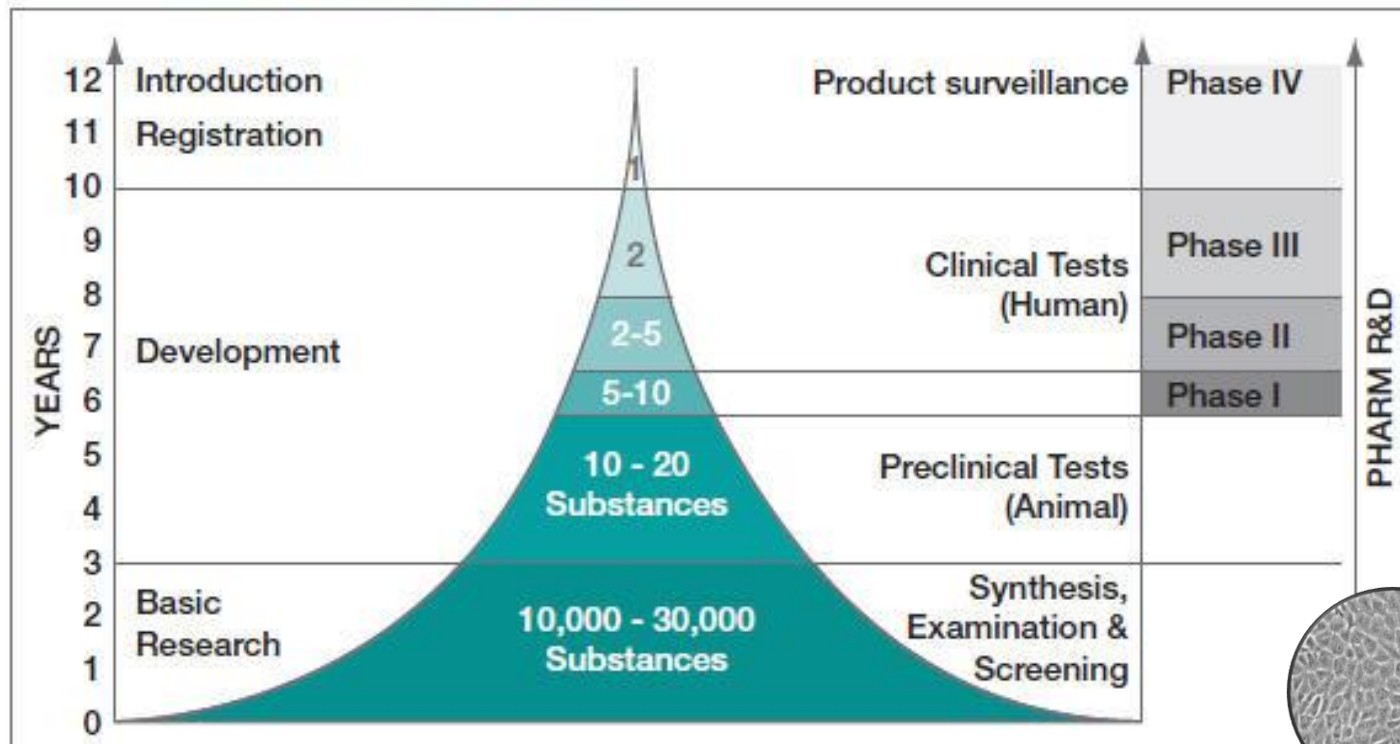
Wyss Institute for Biologically Inspired Engineering, Harvard University
Active Matter | Kavli Institute for Theoretical Physics | May 15th, 2014



Traditional drug development

Time: 10 years
Cost: \$1 Billion/drug

Figure 1: The drug development pyramid

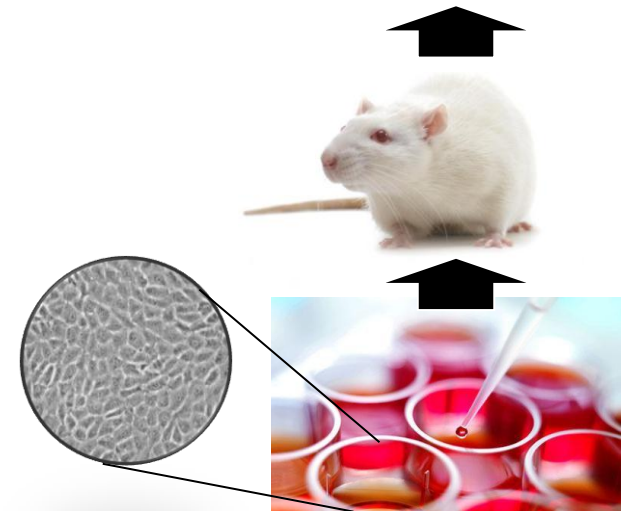
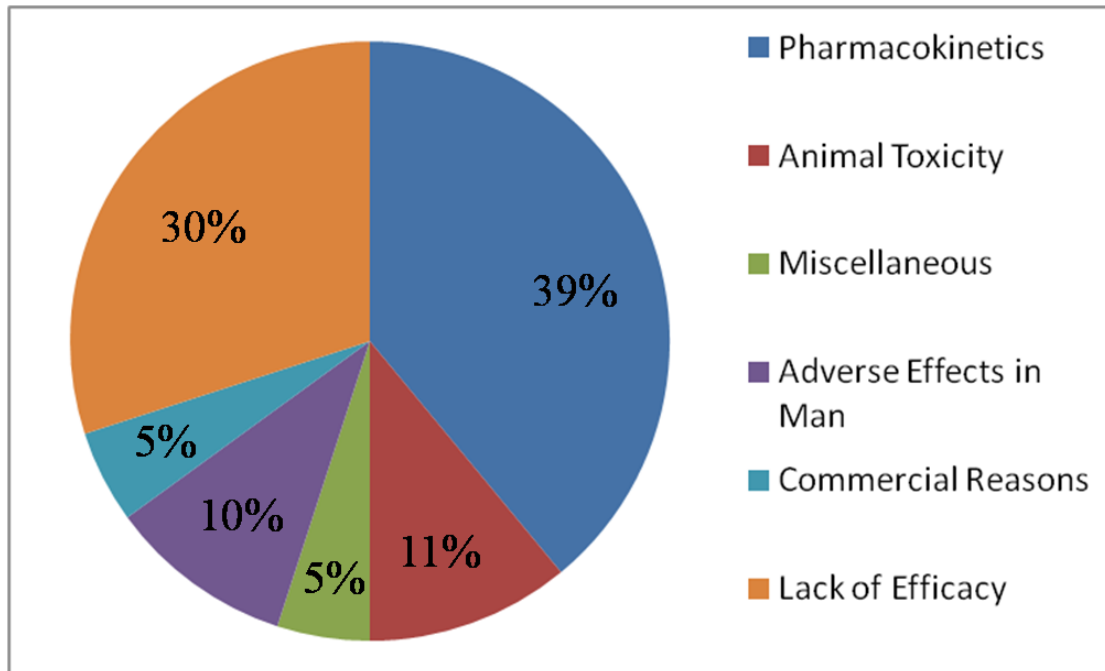




Traditional drug development

Time: 10 years
Cost: \$1 Billion/drug

Reasons for drug failure: poor translation between test platforms and humans





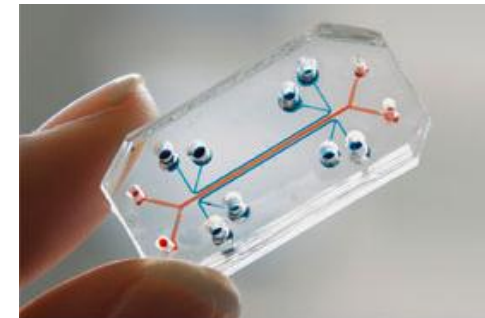
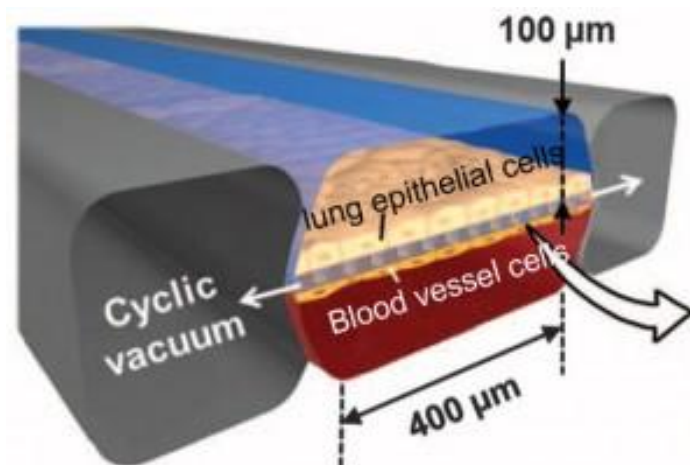
The quest for a better *in-vitro* model of human disease and drug response

Organ-on-a-chip

- 3-D microfluidic cell culture chip from human cells
- simulates characteristic mechanics and physiological responses of entire organs

→ “human” platform for drug testing and disease models

Example: lung-blood barrier function





Organ-on-a-chip design

Biology

Identify relevant structure-function relationships

Structure and motion of building blocks (cells, matrix)



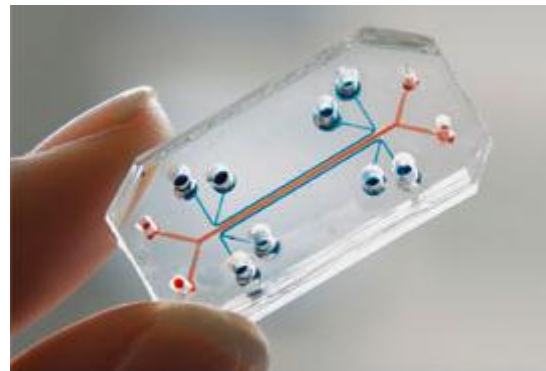
Emergent functions



Quantitative metrics of organ fitness

Engineering

Design, build and test structure-function relationships





Organ-on-a-chip design

Biology

Identify relevant structure-function relationships

Structure and motion of building blocks (cells, matrix)



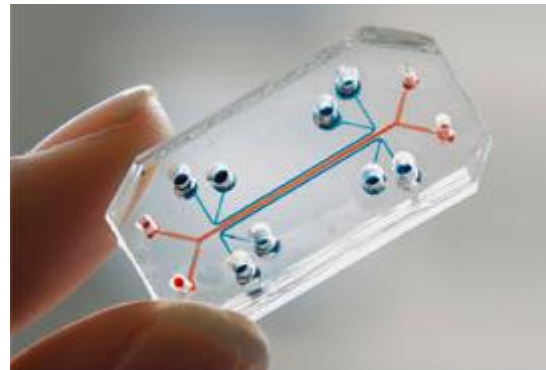
Emergent functions



Quantitative metrics of organ fitness

Engineering

Design, build and test structure-function relationships



Example lung chip

Metrics of fitness are

- Structural integrity
- Absorption air → blood



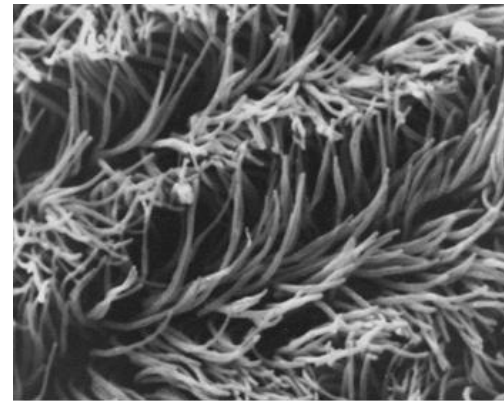
2 case studies

Muscle-powered fluid transport ...relevant to cardiovascular system



with
John Dabiri (Caltech)
Kit Parker (Harvard)
Donald Ingber (Harvard)

Cilia-powered fluid transport ... relevant to respiratory organs, brain, Fallopian tube

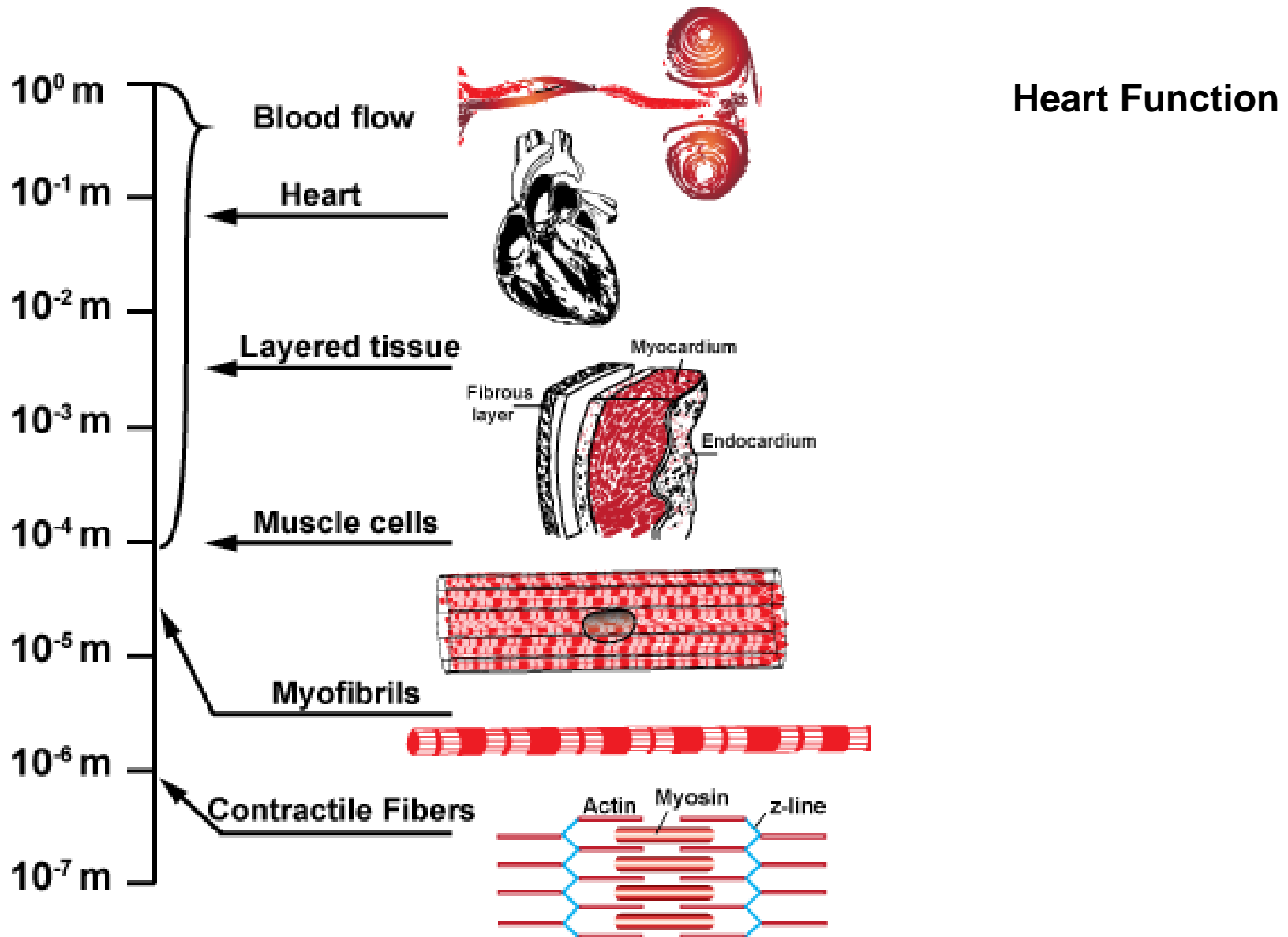


2 μ m

with
Eva Kanso (USC)
Margaret McFall-Ngai (UW Madison)
Edward Ruby (UW Madison)
John Dabiri (Caltech)
Scott Fraser (USC)

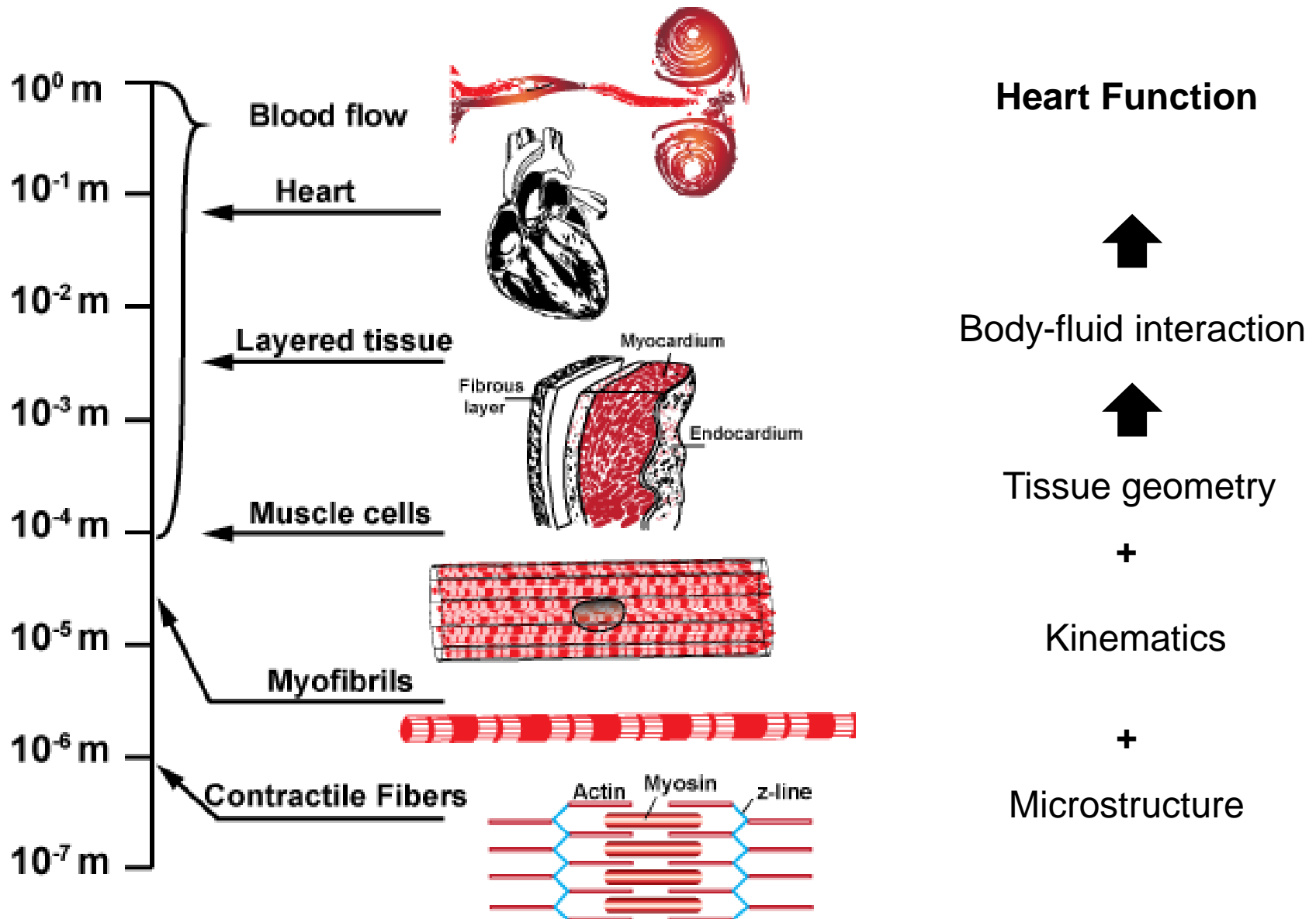


Case study: human heart





Case study: human heart



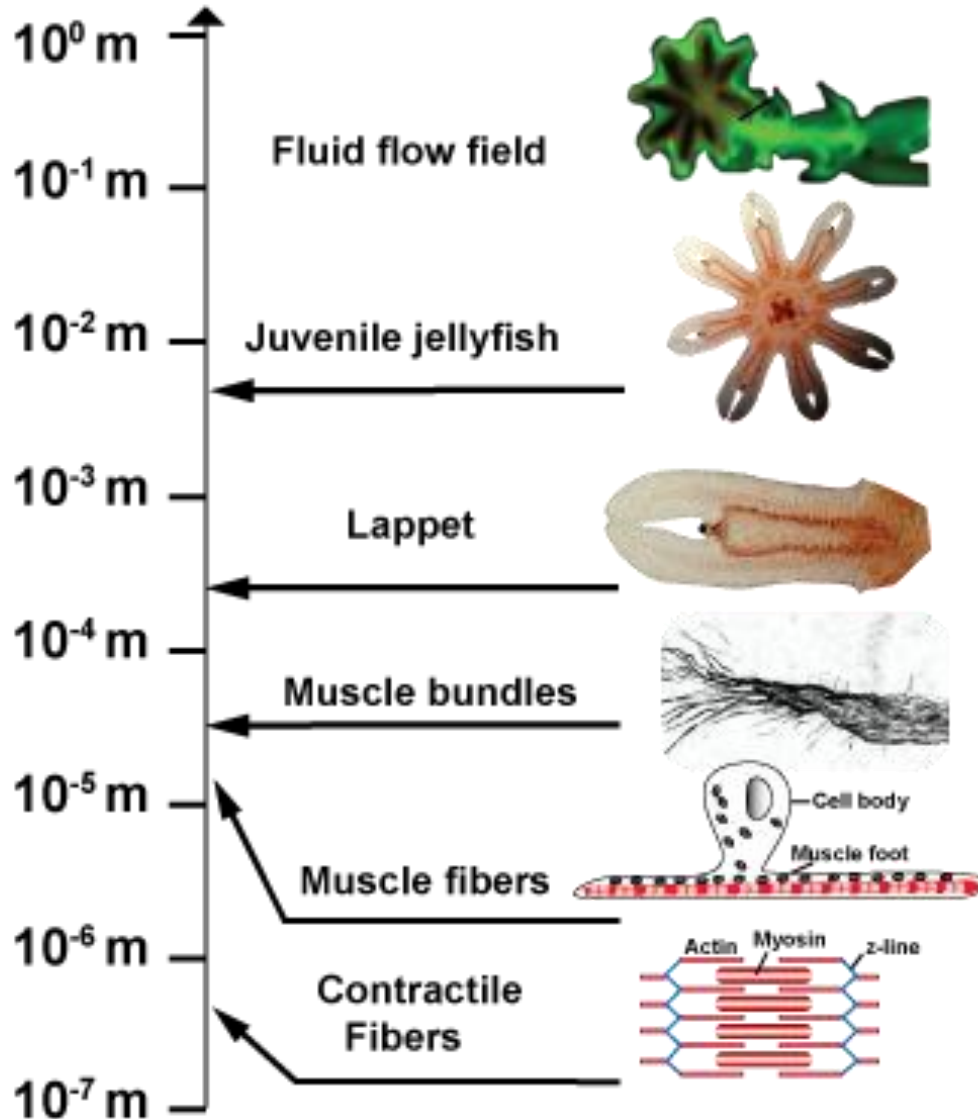


A simplified heart: jellyfish





A simplified heart: jellyfish



Jellyfish Function

Body-fluid interaction

Tissue geometry

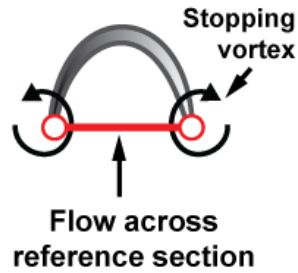
Kinematics

Microstructure

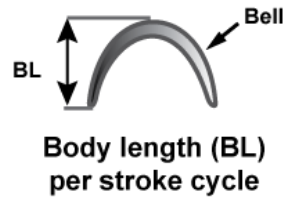


Measuring jellyfish fitness

Feeding metric



Propulsion metric

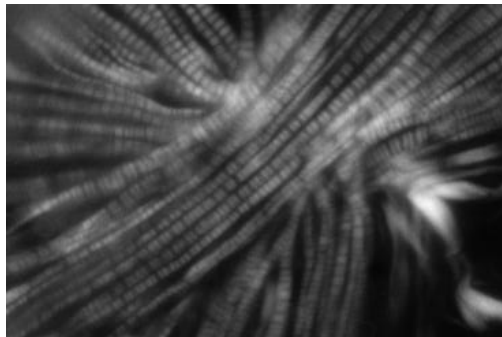


Jellyfish function

- Feeding flux
- Propulsion



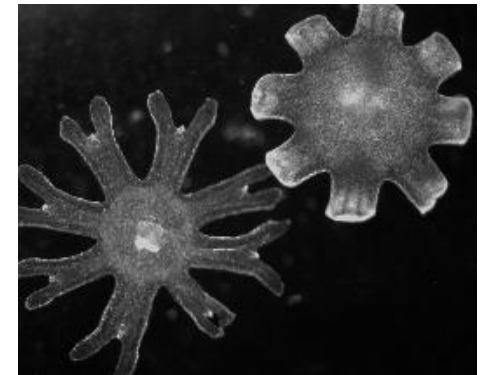
Fluid transport



Muscle fiber alignment



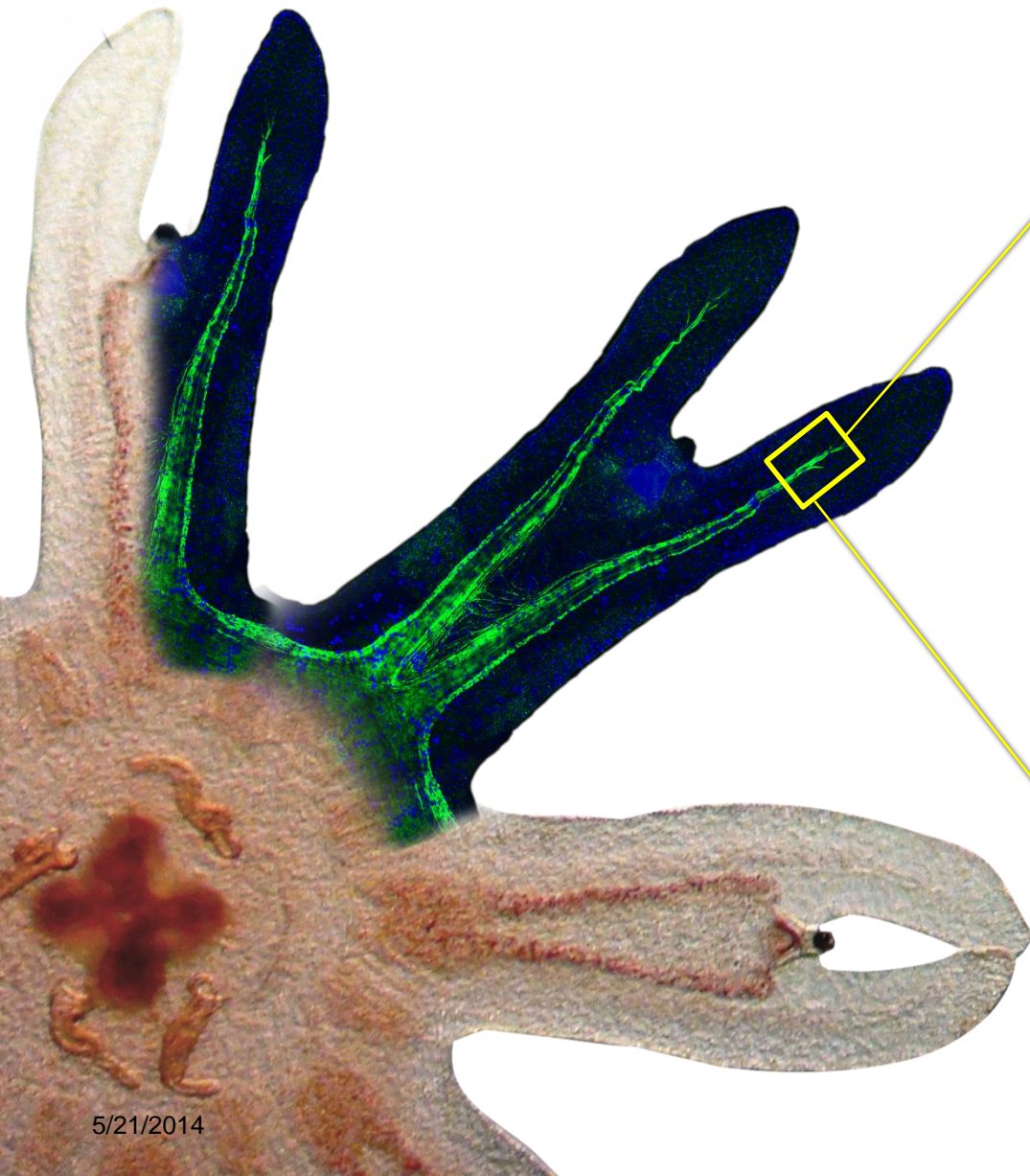
Contraction kinematics



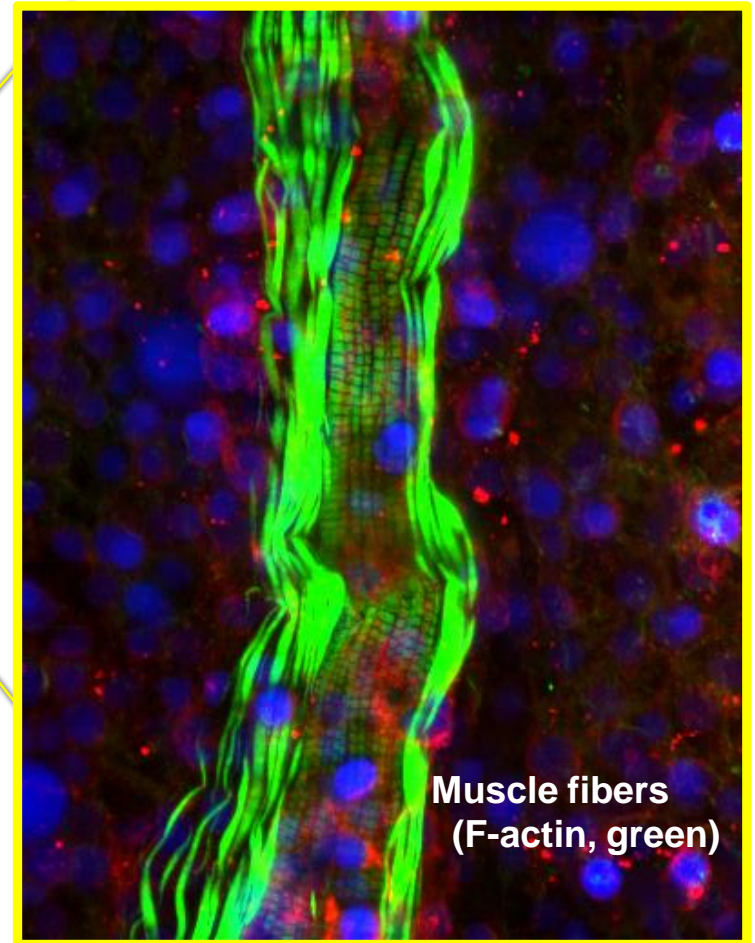
Bell geometry



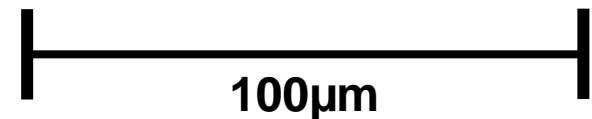
Muscle fiber alignment



Anisotropic muscle layout

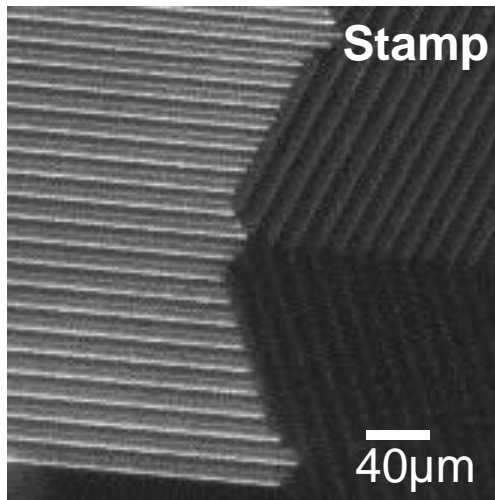
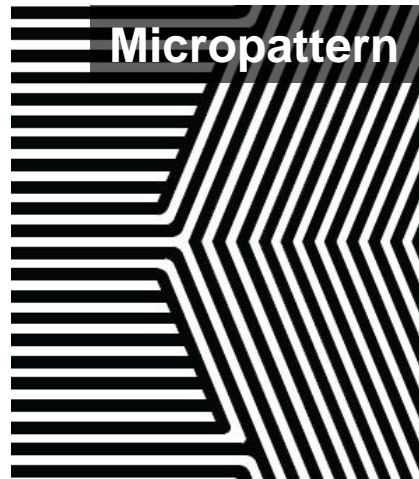


Muscle fibers
(F-actin, green)

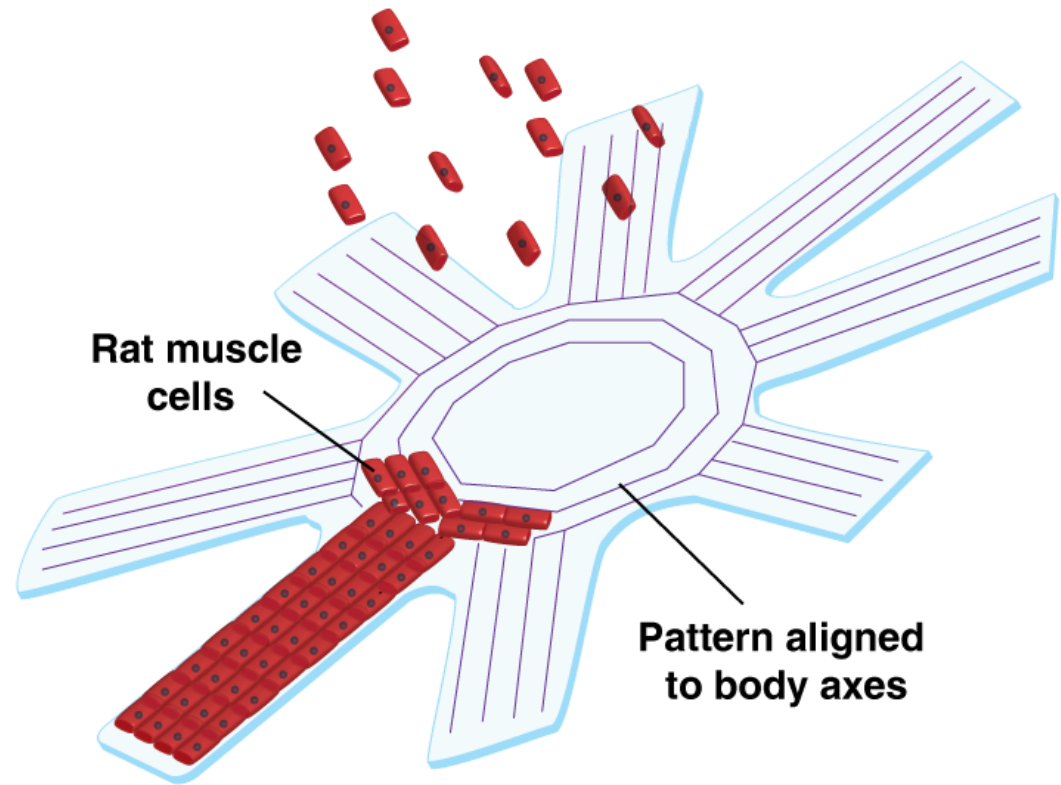




Muscle fiber alignment

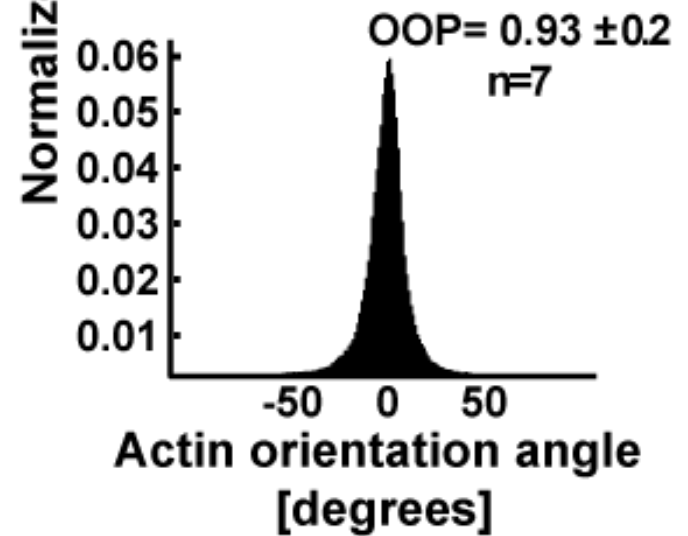
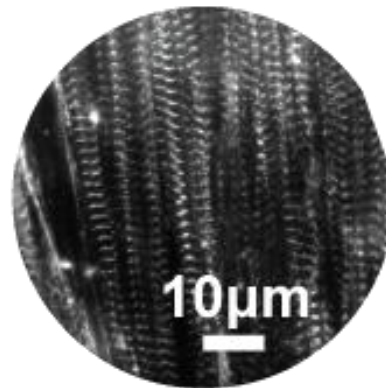
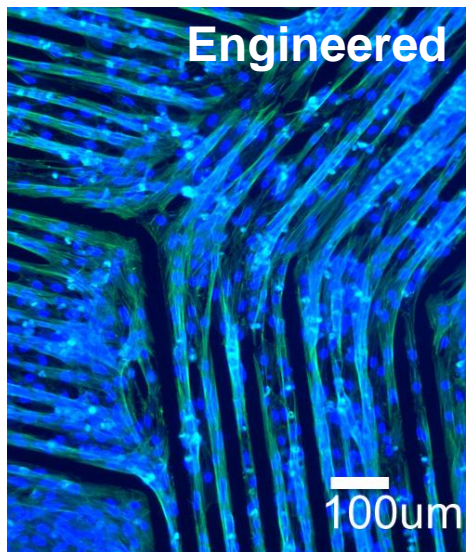
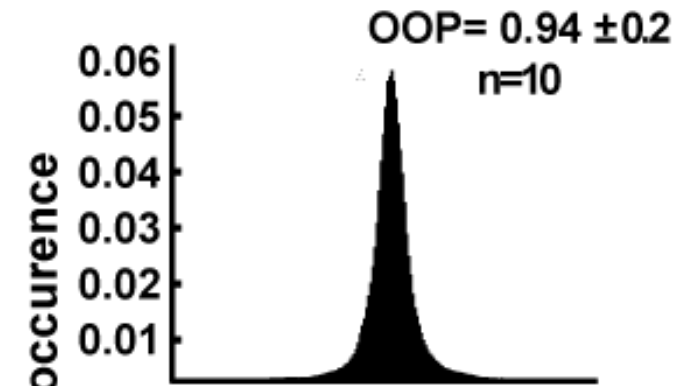
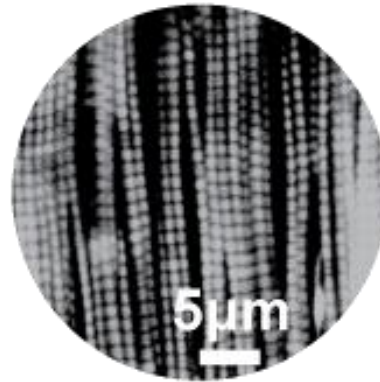
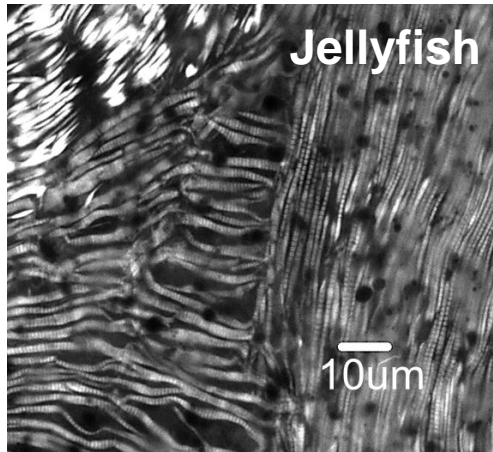


Anisotropic muscle layout

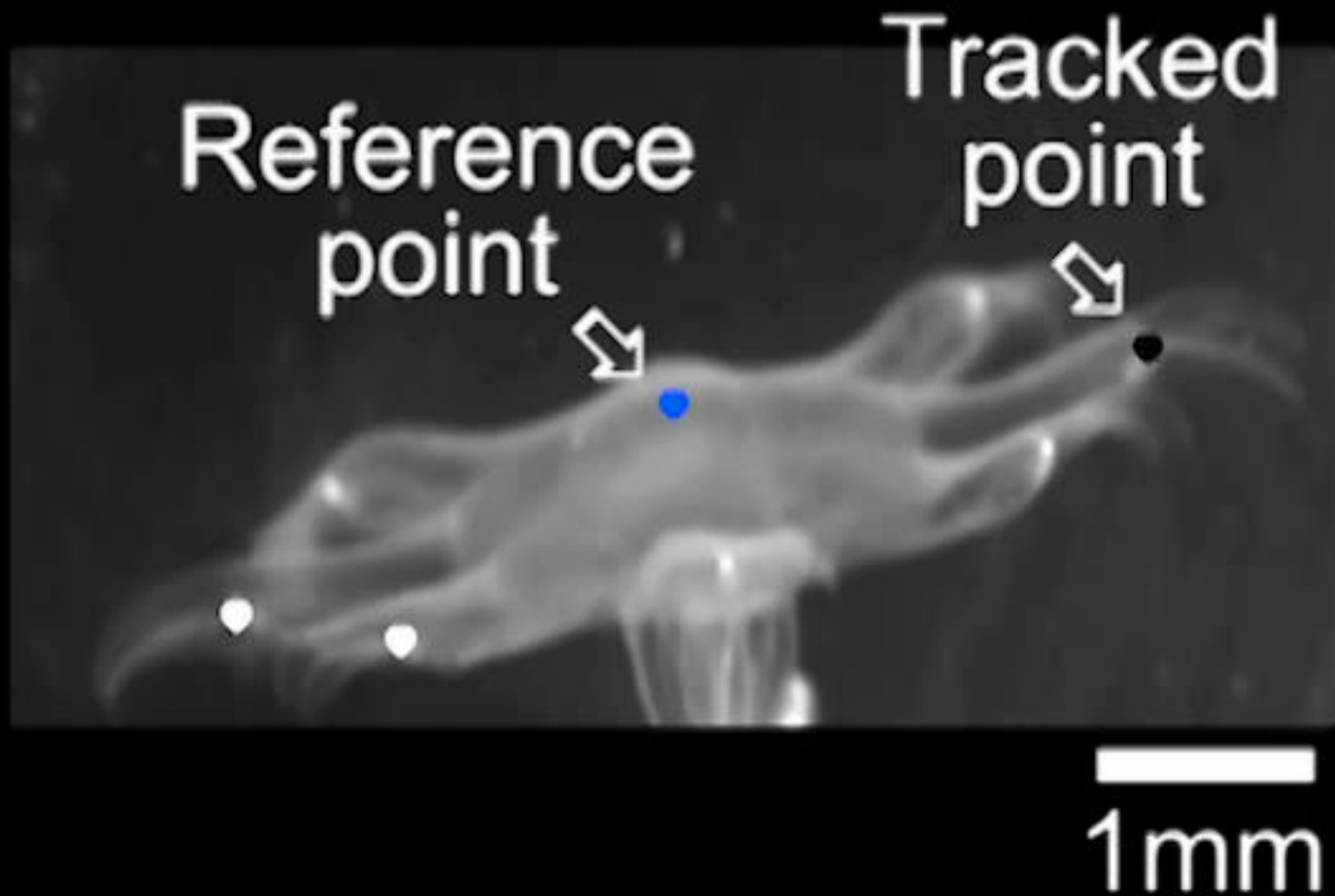




Muscle fiber alignment



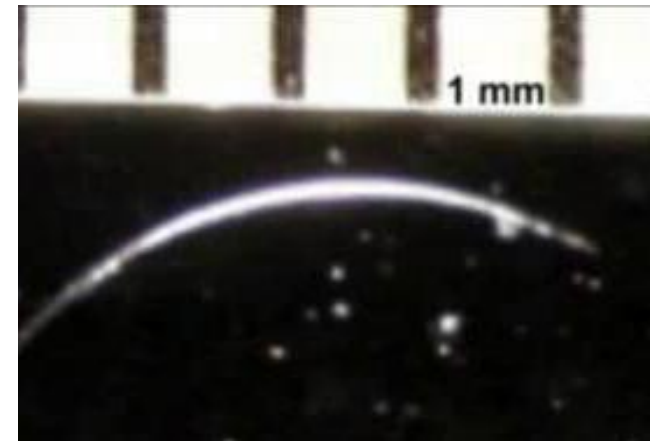
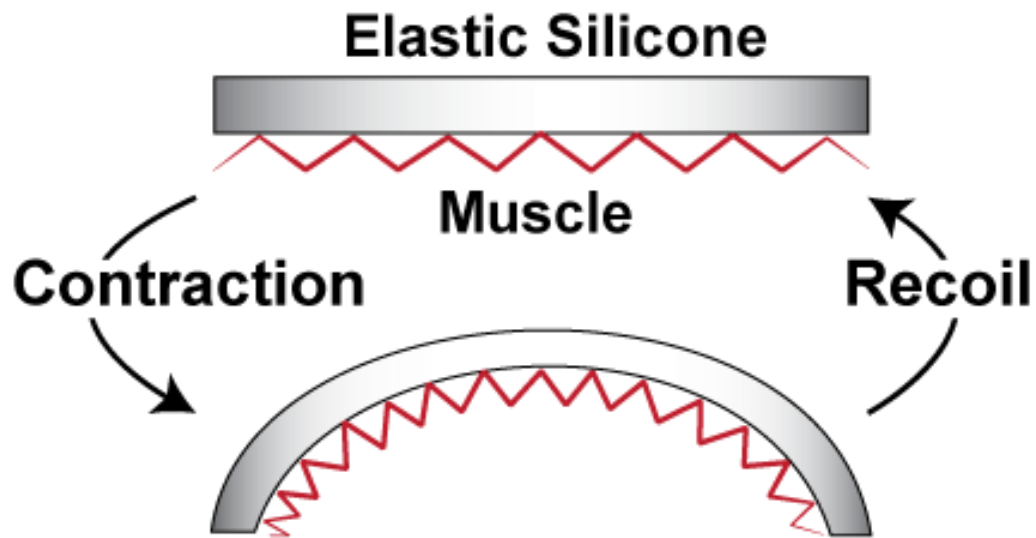
Bell kinematics





Bell kinematics

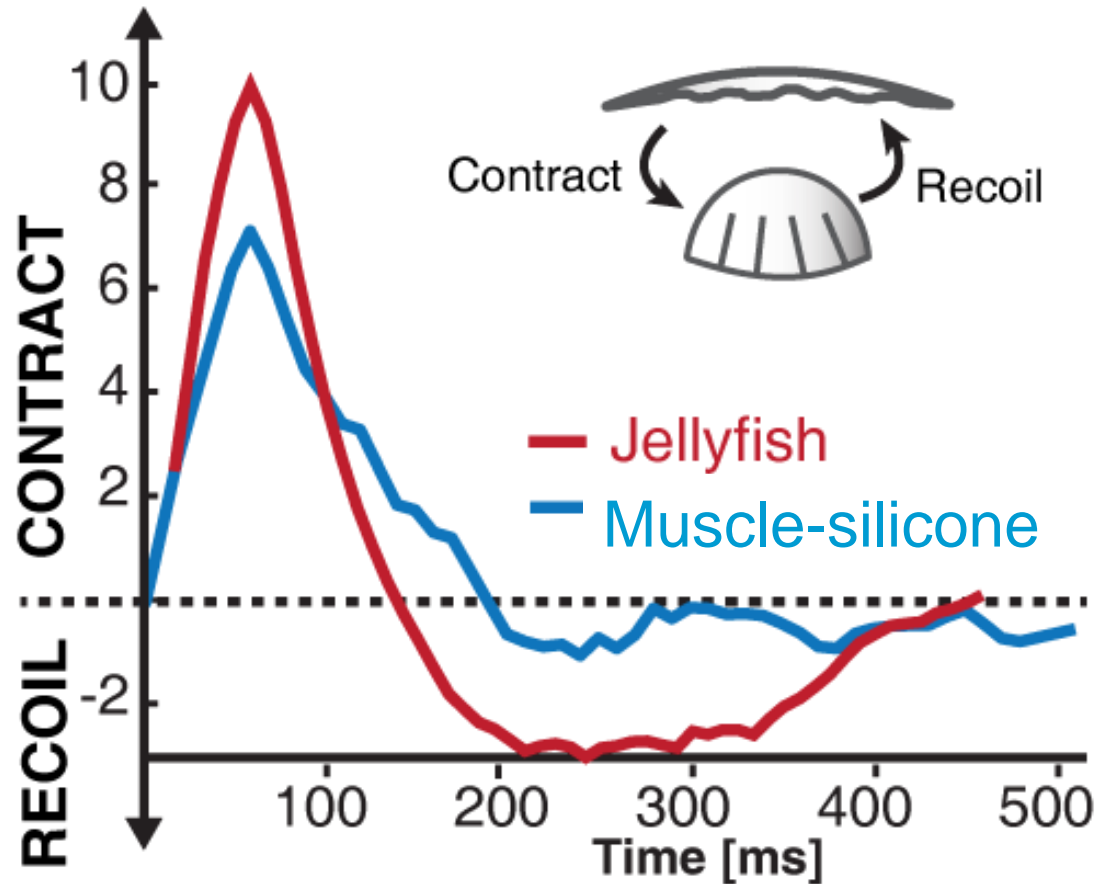
Muscle-elastomer composite





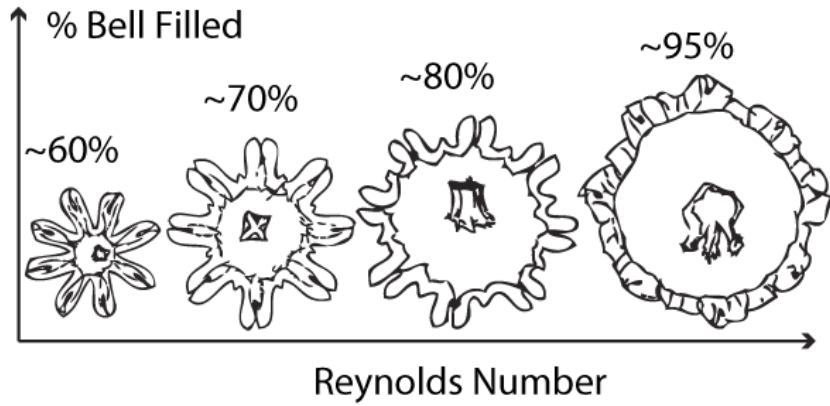
Bell kinematics

Angular velocity [r/sec]





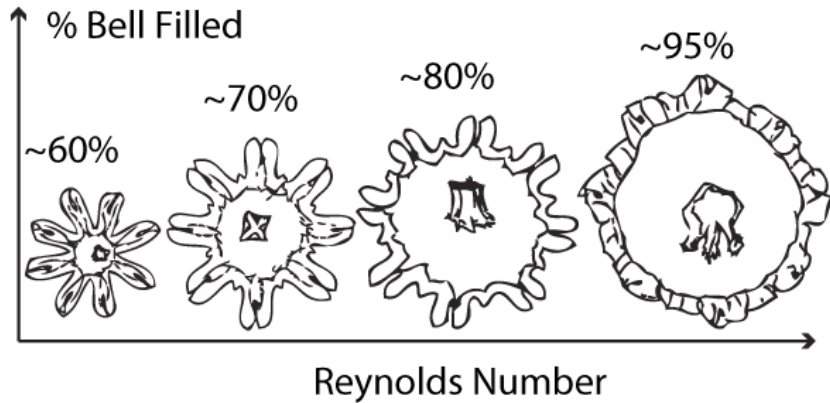
Body geometry



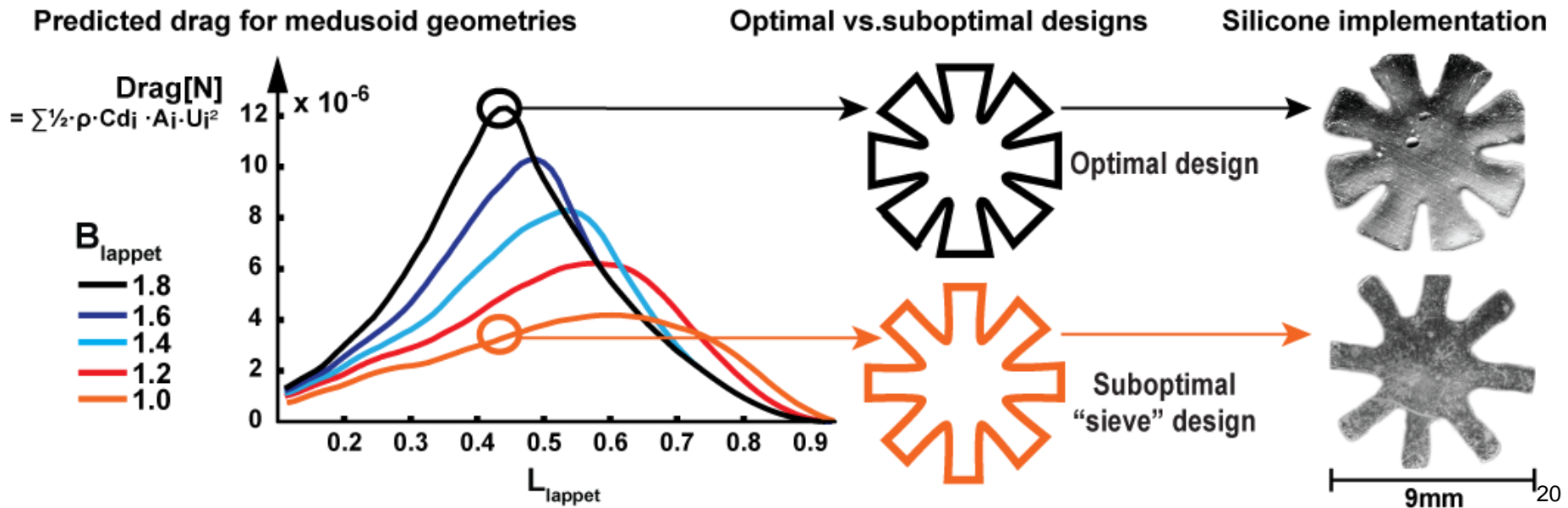
Feitl, 2009; Nawroth, 2010



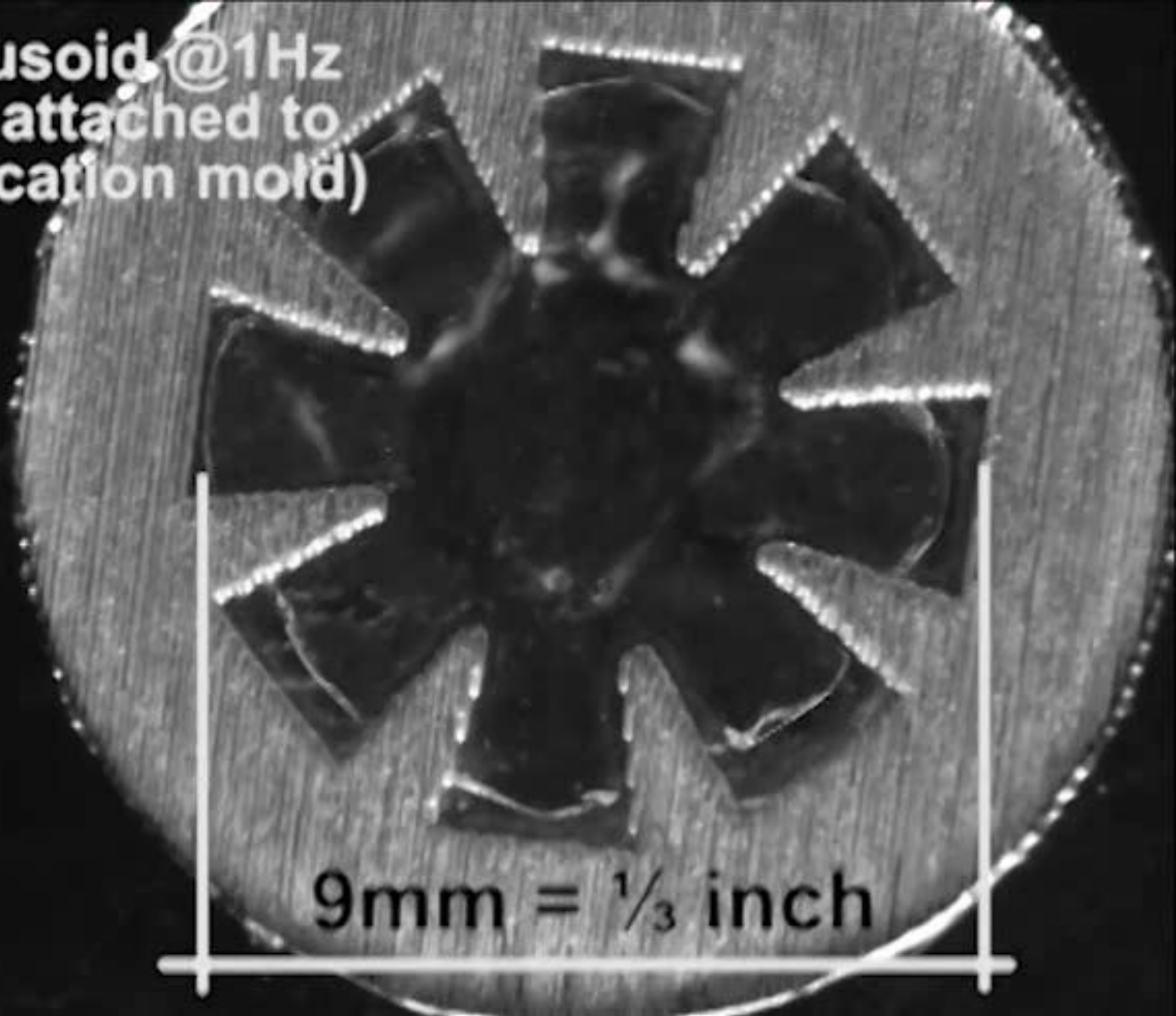
Body geometry



Feitl, 2009; Nawroth, 2010



Medusoid @ 1Hz
(still attached to
fabrication mold)



9mm = $\frac{1}{3}$ inch

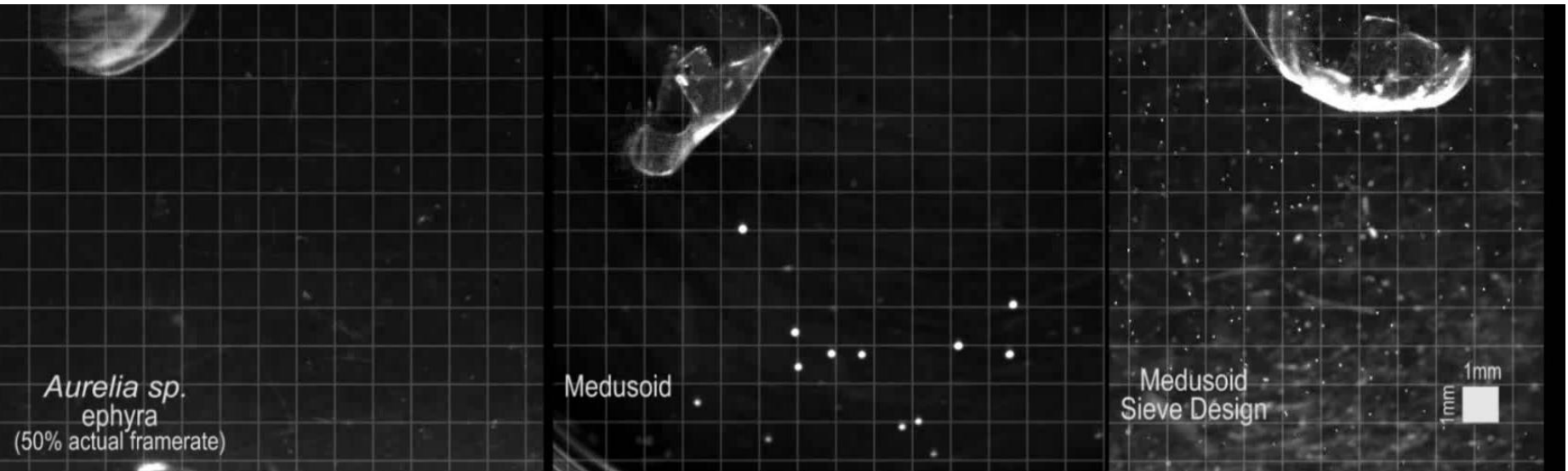


Real and artificial jellyfish propulsion

Control: Jellyfish

Optimal design

Suboptimal design

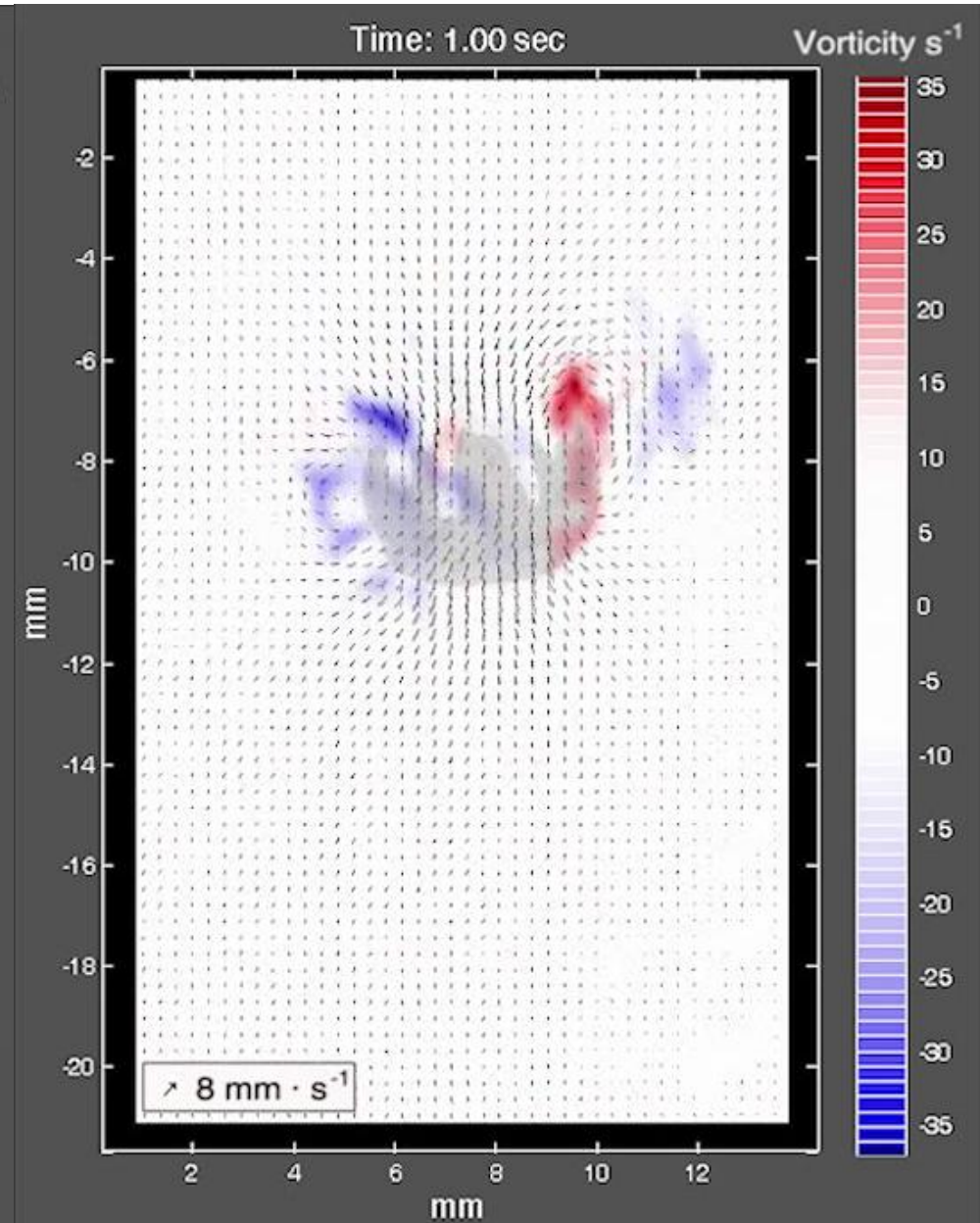
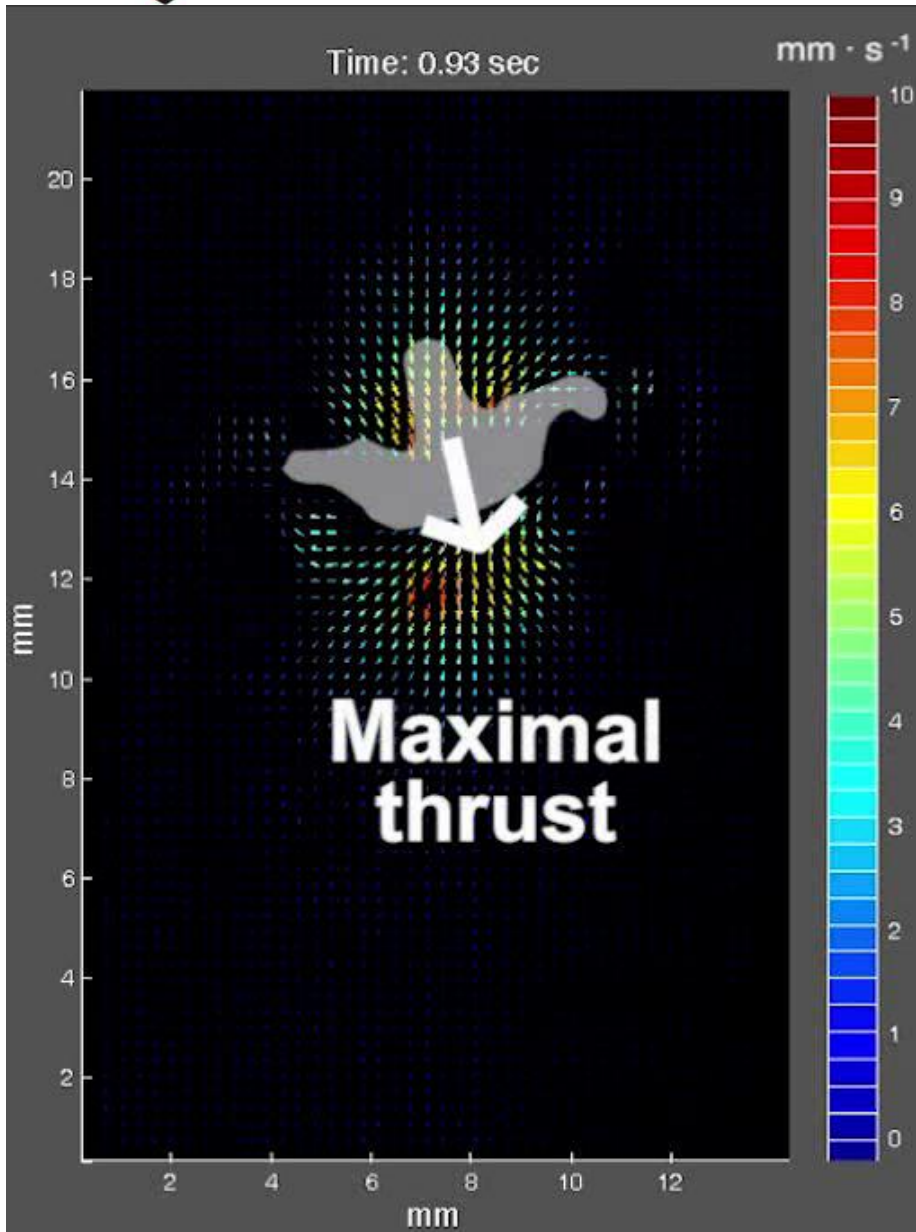


Quantifying feeding and propulsion currents



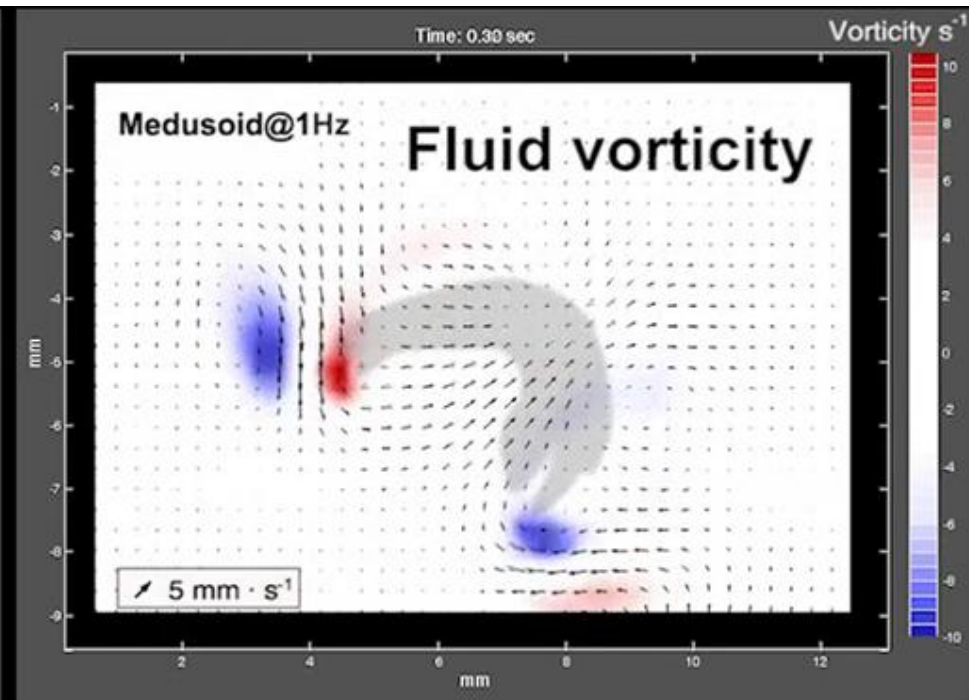
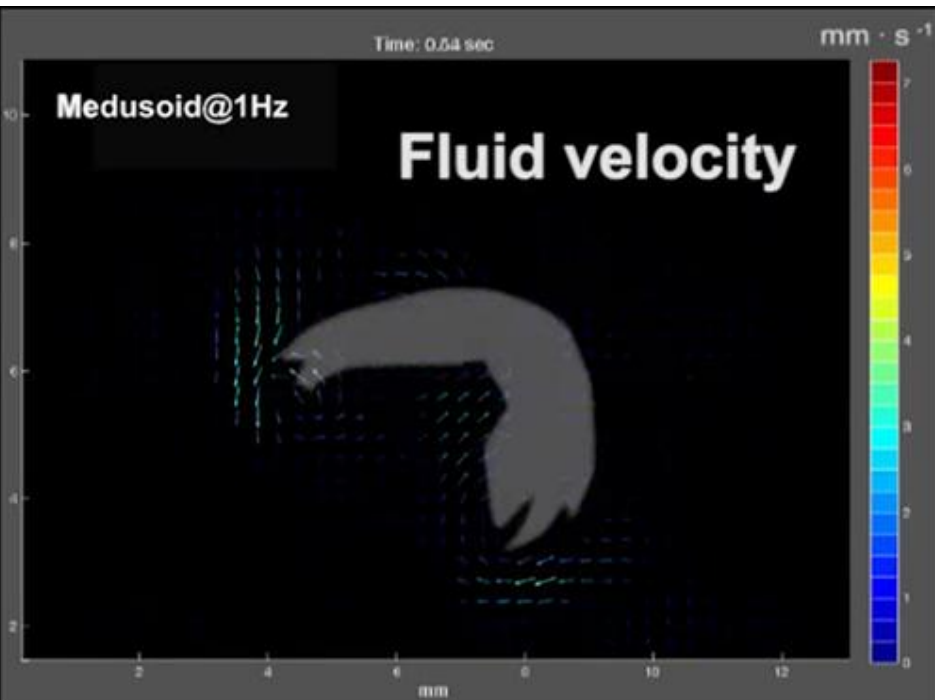


Control: Flow field in jellyfish





Flow field in optimal Medusoid





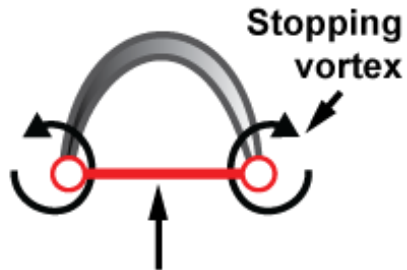
Fitness metrics

a Propulsion metric



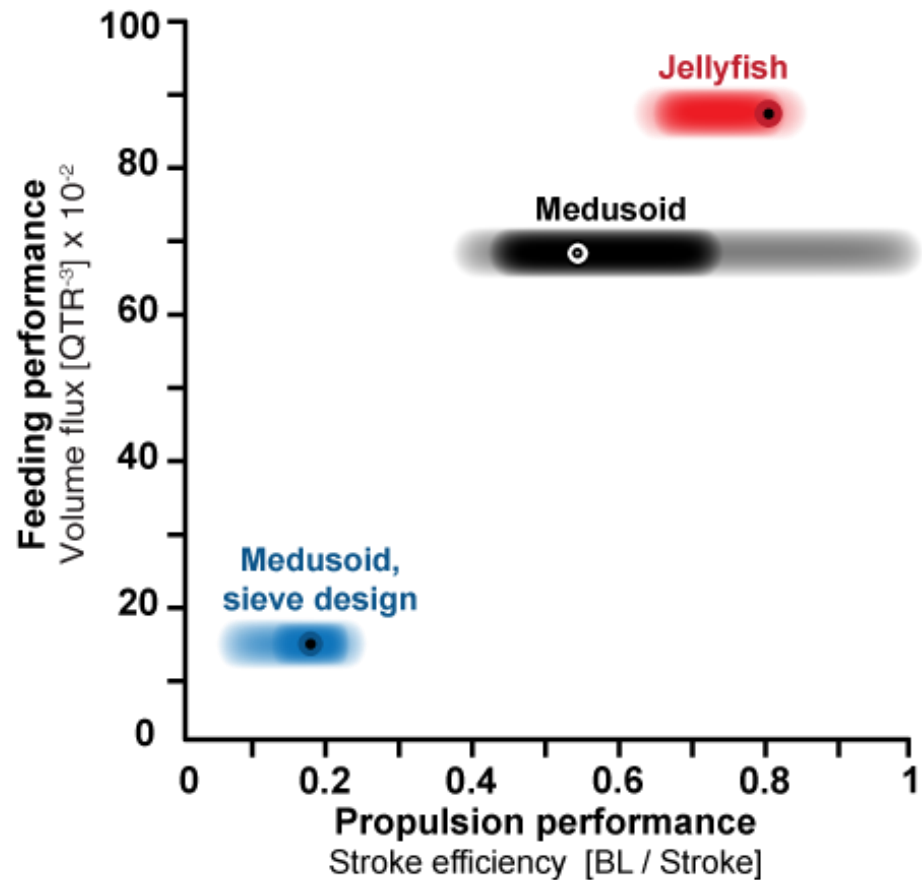
Body length (BL)
per stroke cycle

b Feeding metric



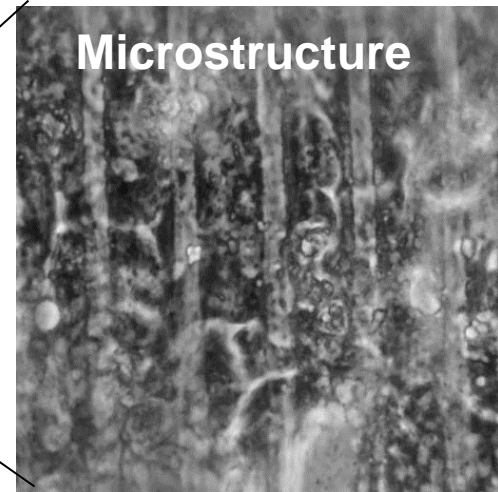
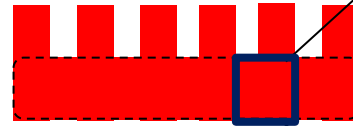
Flow across
reference section

c Performance matrix

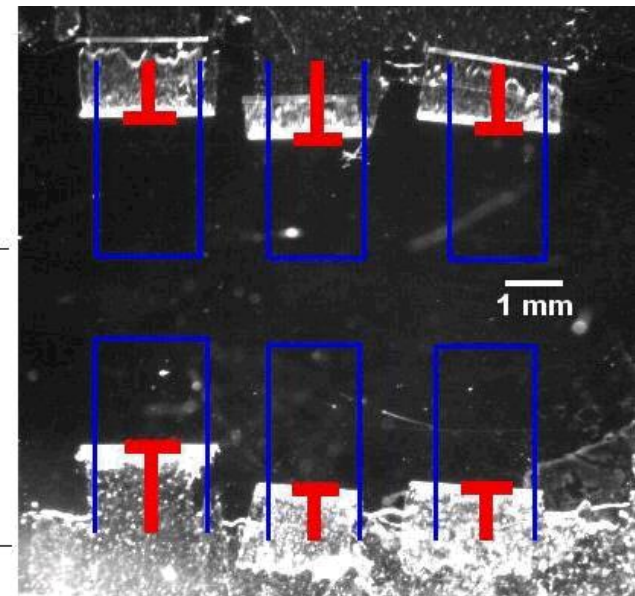
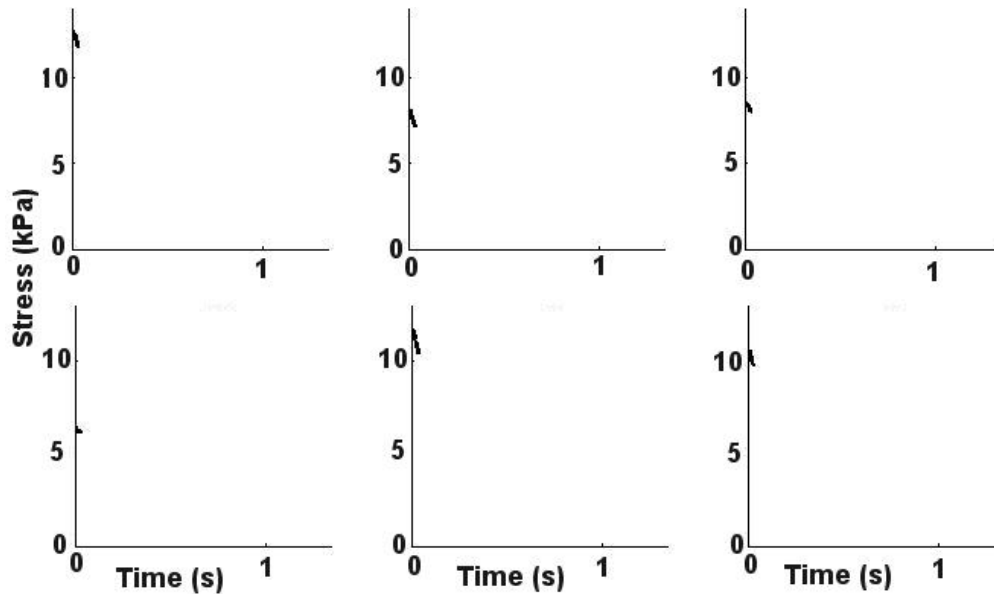




Heart-on-a-chip: further reduction



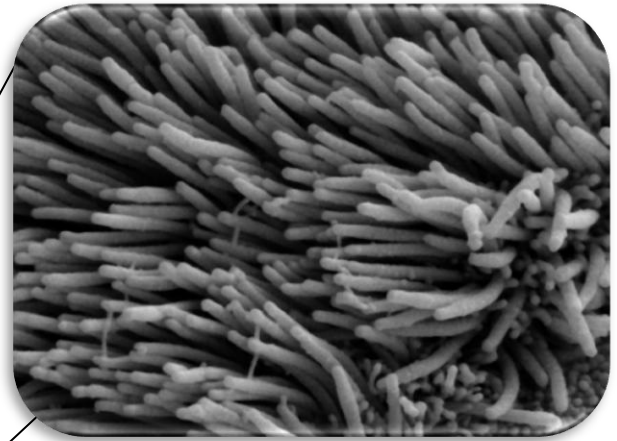
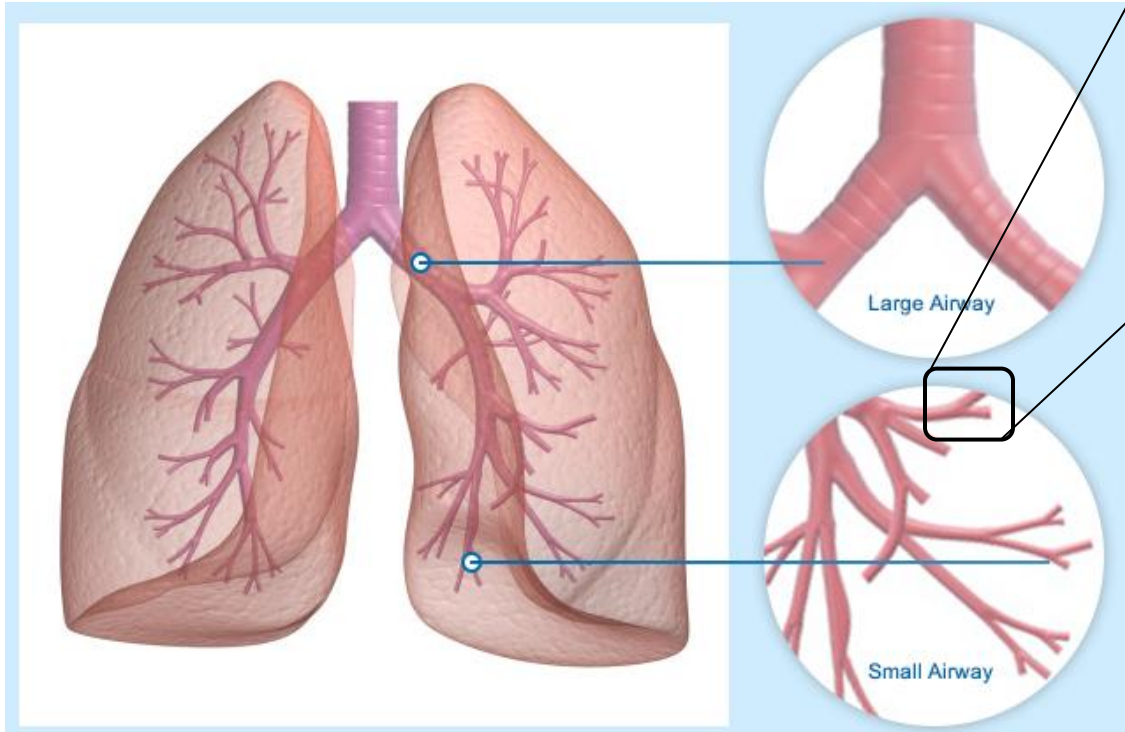
Tissue contractile stress





Case study: ciliated epithelium

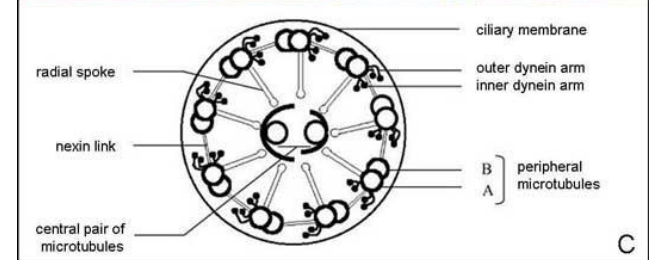
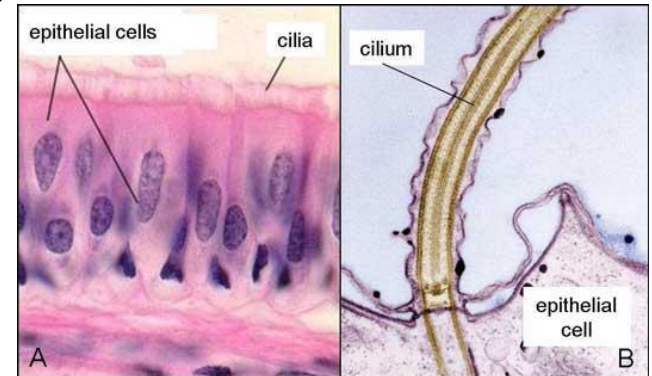
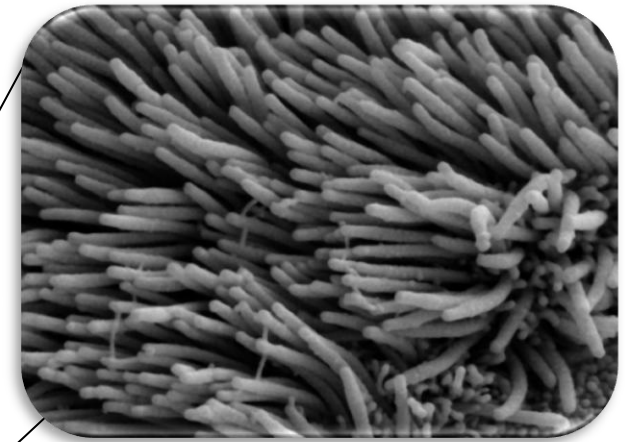
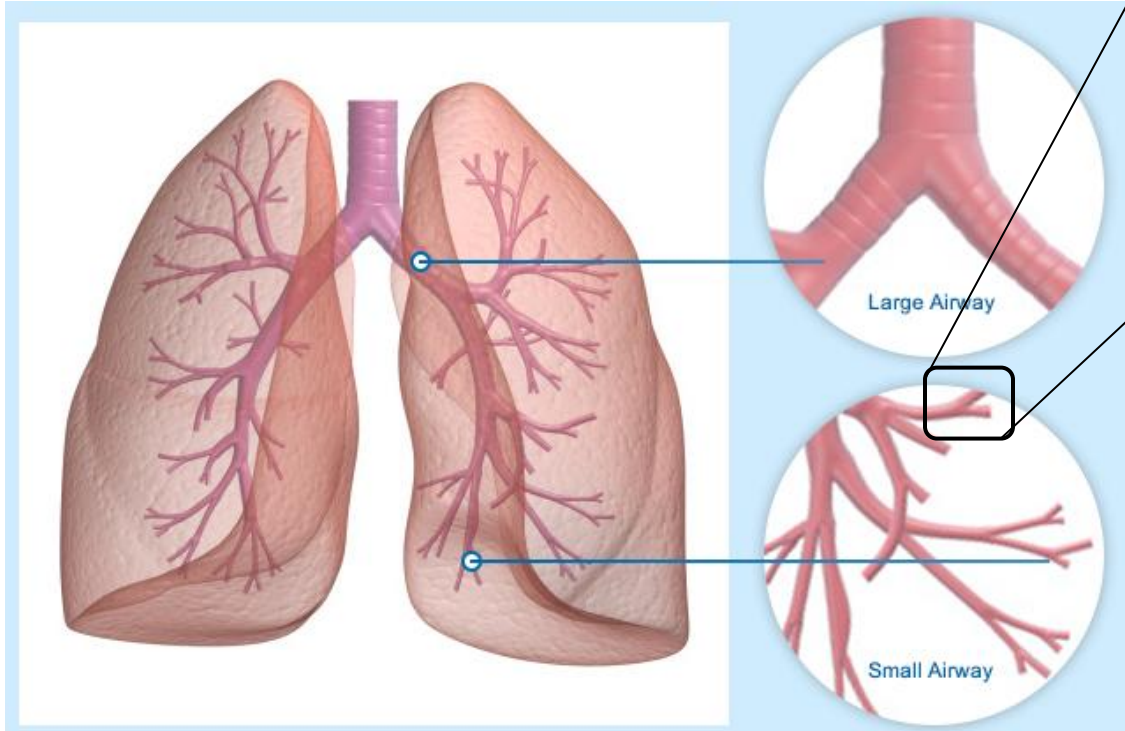
e.g., respiratory epithelium





Case study: ciliated epithelium

e.g., respiratory epithelium



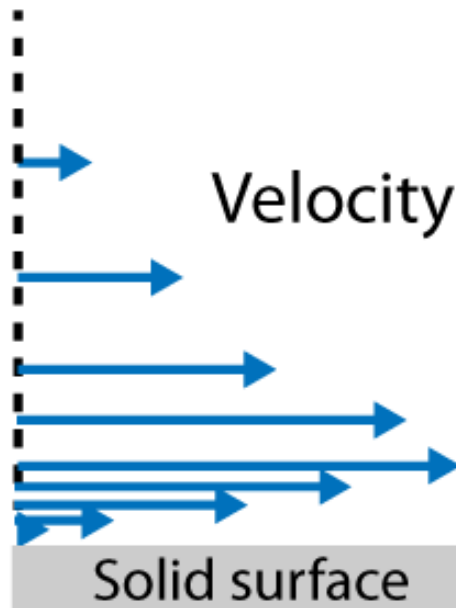


Cilia-powered fluid transport

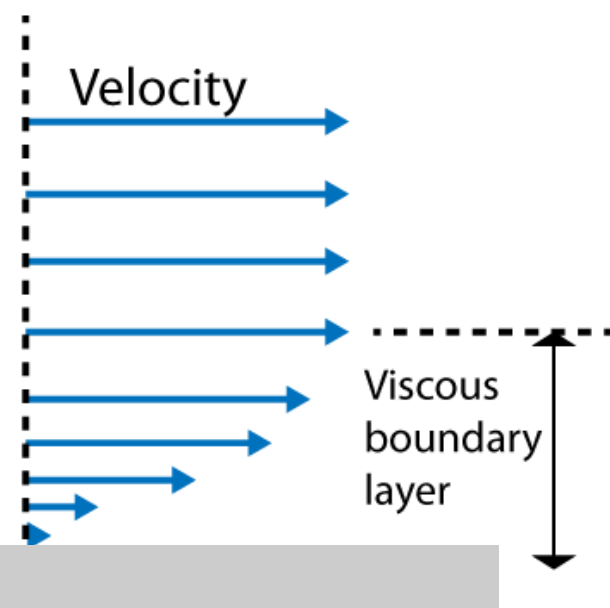
Ciliated epithelium (Paramecium)



Cilia flow profile



Bulk flow profile



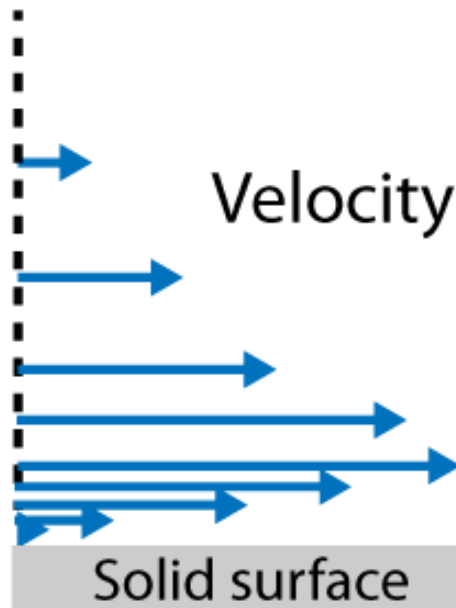


Cilia-powered fluid transport

Ciliated epithelium (Paramecium)



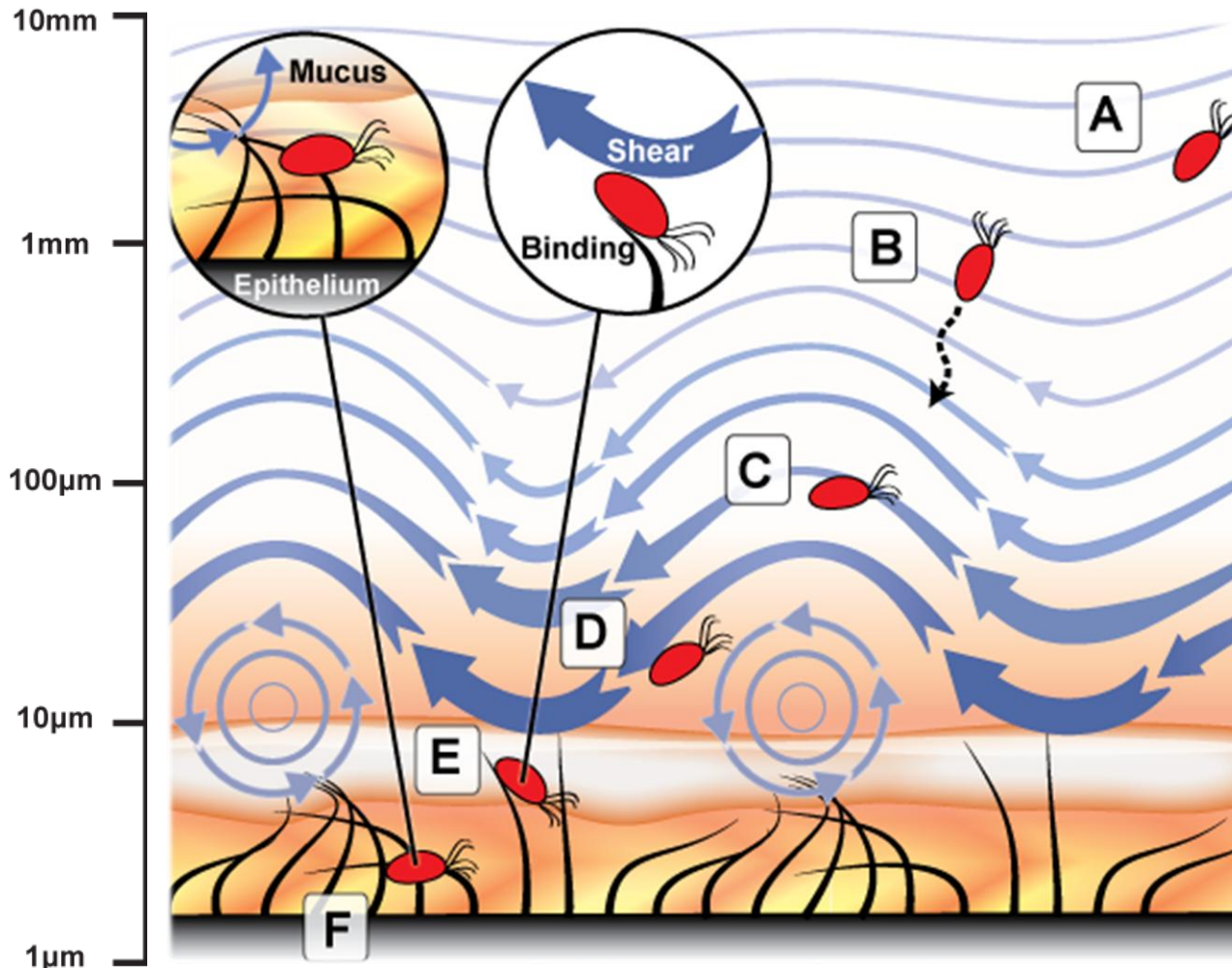
Cilia flow profile



- Particle capture & clearance
- Transport of fluids, solutes and cells



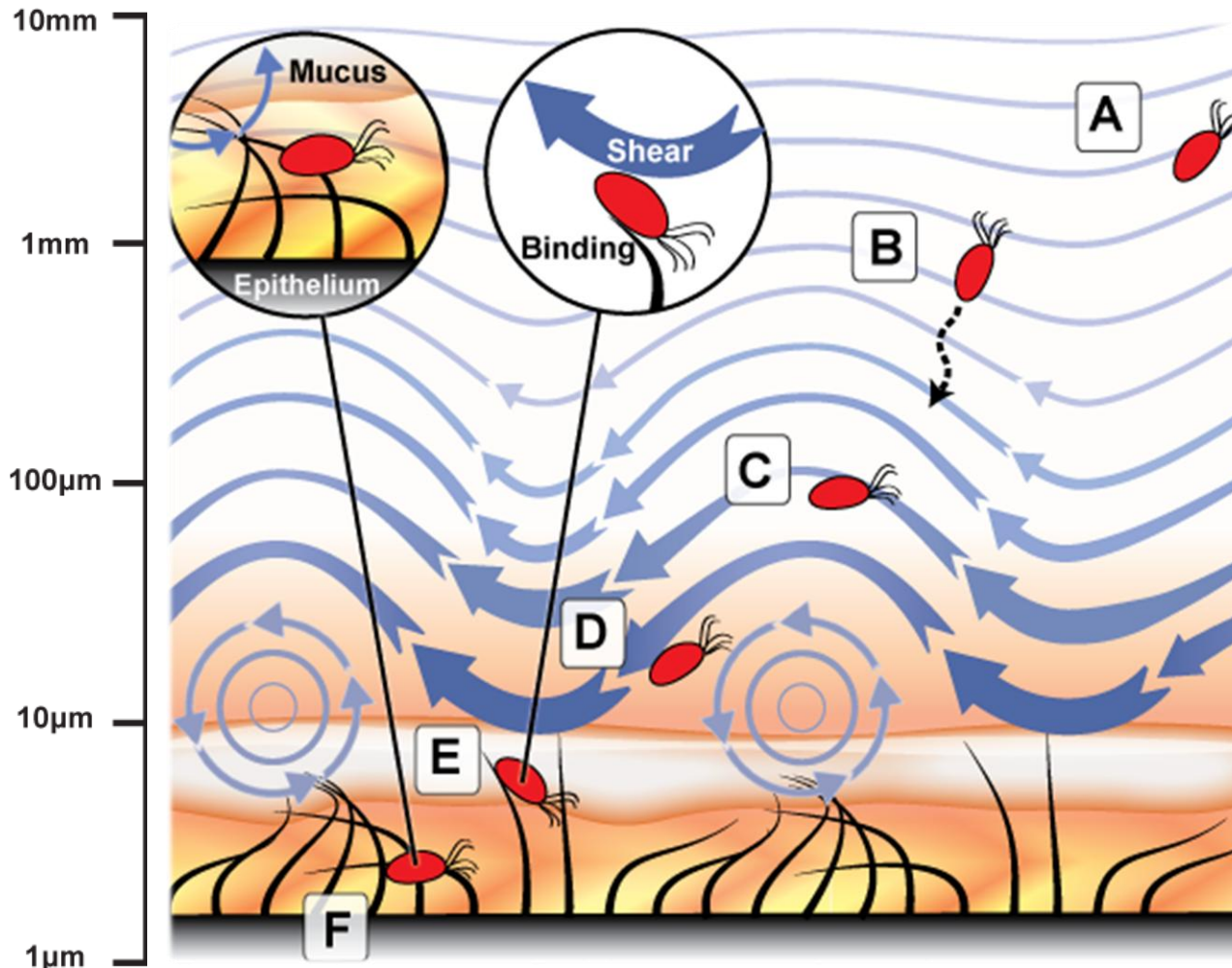
Structure-function relationships of ciliated surfaces



Cilia fitness
Selective bacterial recruitment



Structure-function relationships of ciliated surfaces



Cilia fitness

Selective bacterial recruitment



Fluid transport and mixing



Surface geometry

+

Metachronal wave

+

Ciliary structure and kinematics

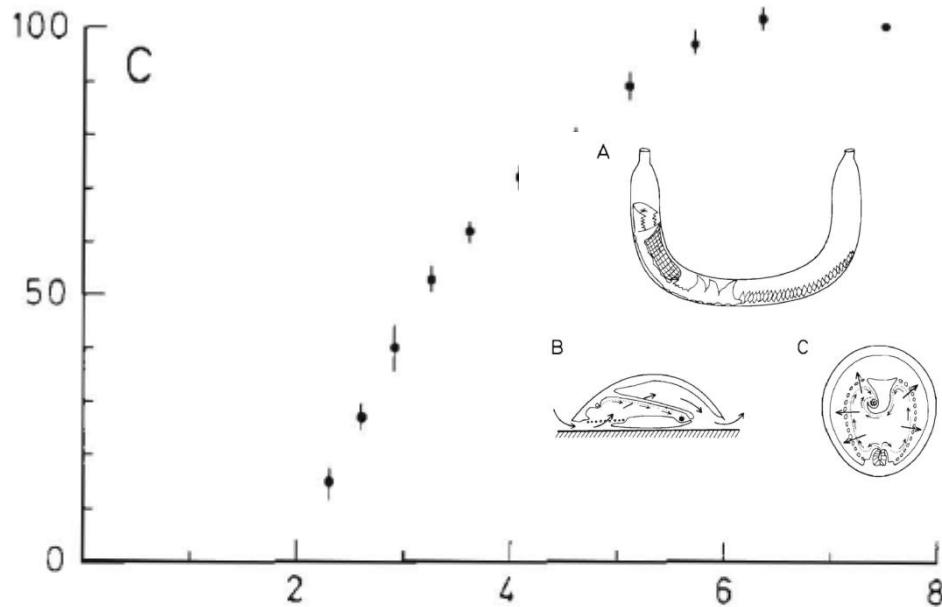


A master of selective bacterial recruitment: The Hawaiian bobtail squid



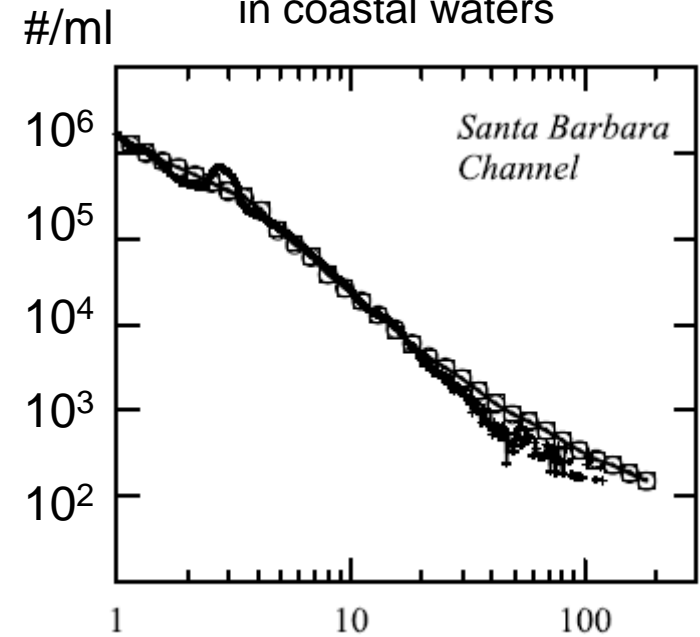
Internal ciliated organ captures 1 μm bacteria species (*Vibrio fischeri*) from huge microbial background (0.5%), and wide range of particle sizes

Retention efficiency in typical ciliary filter feeders



Particle diameter (μm)
Jorgensen, 1994

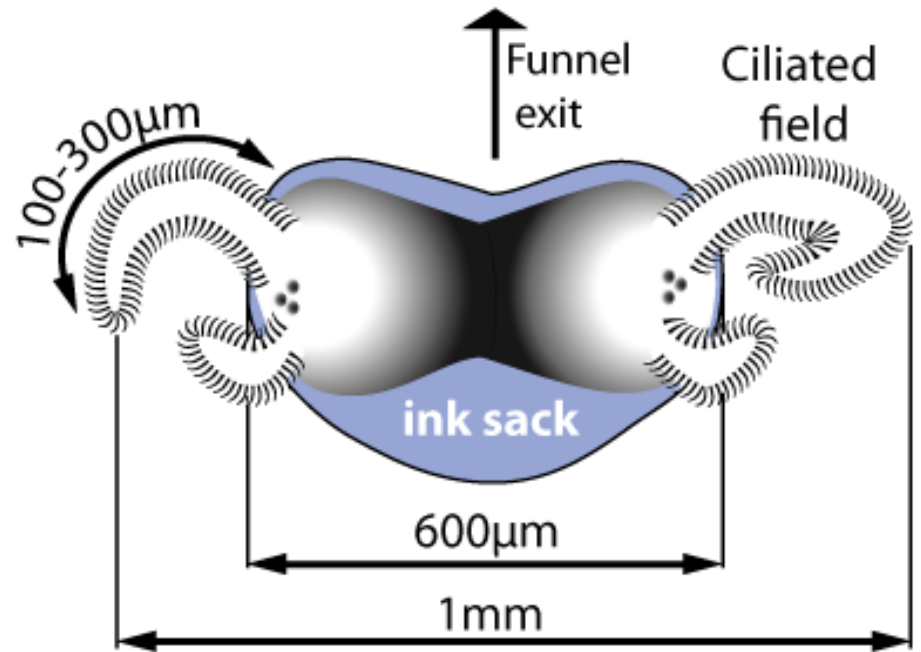
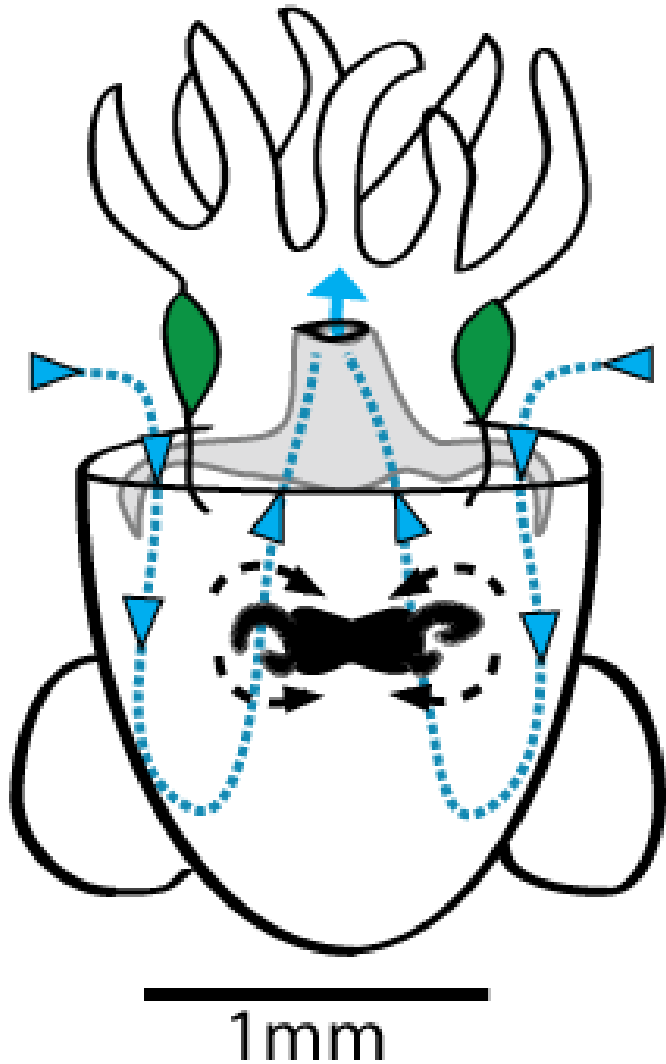
Particle size distribution
in coastal waters



Largest dimension in μm
Reynolds et al., 2010



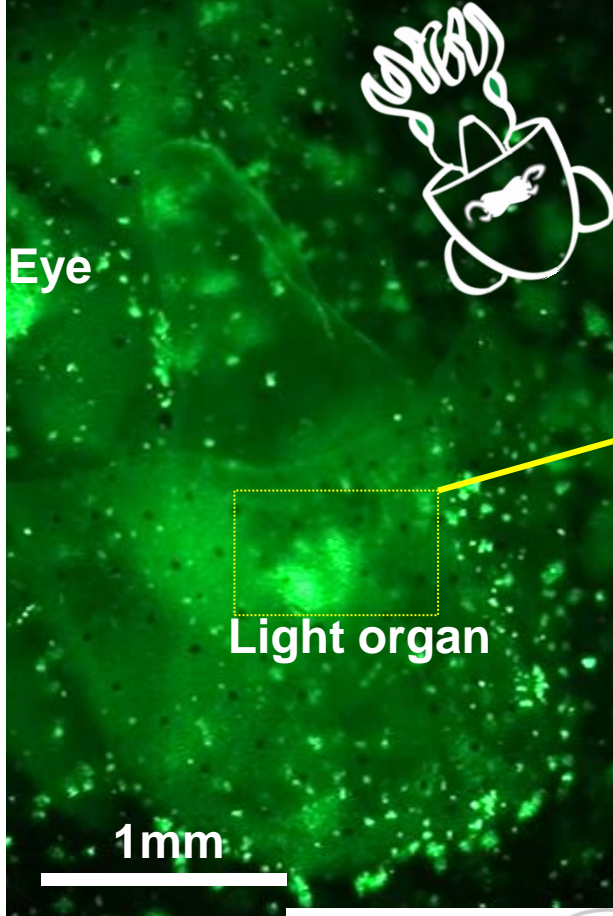
The squid ciliated organ



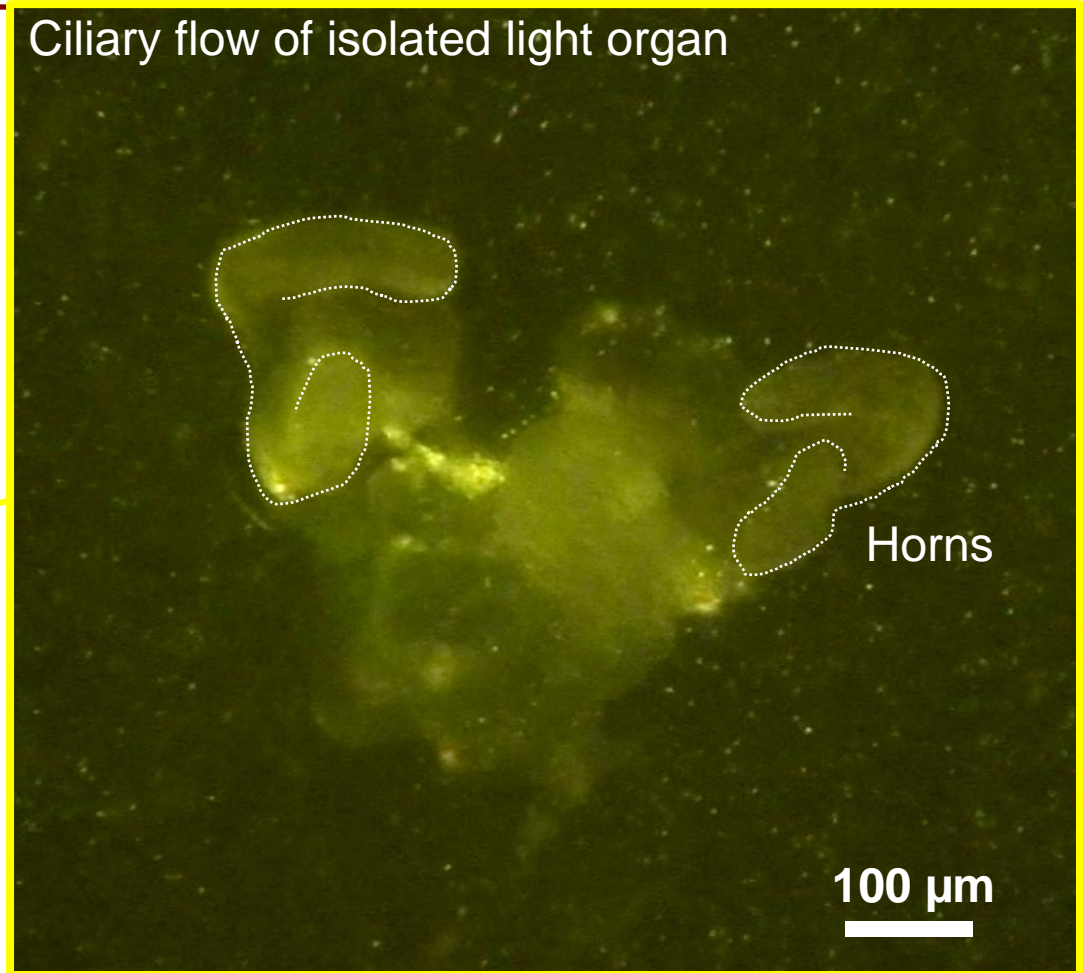


The light organ is subject low Reynolds number flow

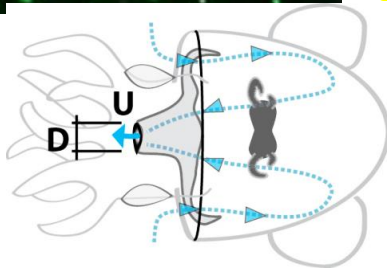
Light organ in mantle flow



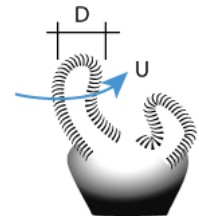
Ciliary flow of isolated light organ



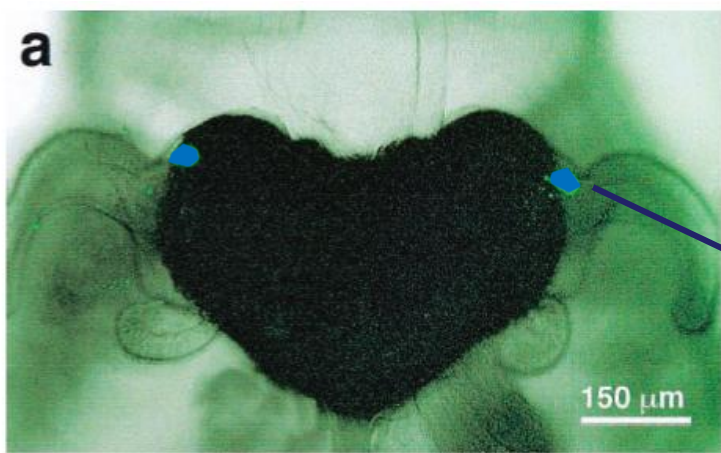
$$Re = \frac{UD}{\nu} = 0.2$$



$$Re = \frac{UD}{\nu} = 0.004$$

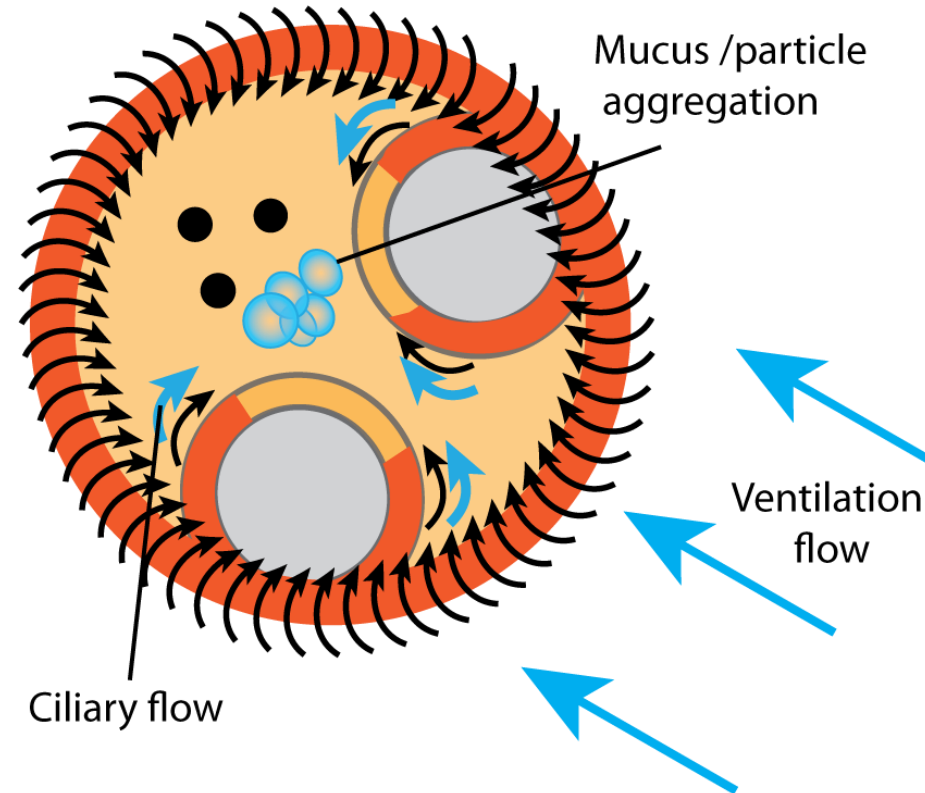
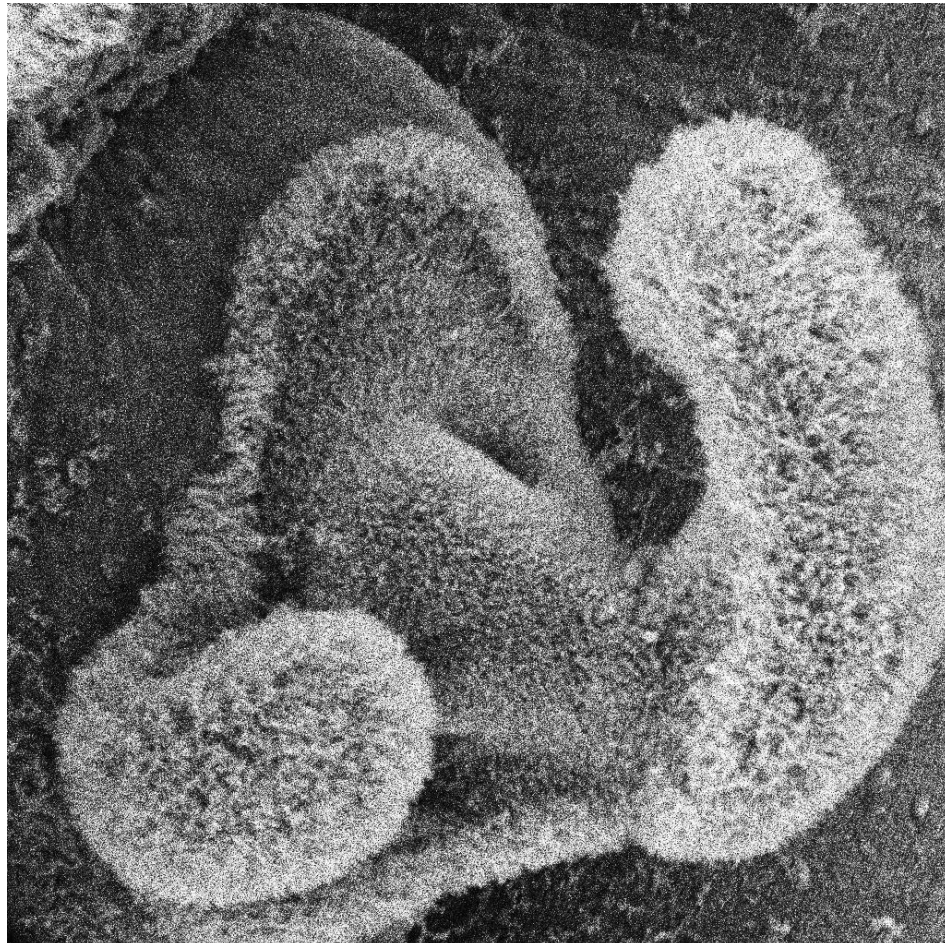


a

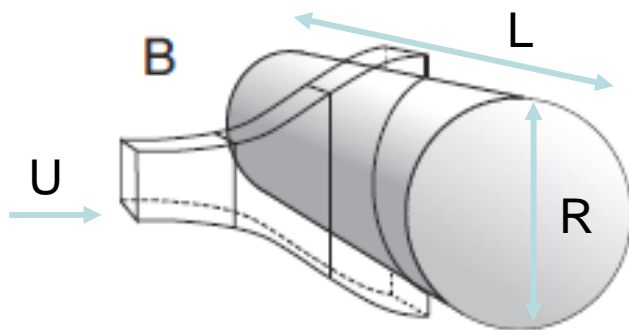
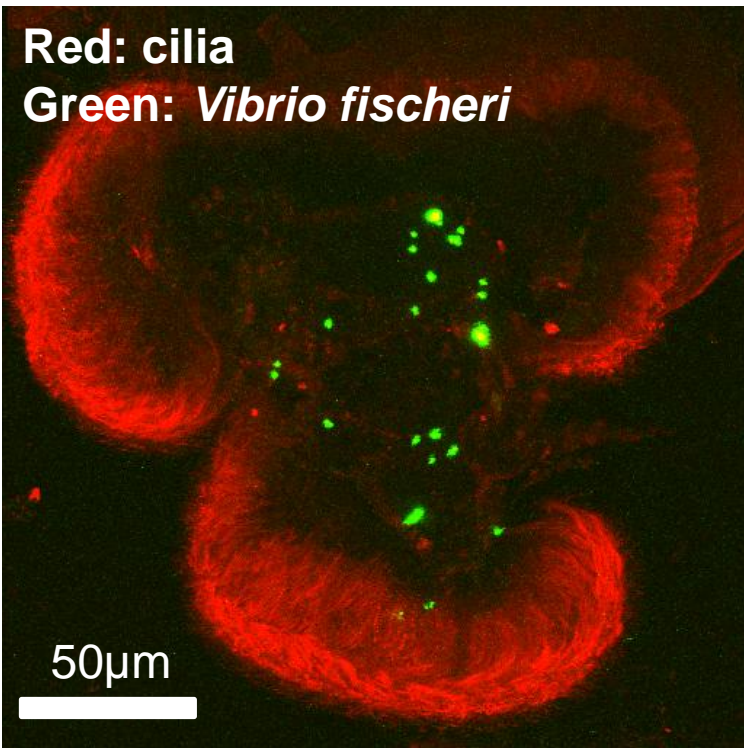


Mucus aggregation

The mucociliary trap



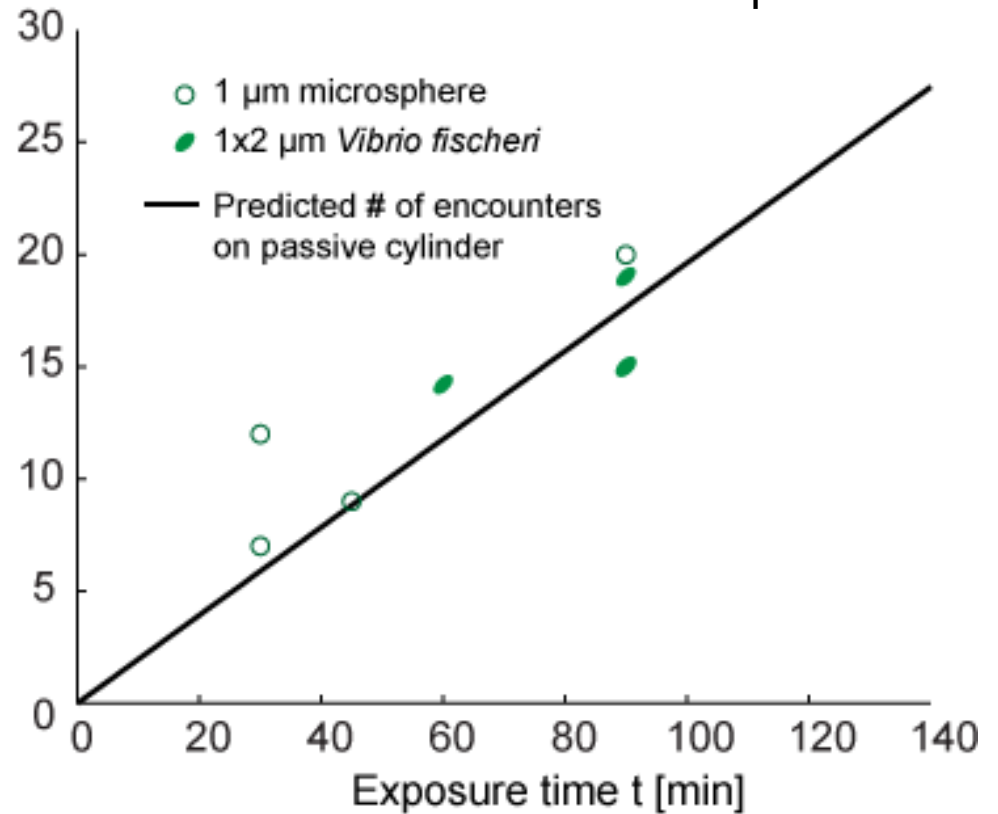
The mucociliary trap: not such a good trap?



Encounter model for passive cylinder in flow
(Humphries, PNAS, 2009)

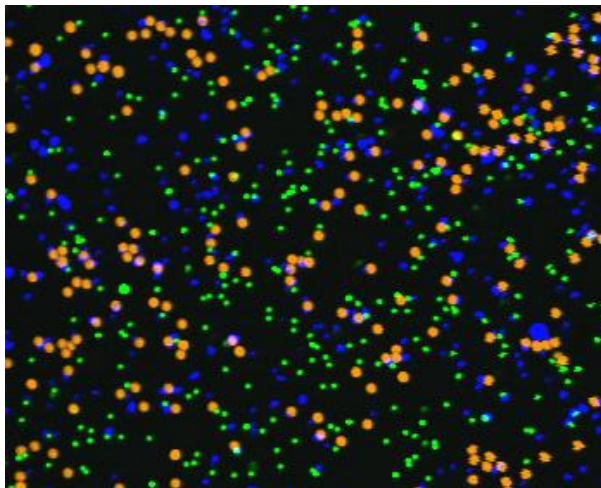
captured in mucus

10^7 particles/ml

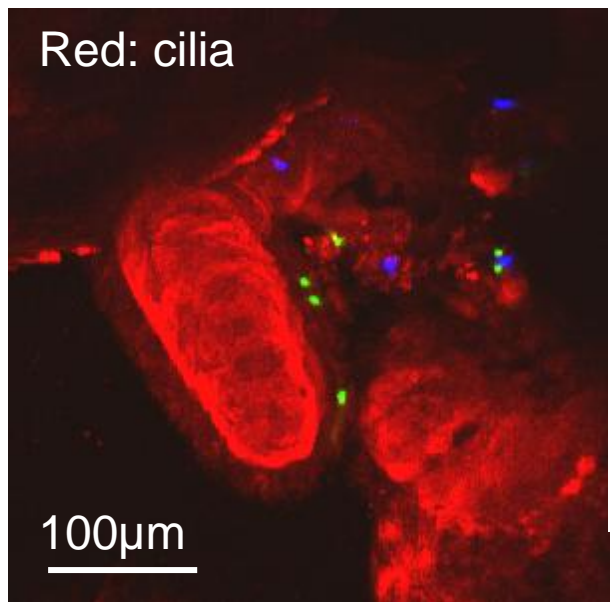


Capture rate similar to predicted encounter rate of passive cylinder in flow
→ **No obvious increase in capture rate compared to chance**

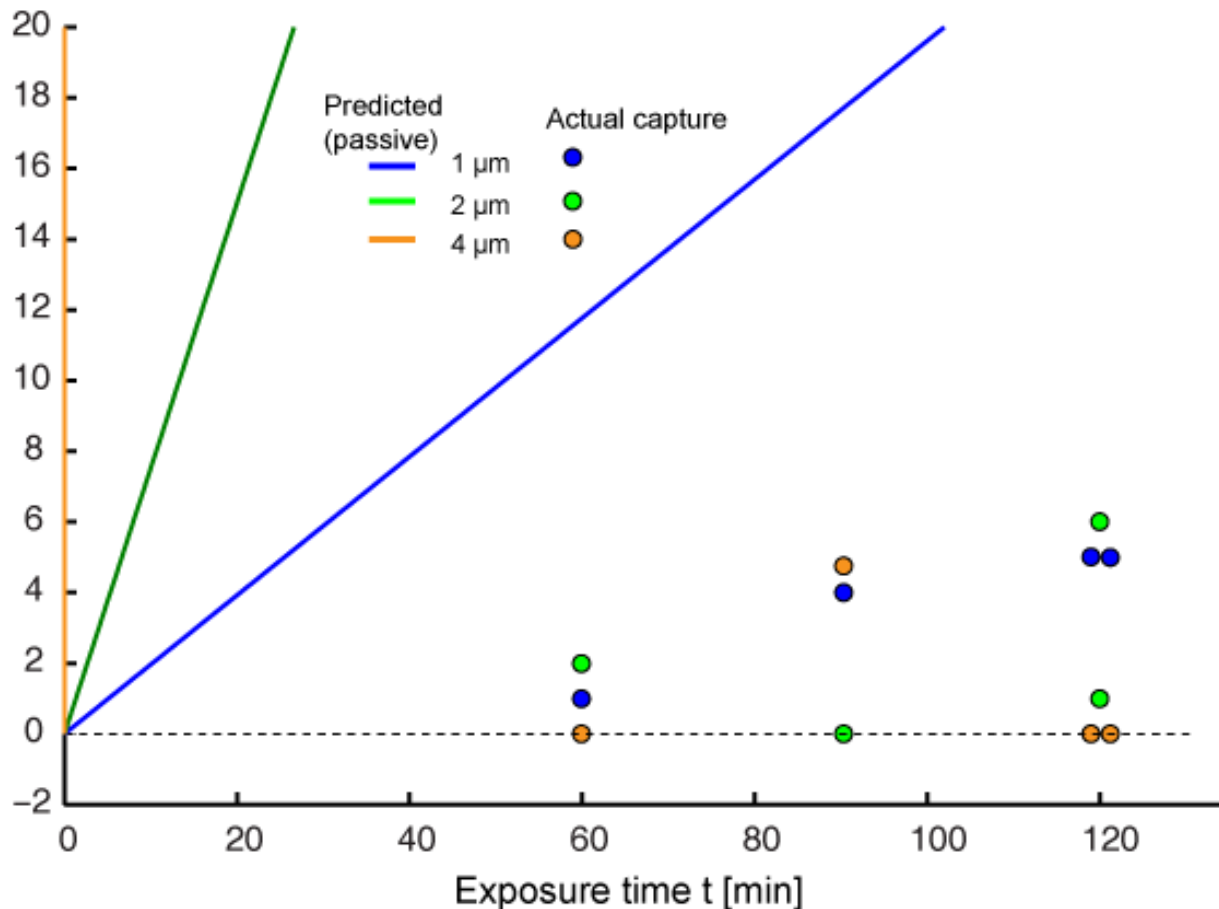
The mucociliary trap - not for everyone: Evidence for size-biased capture



1 μm (blue), 2 μm (green),
4 μm (orange);
 10^7 particles/ml each

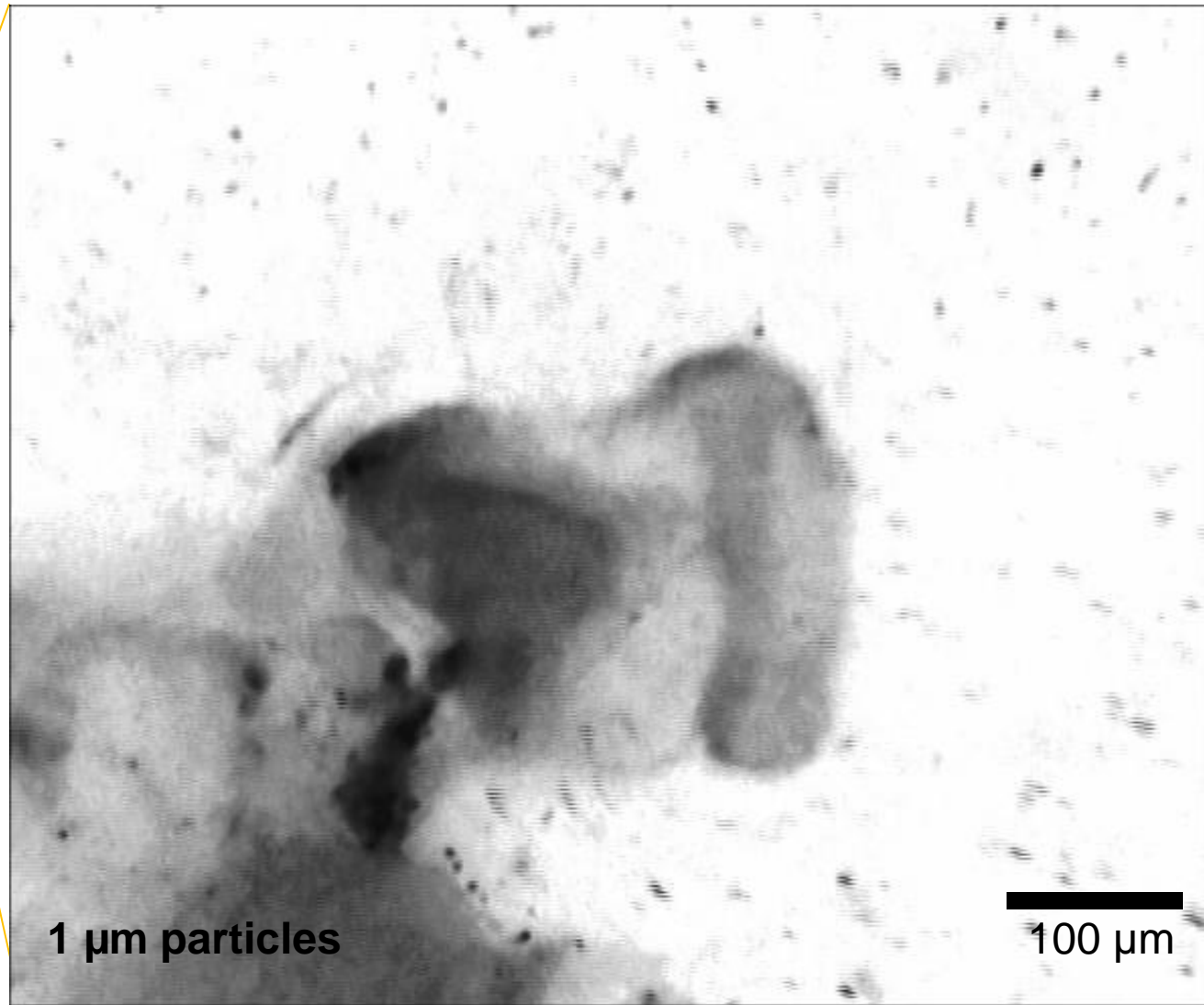
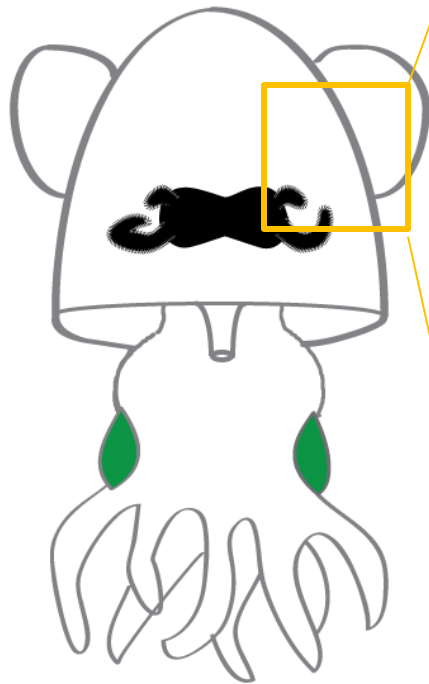


of encounters (captures)



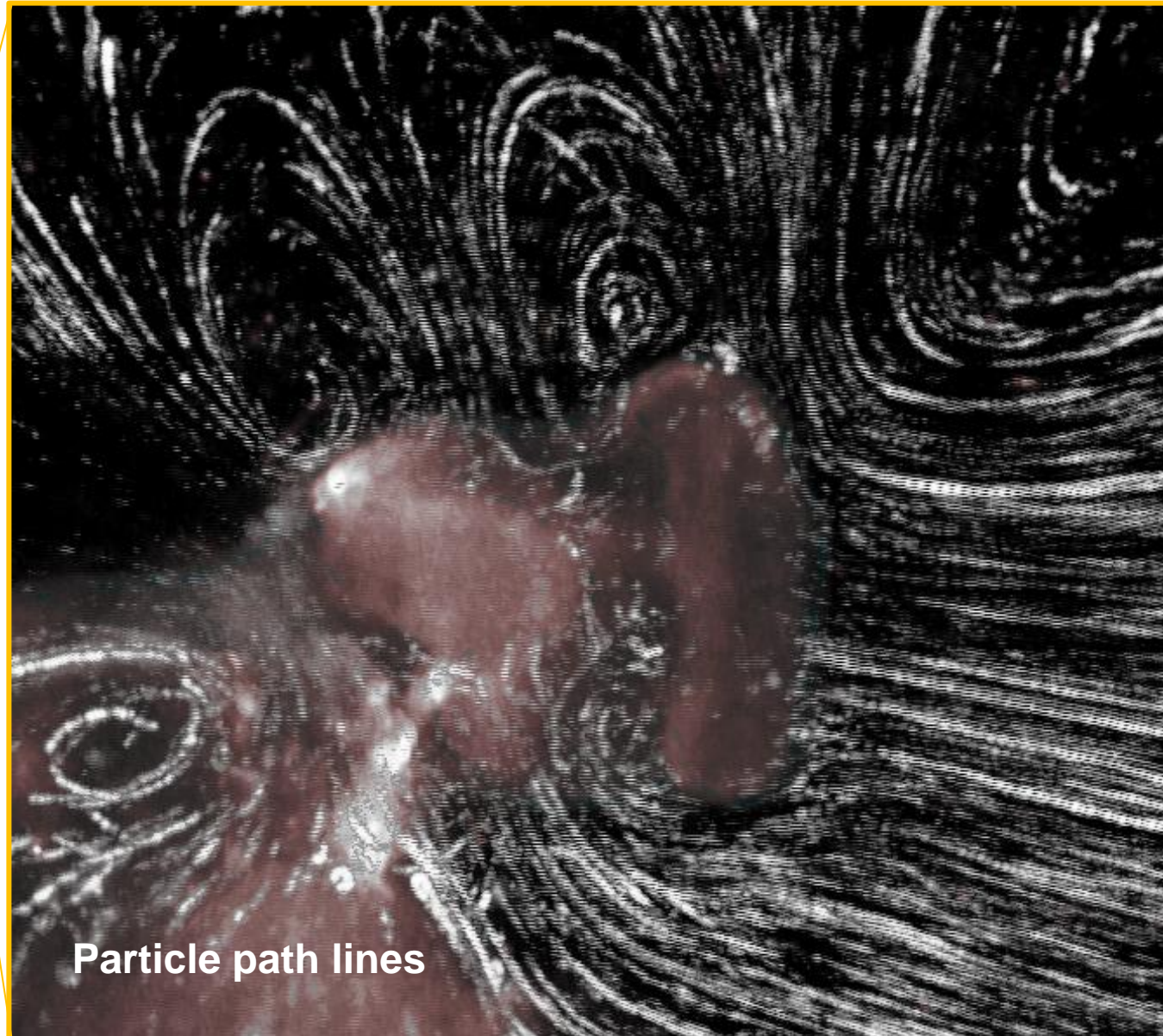
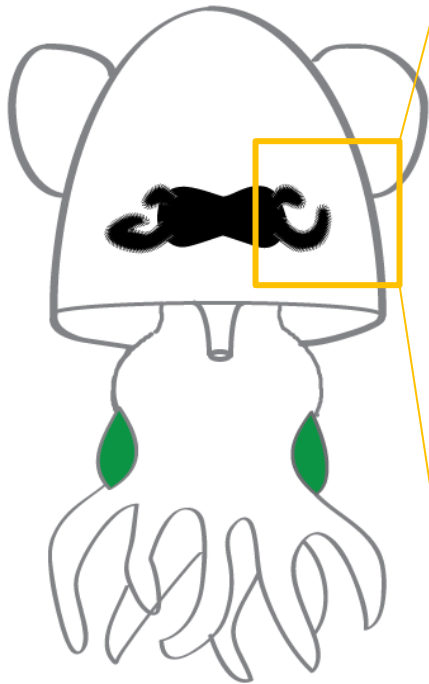


Cilia-generated flow field





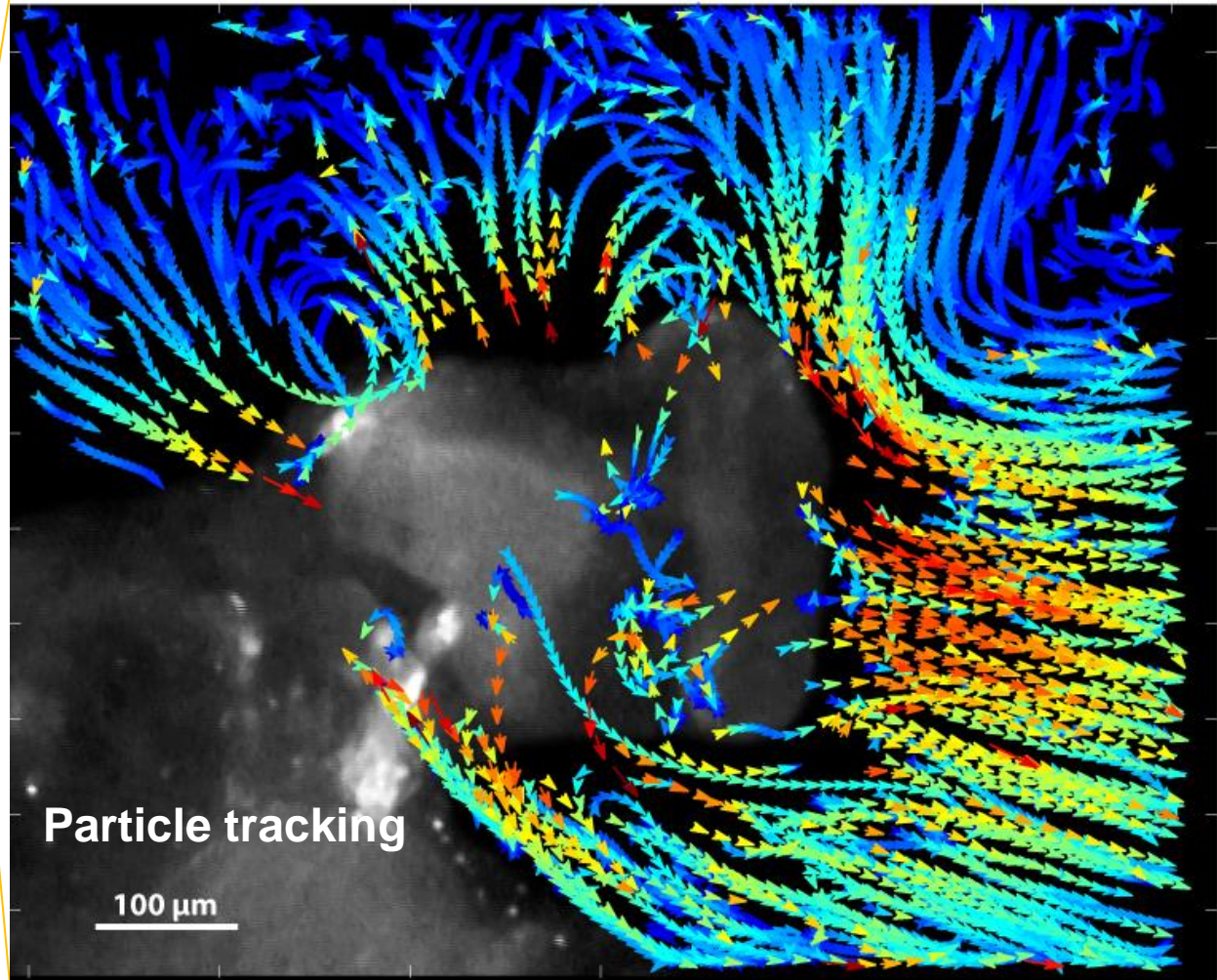
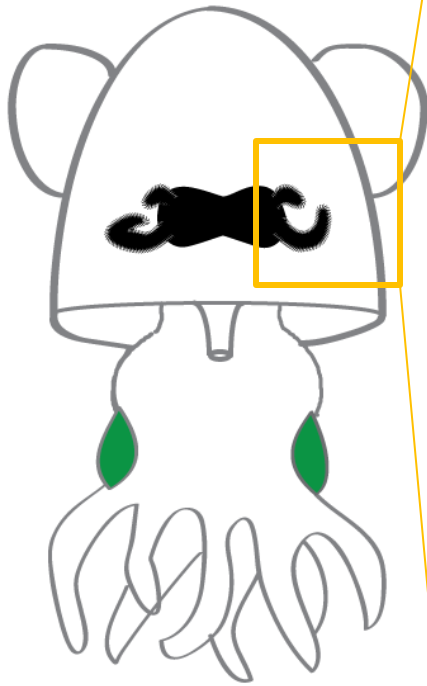
Cilia-generated flow field



Particle path lines



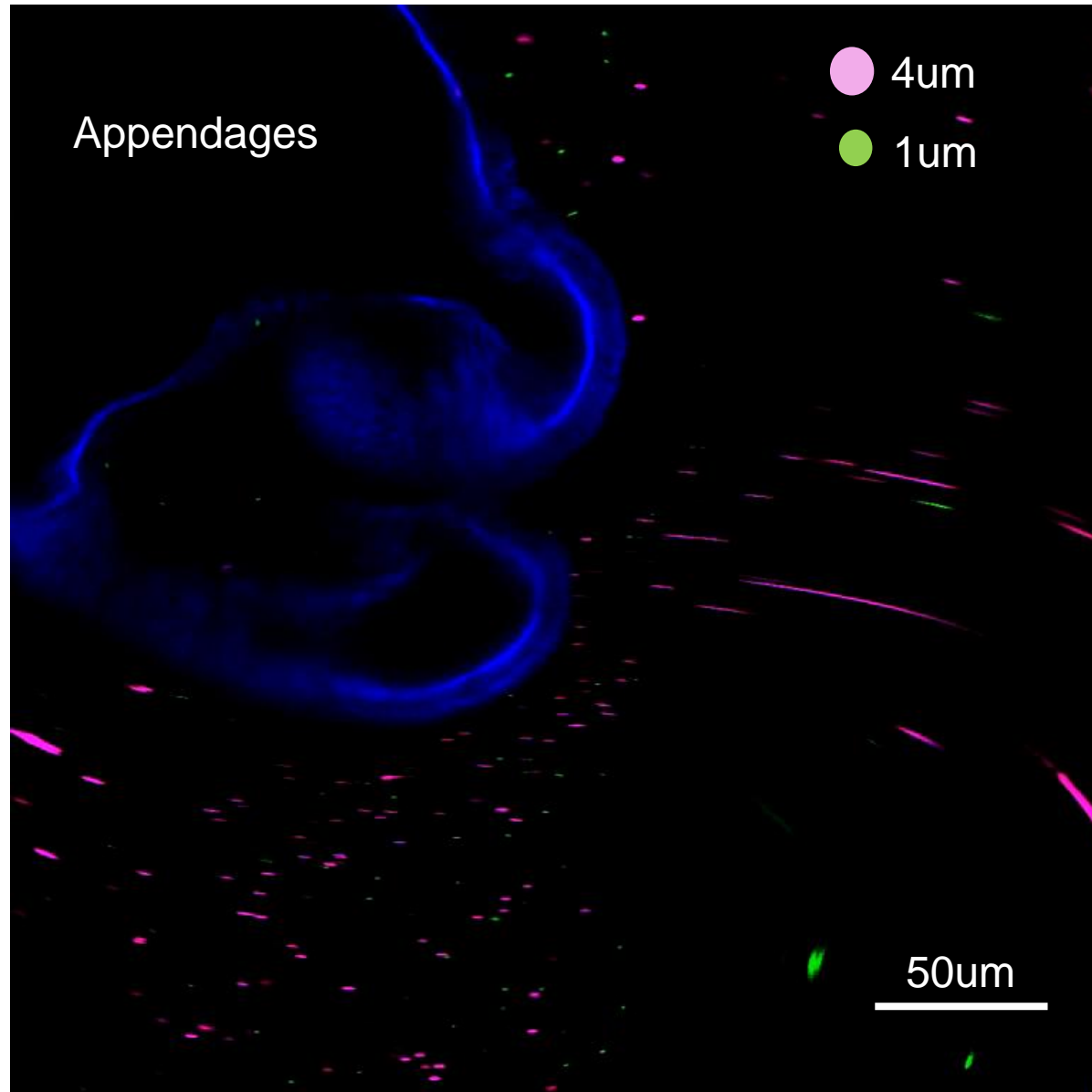
Cilia-generated hydrodynamic sieve



Ratio advected particles:captured particles $\approx 50:1$

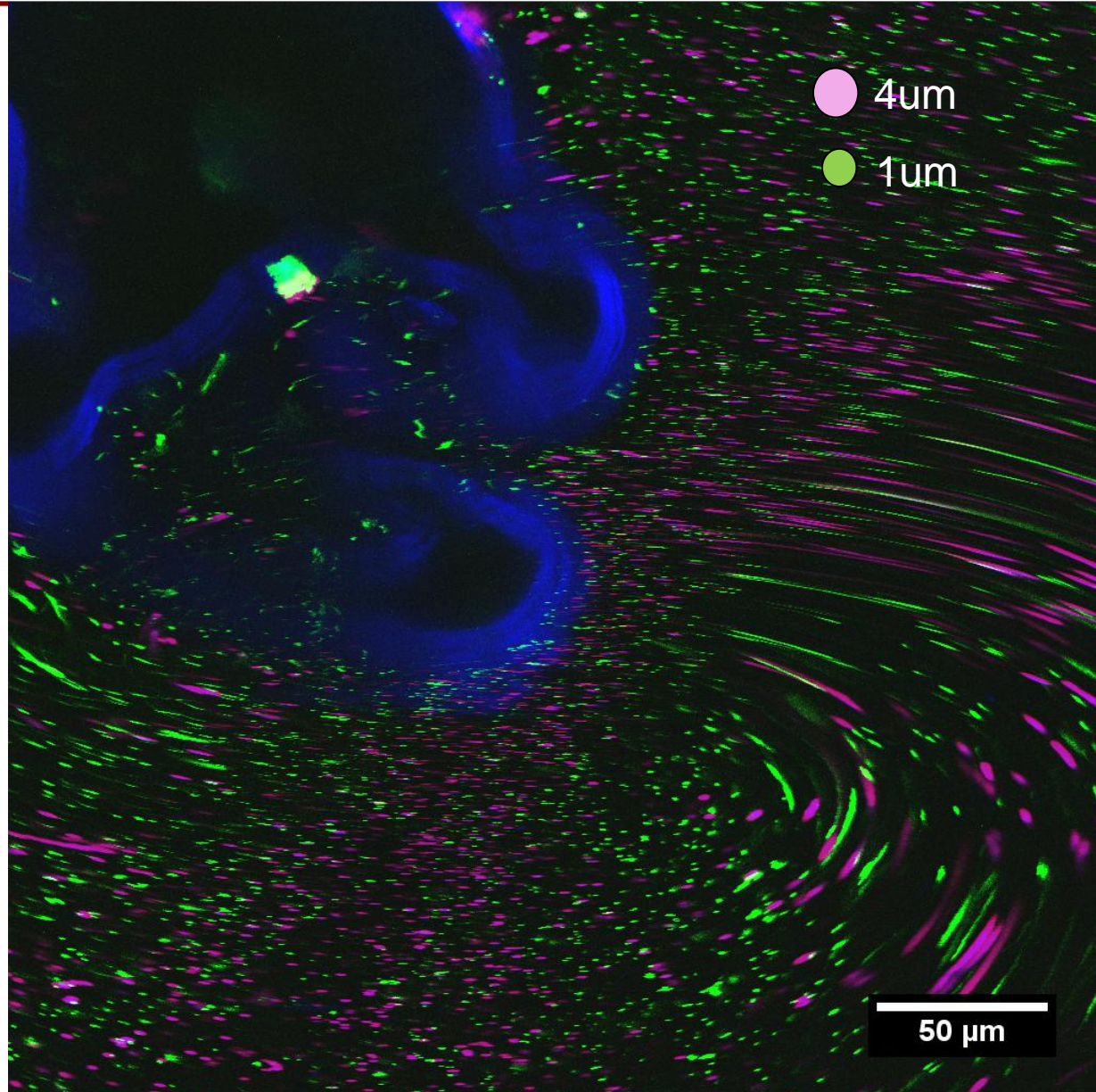


In vivo capture





In vivo capture

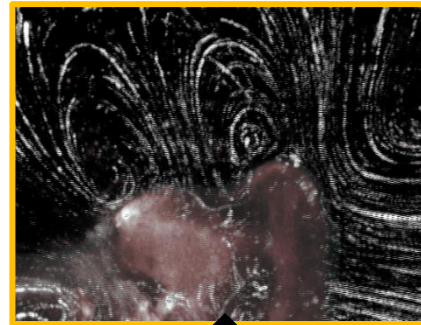




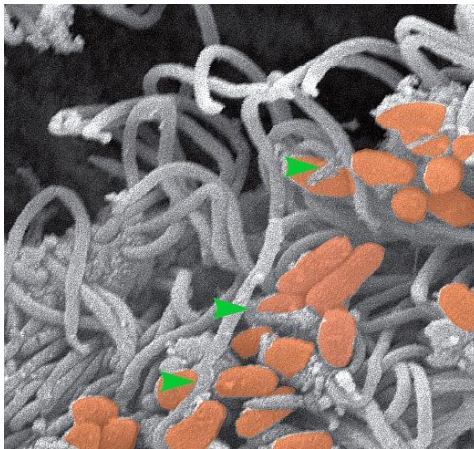
Current work: Identifying structure-function relationships...

Cilia fitness

Capture and aggregation of bacteria (-sized particles)

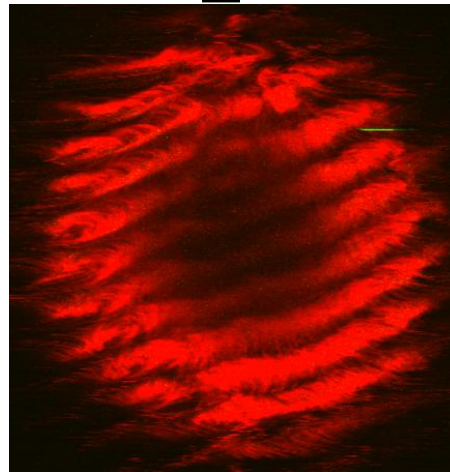


Fluid transport
and mixing



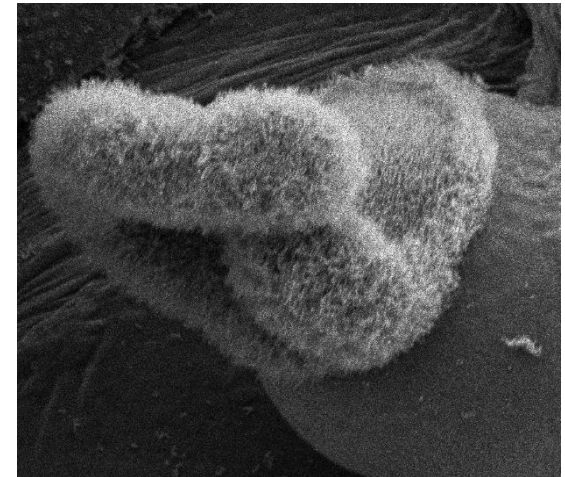
Ciliary structure and
kinematics

+



Metachronal wave

+



Surface geometry



Current work: Identifying structure-function relationships...

...using a variety of methods and approaches

