

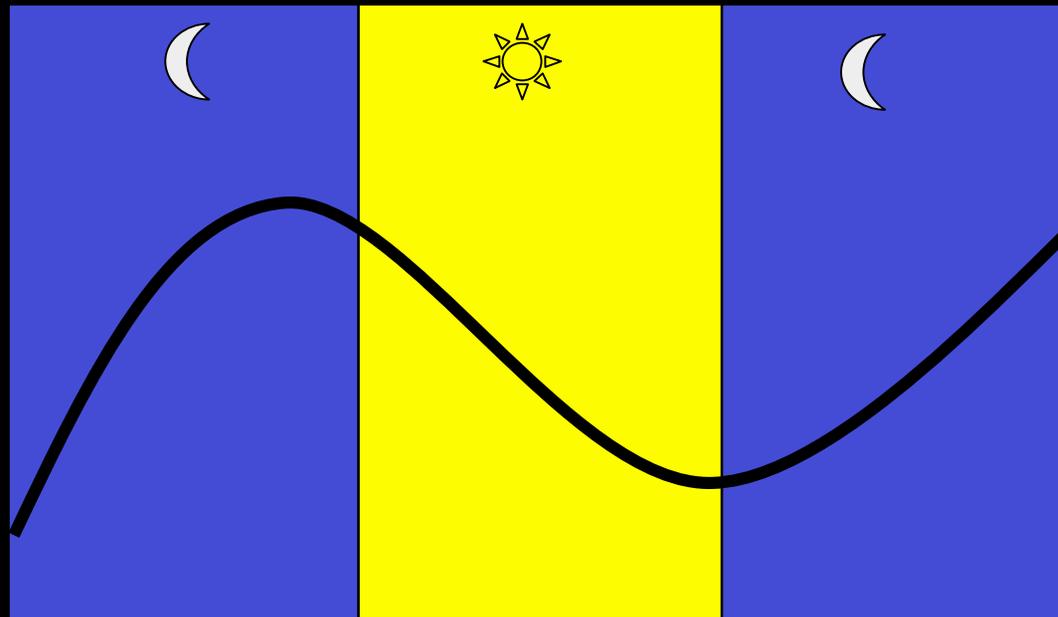
Complexity in the *Neurospora* circadian clock



1. Overview of FWO
2. FLO's
3. Output
4. Frequency Demultiplication

The “pacemaker” involves the interactions of multiple feedback loops that can program phase-dependent oscillations.

Endogenous Circadian Clocks Allow Anticipation of Environmental Cycles



↑
Make protective proteins
BEFORE sunrise

Neurospora crassa

376

Frank-Roman Lauter

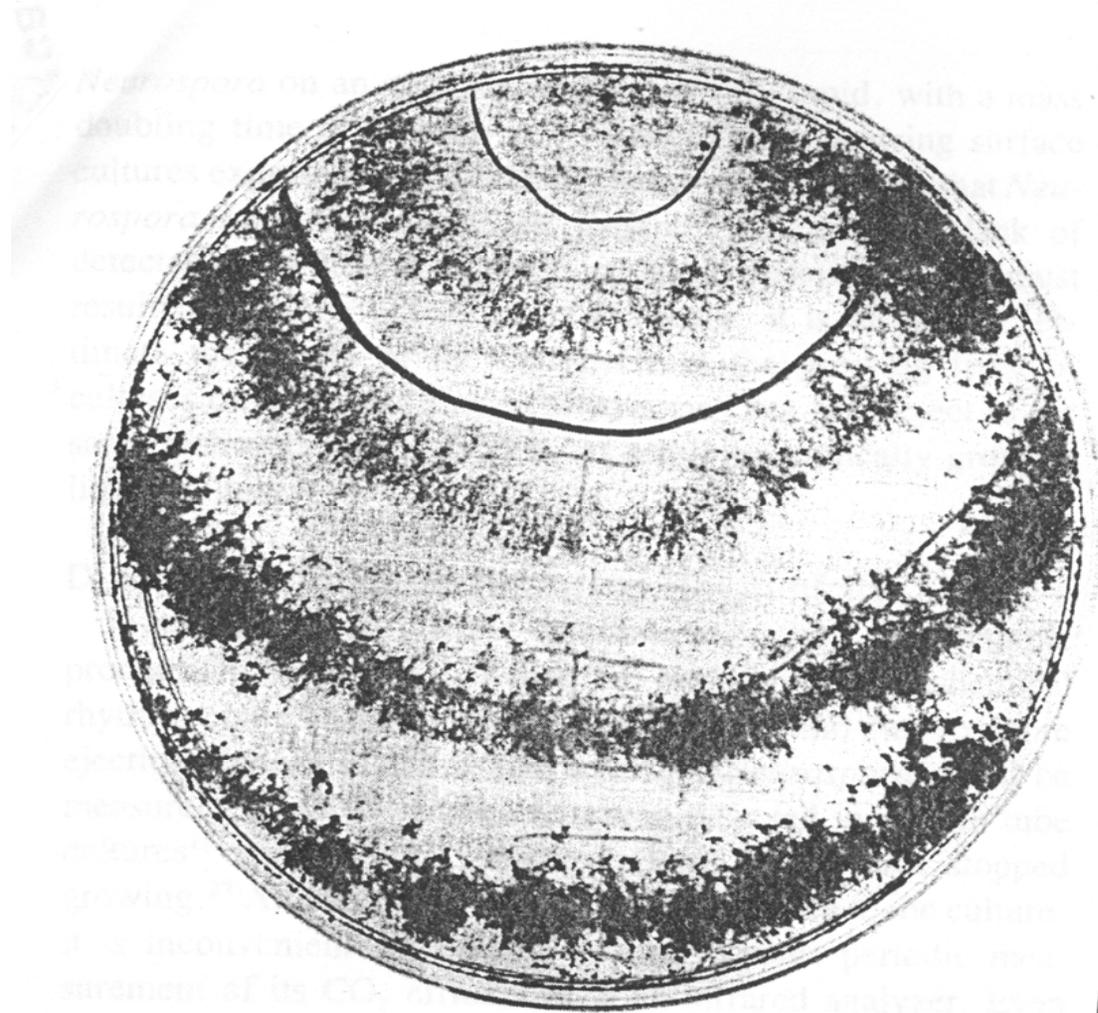


Figure 1. Bread spoiled by the ascomycete *Neurospora crassa*. (Picture courtesy of David Perkins)



- Haploid genome with 7 chromosomes, sequenced.
- Reproduces both asexually and sexually.
- Easy to manipulate both genetically and biochemically.
- Easy to monitor the circadian rhythm of asexual spore development.

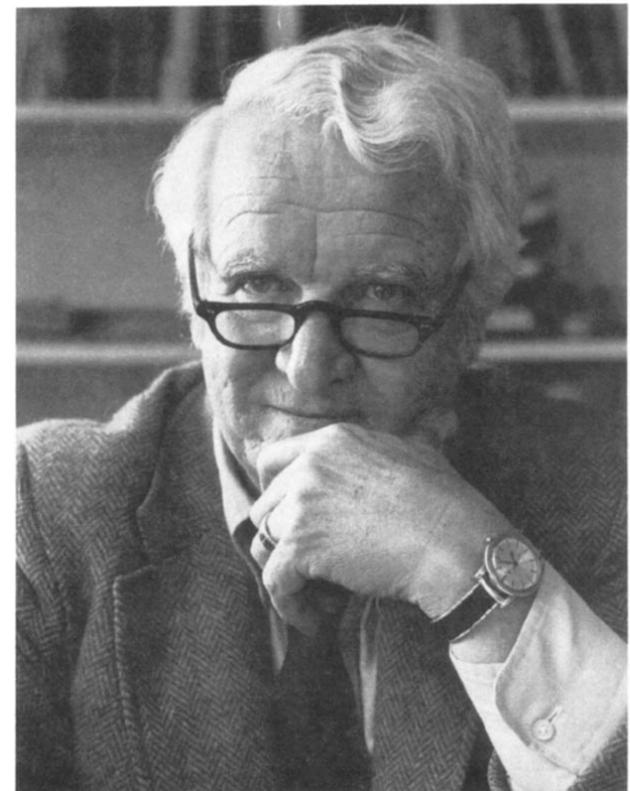
William Brandt (1953) *Mycologia*: Patch mutant (proline auxotroph), first report of a rhythm in *N. crassa*.



Pittendrigh CS, Bruce BVG, Rosenzweig NS, Rubin ML.
1959. A biological clock in *Neurospora*. Nature 184: 169-170.

Showed that the developmental rhythm
is circadian (temp compensated,
entrainable by light/dark cycles)

**“the Neurospora system
appears potentially valuable for
an attack on the basic problem
of the nature of oscillations
that constitute the circadian
clock”**



Colin S. Pittendrigh

Malcolm Sargent identified the **band (bd)** mutation (1972) and developed conditions, using **race tubes**, to promote the best expression of the rhythm. He also showed that the rhythm can be reset by light and temperature pulses.



Asexual Development Cycle

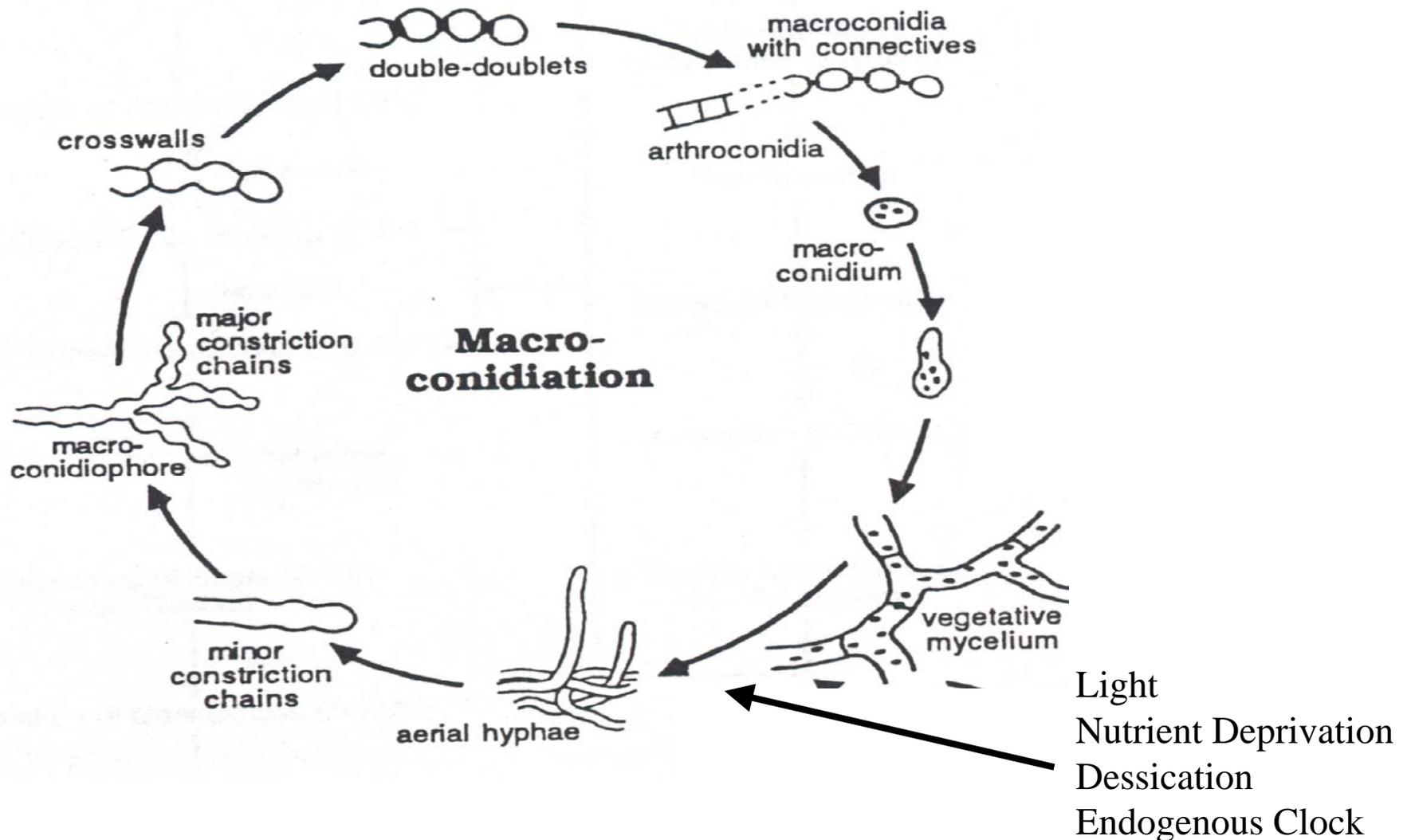
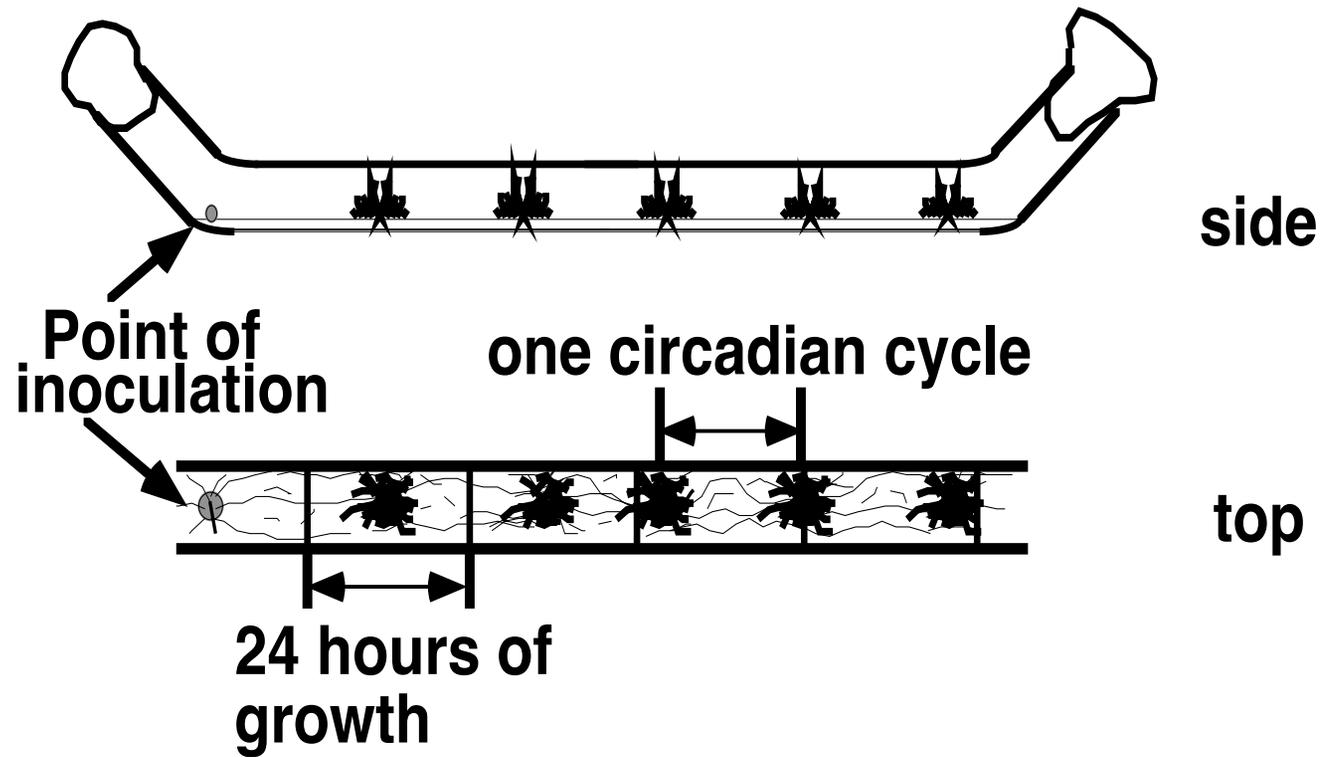
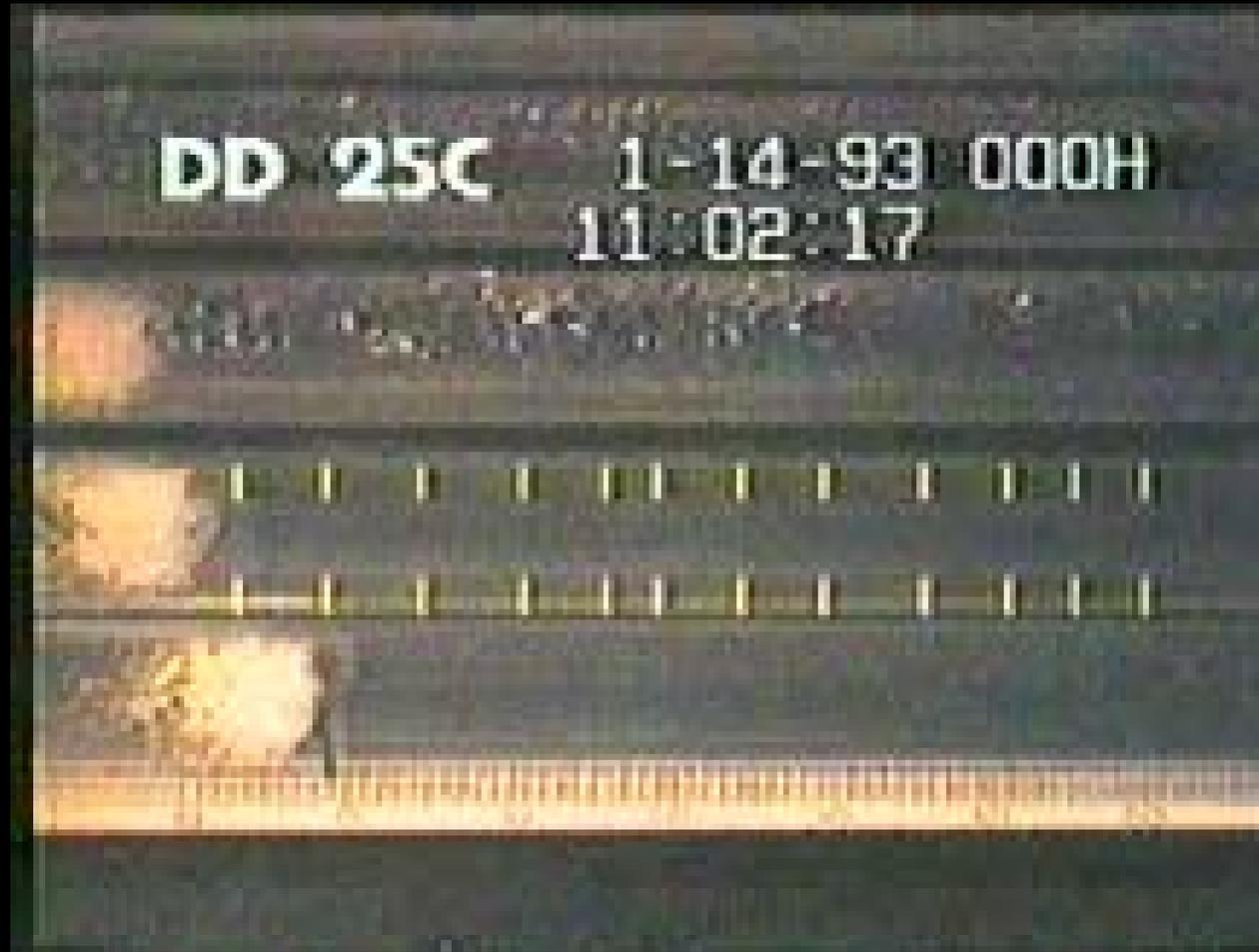


Diagram of a race tube

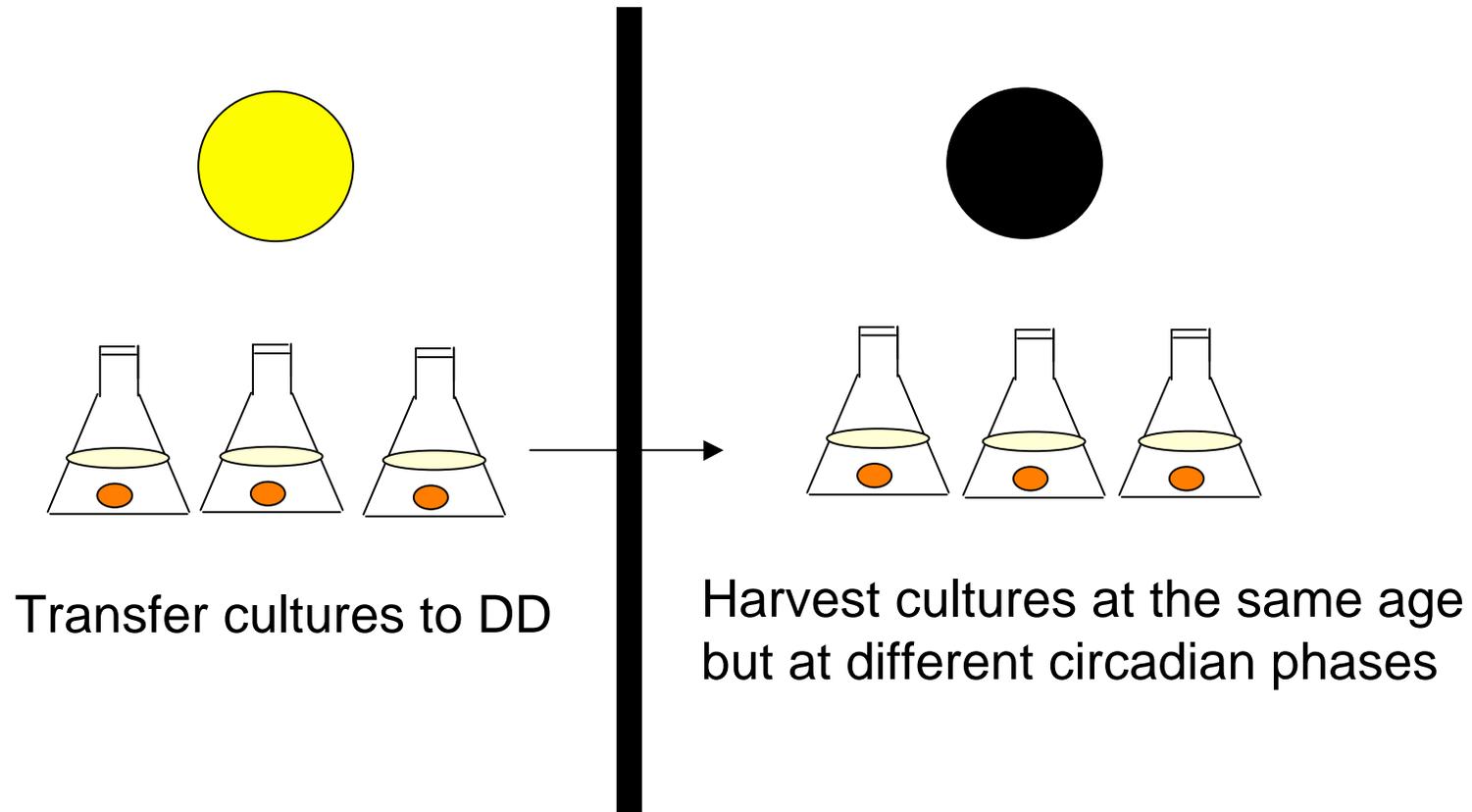


******band (bd) strain******



Time lapse video of Neurospora Circadian Conidiation Rhythms by Van Gooch and Jason Thoen.

Development of **liquid culture** methods (Nakashima) to monitor cycling mRNAs and proteins.



* Luciferase and gfp variants

Jerry Feldman (1960s-1970s) was the first to identify mutations that alter the circadian rhythm of development in *Neurospora*.



Jay Dunlap and Jennifer Loros (1989) cloned the first clock gene (*frequency*) in *Neurospora* (McClung et al, 1989).

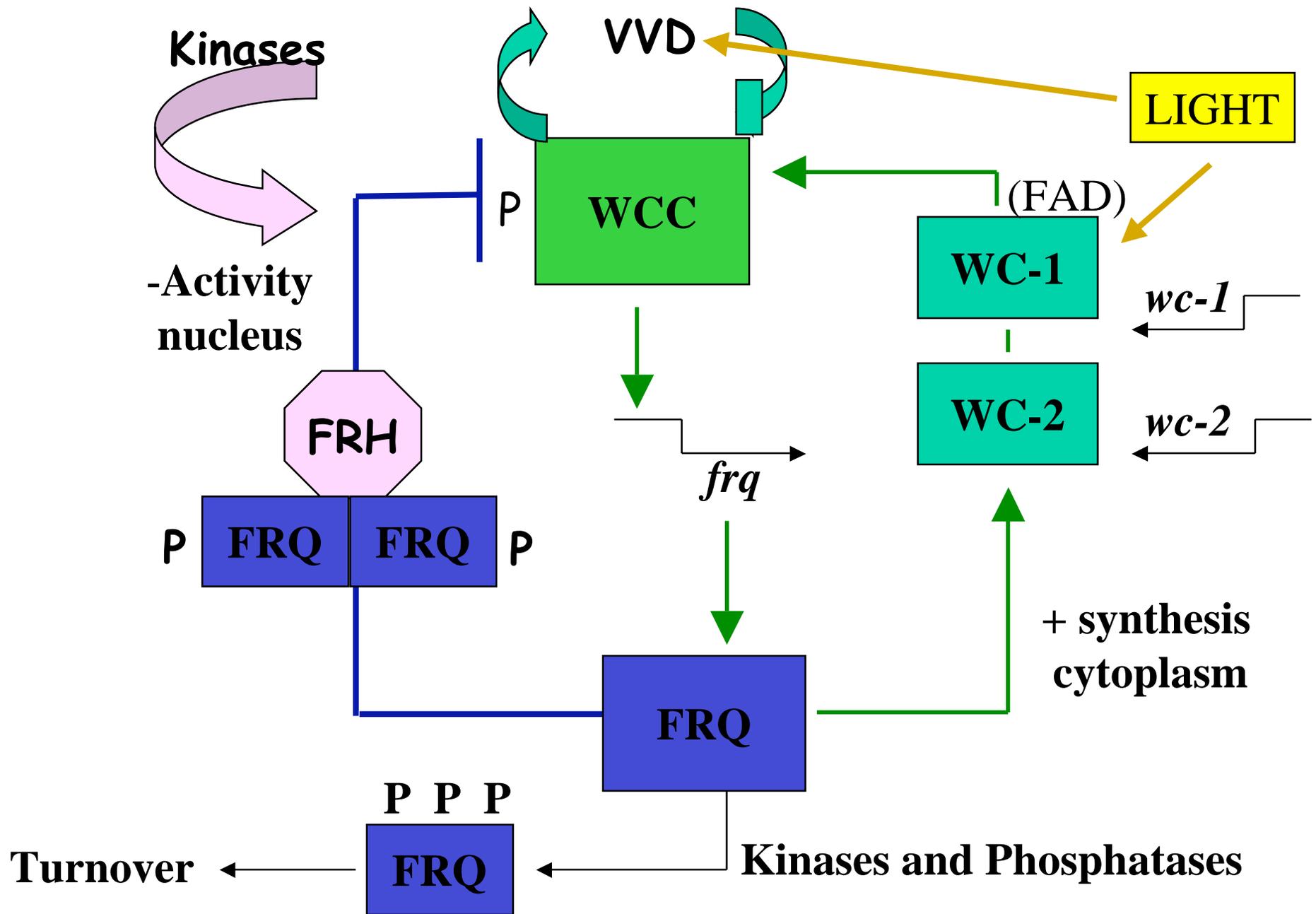
First global screen for ccgs

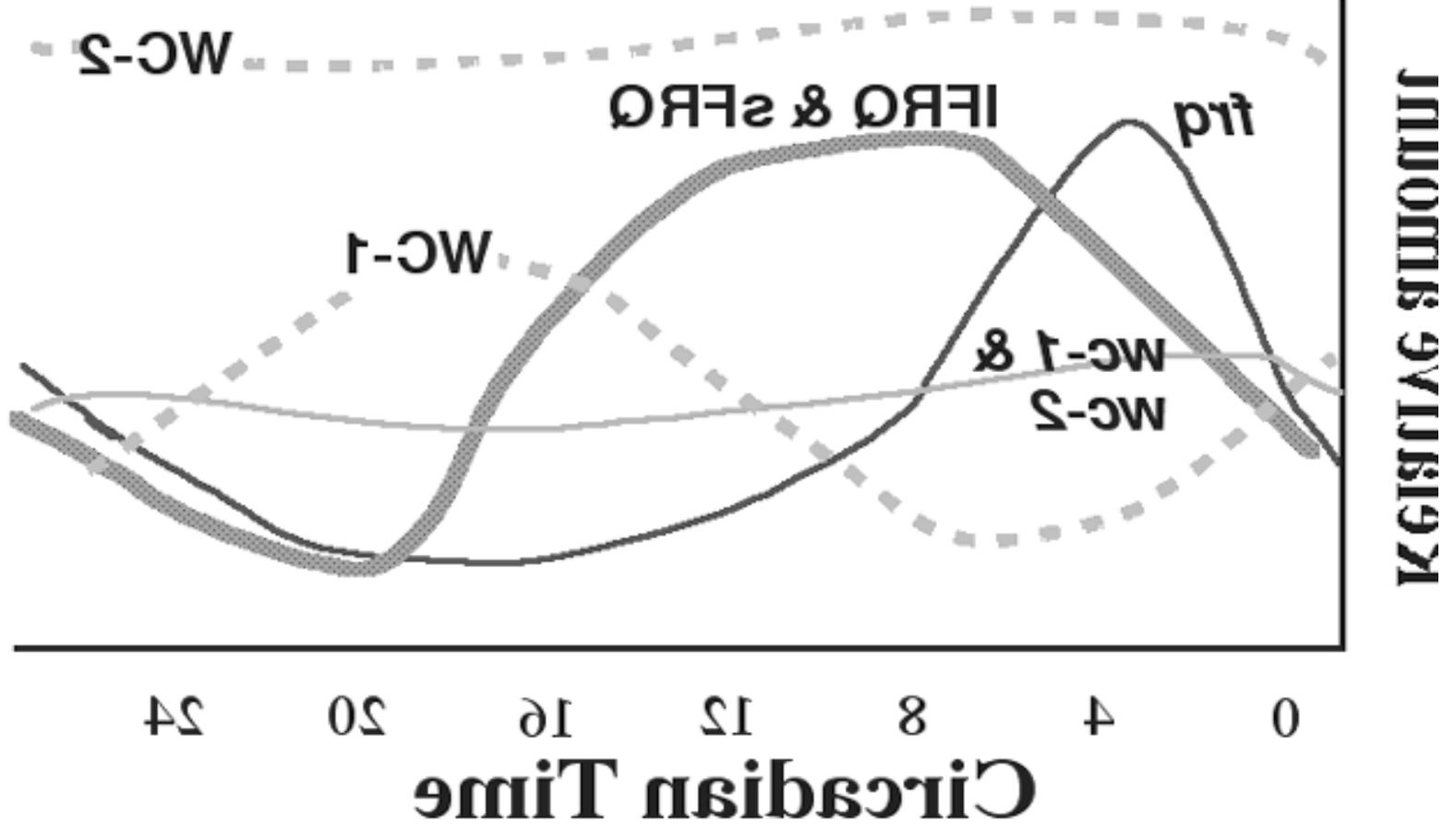
(Loros et al, 1989)



The beginnings of molecular/genetic analysis of the *N. crassa* clock

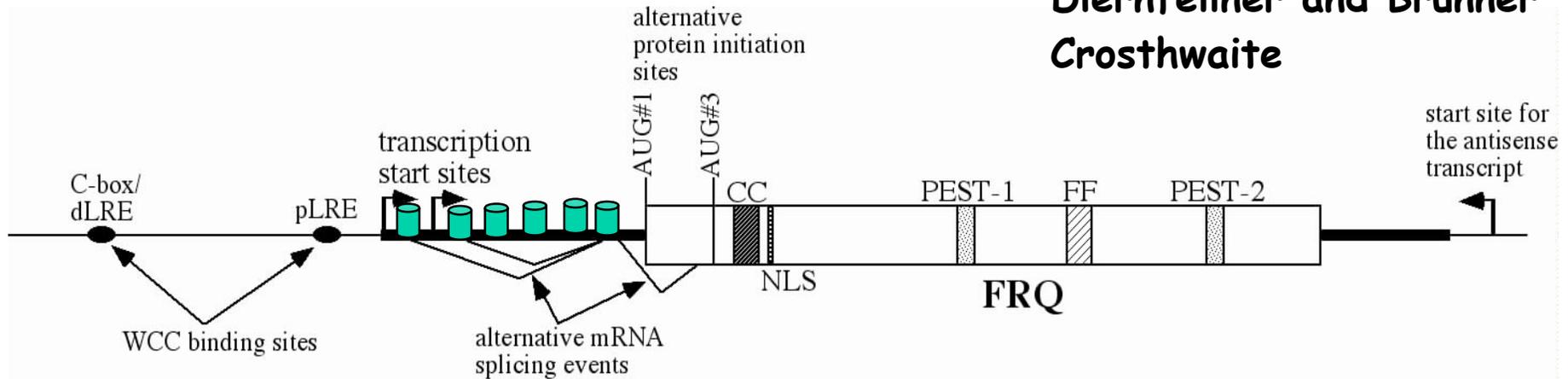






The effects of temperature on the FWO:

Liu, Merrow, Loros, Dunlap
Colot, Loros, Dunlap
Diernfellner and Brunner
Crosthwaite



Temperature increase - *frq* mRNA levels go up, more FRQ protein made.

Transcription initiation more often at the downstream site of transcription.

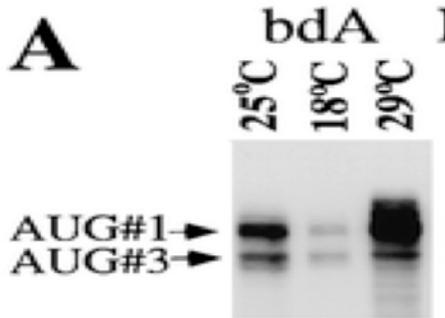
The increase in FRQ protein levels depends on the 5' UTR and 6 uORFs. Initiation at uORFs is more efficient at low temp.

The ratio of I and sFRQ are temperature-dependent as well. See more IFRQ as the temp increases. Strains that can only express IFRQ are arrhythmic at high temps, whereas those that can only express sFRQ lose rhythmicity at low temp. Intron 6 contains the translation initiation site of IFRQ - very little splicing at high temp.

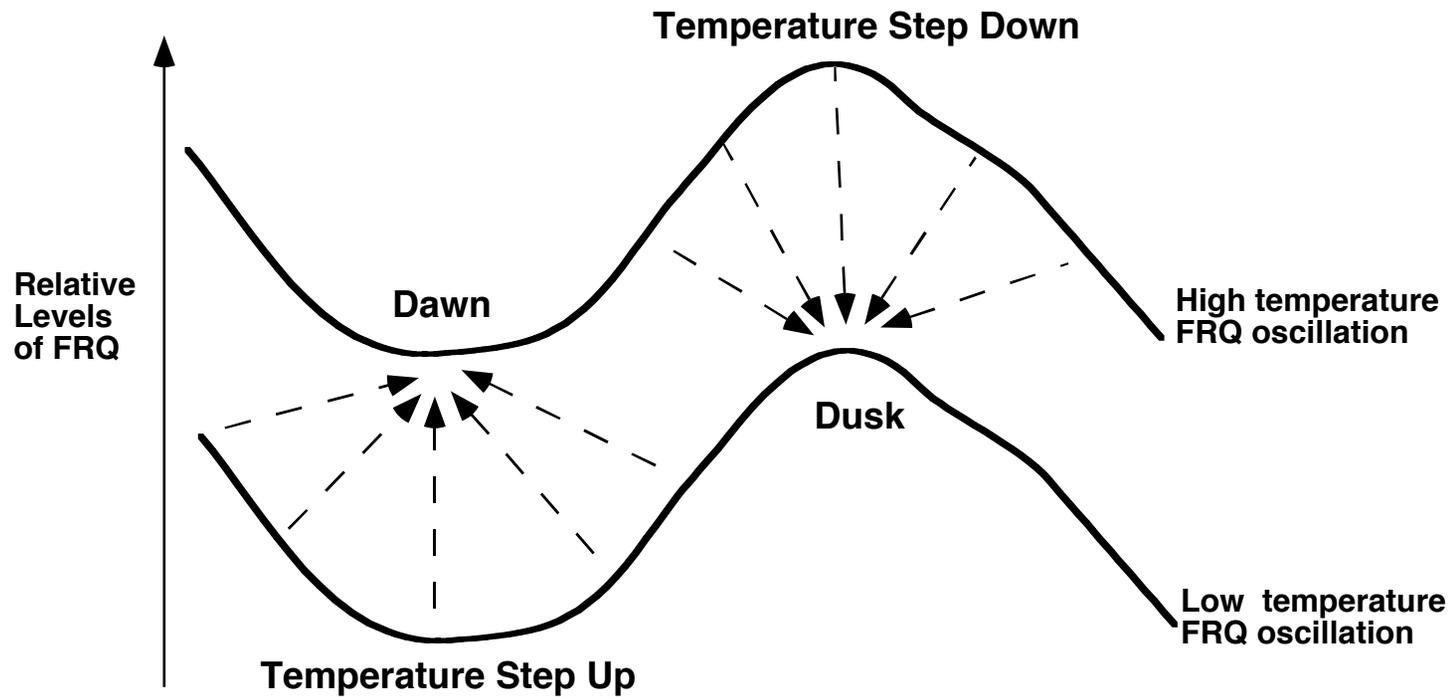
Helps to explain temperature entrainment, not compensation

Temperature resetting of the *Neurospora* clock

Posttranscriptional mechanism

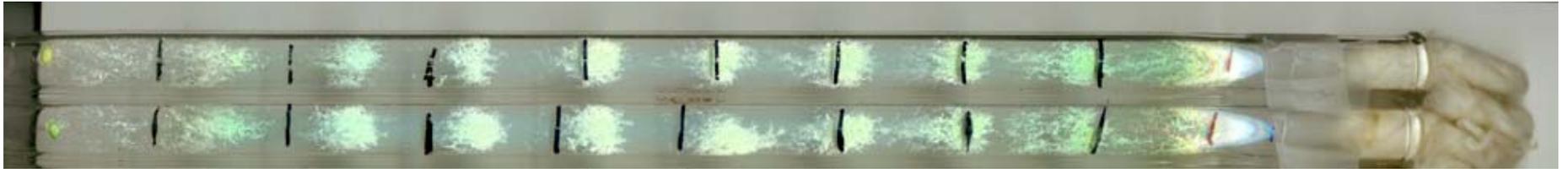


Liu Y, Garceau NY, Loros JJ, Dunlap JC. Cell. 1997 May 2;89(3):477-86

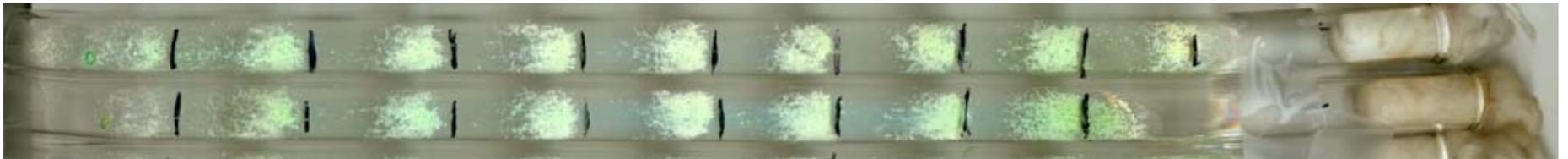


The effects of light on the FRQ/WCC oscillator

DD



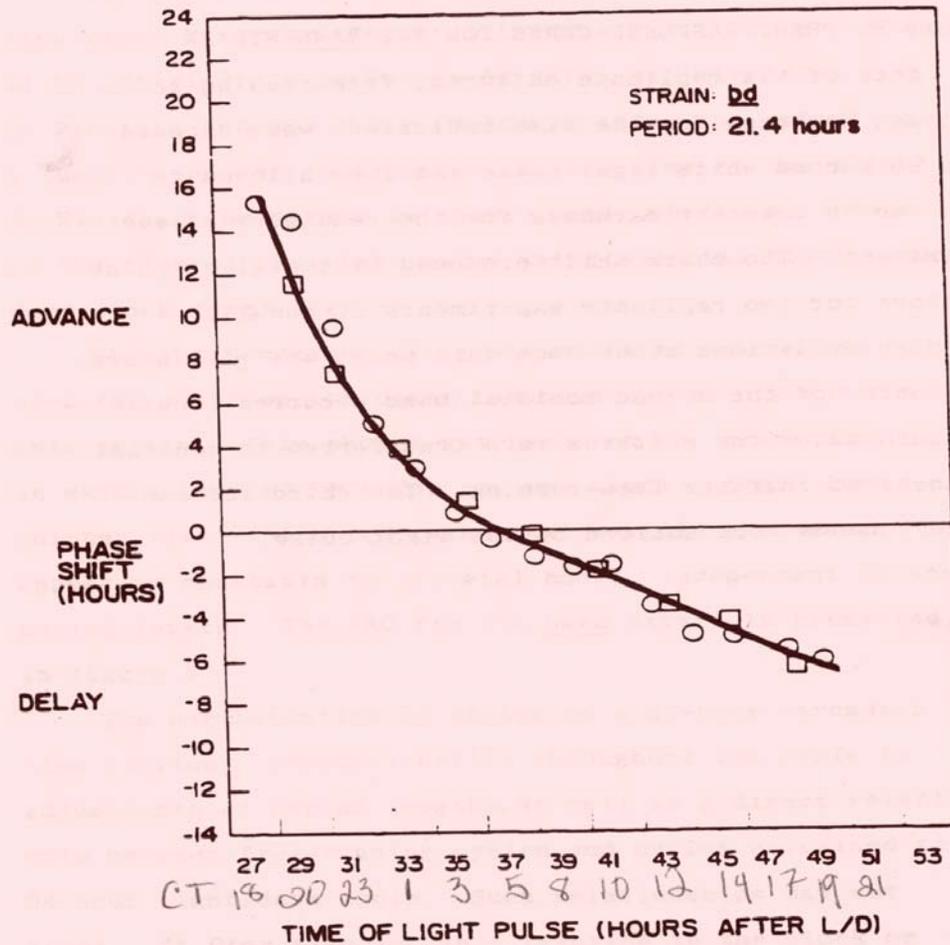
LD 12:12



Constant light

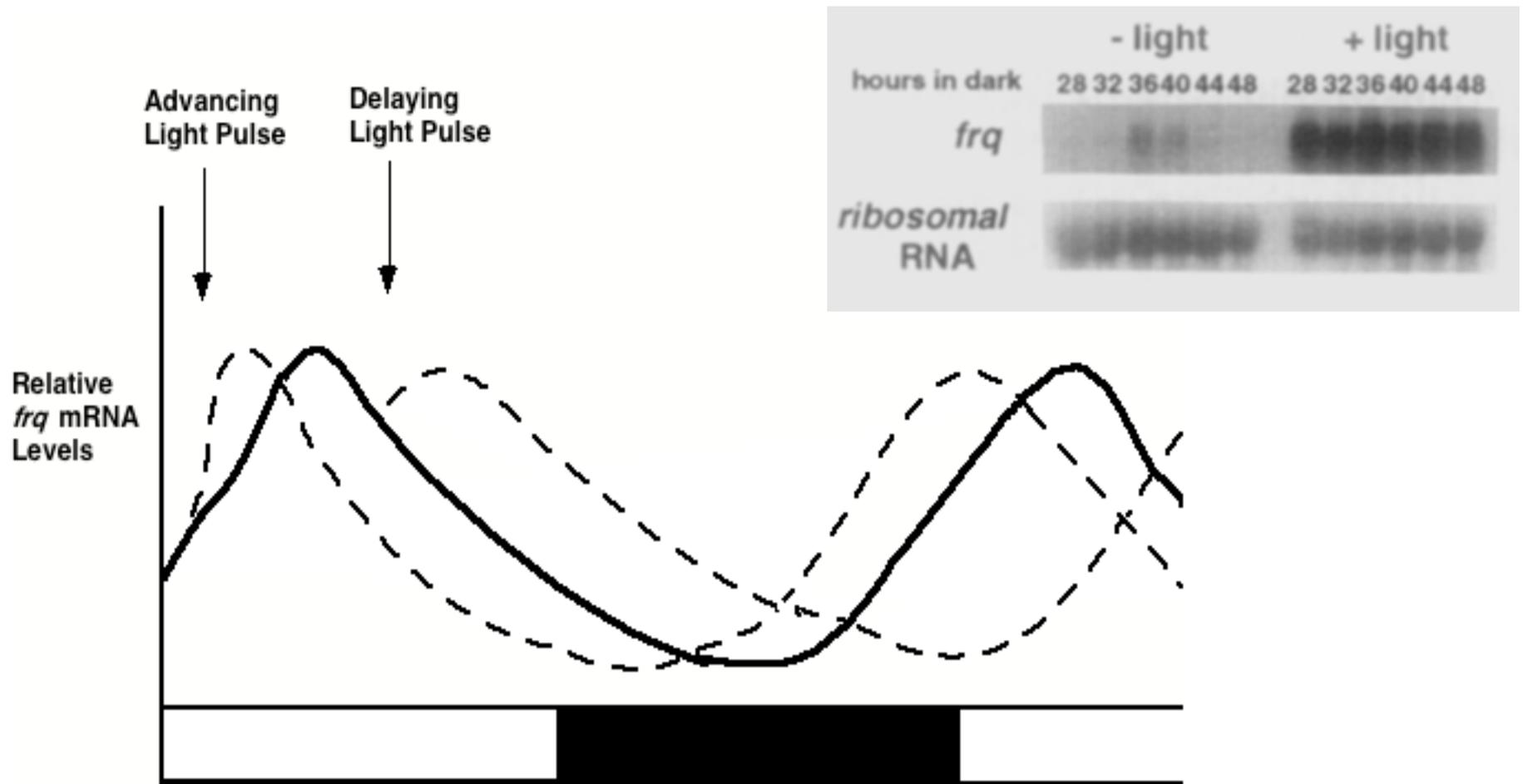


90 sec white light



Dharmanda and Feldman

How Light Resets the Clock



Shows how the same signal can elicit a delay and an advance of the clock

Crosthwaite SK, Loros JJ, Dunlap JC Cell. 1995 Jun 30;81(7):1003-12.

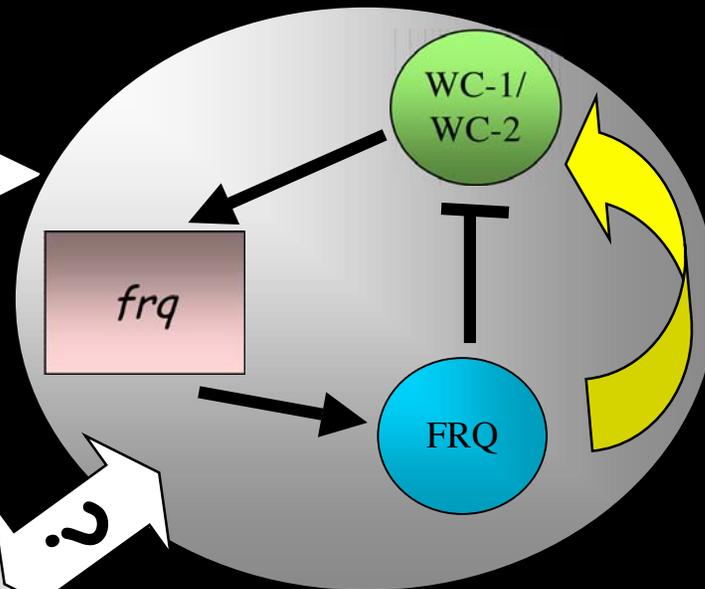
The *Neurospora* Circadian Clock System

Input Signals

Light

Temp

Oscillator(s)



Rhythmic Output

→ Development



? → Stress Responses

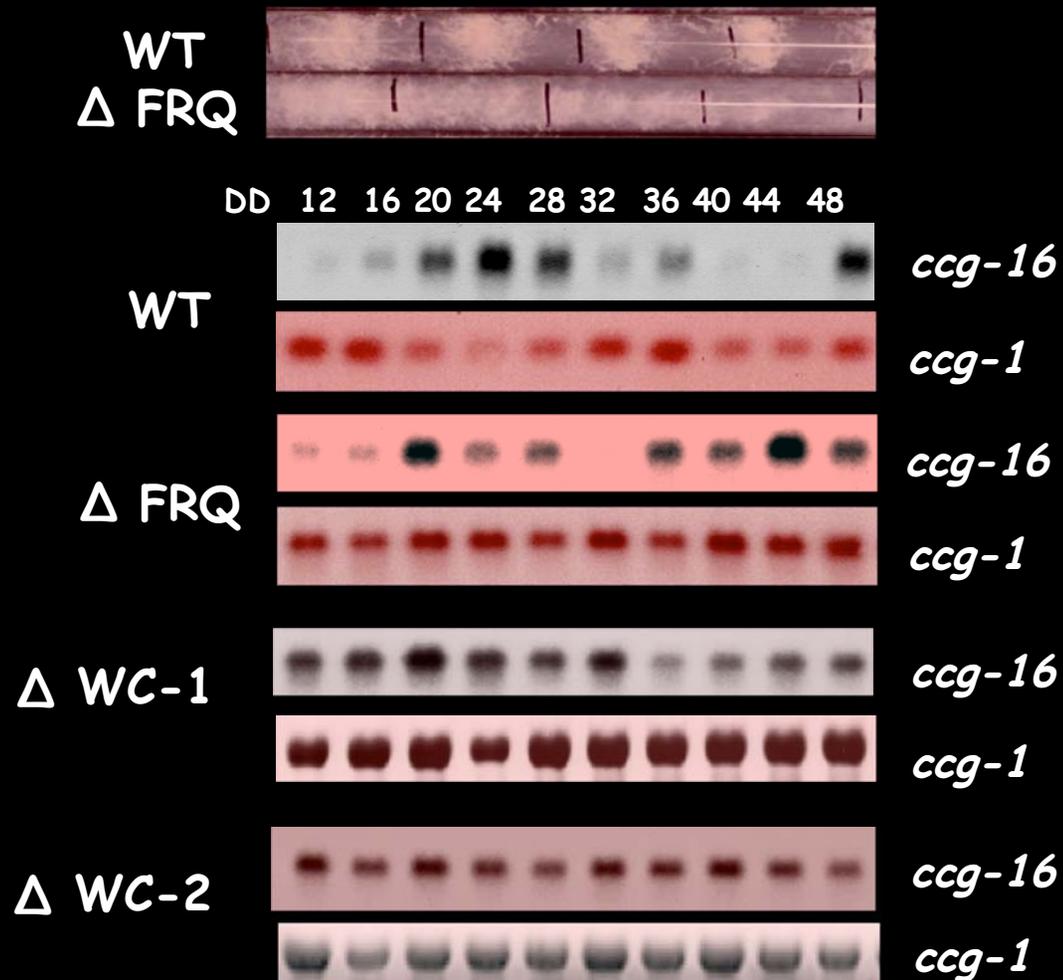
→ Gene Expression
(ccg)
up to 20%

Other
Oscillators?

Evidence for multiple oscillators within Neurospora cells

- **Genes/proteins** that are rhythmic in the absence of a functional FRQ/WCC oscillator. Peter Ruoff - nitrate reductase
- **Developmental rhythms** can be observed in strains that do not have a functional FRQ/WCC oscillator under certain growth conditions and in certain mutant backgrounds. Jennifer Loros, Stu Brody, Pat Lakin-Thomas, Martha Merrow, Till Roenneberg, Yi Liu

The evening-specific *ccg-16* gene is regulated by a FRQ-less Oscillator (FLO)

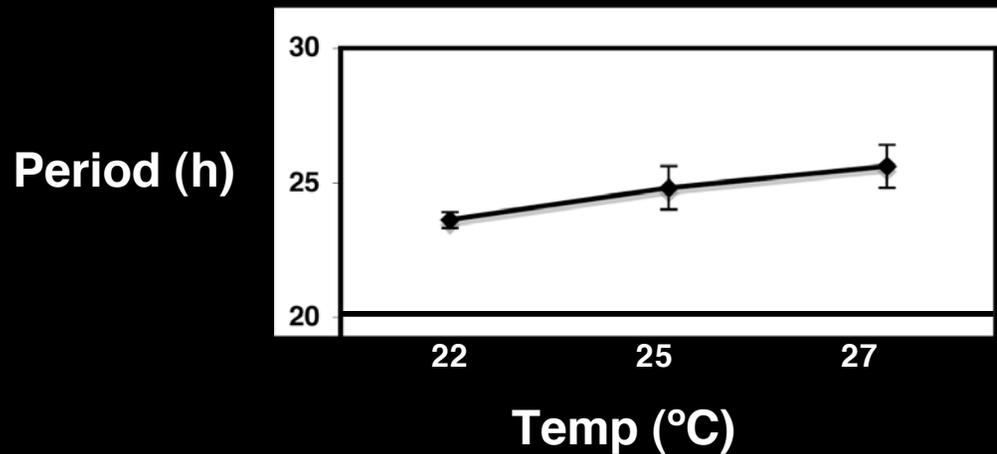


* WT = *bd*

Phase

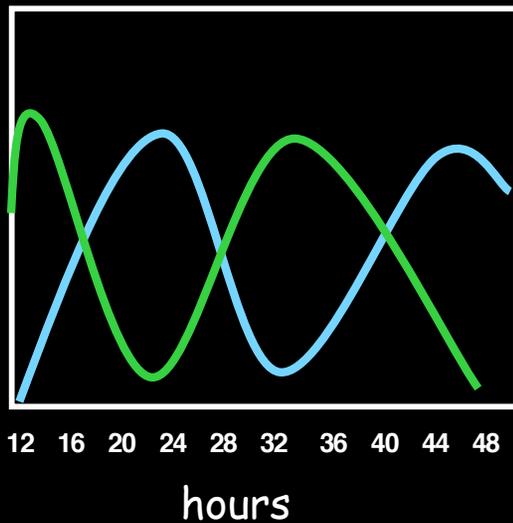
The *ccg-16* mRNA rhythm is temperature compensated

Δ FRQ



$$Q_{10} = 0.9$$

The FLO is synchronized by temperature shifts



Δ FRQ

12 16 20 24 28 32 36 40 44 48

DD30 to DD25

LL30 to LL25

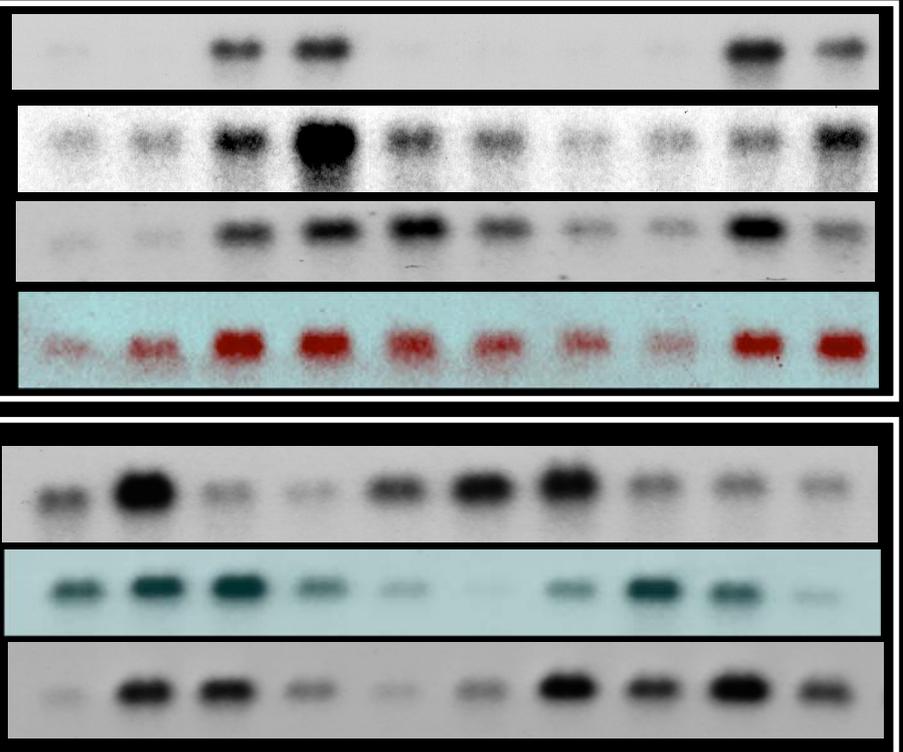
DD30 to LL25

LL30 to DD25

DD25 to DD30

LL25 to LL30

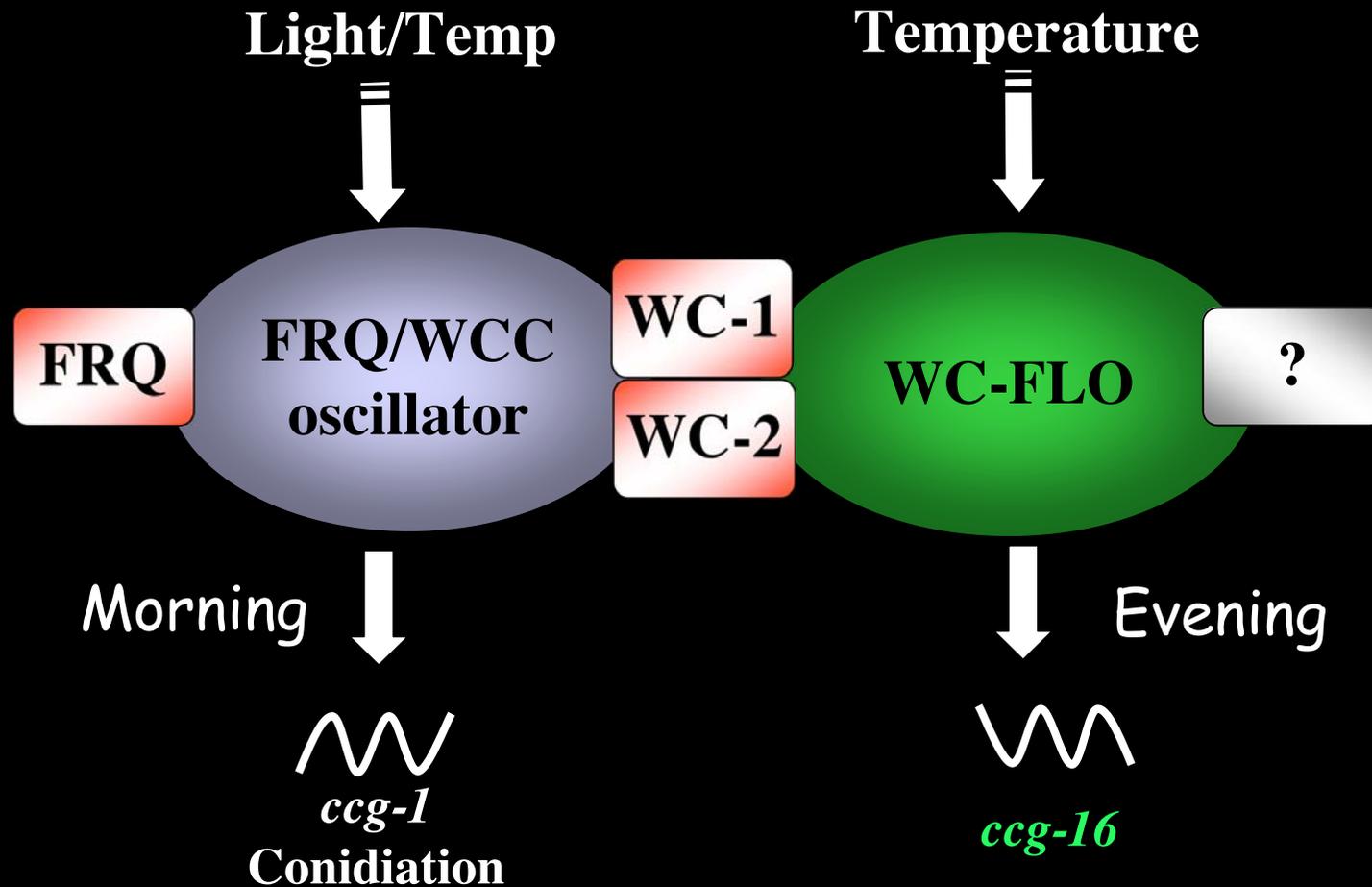
DD25 to LL30



A circadian WC-FLO:

- a. The WC-FLO programs a free-running rhythm in the absence of a functional FRQ oscillator.
- b. The WC-FLO is temperature compensated.
- c. The WC-FLO is responsive to temperature shifts.

Evidence of a 2nd Circadian Oscillator



Pitt E&M oscillators

Evidence for a 3rd oscillator

Continuous bright light (1200 Lux)

Strain

Period 22°C

WT



LM1



18 h \pm 1.2

Δ WC-1, LM1



23 h \pm 1.8

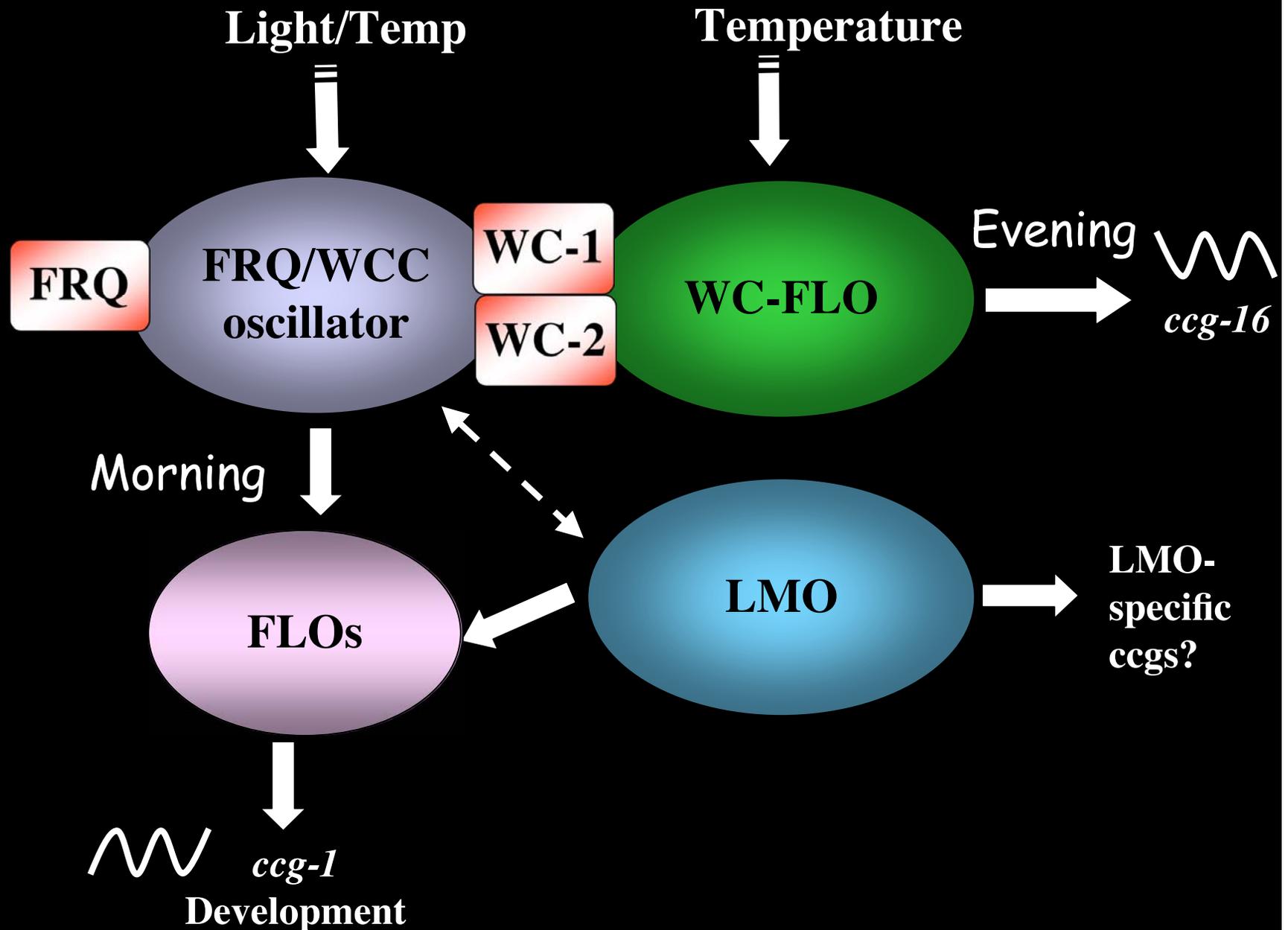
Δ FRQ, LM1



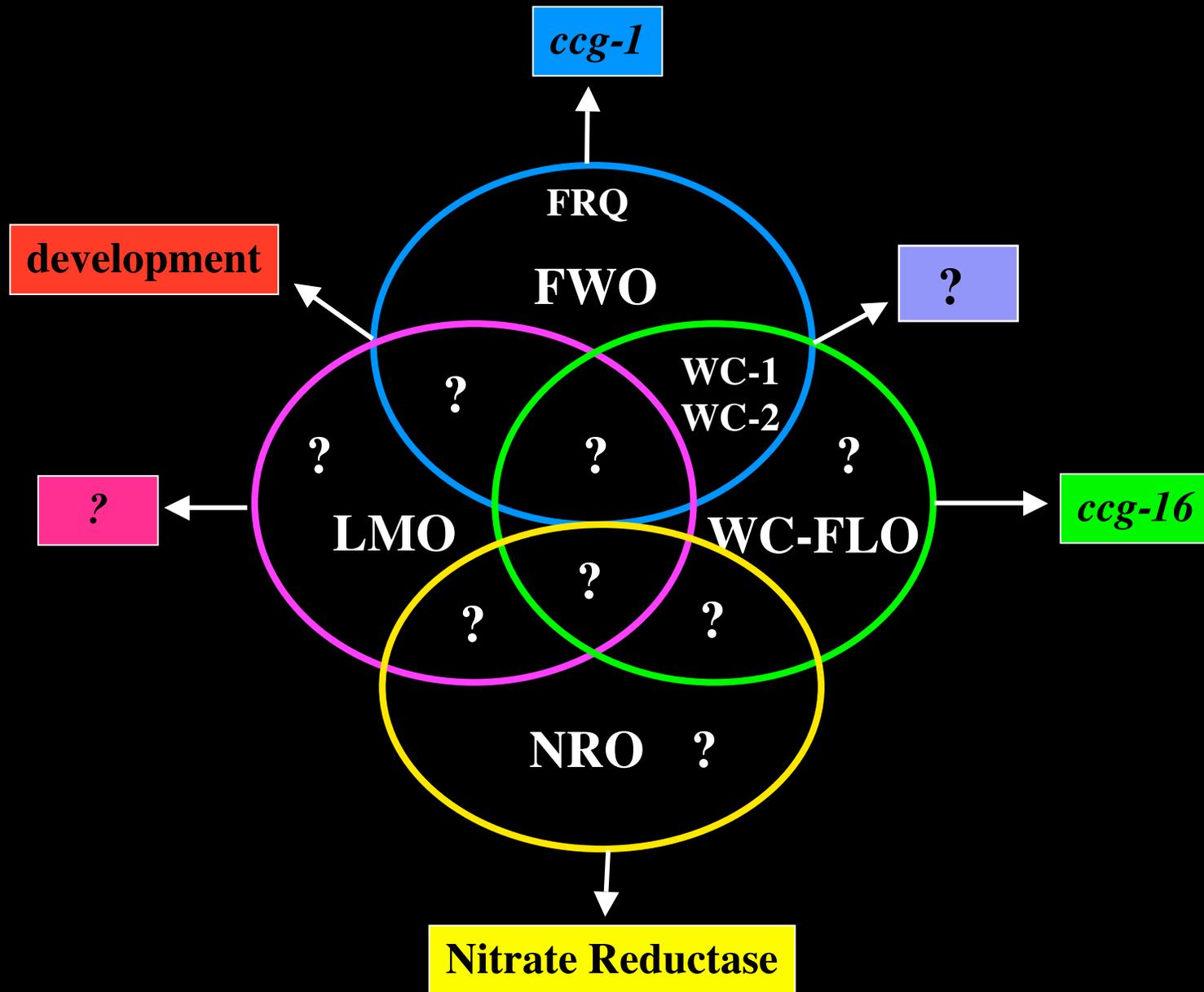
23 h \pm 1.6

Huang, H., Seo, K., unpublished

Working Model of the Organization of the Neurospora Clock



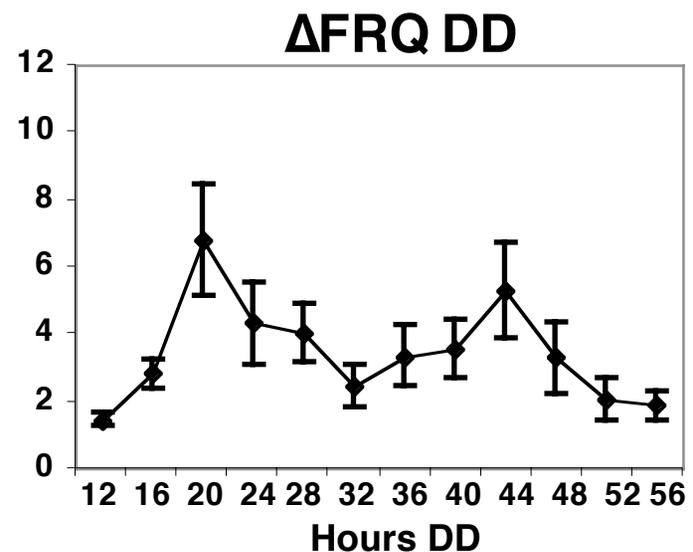
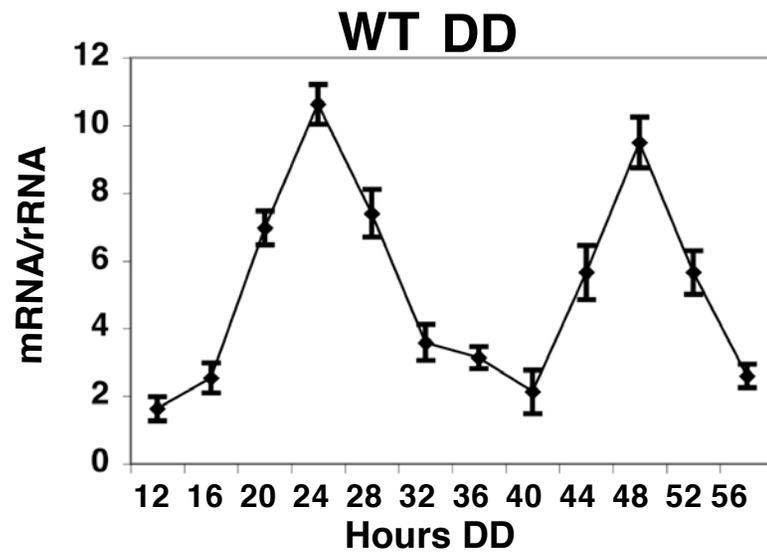
Multilayered Organization of a Circadian Clock



Why multiple oscillatory loops?

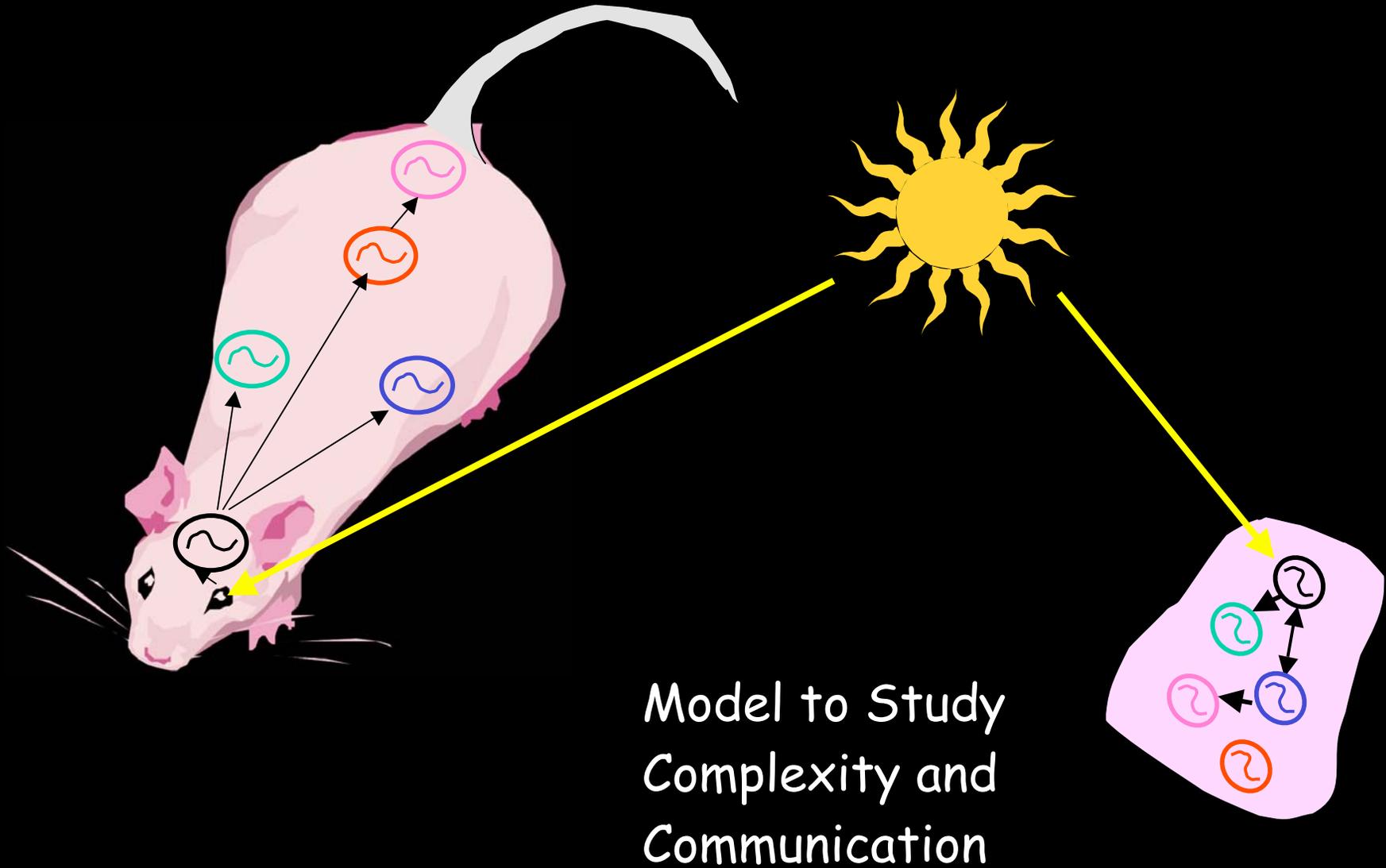
- Different inputs/outputs
- Provide precision?

The FRQ/WCC oscillator affects the phase and amplitude of *ccg-16* rhythms



$N \geq 5$

Clock organization and oscillator complexity



Output pathways

How do oscillators signal through output pathways to control overt rhythmicity?

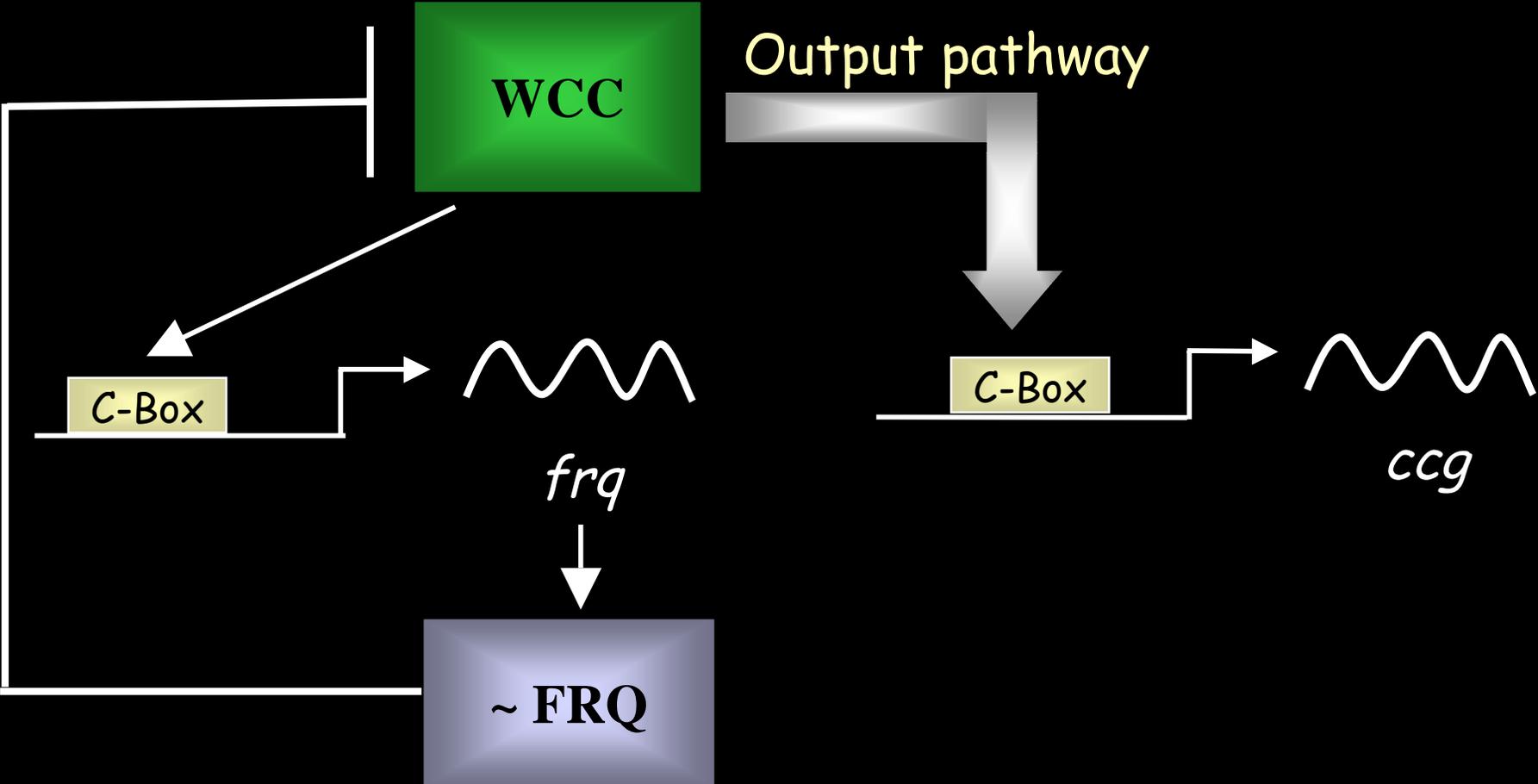
Focus on the FRQ/WCC oscillator

182 clock-controlled genes (ccgs)/~1400
that peak in expression at different times of day

Category	#
Cell Division	2
Cell Signaling/Communication	17
Cell Structure/Cytoskeleton	10
Cell Defense	6
Development	10
Gene Expression	5
Metabolism	32
Protein Synthesis	33
Protein Processing	10
Unclassified	<u>57</u>
Total	182

Full genome arrays in progress

18 ccgs contain a Clock (C) - box in their promoter

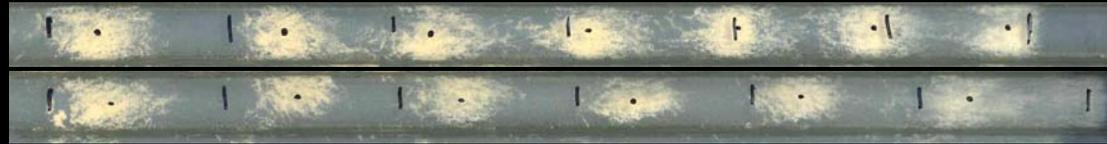


Knockout of candidate genes

SHK-1 kinase binding protein (*skb-1*)

WT

Δ *skb-1*



Period

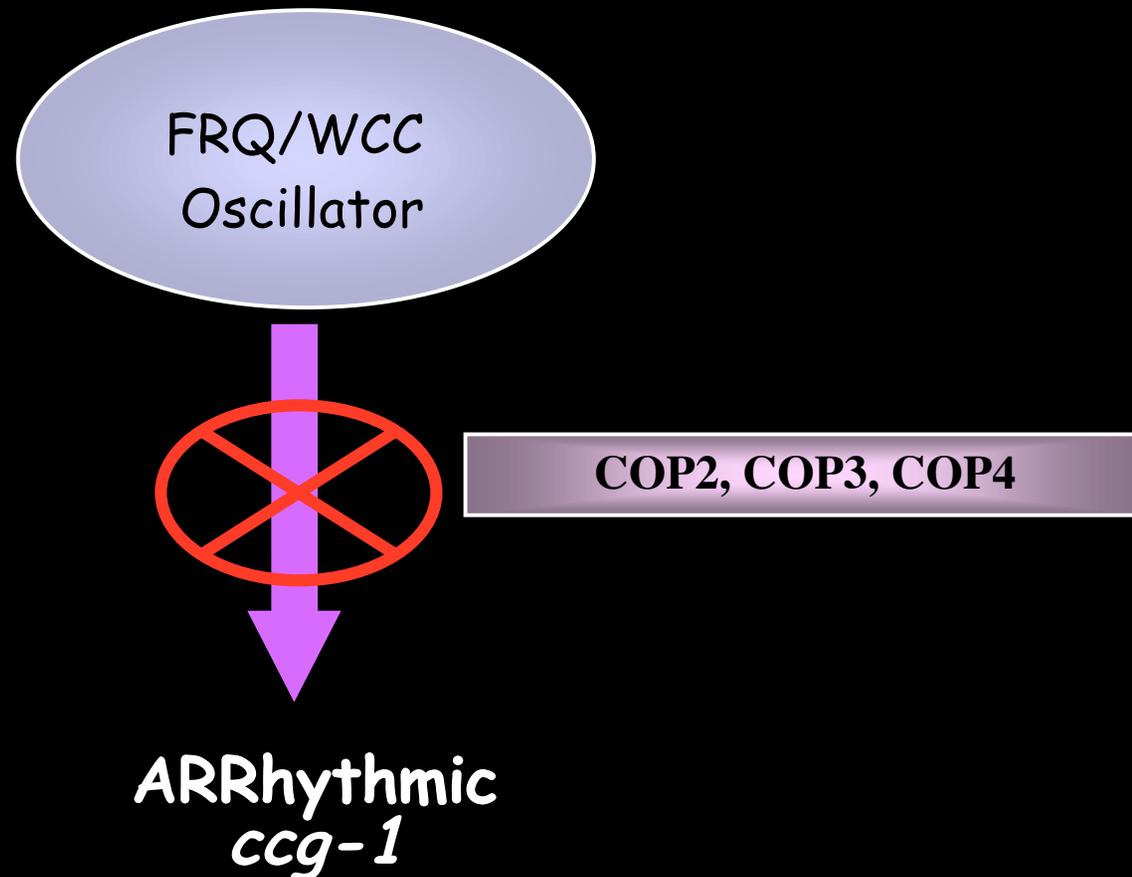
22 h

24 h



microarrays

Genetic selection for mutations in the output pathways
regulating rhythmic *ccg-1* expression



Vitalini et al, 2004 *Genetics*; Vitalini et al, PNAS submitted

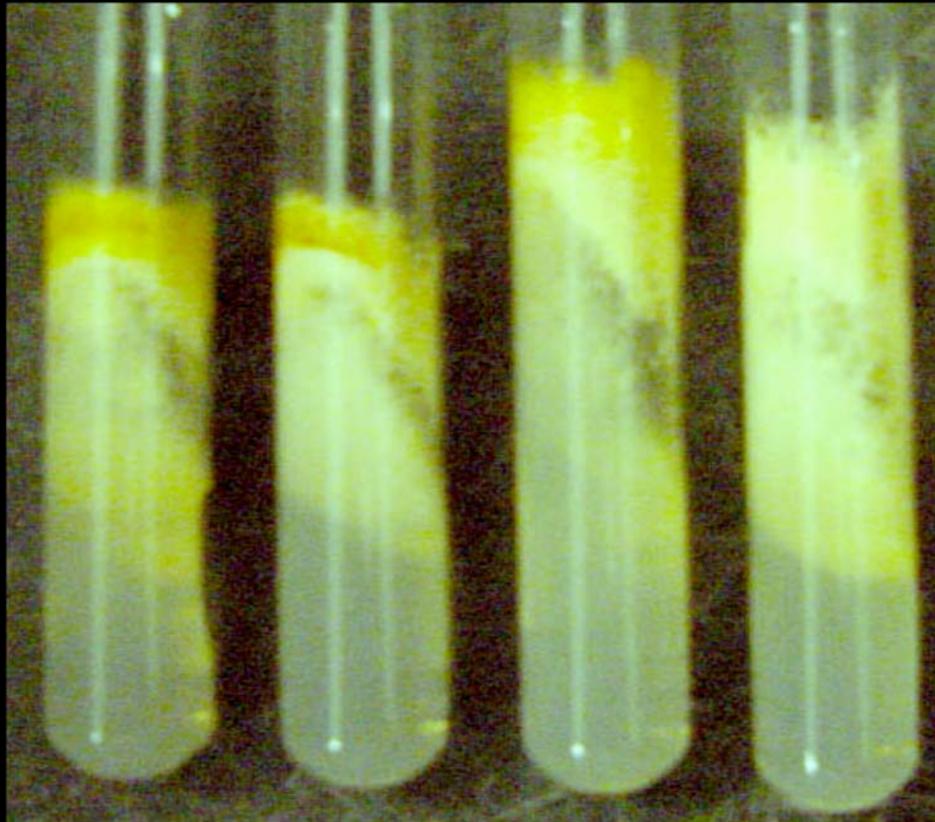
The *COP* mutants have a phenotype

COP2

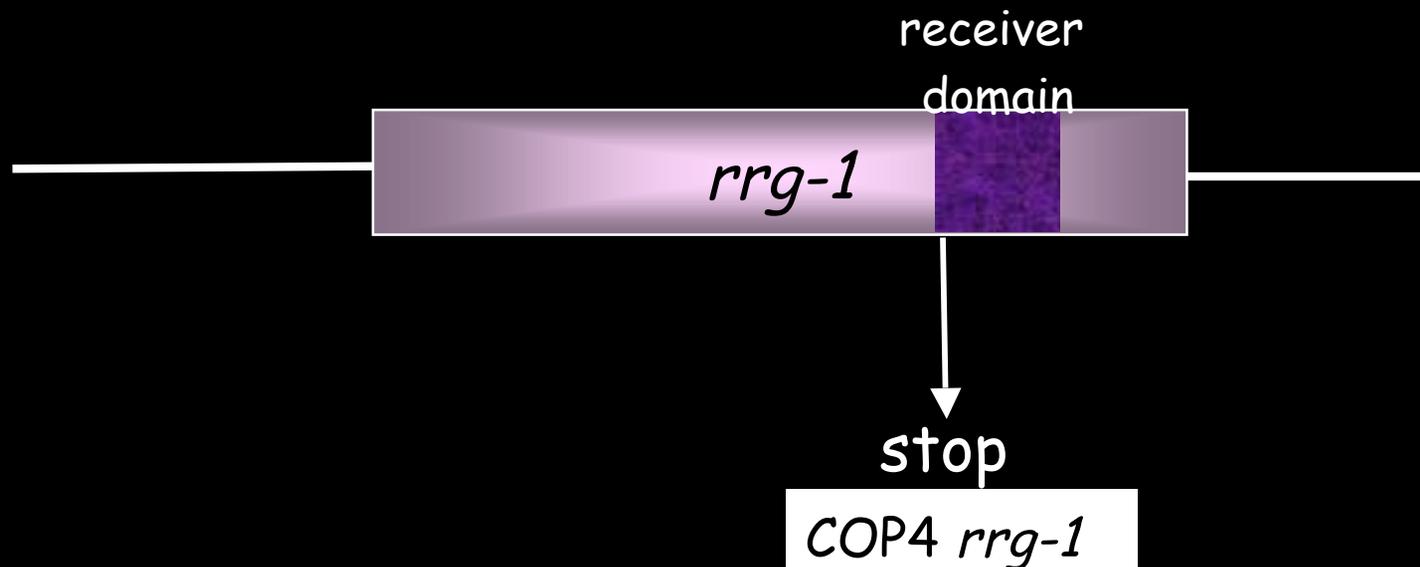
COP3

COP4

Parent

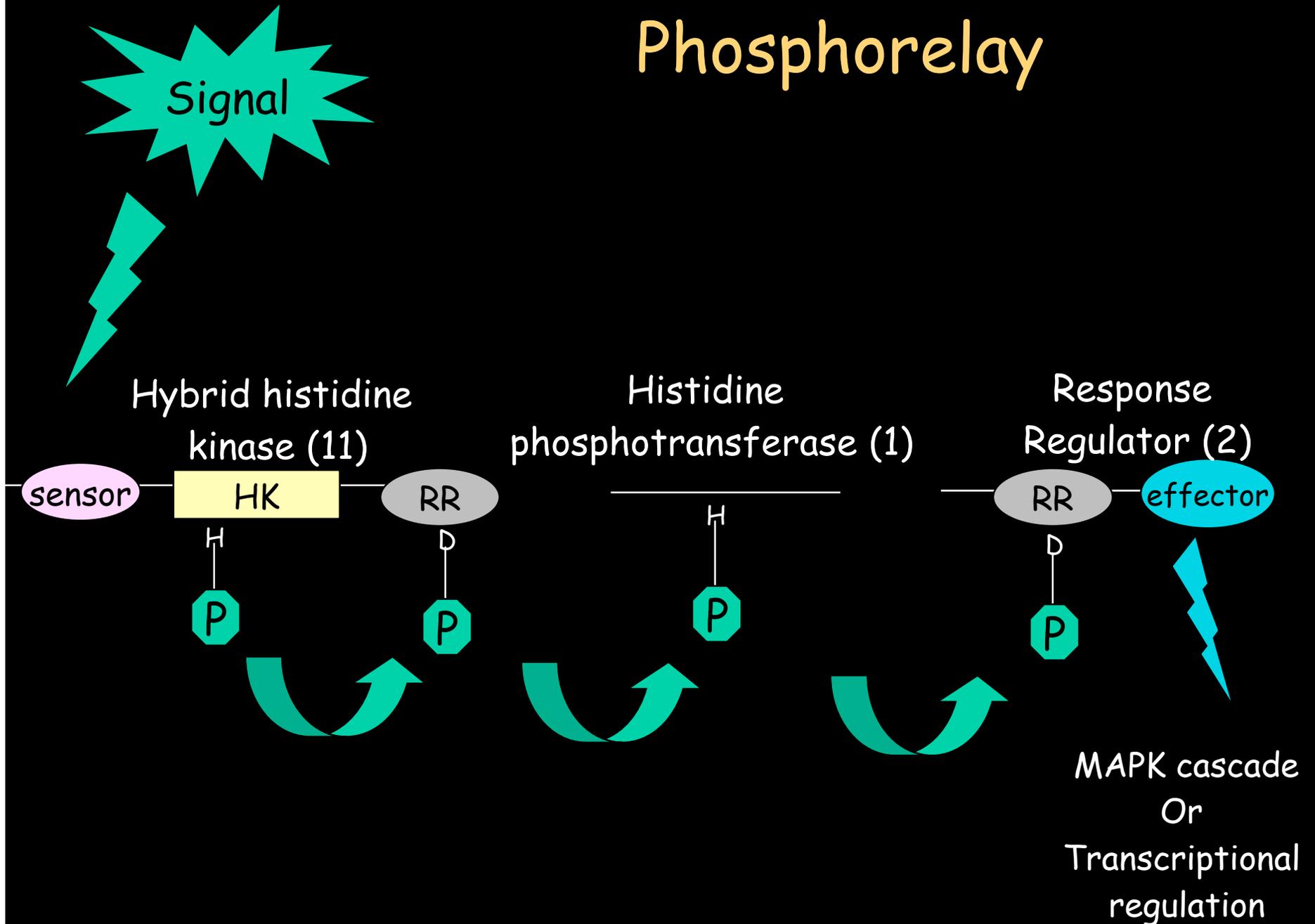


Genetic mapping localized *COP4* to the *rrg-1* gene
that encodes a response regulator



Jones, CA., Greer-Phillips, S., and Borkovich, KA. 2007 MBC in press

Phosphorelay



Phosphorelay



Sensor Histidine Kinase



Histidine Phosphotransferase



Response Regulator



COP4
COP3

MAPKKK



MAPKK



MAPK



COP2

Osmotic stress resistance, Conidial integrity, Sexual Dev't
Rhythmic gene expression

rrg-1 is not required for FRQ/WCC oscillator function

OS-1



HPT-1



RRG-1



OS-4



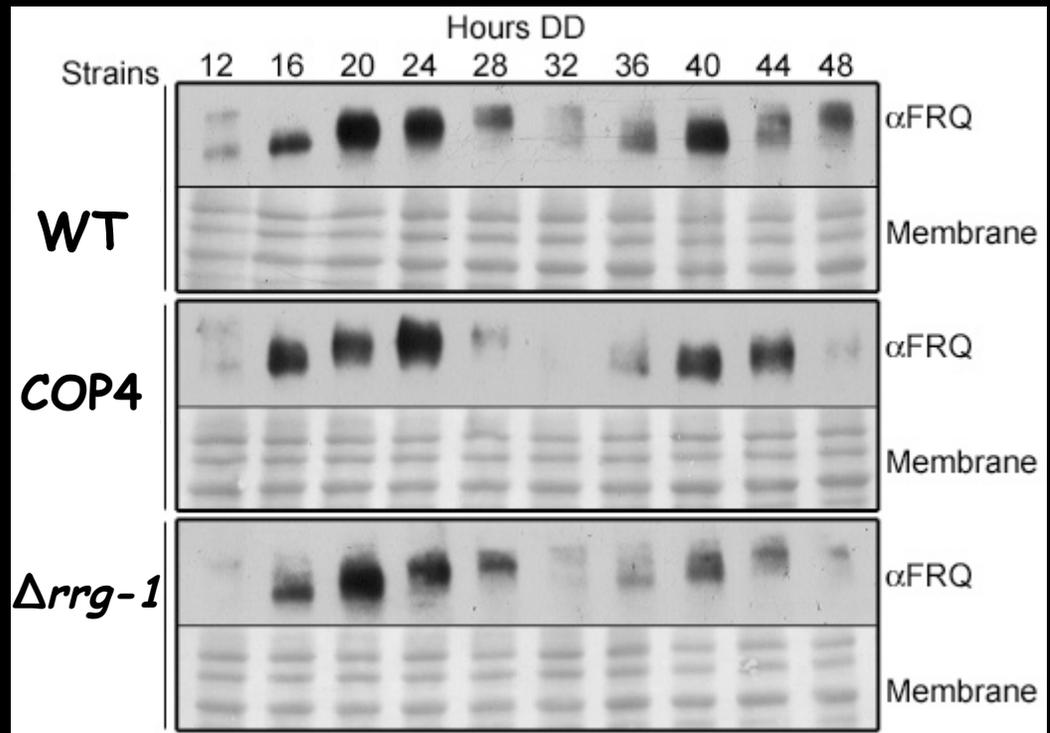
OS-5



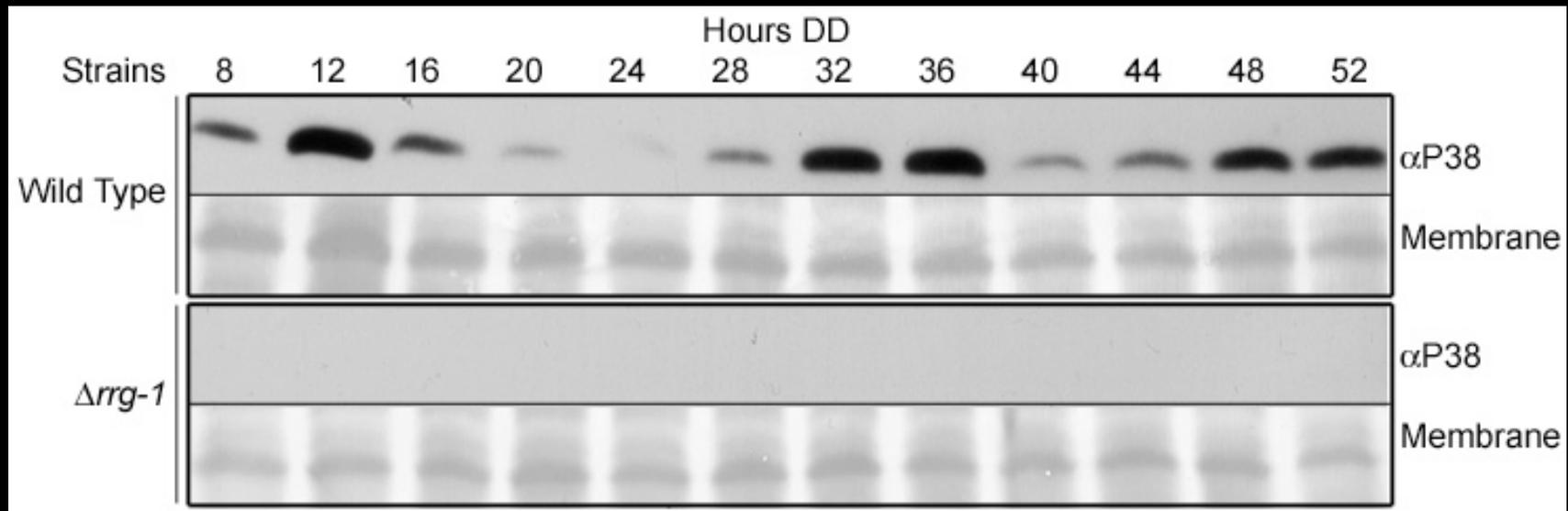
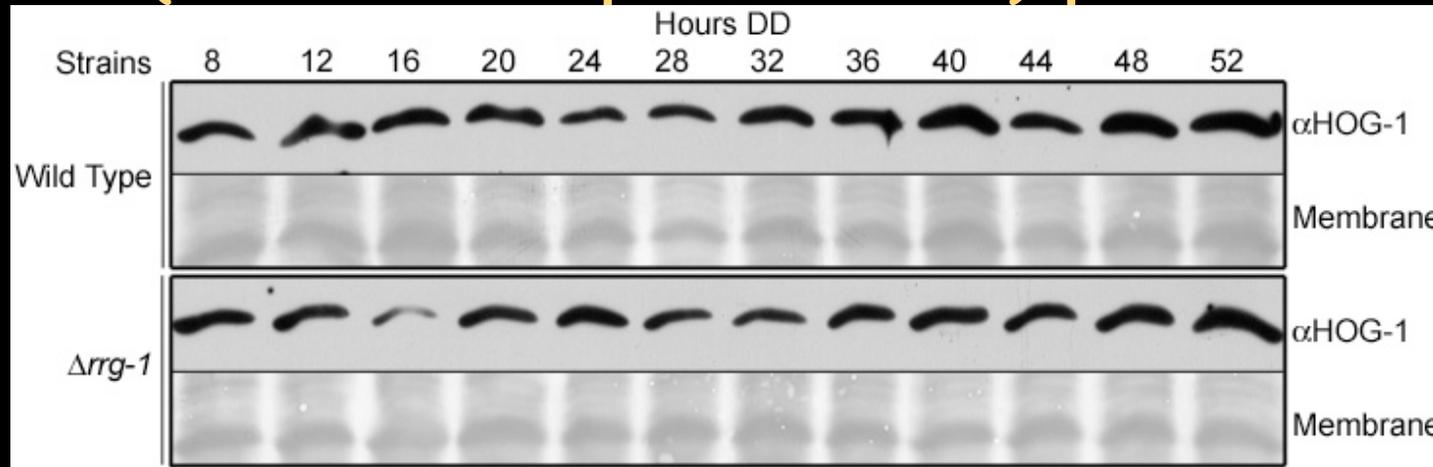
OS-2



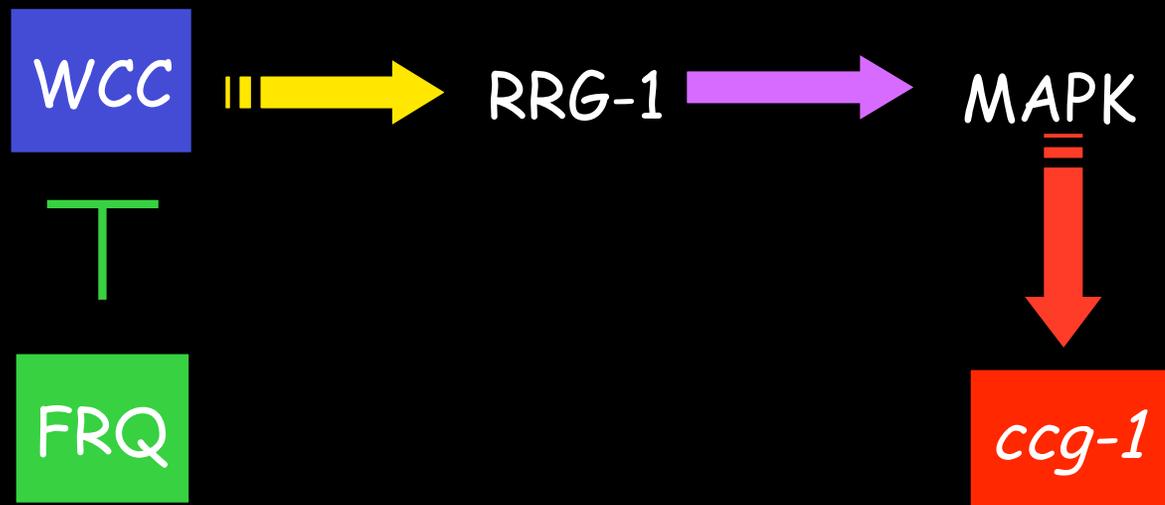
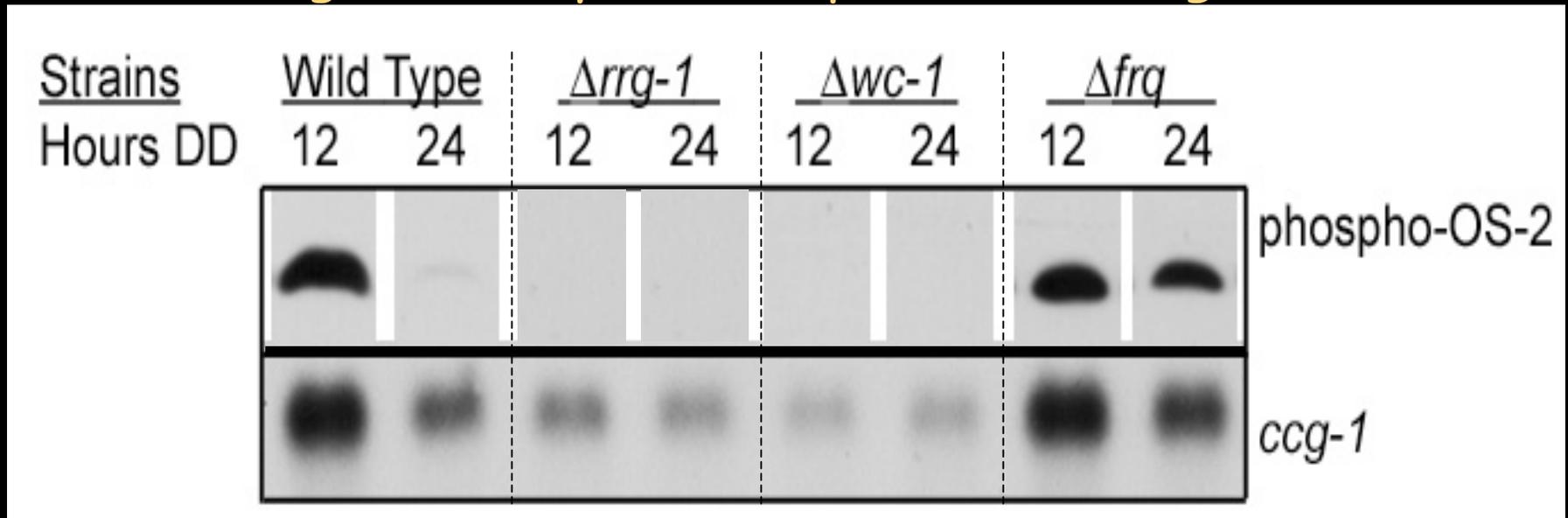
Rhythmic *ccg-1*



Circadian rhythm in phosphorylated OS-2 (conserved p38 MAPK) protein

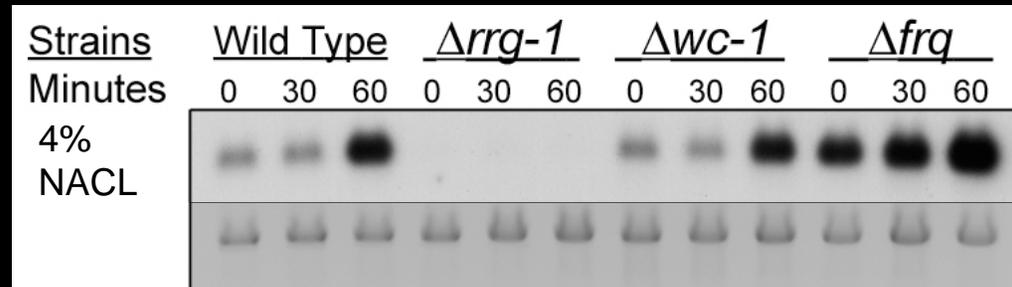


The HOG pathway is an output of the FRQ/WCC that regulates rhythmic expression of *ccg-1*

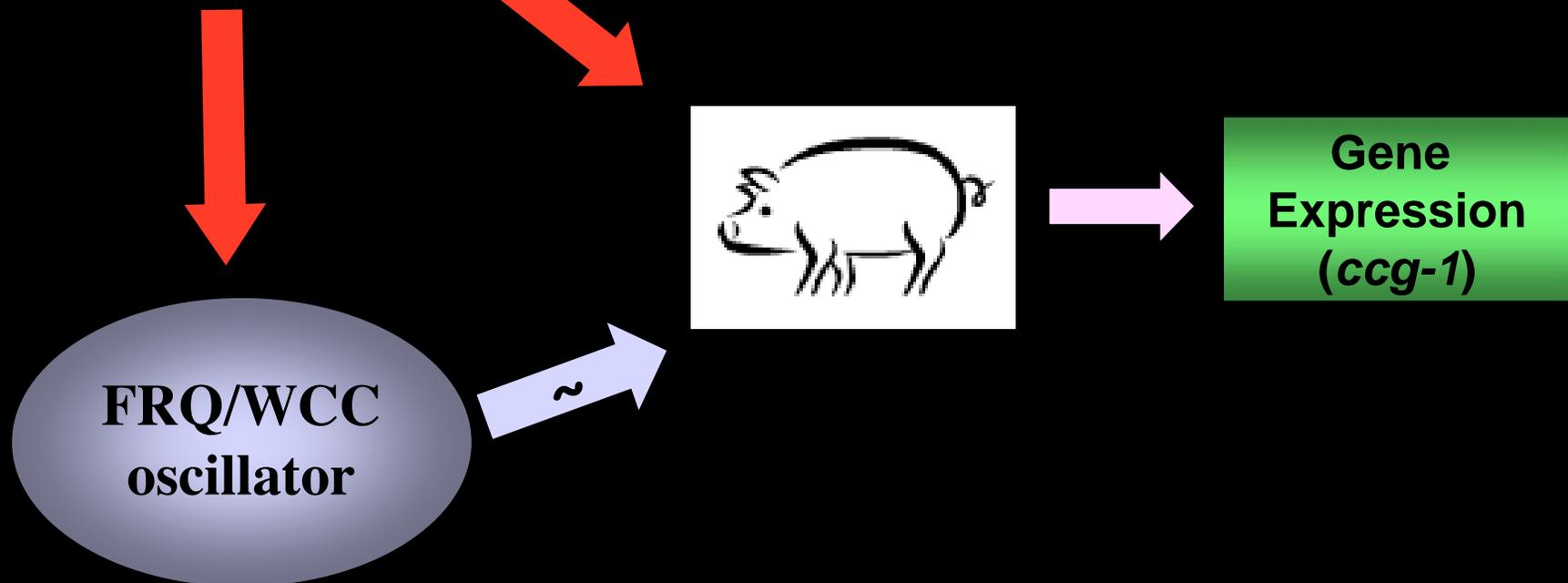


The FRQ/WCC oscillator is not necessary for the osmotic stress response

Osmotic Stress



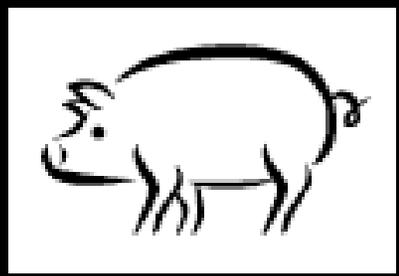
ccg-1
rRNA



Osmotic Stress

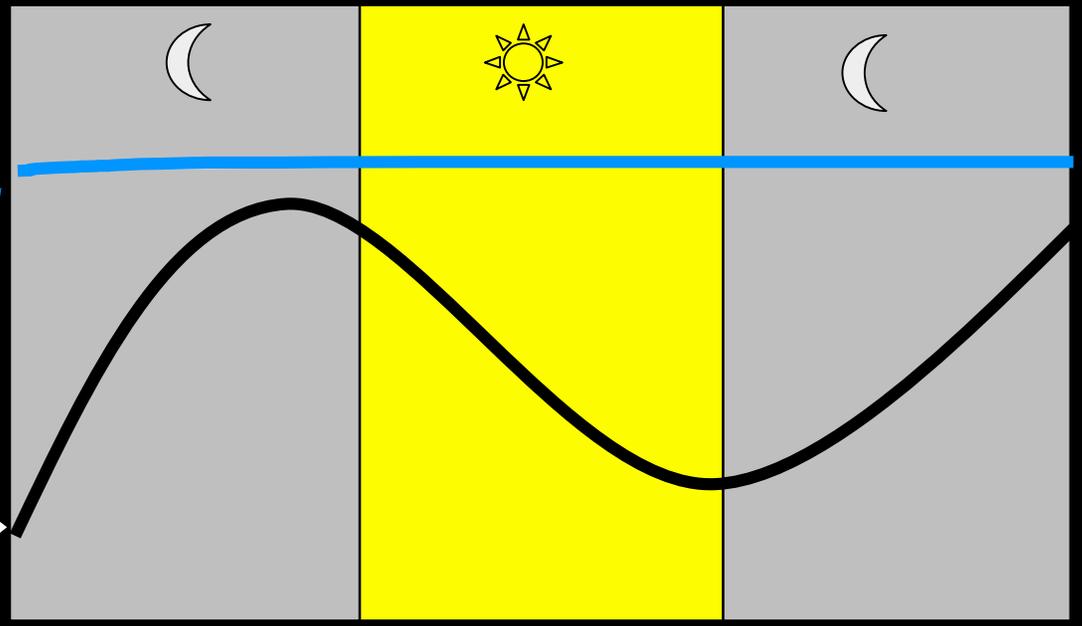


Control of the HOG pathway by the clock suggests that the rhythmic activity of the pathway allows anticipation and preparation for osmotic stress.



Transcription factors

Gene Expression



DD 12
Dawn

DD 24
Dusk

Summary:

Multiple (overlapping) oscillator loops exist within a single cell and function to regulate distinct biological rhythms, and may add to the precision of the clock.

- Important model for dissecting a multi-oscillator clock

Output pathways - The conserved HOG/P38MAPK pathway is an output pathway for the circadian clock.

- Circadian oscillators can hijack established signaling pathways
- P38 kinases mediate a variety of cellular behaviors including apoptosis, cell cycle, differentiation, inflammation, and tumorigenesis

Frequency Demultiplication

A work in progress....

Louis Morgan

*Oscillators can only be entrained within a given range of periods (limits of entrainment). When the period of the entraining cycle is close to half (third, fourth, etc.) of the oscillators endogenous period, the oscillator may entrain to every second entraining cycle (skipping a beat - known as frequency demultiplication).

In *N. crassa*, temperature, but not light cycles produce frequency demultiplied rhythms, suggesting the possibility that different mechanisms are used for entrainment.

i.e. does temp entrainment involve both the FWO and a FLO, whereas light involves only the FWO?

Or does light drive (mask) the rhythm

Frequency demultiplication was first discovered by Van der Pol in simulations of heart rhythms by electrical circuits, suggesting:

**common features
between electrical and biological oscillators.**

van der Pol, B. and van der Mark, J., "Frequency demultiplication", *Nature*, 120, 363-364, (1927)

Warm to cold
Dark to light



WT	DD		22
	LL		AR
	LD 12:12		24
	LD 6:6		12
	TE 12:12		24
	TE 6:6		24
	TE 3:3		22
<i>frq¹⁰</i>	DD		AR
	LL		AR
	LD 12:12		AR
	TE 12:12		24
	TE 6:6		12
	TE 18:18		AR

Is an intact FRQ/WCC oscillator (and not just FRQ) required for FD?

wc-1 and *wc-2* null strains
also do not undergo FD

In LL, *frq* mRNA is constantly elevated (10X the DD peak)

TE 12:12 LL



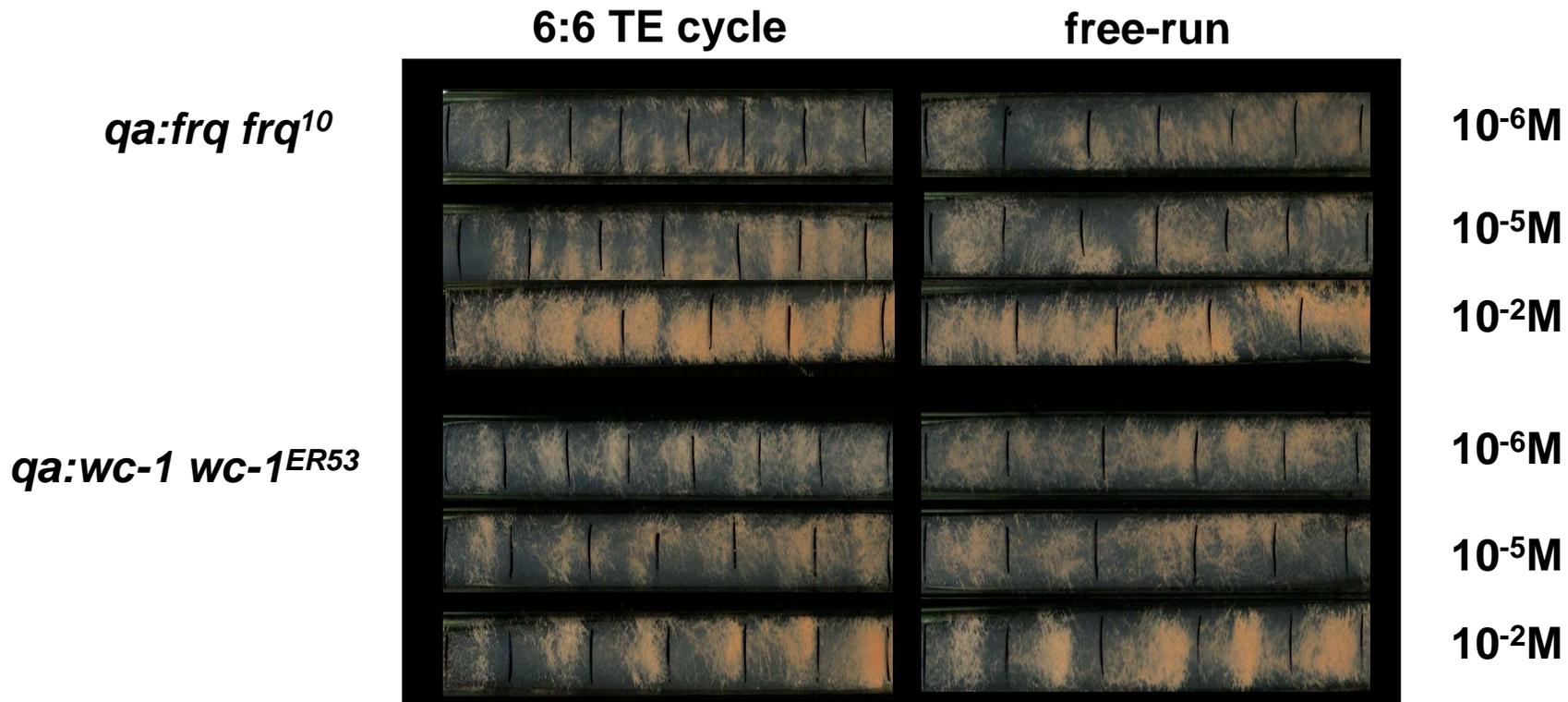
24

TE 6:6 LL

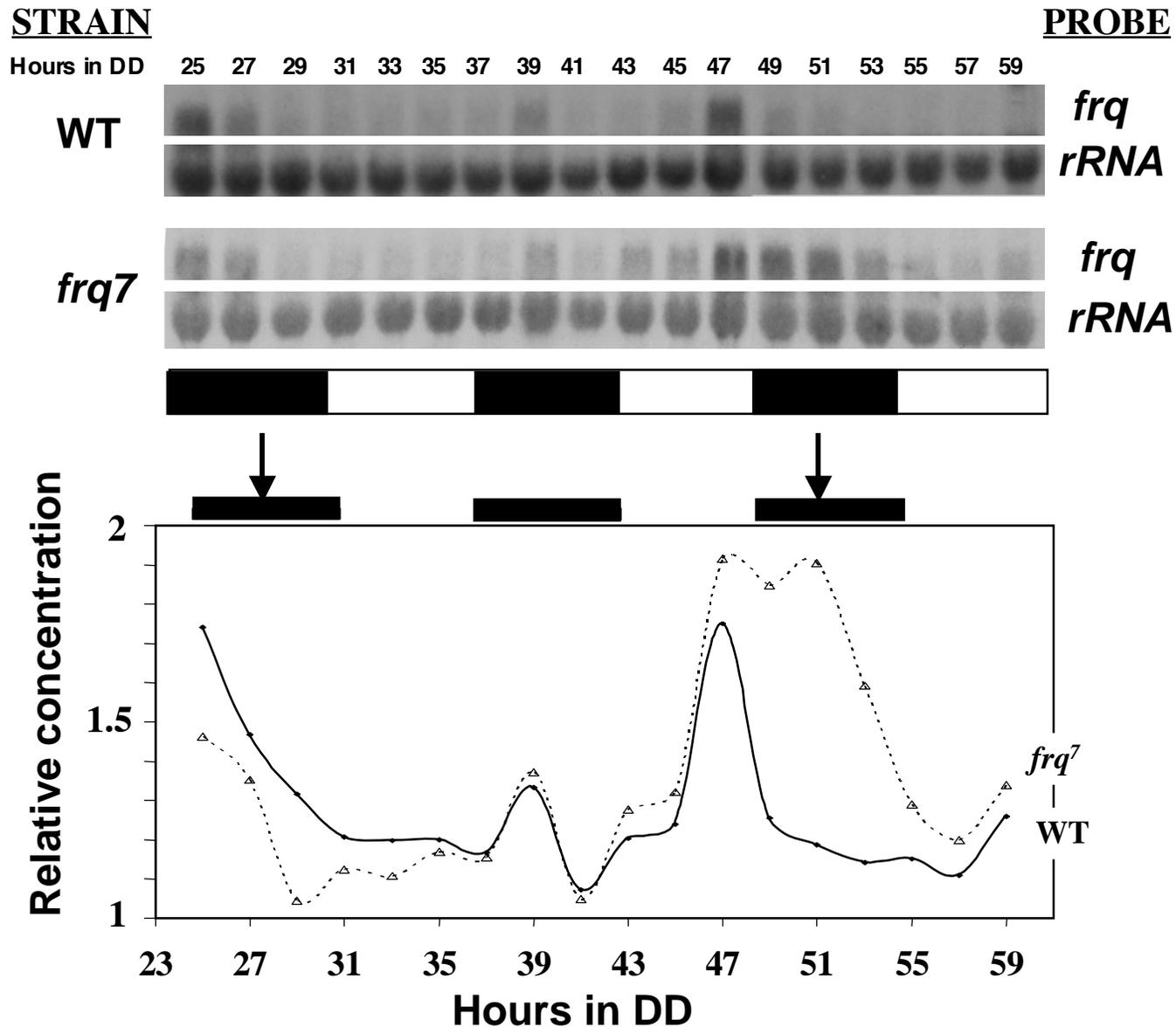
12

Suggests that saturation of the FRQ/WCC oscillator by LL blocks frequency demultiplication, but does not block the effects of temperature cycles on the conidiation rhythm generated by the FLO.

frq mRNA rhythms are required for overt rhythmicity
Are *frq* mRNA rhythms required for FD?



Lower threshold of inducer needed to cause FD vs. generate a free running rhythm in *qa:wc-1* suggests that the FLO can amplify the signal coming from the FRQ/WCC oscillator



Induction of *frq* by temperature is phase-dependent

The period of the FRQ/WCC oscillator determines the period in temperature entrainment cycles

No defects in 12:12 LD or 12:12 TE cycles

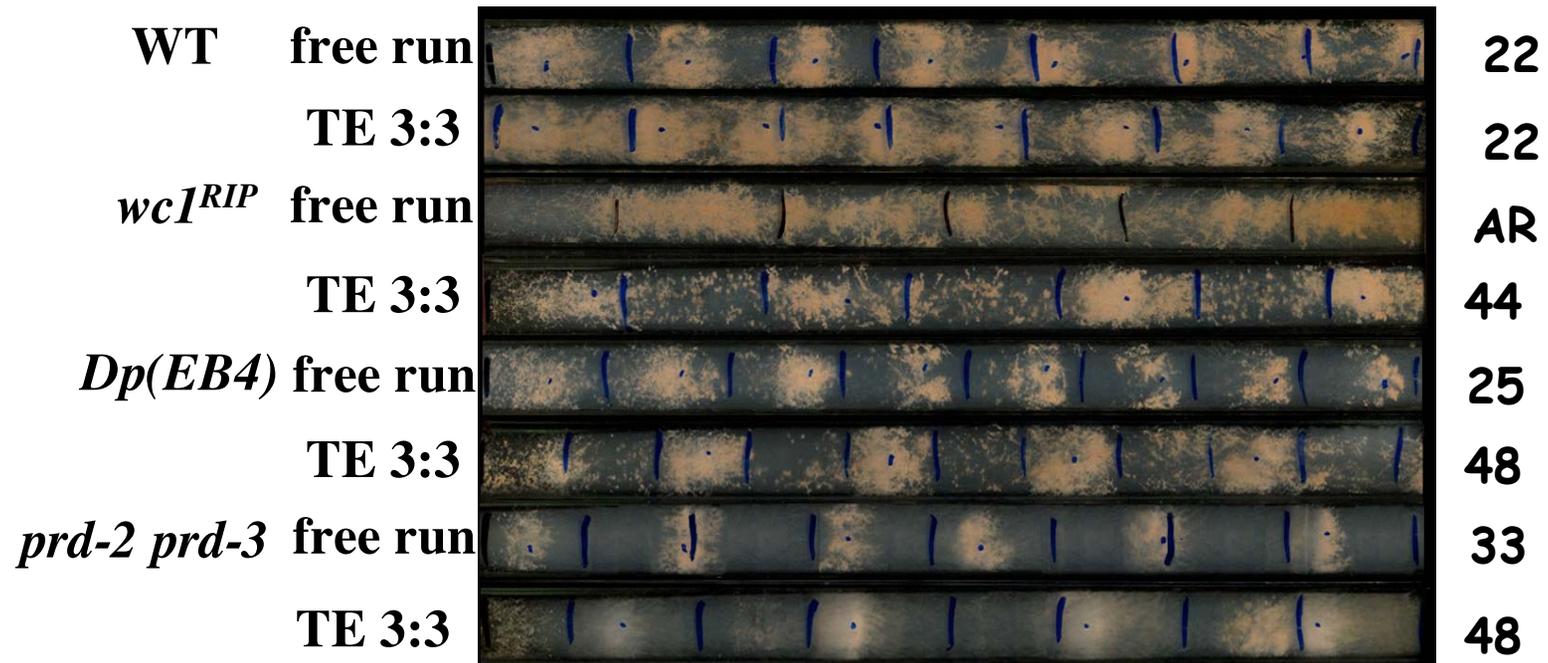
Free running period

TE 4:4



If the strains free-running period is close to 24 h (20-28) an ~ 24h rhythm was observed in TE 6:6 cycles. A 12 h rhythm if outside of this range. A rhythm close to the FRP was observed in shorter cycles, such as 4:4. But, some TE cycles caused arrhythmic behavior (*frq²*) or over de-multiplication!

Some strains "over-demultiply" in short TE cycles and adopt a rhythm longer than their free-running rhythm.



Light/dark cycles set the period of the FRQ/WCC Oscillator



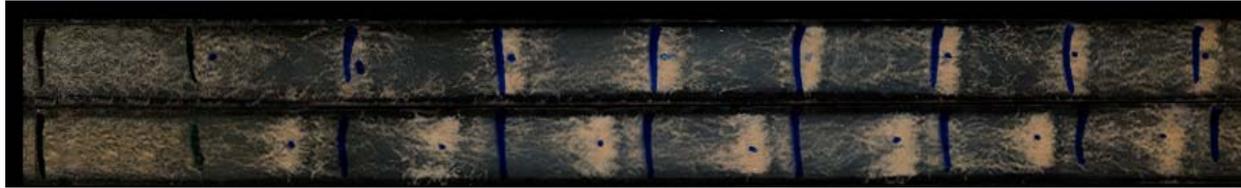
In 4:4 TE cycles alone, WT is 22 h. Thus, the period of the demultiplied rhythm is controlled by the LD cycle period.

Time delay - synchronization of the oscillators is needed for FD?

Phase-locking dynamics

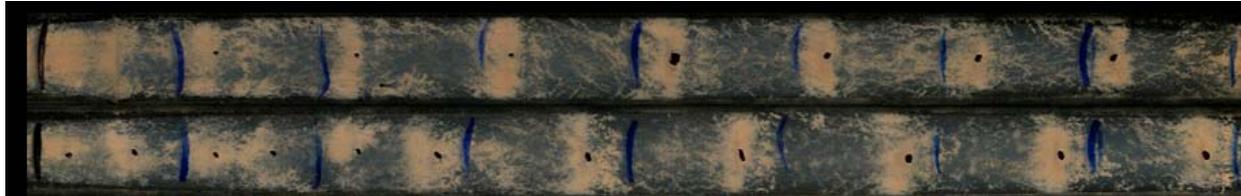
The adopted phase is dependent on initial conditions. WT strain grown in 6:6 TE cycles was transferred from LL 22°C to either the cold phase of the cycle (top) or warm phase of the cycle (bottom).

WT cold
WT warm

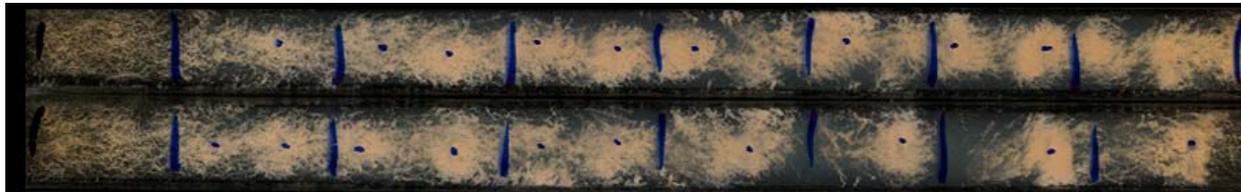


Delayed period doubling is shown between replicate cultures of the WT and *frq²* strains grown in 6:6 TE cycles.)

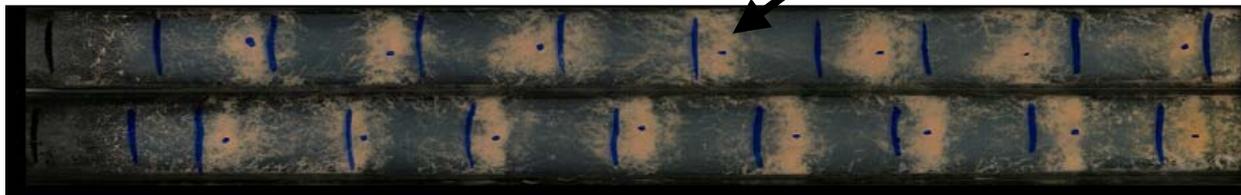
WT



frq²



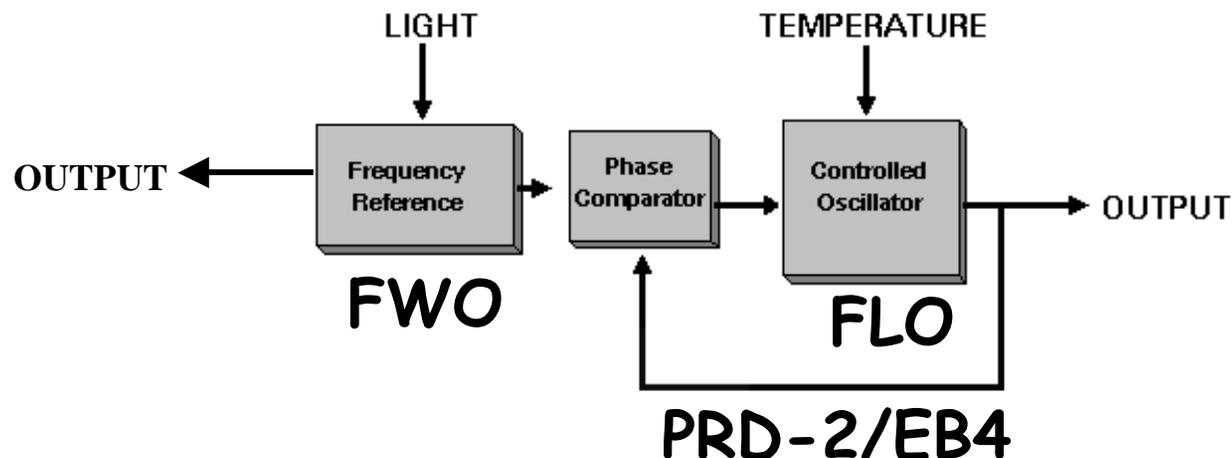
frq⁷



The *frq⁷* strain appears to temporarily slip out of a phase-locked condition before returning to the 24-hr entrained rhythm (black arrow); a WT strain in the same assay is shown for comparison.

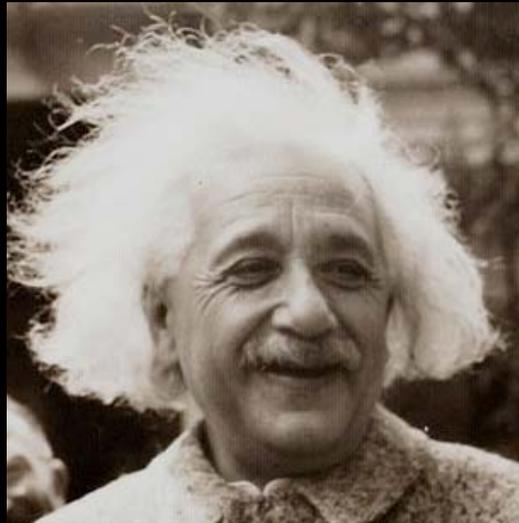
Summary:

1. Loss of oscillation of *frq* mRNA results in loss of FD, but not 1:1 entrainment in temperature cycles - consistent with presence of a temperature responsive FLO. Implies that the FRQ/WCC oscillator controls the FLO when it is intact.
2. The time delay before the onset of FD suggests that the phase-locking of the FRQ/WCC oscillator and the FLO occurs after a temperature induced oscillation is manifest in the FLO - suggests a mechanism is present which controls the coupling between the FRQ/WCC oscillator and the FLO.
3. Over-demultiplication seen in *prd-2* and EB4 mutations suggest that these have defects in the oscillator coupling mechanism.
4. Demultiplied oscillations tend to be attracted towards the natural frequency of the system (their FRP).



“The only reason for time is
so that everything doesn't
happen at once.”

Albert Einstein



Acknowledgements

Output

Genetic Selection

Michael Vitalini

Teresa Lamb

Chuck Goldsmith

Irene March

Louis Morgan

Arrays

Alejandro Correa

Zachary Lewis

Andrew Greene

Johnny Fazzino

FLO

ccg-16

Renato de Paula

Lindsay Bennett

Zachary Lewis

Andrew Greene

LM mutations

Kyung Seo

Howard Huang

Xiaoguang Liu

Kaitlyn Beasley

NIH GM58529

NIH P01 NS39546

Collaborators

A&M

Dan Ebbole

Terry Thomas

Wayne Versaw

Bill Park



Center for Research
on Biological Clocks
Texas A&M
University System

Kathy Borkovich

Carol Jones

UC Riverside

FRQ is a central component of the core Neurospora feedback loop

(Aronson et al, 1994)

- Loss of function mutations of *frq* result in loss of normal free running rhythms in conidiation.
- Partial function mutants alter period (16-29 h) and temperature compensation.
- FRQ protein and *frq* mRNA accumulation are rhythmic in DD and rhythmic expression is essential for conidiation rhythms.
- Overexpression of *frq* at an ectopic locus represses transcription from the endogenous locus - feedback loop



WC-1 and WC-2; regulate all known blue light responses in *Neurospora* (G. Macino) and are central oscillator components (Crosthwaite et al, 1997).



Role in DD - null mutants are arrhythmic in DD

WC-1: A blue-light photoreceptor, binds the flavin chromophore FAD, protein accumulation is rhythmic.

WC-2: mRNA and protein are abundant and not rhythmic.

WC-1/WC-2 heterodimerize through their PAS domains to form a complex (WCC) which can activate *frq* transcription - WCC binds to the *frq* promoter at 2 sites.

How does the negative feedback loop keep from winding down?

Interlocked loops - observed in all eukaryotic circadian oscillators

Lee K, Loros JJ, Dunlap JC.

Interconnected feedback loops in the Neurospora circadian system. Science. 2000 Jul 7;289(5476):107-10.



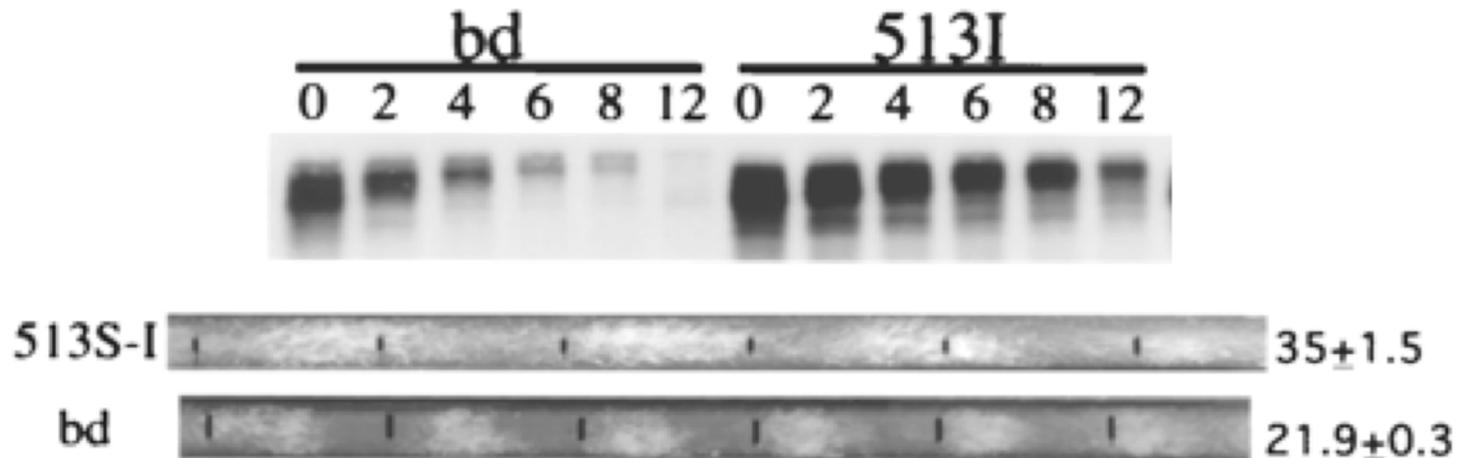
FRQ turnover



Yi Liu

Several kinases can phosphorylate FRQ and mutation of the kinases affects the period and stability of FRQ.

Ser 513 critical for FRQ function - a single substitution of isoleucine stabilizes FRQ and lengthens the period from 22 to 35 h.

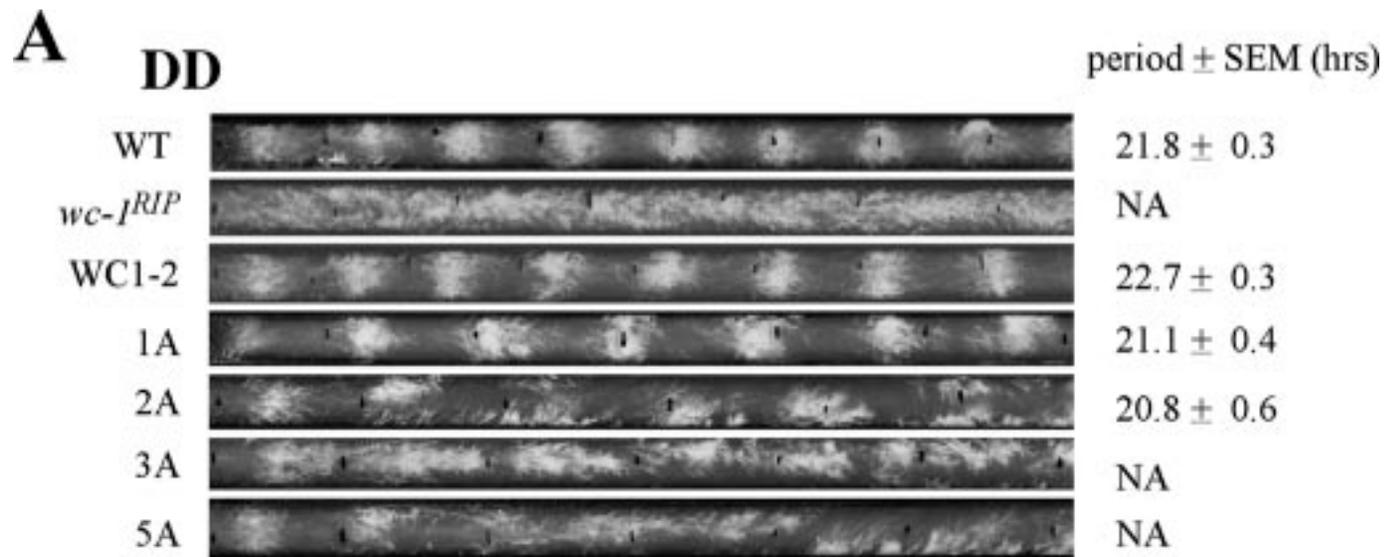


He Q, Shu H, Cheng P, Chen S, Wang L, Liu Y.

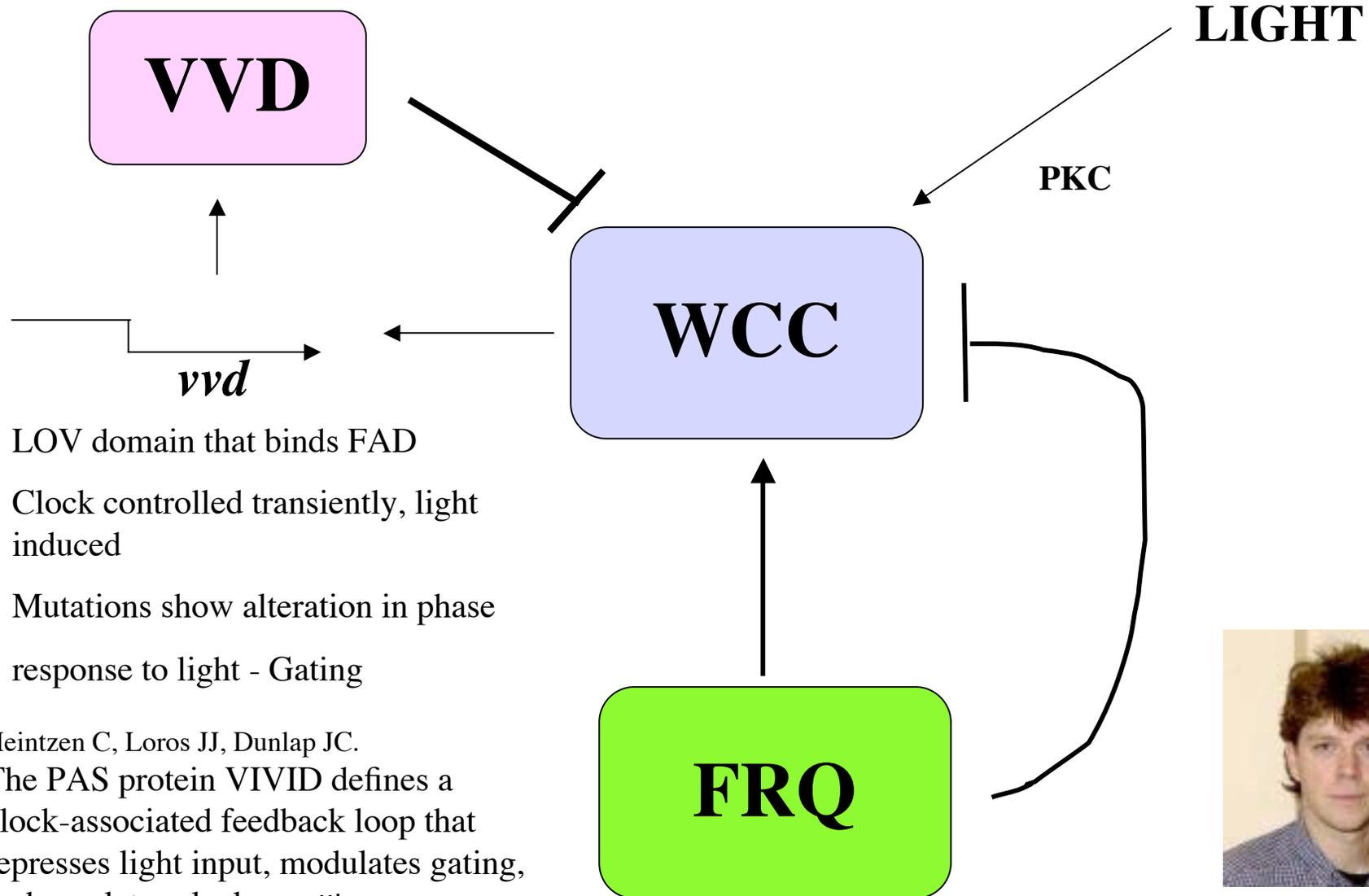
Light-independent phosphorylation of WHITE COLLAR-1 regulates its function in the Neurospora circadian negative feedback loop.

J Biol Chem. 2005 Apr 29;280(17):17526-32.

Identified 5 P sites on WC-1 and found that mutation show short period, low amplitude, or arrhythmic conidiation rhythms in DD.



VIVID



Heintzen C, Loros JJ, Dunlap JC.
The PAS protein VIVID defines a
clock-associated feedback loop that
represses light input, modulates gating,
and regulates clock resetting
Cell. 2001 Feb 9;104(3):453-64.