

Clocks and Dominoes: Studies into Positive Feedback and the Continuum from Early Embryo to Somatic Cell

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Kavli Institute for Theoretical Physics - Biological Clocks

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INDIANA UNIVERSITY
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the biocomplexity institute

1. Digital Responses In Vivo: Mitotic Toggle Switches?

2. The Embryonic Mitotic Oscillator: Positive Feedback and the Emergent Properties of the CDK1/Cyclin B Module

3. Positive Feedback and Somatic Mitosis: To Enter, To Exit, To Not Oscillate?

Digital Biology Converted from Graded Stimuli



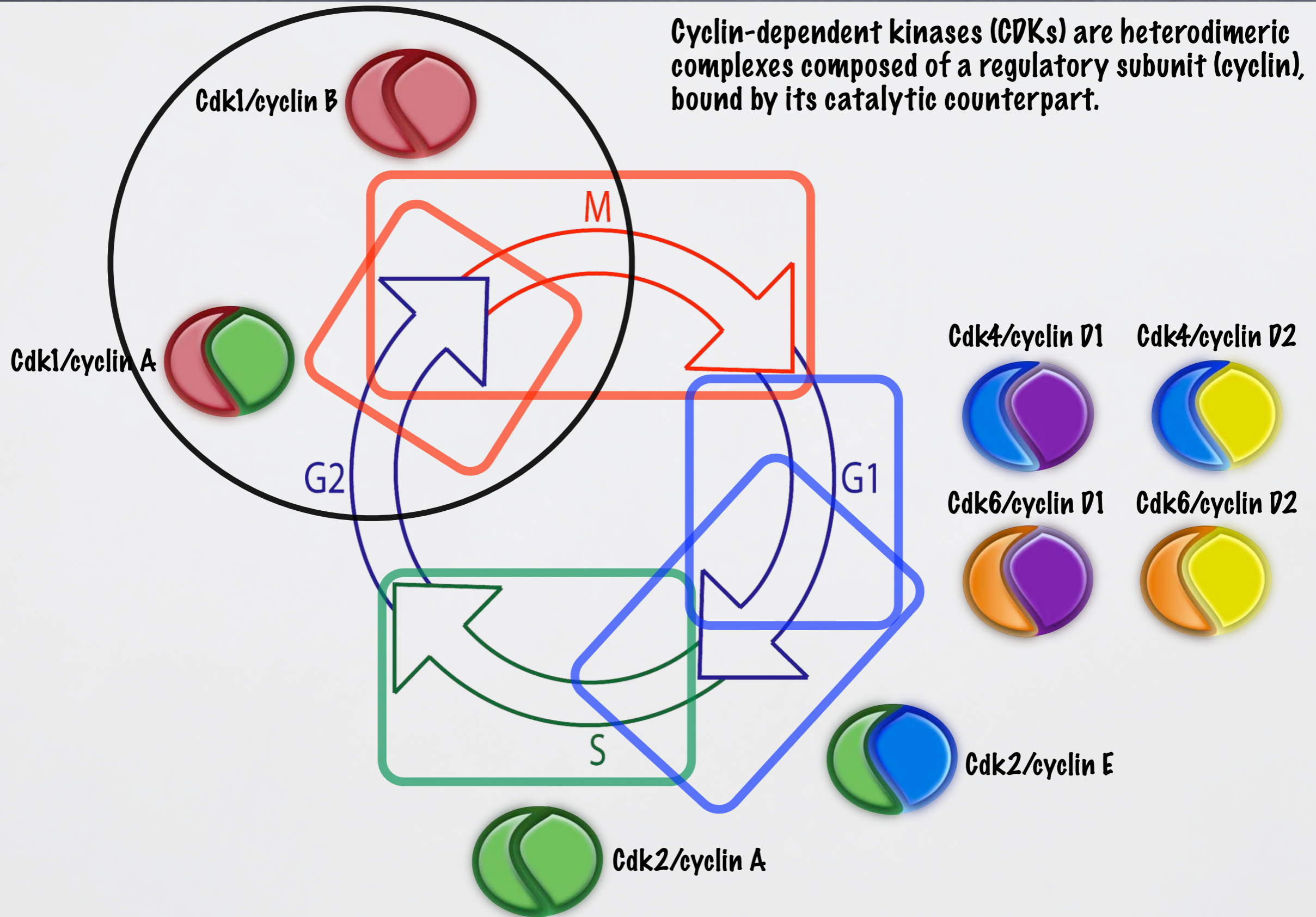
Cyclin



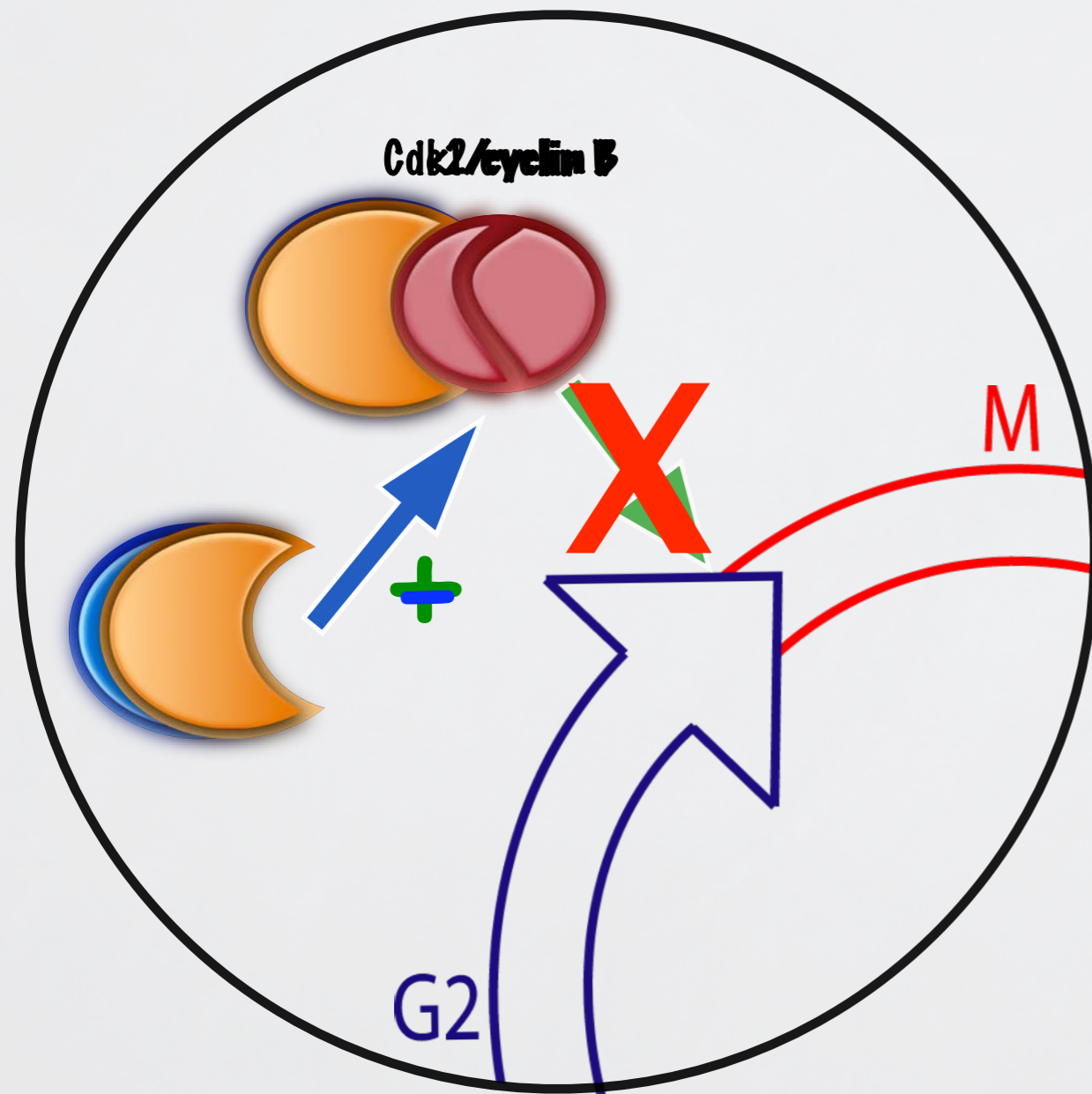
All-or-none character.

Cells must avoid "half-way" completion of mitosis; returning to a "G2 state".

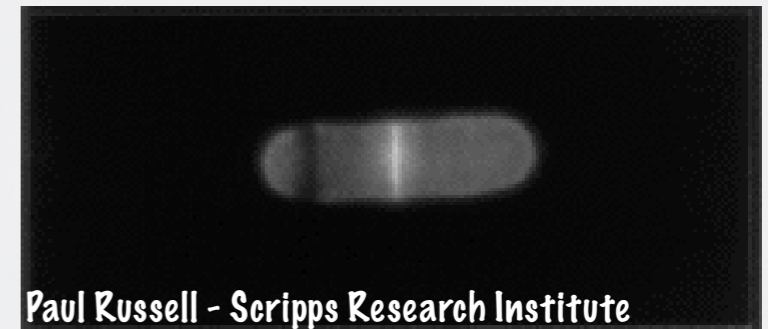
What About Mitosis, its Inputs and Outputs?



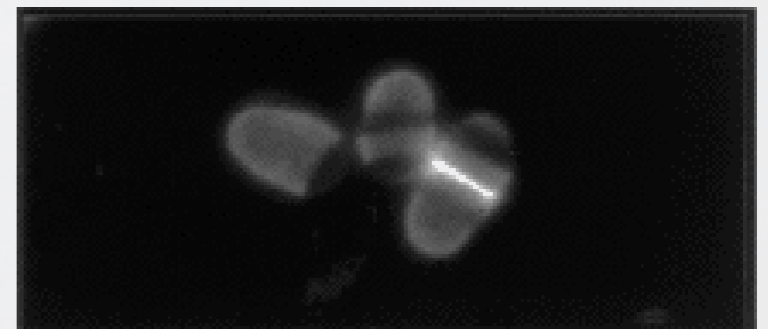
What About Mitosis, its Inputs and Outputs?



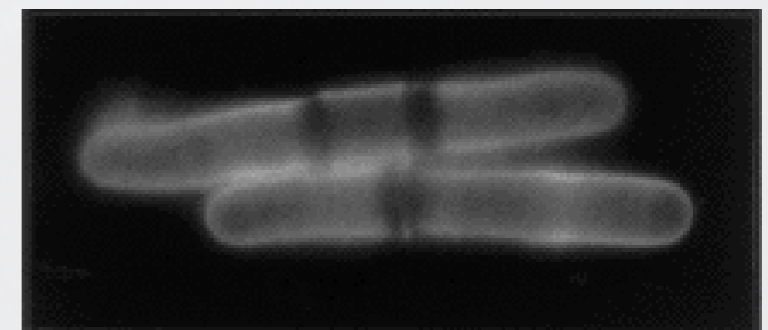
WT *S. pombe*



wee1-50



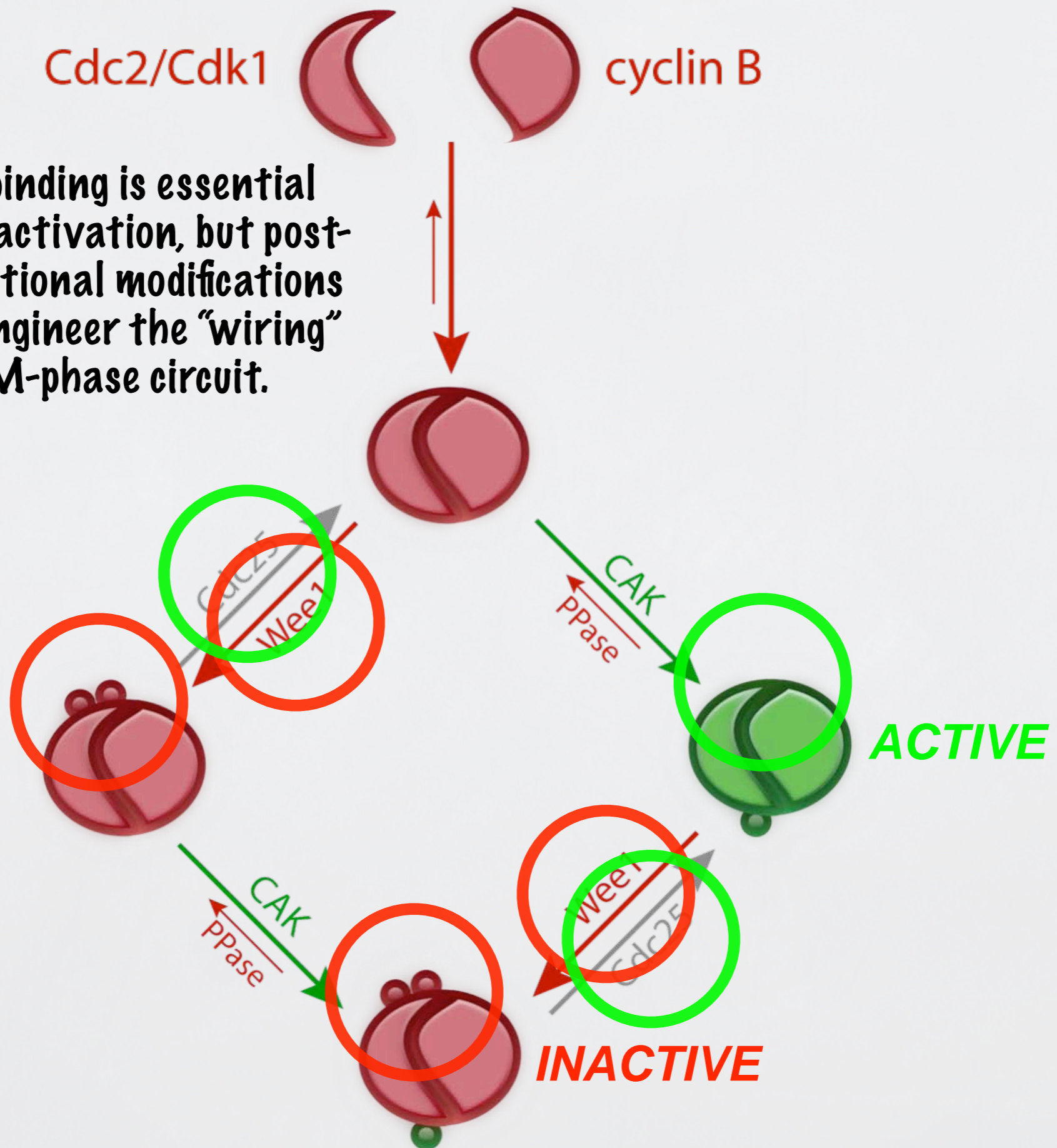
cdc25-22



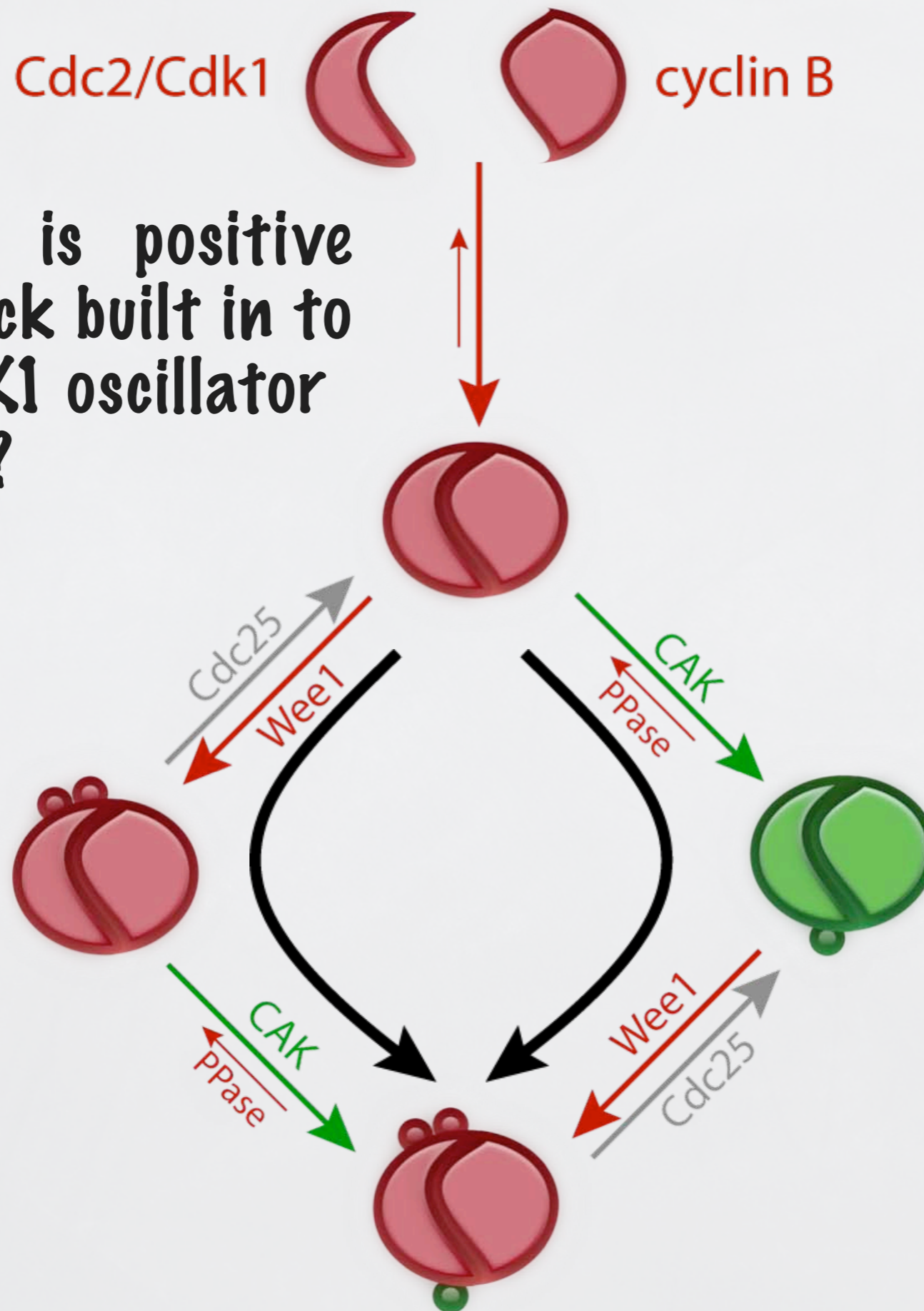
Cdc2/Cdk1

cyclin B

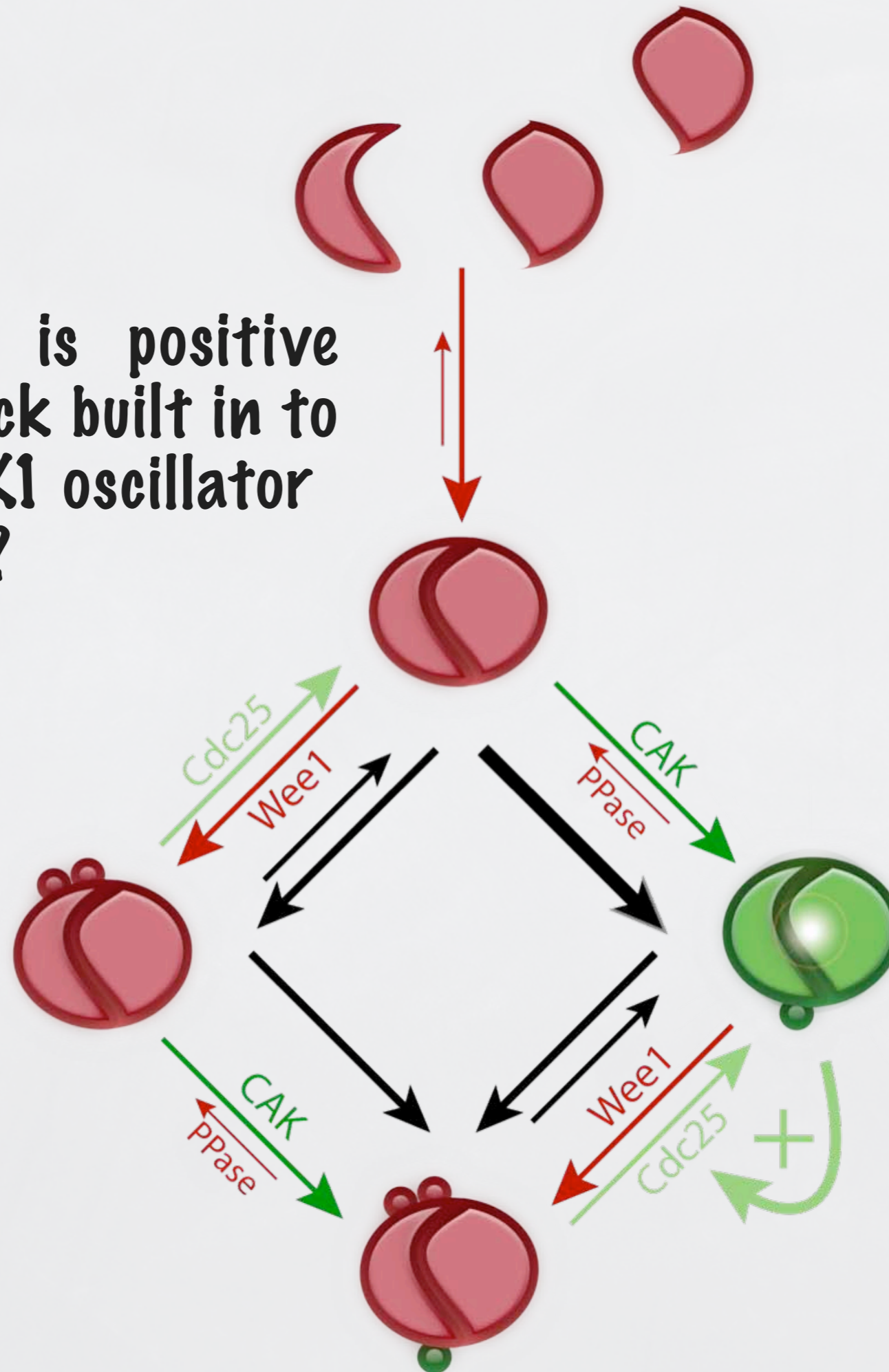
Cyclin binding is essential to CDK activation, but post-translational modifications truly engineer the "wiring" of the M-phase circuit.



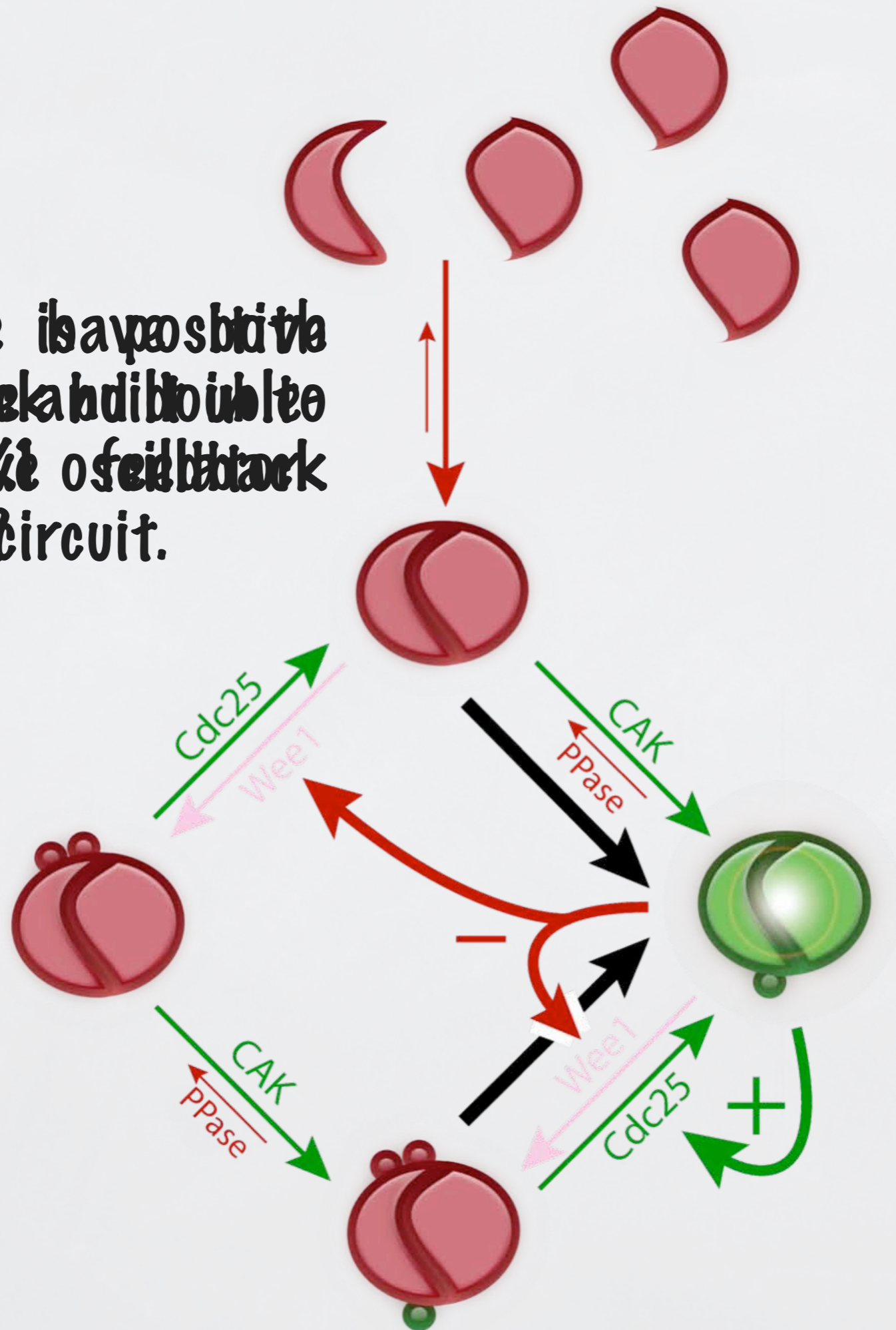
Where is positive feedback built in to the CDK1 oscillator circuit?



Where is positive feedback built in to the CDK1 oscillator circuit?

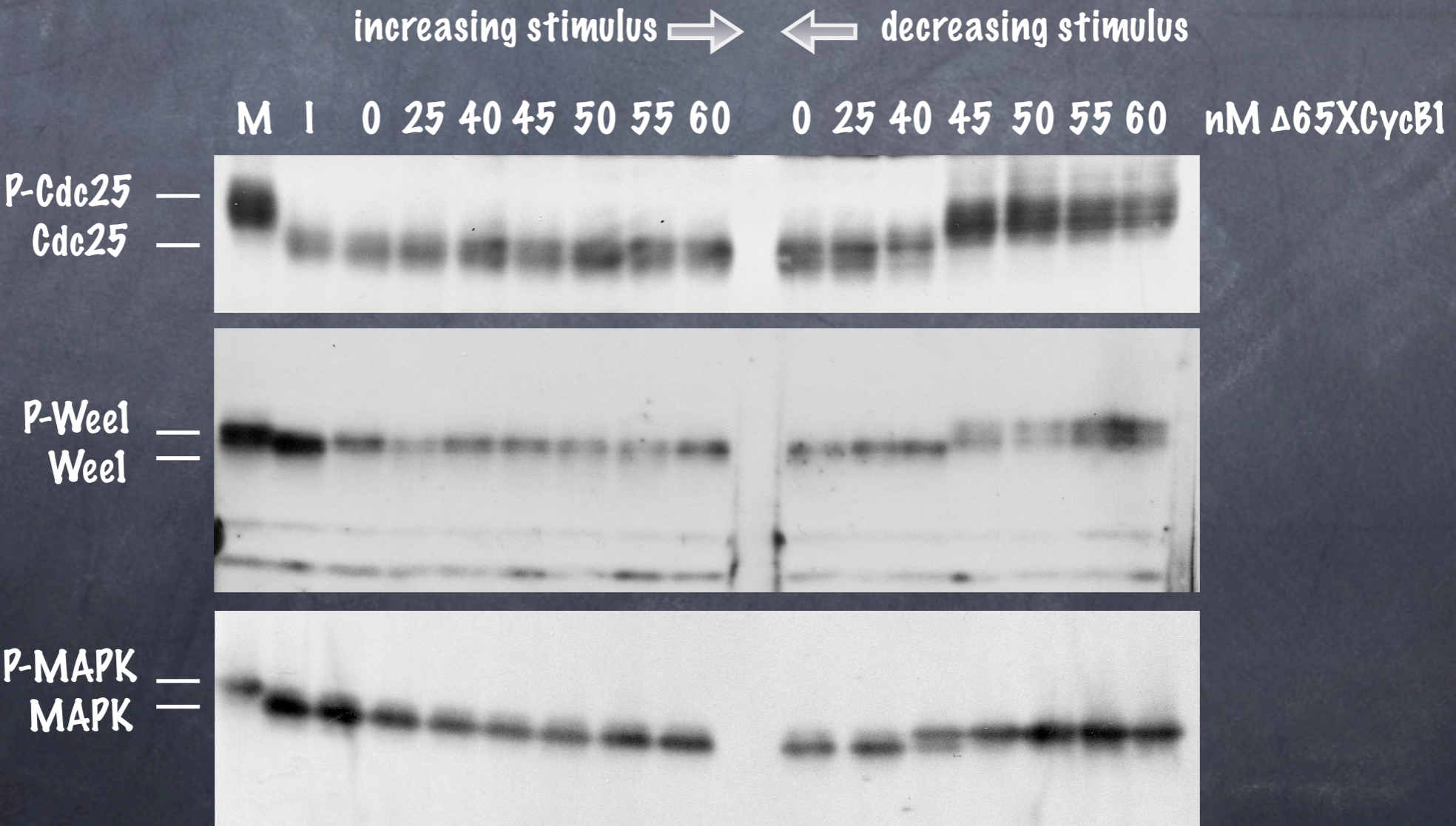
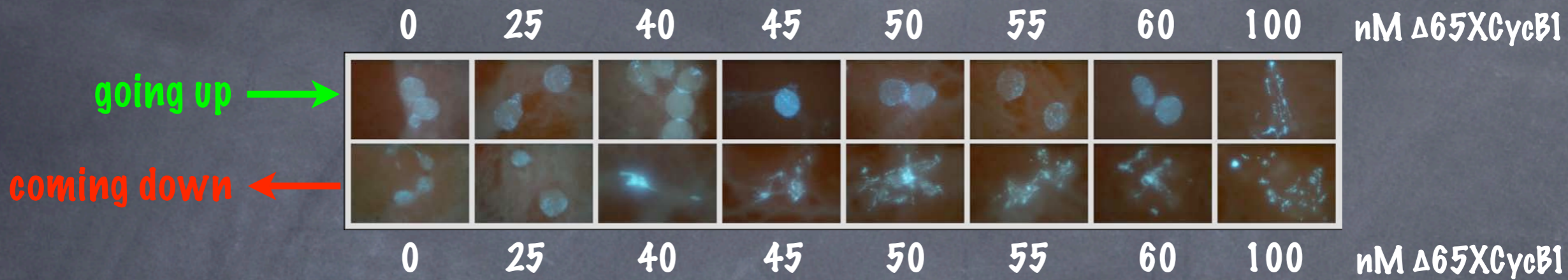


Where is a positive feedback and a negative feedback in this circuit.

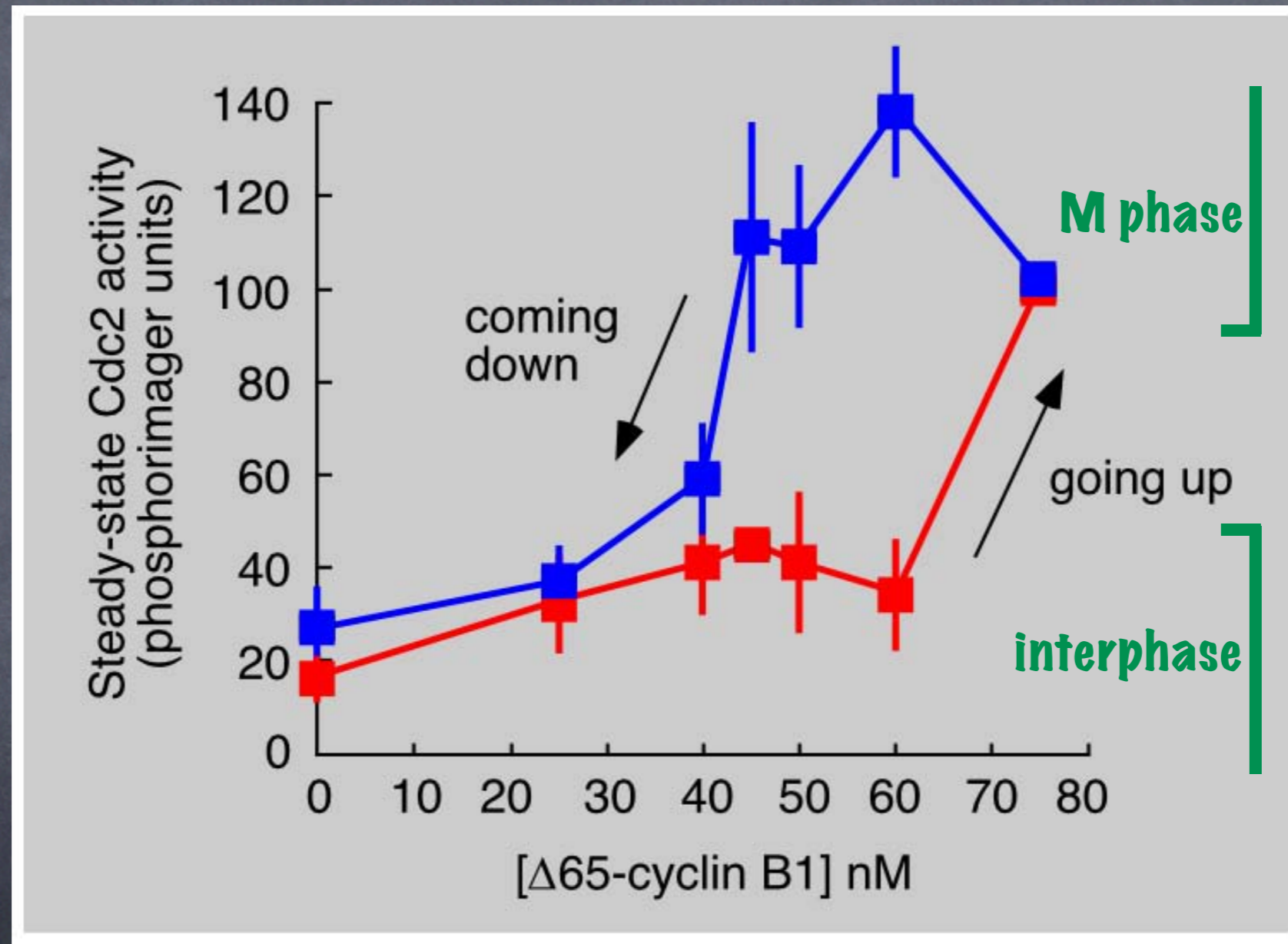


What is the steady-state response of CDK1 to cyclin B?

Cell Biological and Molecular Markers Exhibit Hysteresis



Hysteresis in CDK1 Activation



positive feedback



hysteresis



bistability

Testing a Mathematical Model of the Yeast Cell Cycle

Frederick R. Cross,* Vincent Archambault, Mary Miller, and Martha Klovstad

The Rockefeller University, New York, New York 10021

Submitted May 25, 2001; Revised September 25, 2001; Accepted October 10, 2001
Monitoring Editor: Mark J. Solomon

We derived novel, testable predictions from a mathematical model of the budding yeast cell cycle. A key qualitative prediction of bistability was confirmed in a strain simultaneously lacking *cdc14* and *C1* cyclin. The model predicts that the transition from interphase to mitosis is irreversible.

Hysteresis drives cell-cycle transitions in *Xenopus laevis* egg extracts

Wei Sha*, Jonathan Moore†, Katherine Chen*, Antonio D. Lassaletta*, Chung-Seon Yi*, John J. Tyson*, and Jill C. Sible*‡

*Department of Biology, Virginia Polytechnic Institute and State University, Blacksburg, VA 24061-0406; and †Cancer Research UK London Research Institute, Clare Hall Labs, South Mimms, Herts, EN6 3LD, United Kingdom

Edited by Thomas D. Pollard, Yale University, New Haven, CT, and approved November 21, 2002 (received for review September 3, 2002)

Cells progressing through the cell cycle must commit irreversibly to mitosis without slipping back to interphase before properly segregating their chromosomes. A mathematical model of cell-cycle progression in cell-free egg extracts from frog predicts that irre-

activated by removal of the inhibitory phosphate groups. Because Cdc2 activates cyclin proteolysis, the rate of cyclin degradation in M phase exceeds its rate of synthesis, and cyclin concentration falls. However, according to the model, the extract

Building a cell cycle oscillator: hysteresis and bistability in the activation of Cdc2

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*Department of Molecular Pharmacology, Stanford University School of Medicine, Stanford, CA 94305-5174, USA

†Department of Mathematics, Rutgers University, New Brunswick, NJ 08903, USA

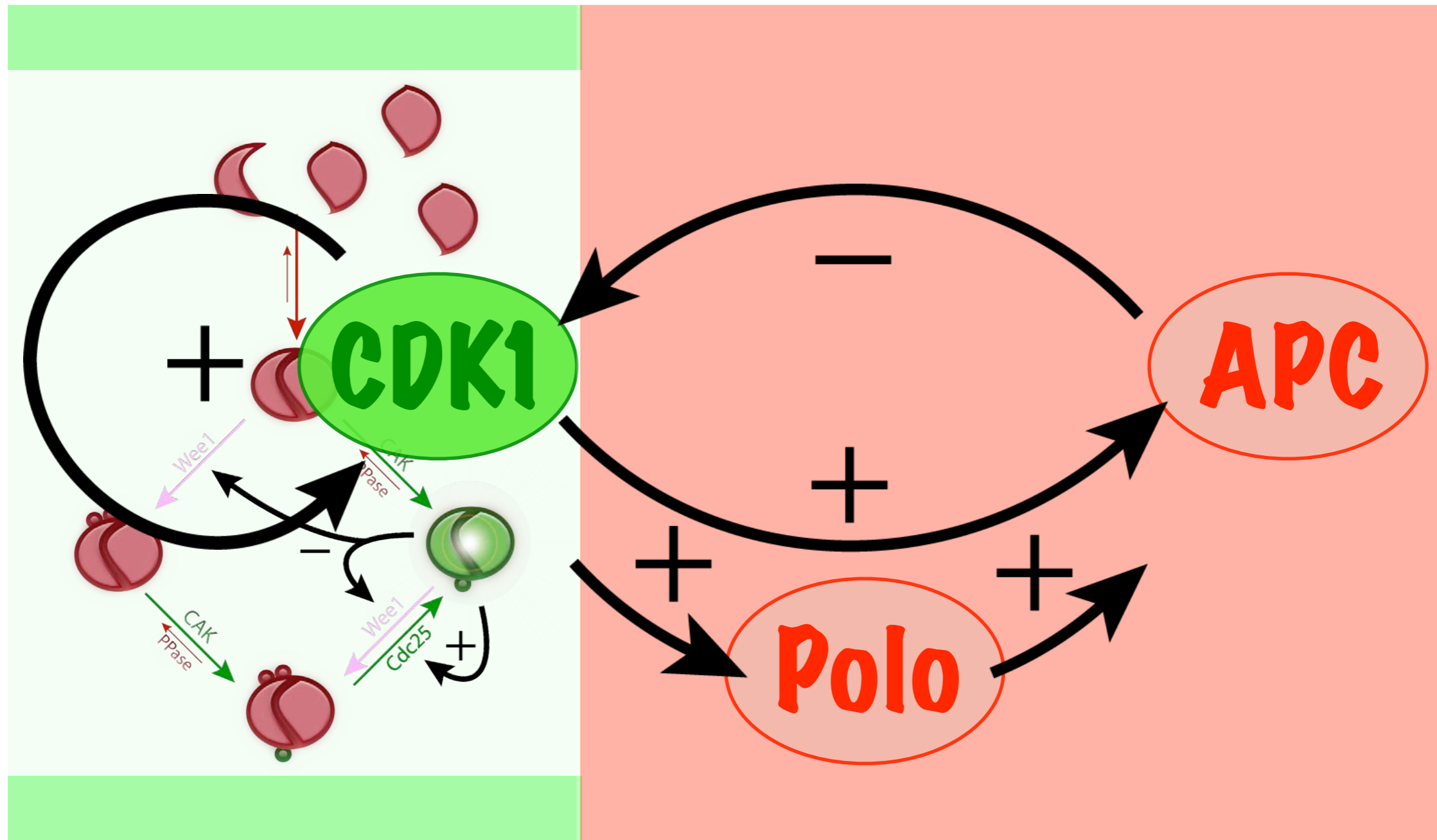
‡e-mail: james.ferrell@stanford.edu

Published online: 10 March 2003; DOI: 10.1038/ncb954

In the early embryonic cell cycle, Cdc2–cyclin B functions like an autonomous oscillator, whose robust biochemical

feedback²³, nor is it the only useful systems-level property that can arise from positive feedback loops (for example, sensitivity amplification²⁴ is a more robust property of positive feedback systems

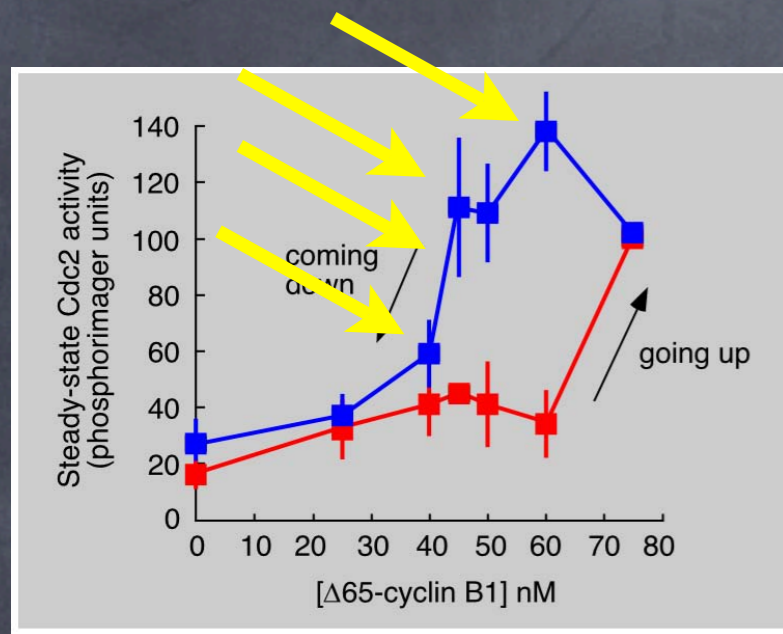
To oscillate, CDK1 must be inactivated



positive feedback

negative feedback

A Cell Biologist's Motivation Through Modeling

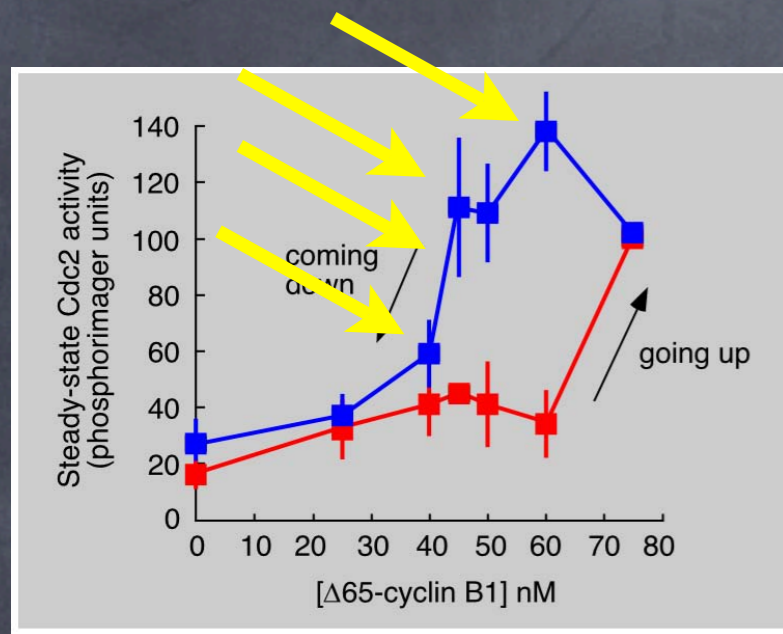


CDK1 activity is sustained and cell extracts remain in M phase at steady state, even as cyclin B levels are reduced.

What about the anaphase-promoting complex-mediated negative-feedback loop?

If CDK1 activity is reduced, how is APC activity maintained and remaining cyclin degraded?

A Cell Biologist's Motivation Through Modeling



CDK1 activity is sustained and cell extracts remain in M phase at steady state, even as cyclin B levels are reduced.

Some simplified possibilities:

- 1) The CDK1 \rightarrow APC activation loop is long enough for CDK1 inactivation to not cause immediate APC inactivation. **Does not force the APC system to sustain activity.**
- 2) The degradation of an inhibitor brings about APC activation, so CDK1 activity is not required to maintain APC activity. **What inhibitor, and what is degrading it?**
- 3) There is positive feedback intrinsic to APC activation, separate from the CDK1 module.

Report

Rewiring the Exit from Mitosis

Andrea Ciliberto^{1,3}

Anna Lukács¹

Attila Tóth¹

John J. Tyson²

Béla Novák^{1,*}

¹Molecular Network Dynamics Research Group of Hungarian Academy of Sciences and Budapest University of Technology and Economics, Budapest, Hungary

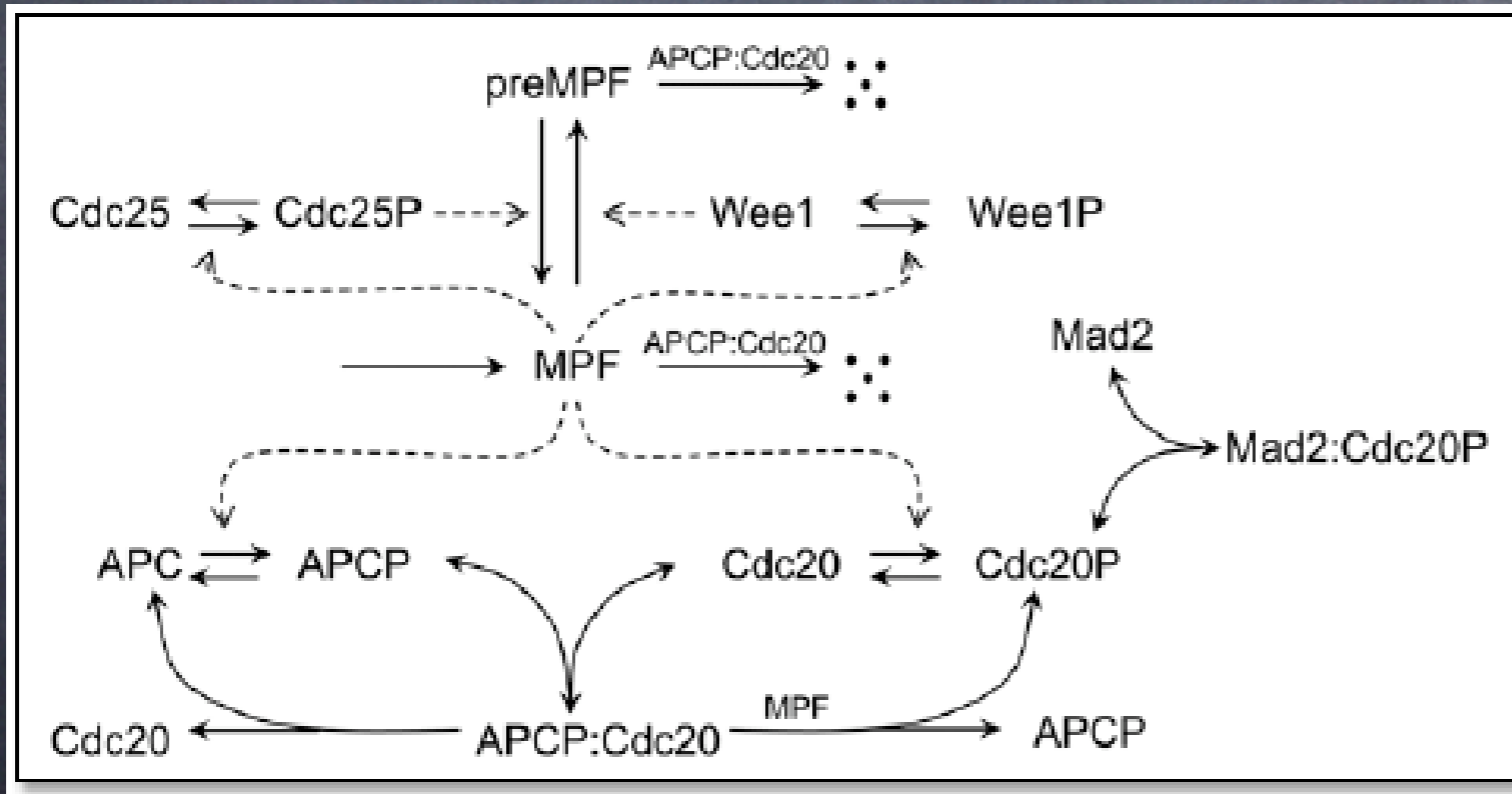
²Department of Biology, Virginia Polytechnic Institute and State University, Blacksburg, Virginia USA

³Current address: FIRC Institute of Molecular Oncology, Milan, Italy

*Correspondence to: Bela Novak, Molecular Network Dynamics Research Group

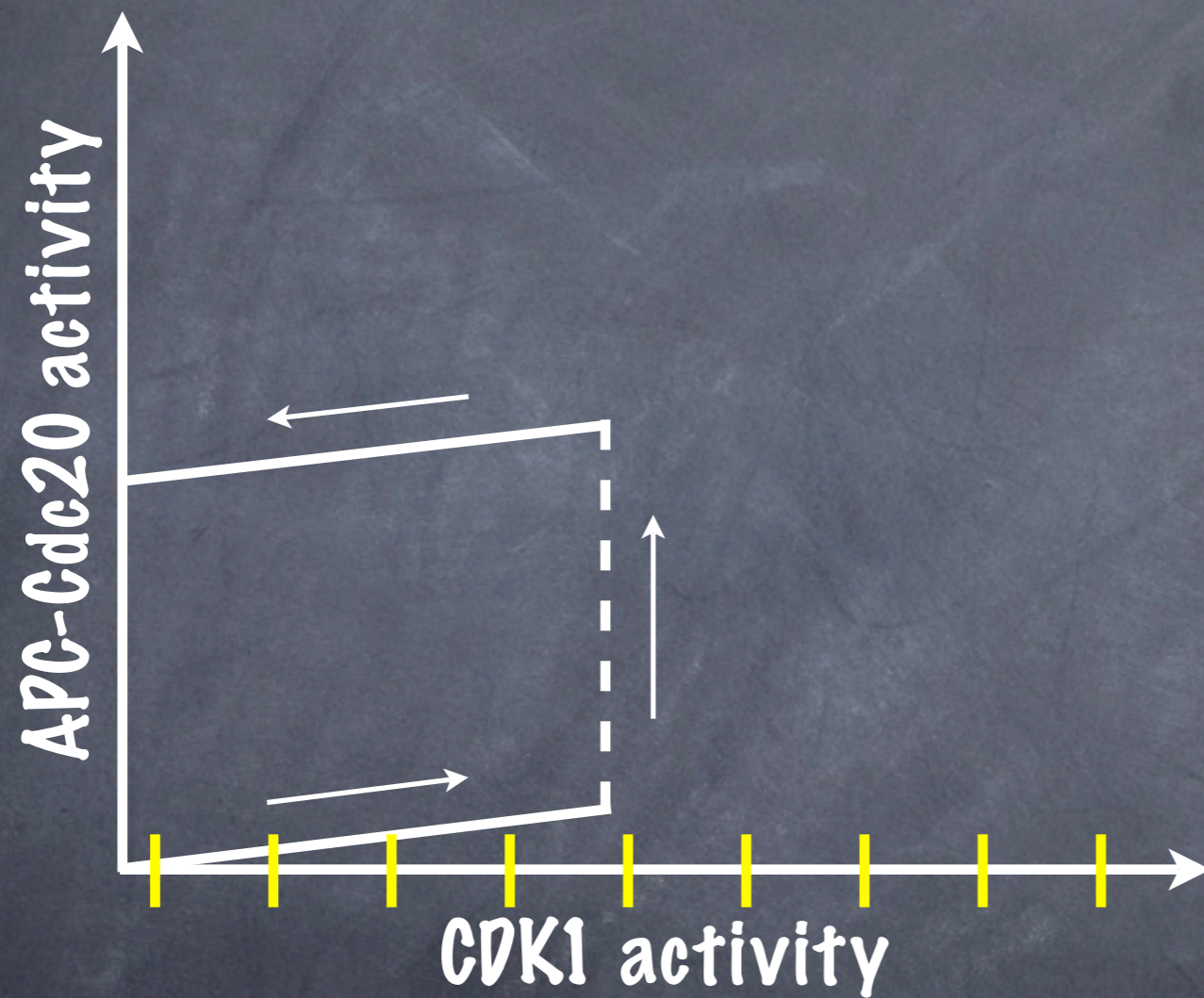
ABSTRACT

The mitotic cell cycle can be described as an alternation between two states. During mitosis, MPF (mitosis promoting factor) is high and keeps inactive its numerous molecular antagonists. In interphase, MPF is inactivated, and the antagonists prevail. The transition between the two states is ensured by 'helper' molecules that favor one state over the other. It has long been assumed that active MPF (a dimer of cyclin B and cyclin-dependent kinase 1) induces exit from mitosis by activating APC:Cdc20, a ubiquitin ligase responsible for cyclin B degradation. The molecular details have not been fully worked out yet, but recent results show that MPF and the ubiquitin ligase are not involved in a simple negative feedback loop. While it is proven that MPF activates APC, new data suggest that MPF inhibits Cdc20, i.e., that MPF and Cdc20 are antagonists. We introduce this new idea into a published model for cell cycle regulation in *Xenopus laevis*, and study its dynamical behavior. We show that the new wiring permits oscillations with a simpler and smaller network than previously envisaged and that the antagonism between MPF and Cdc20 is a keypoint



Could this double-negative (positive) feedback make APC activation hysteretic?

Hysteresis in APC-Cdc20 Activation??

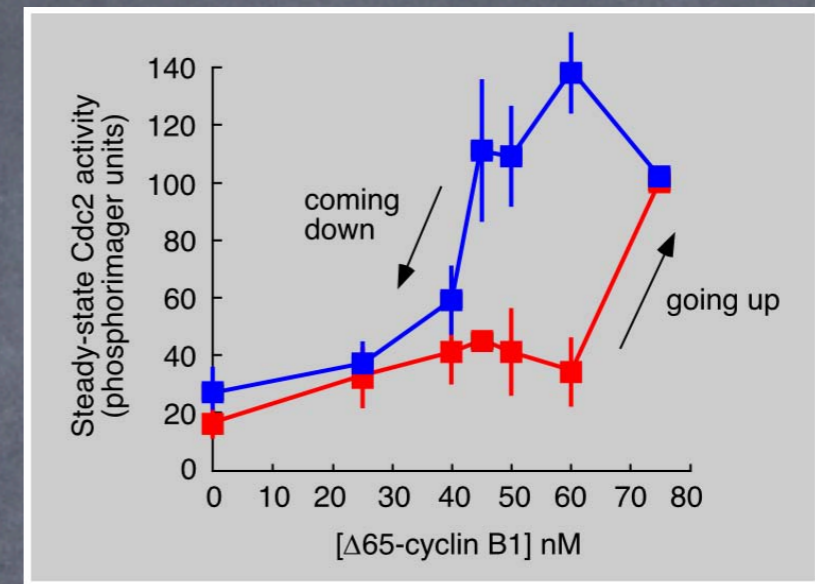


Attaining incremental CDK1 activities will be the experimental challenge here.

Summary-Part I

The CDK1/Wee1/Cdc25 System IS Bistable.

There is hysteresis in CDK1 activation: for some concentrations of cyclin, there are two discrete states; the thresholds for the ON- and OFF-states are different.



Wiring the switch this way ensures either interphase or M phase; there is no "settling" to an intermediate state.

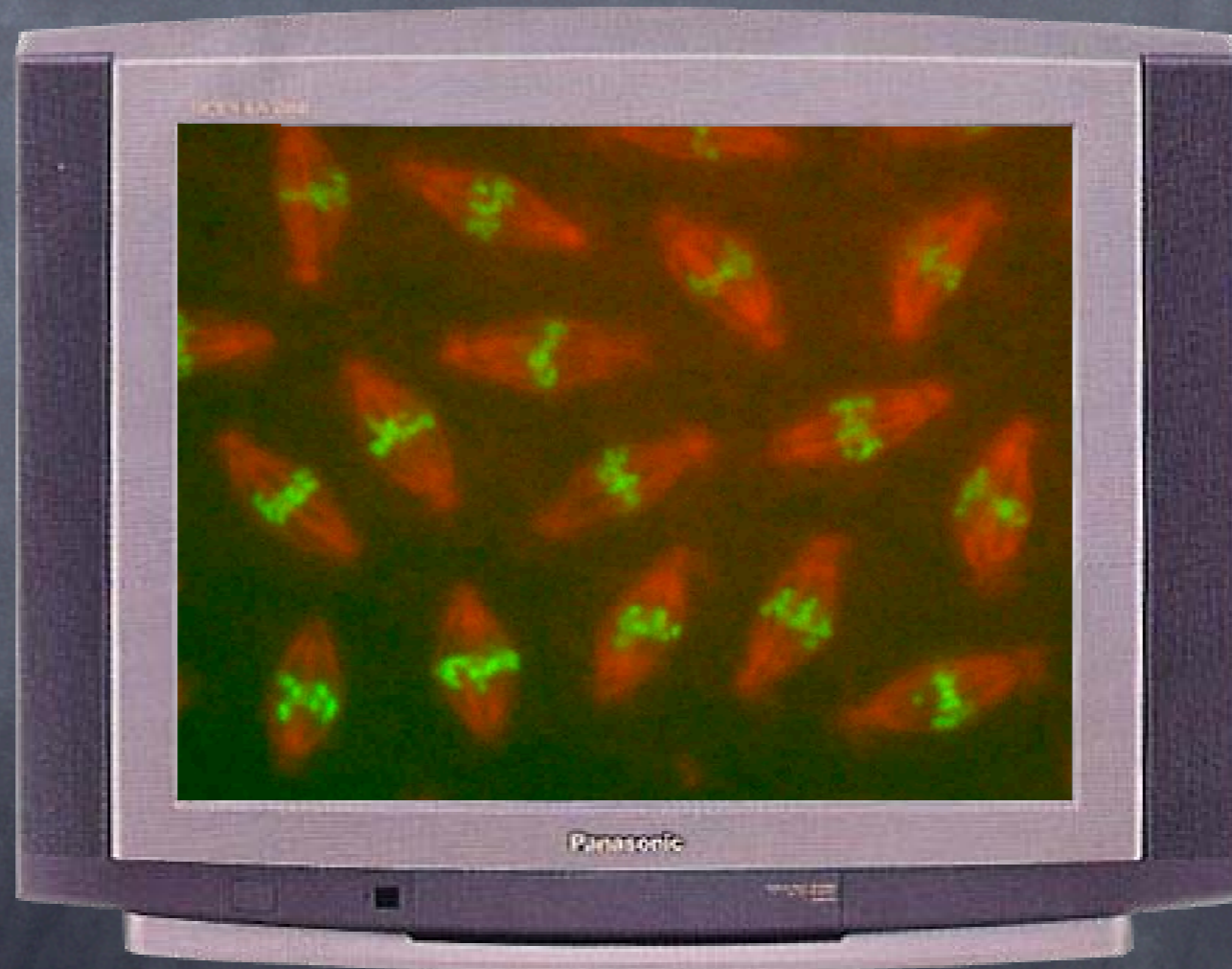
There may be sources of positive feedback in APC activation, and thus, its response may be bistable relative to CDK1 activity.

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Positive Feedback: Simple Relationships Eliciting Complex and Decisive Responses in Many Biological Oscillatory Contexts

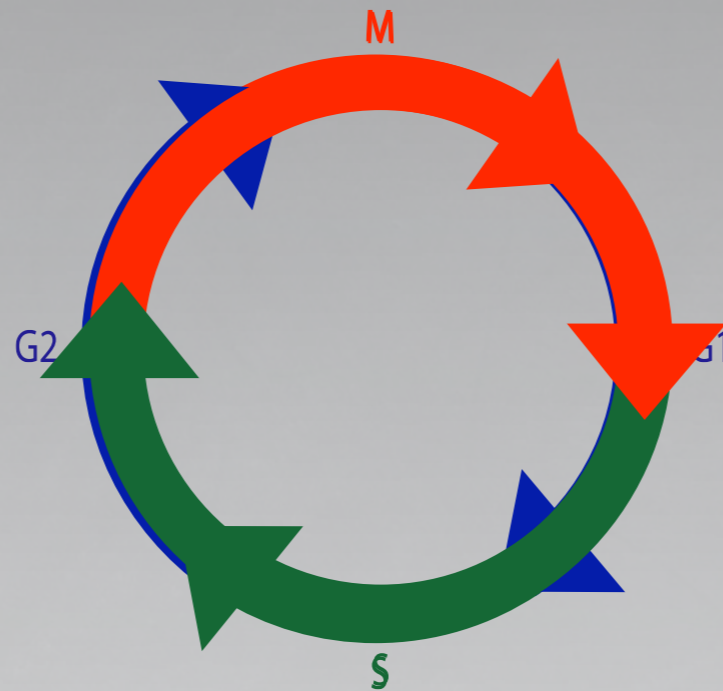


Biological Oscillators Are Diverse in Form and Function

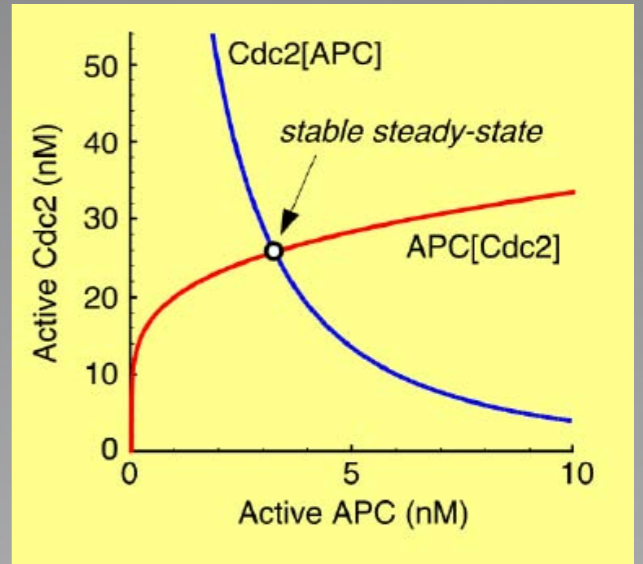
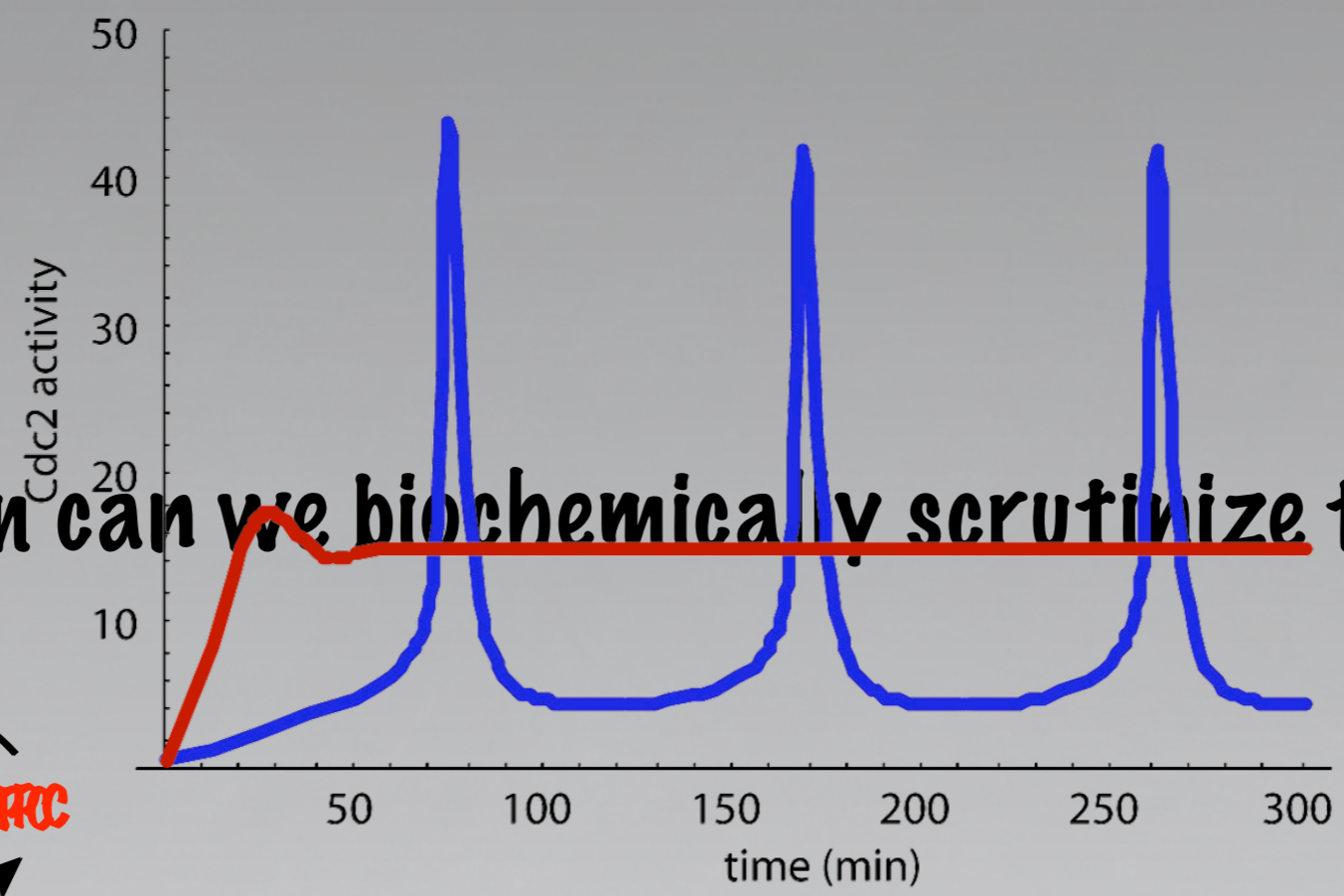
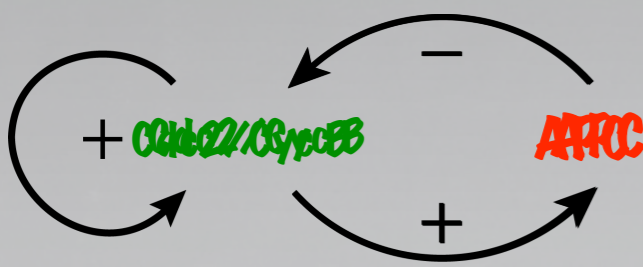
“Pulsatile” - oscillations of CDK1 activity in the embryo;
much like an action potential in a neuron;
no transcription necessary;
driven by rapid modifications of regulatory enzymes.



The somatic cell cycle circuitry has the same core components as an embryo, but the early embryonic cell cycle has no gap/growth phases, so oscillations are very rapid.

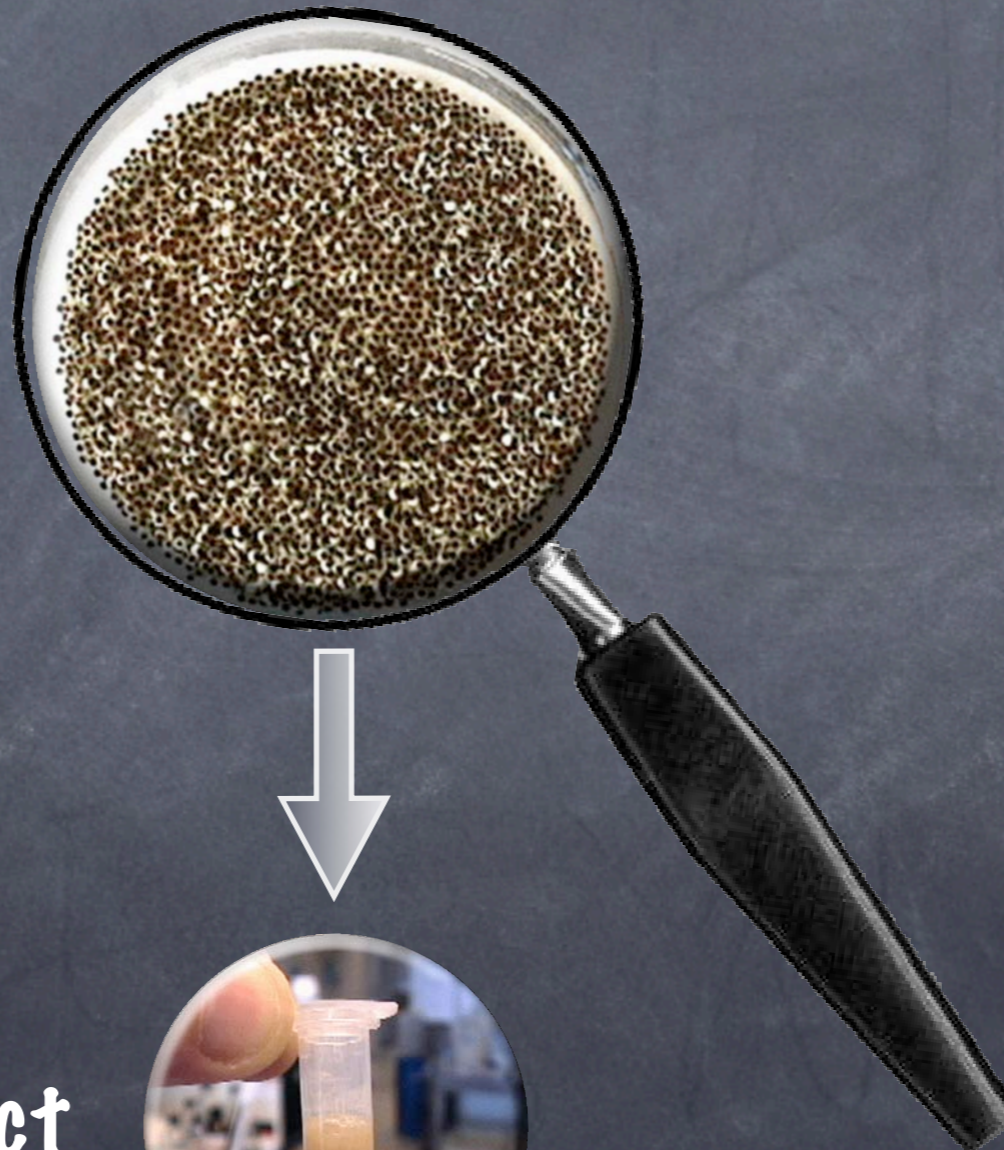
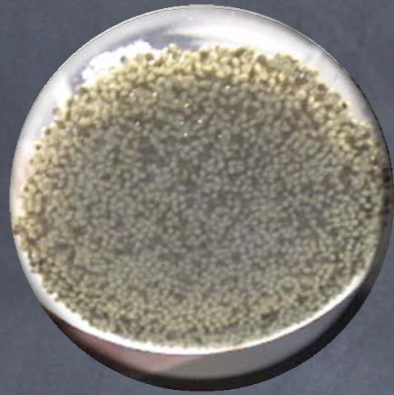


In what system can we biochemically scrutinize these oscillations?



positive feedback
no positive feedback?

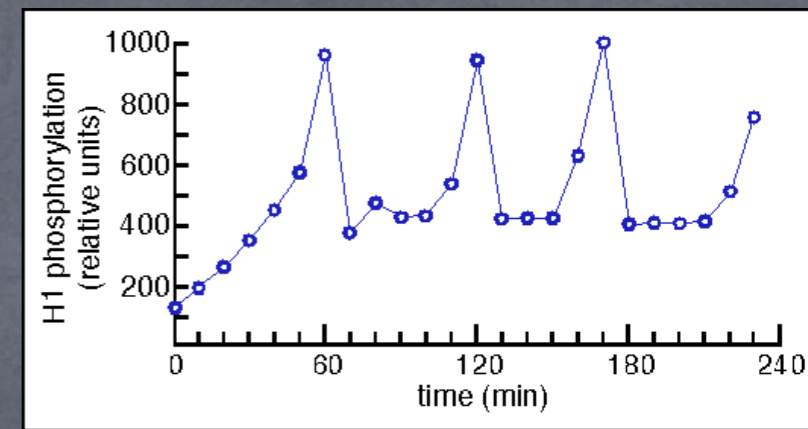
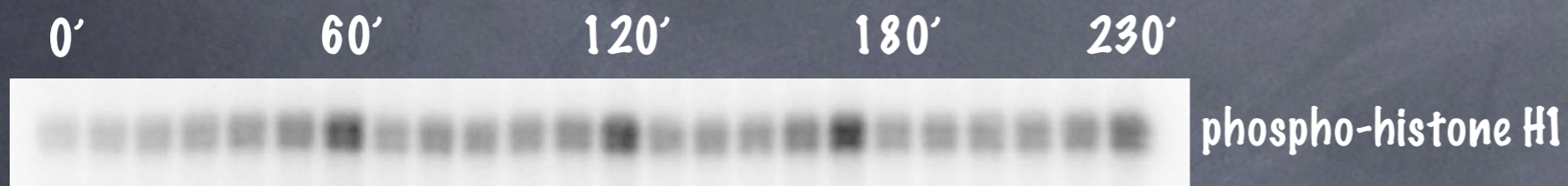
In a model adding positive feedback, however, positive feedback can sustain oscillations.



How to make
a *Xenopus* egg
extract

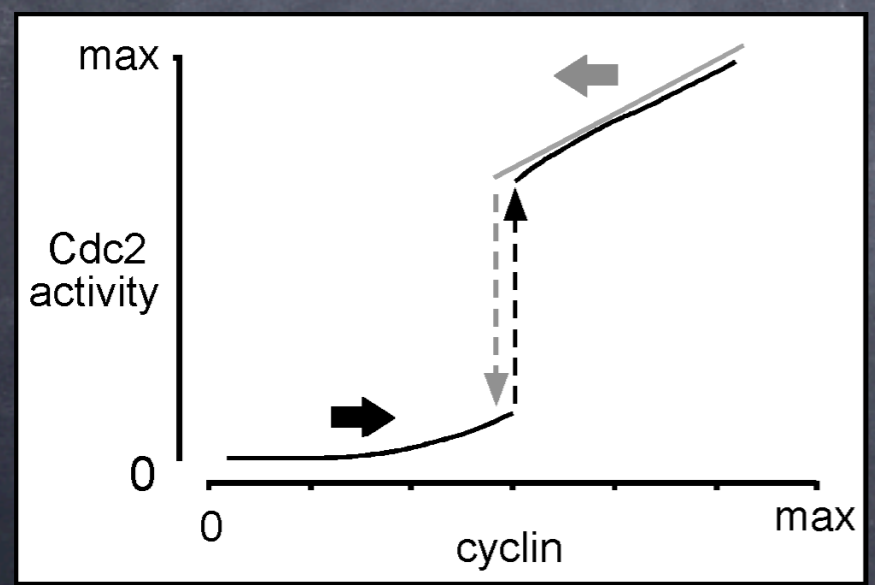
"cycling" egg extract

Oscillations of MPF Activity in Cycling Egg Extracts: What is the Systems-level Logic of This Oscillator?

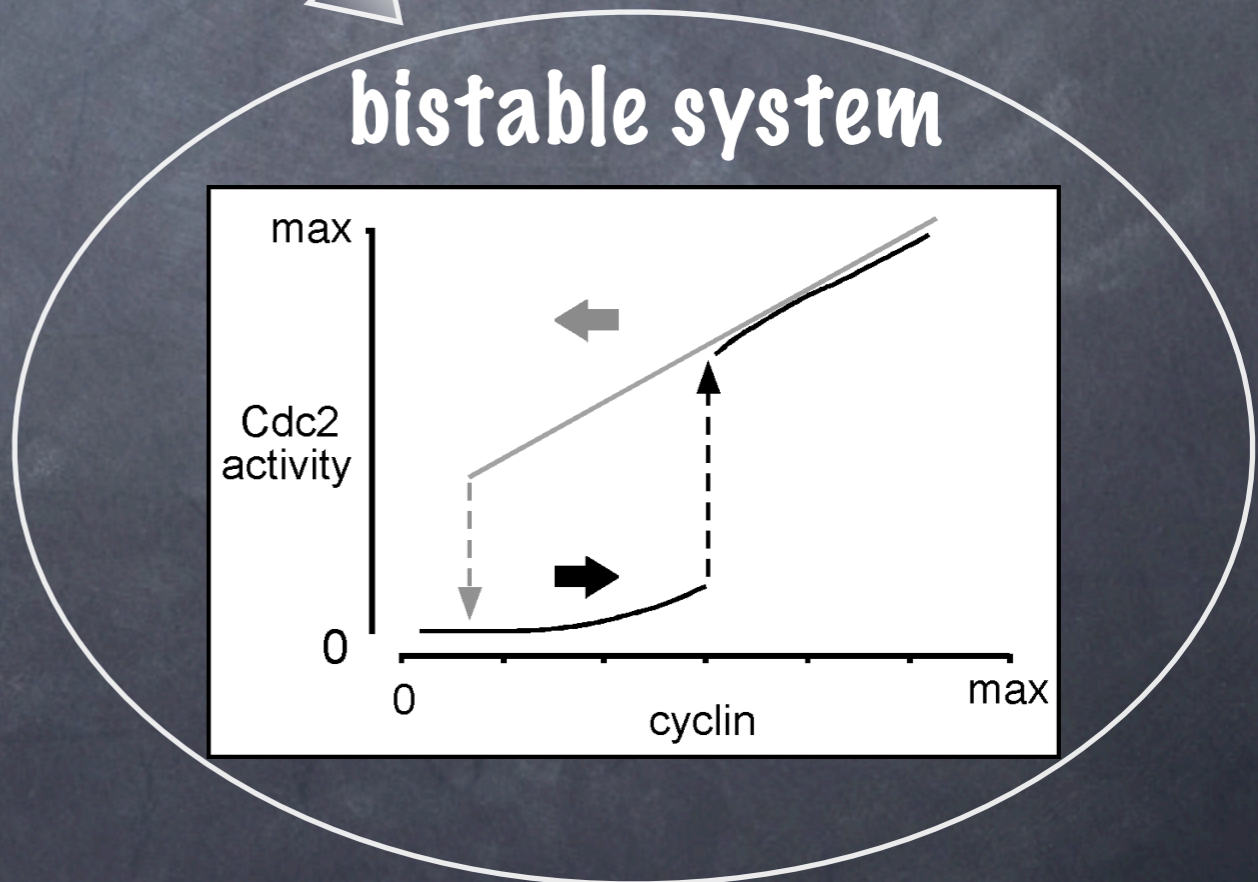


POSITIVE FEEDBACK

monostable system



bistable system

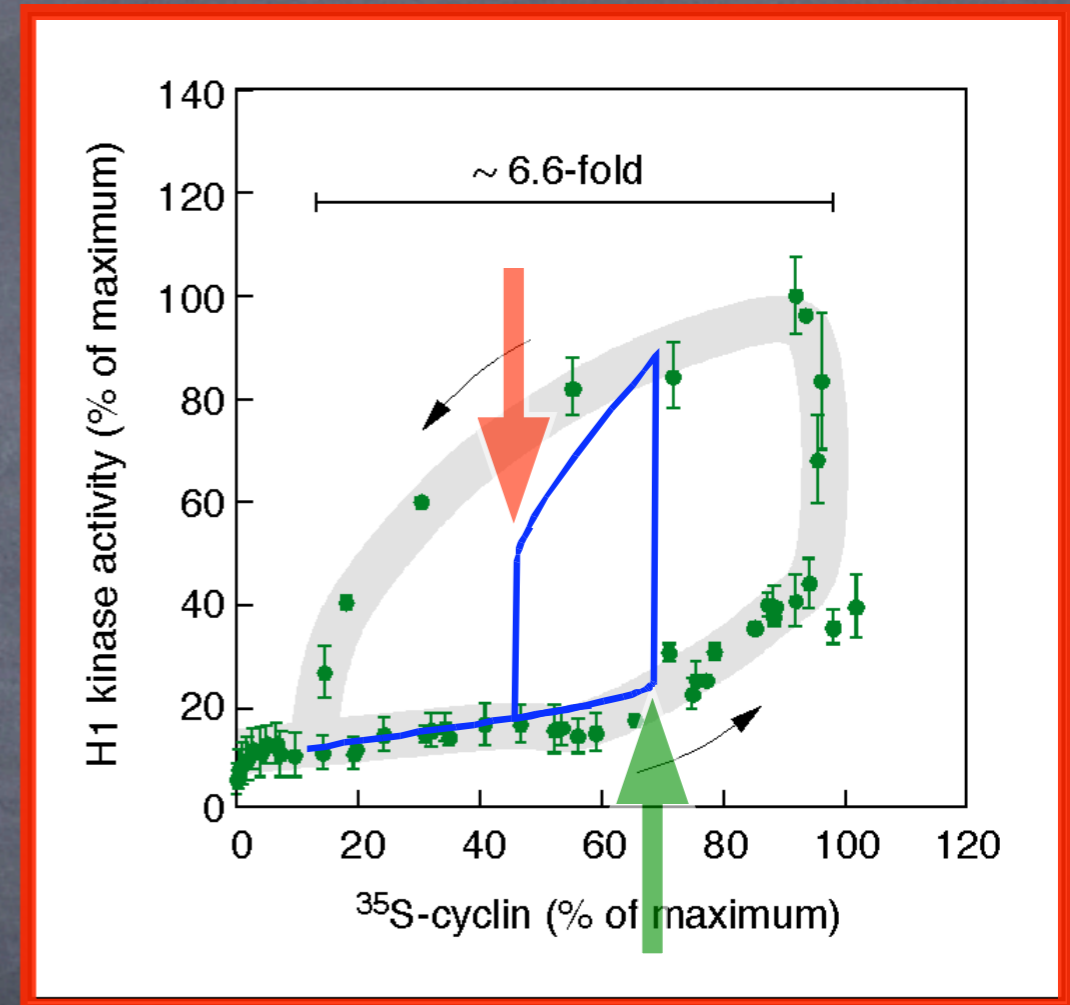
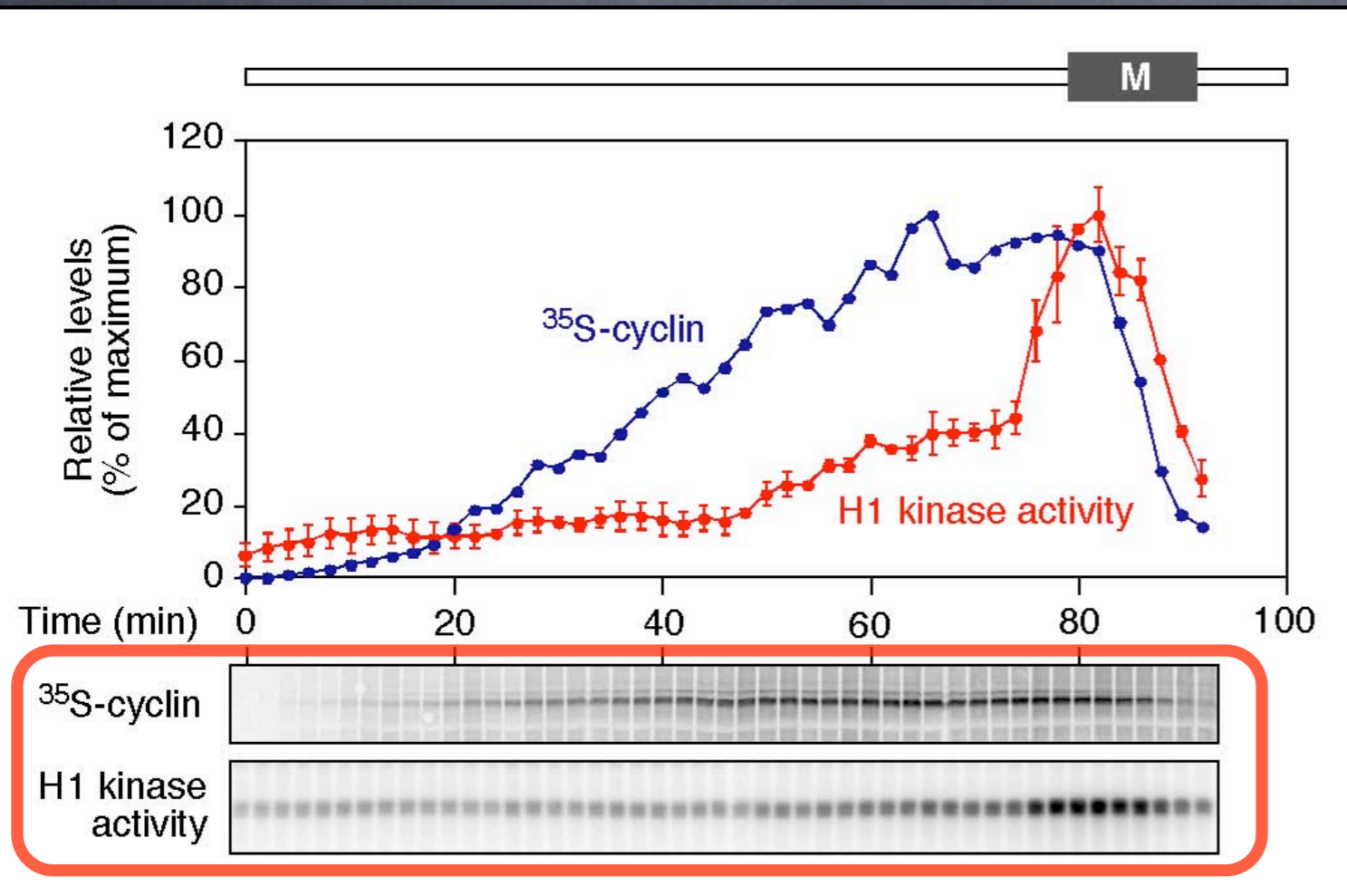


negative feedback oscillator?



relaxation oscillator?

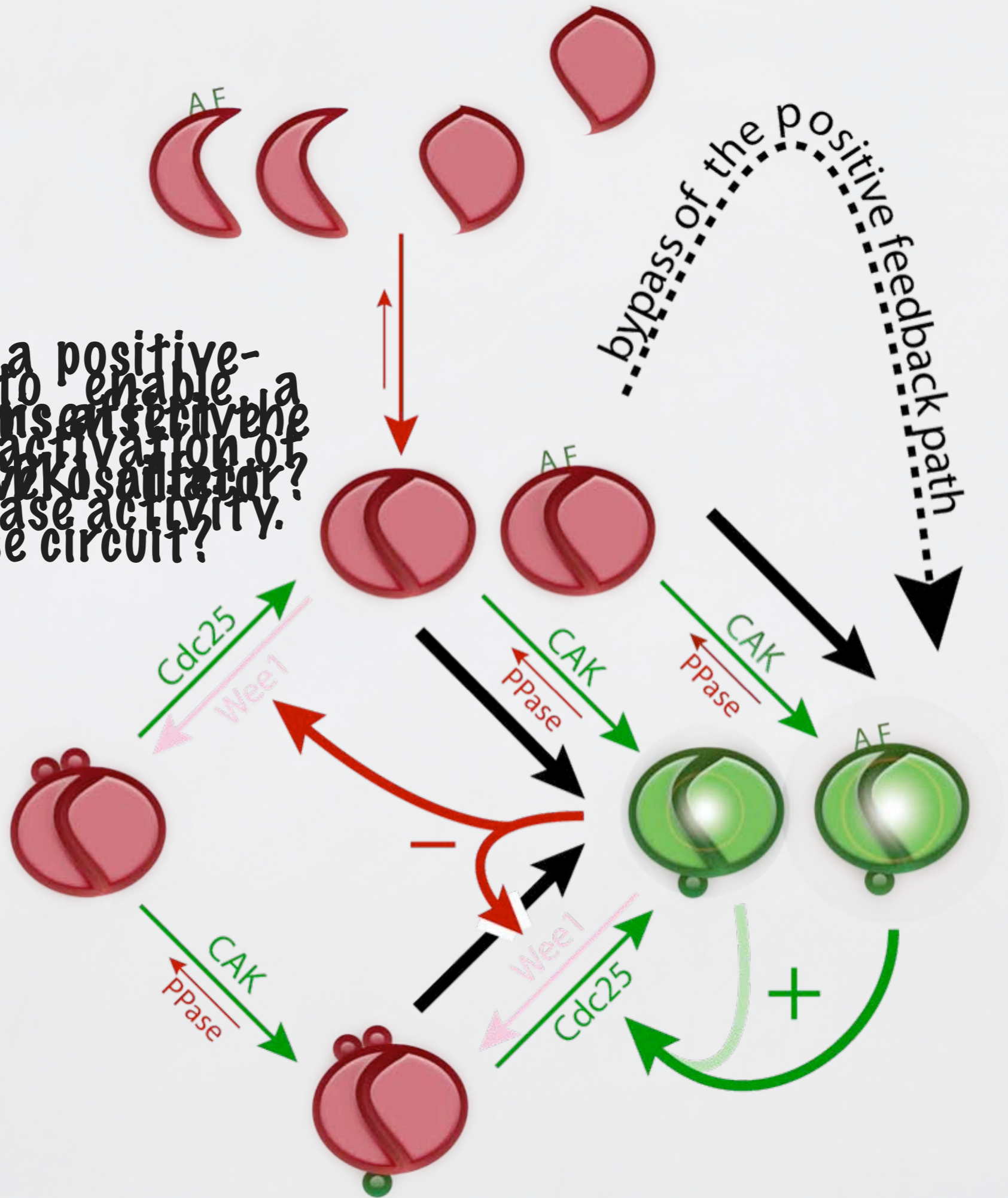
What is the Dynamical Relationship Between Cyclin B, CDK1, & APC?



The stimulus-response of CDK1(Cyclin B) overshoots the steady-state hysteresis loop.

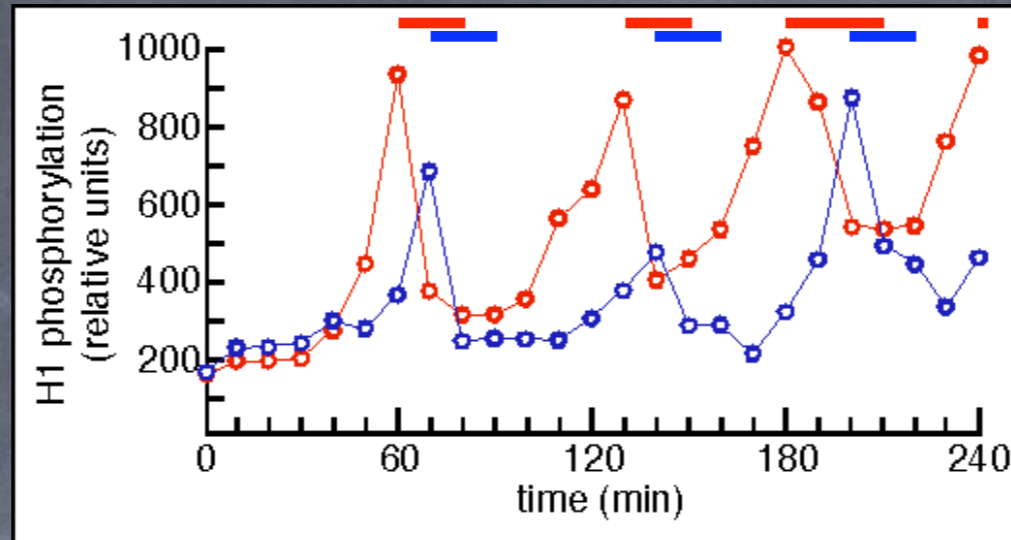
Is hysteresis merely a byproduct of the positive feedback, and the system really never needs to apply it, or does it infer the importance of positive feedback in the circuit?

How might a positive-feedback loop enable a system to overcome the premature activation of Cdk1A? Kinase activity. the M-phase circuit?

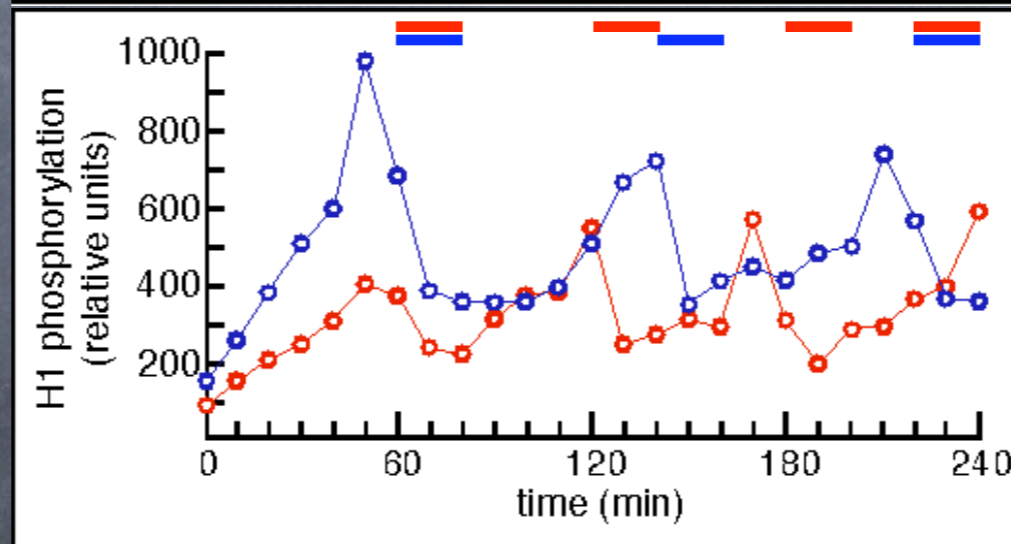


CDK1AF Accelerates Oscillations in a Cycling Egg Extract and Reduces Interphase Length

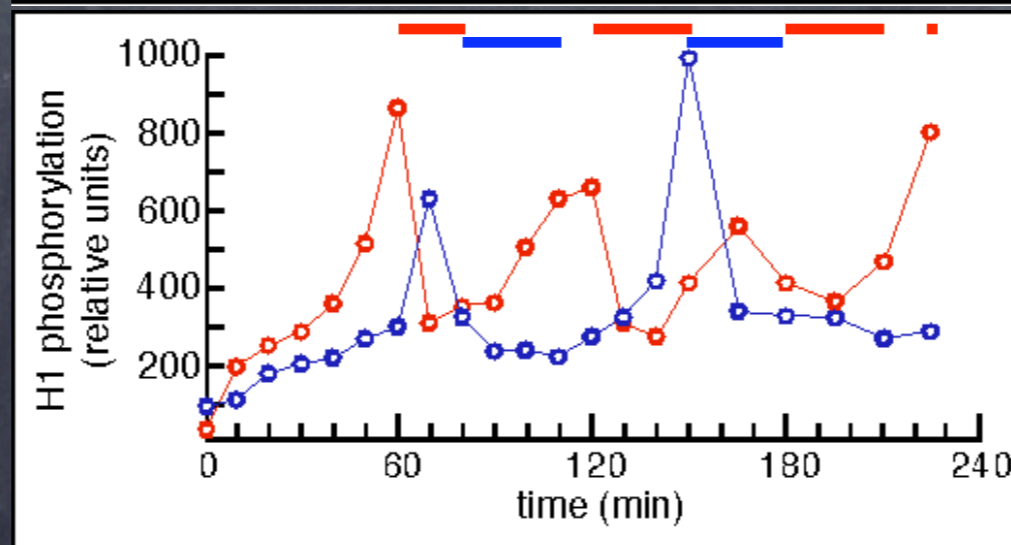
50 nM



100 nM

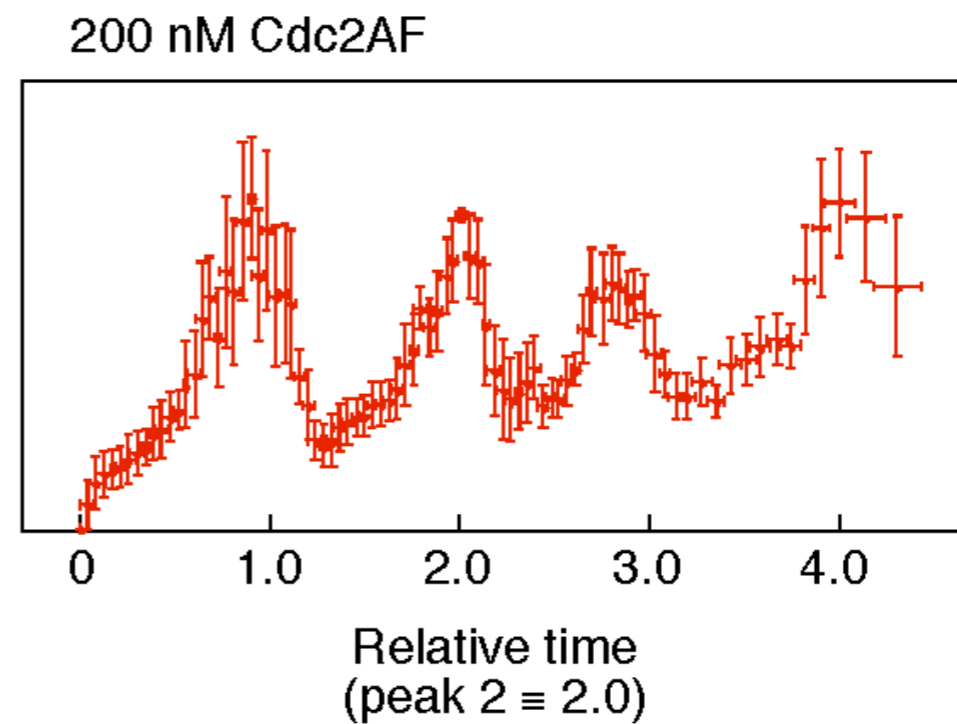
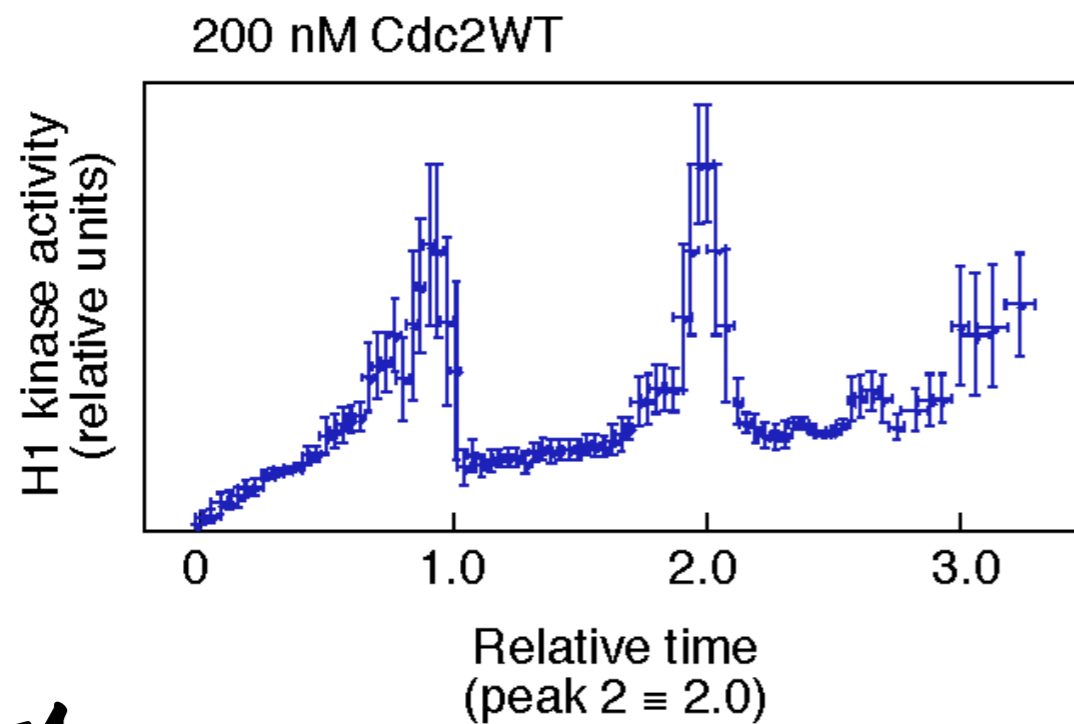


200 nM



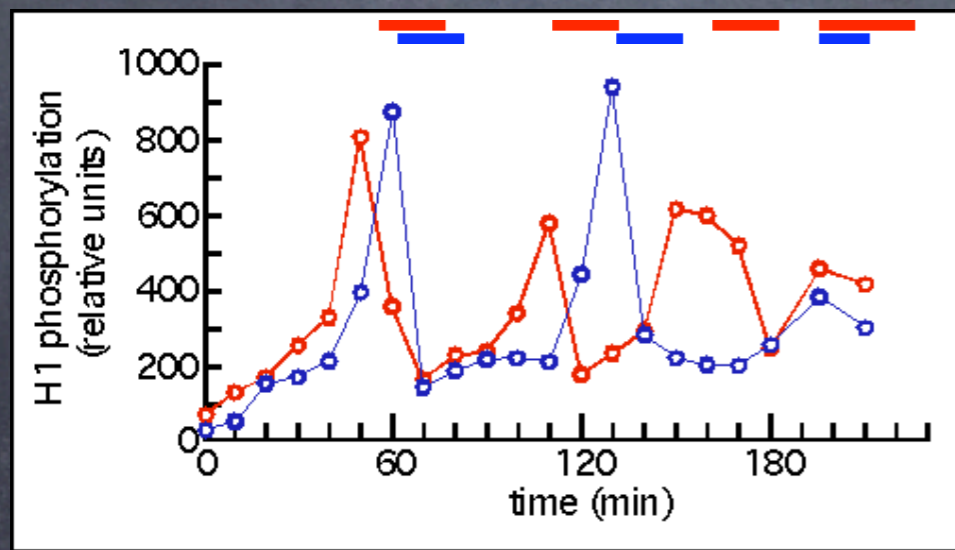
- XCDKIWT
- XCDK1AF

CDK1AF Accelerates Oscillations in a Cycling Egg Extract, Reduces Interphase Length, and Induces Damping

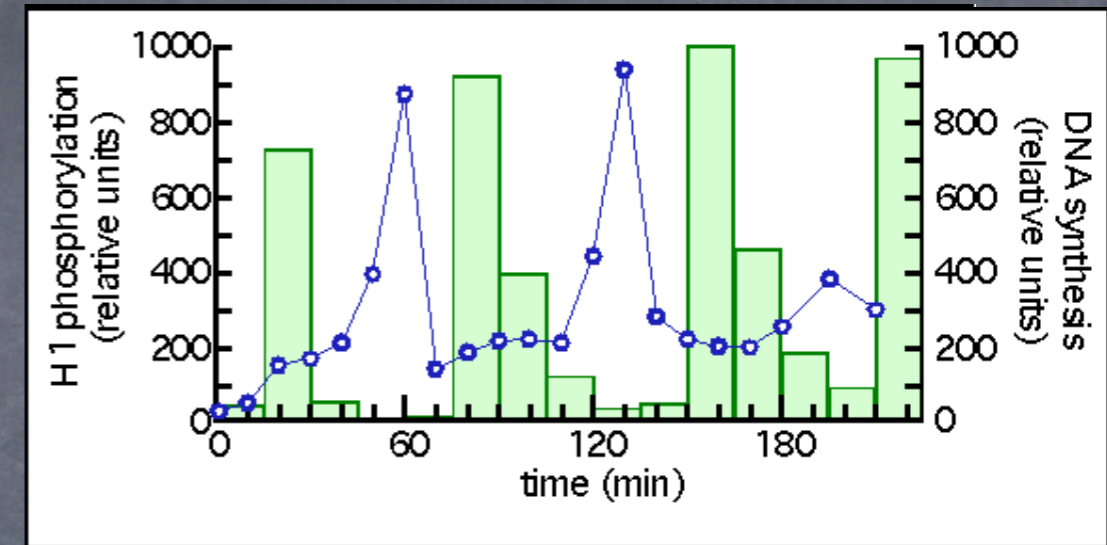


n=4

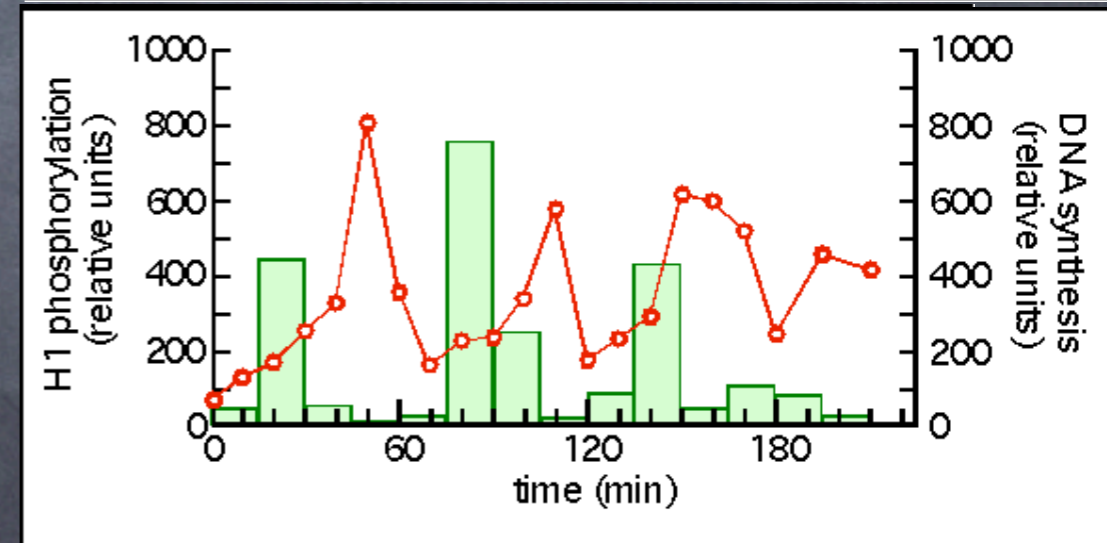
Does a CDK1AF-accelerated Extract Properly Relay Between M- and S-phases?



WT



AF



DNA synthesis is eventually disrupted

- XCDK1WT (200 nM)
- XCDK1AF (200 nM)

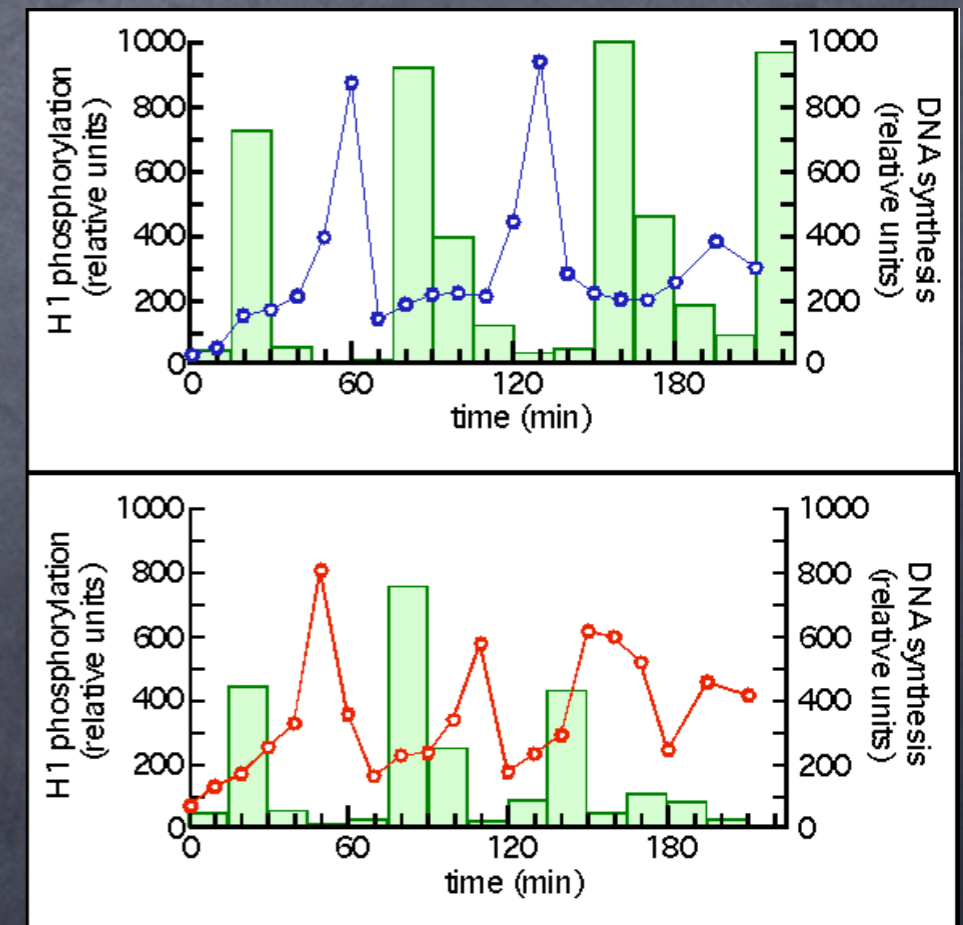
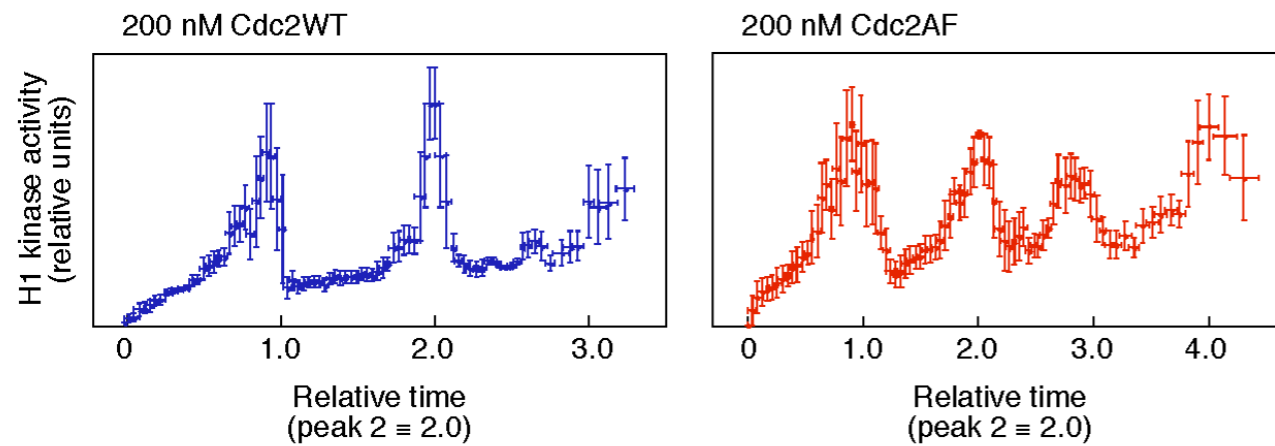
Summary-Part II

Bypassing positive feedback in CDK1 activation:

- accelerates oscillations in a cycling egg extract.
- reduces interphase length.
- induces damping of CDK1 activity.

As a consequence:

- DNA synthesis is inhibited.
- The discreteness between interphase and M-phase periods is compromised.

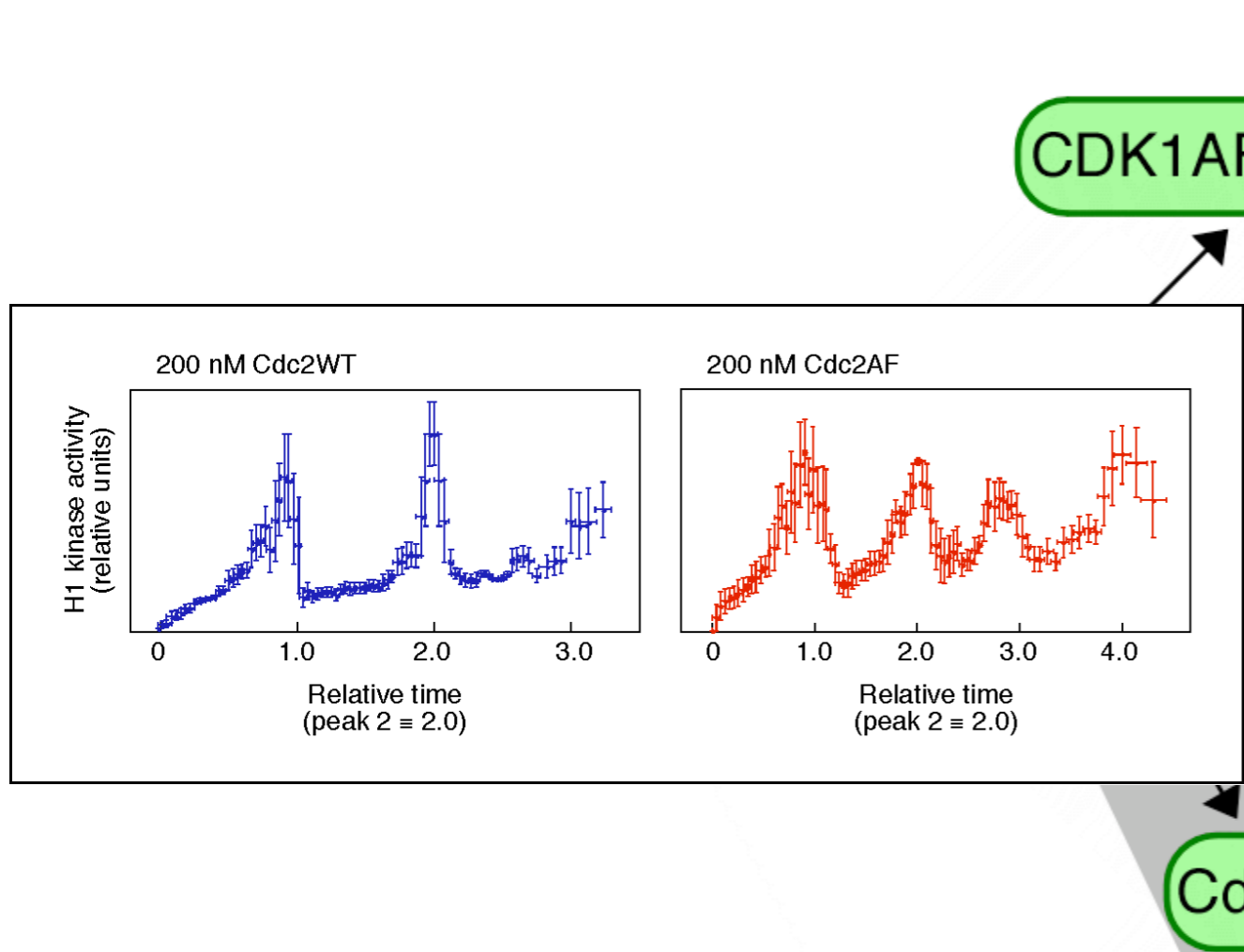


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Dynamical Studies in Embryonic Extracts



Cell, Vol. 122, 565–578, August 26, 2005, Copyright ©2005 by Elsevier Inc. DOI 10.1016/j.cell.2005.06.016

Systems-Level Dissection of the Cell-Cycle Oscillator: Bypassing Positive Feedback Produces Damped Oscillations

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Department of Molecular Pharmacology
Stanford University School of Medicine
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Stanford, California 94305

Summary

The cell-cycle oscillator includes an essential negative-feedback loop: Cdc2 activates the anaphase-promoting complex (APC), which leads to cyclin destruction and Cdc2 inactivation. Under some circumstances, a negative-feedback loop is sufficient to generate sustained oscillations. However, the Cdc2/APC system also includes positive-feedback loops, whose functional importance we now assess. We show that short-circuiting positive feedback makes the oscillations in Cdc2 activity faster, less temporally abrupt, and damped.

cyclin B mRNA cycle faster than control embryos (Hartley et al., 1996). The accumulating cyclin B binds to the cyclin-dependent kinase Cdc2, and, under the proper circumstances, this complex becomes activated and phosphorylates mitotic substrates, driving the transition from interphase to mitosis. The transition from mitosis back to interphase is driven by a negative-feedback loop. Active cyclin B-Cdc2 brings about the activation of the APC, which results in the polyubiquitylation and proteolysis of cyclin (Hershko et al., 1994; King et al., 1995, 1996; Sudakin et al., 1995) (Figure 1A). Although the presence of a negative-feedback loop does not guarantee sustained oscillations of Cdc2 and APC activity, it is difficult to imagine an oscillator that does not contain a negative-feedback loop. Indeed, when APC-resistant cyclin mutants are added to interphase *Xenopus* egg extracts, the extracts approach a steady state of Cdc2 activity rather than oscillating (Murray et al., 1989). This implies that the negative-feedback loop is essential for mitotic oscillations.

Embryos possess a positive-plus-negative feedback CDK1 oscillator. How different is the systems-logic of the CDK1 module in somatic cells? Is it more like a series of dependent, more highly-regulated switches?



Dominoes and Clocks: The Union of Two Views of the Cell Cycle

ANDREW W. MURRAY AND MARC W. KIRSCHNER

We review the recent advances in understanding transitions within the cell cycle. These have come from both genetic and biochemical approaches. We discuss the phylogenetic conservation of the mechanisms that induce mitosis and their implications for other transitions in the cell cycle.

THE CELL CYCLE IS THE SET OF EVENTS THAT IS RESPONSIBLE for the division of a cell into two daughter cells. The recent advances in our

Hartwell and his colleagues (1) and later in the fission yeast by Nurse and his colleagues (2), the result was a description of the cell cycle as a set of dependent reactions. The basis of this dependency is discussed in the accompanying review by Hartwell and Weinert (3). The physiological and embryological approach was championed by researchers who favored marine and amphibian eggs. They argued that eggs and oocytes were the simplest systems for studying the basic processes of the cell cycle, because they were specialized for rapid cell division. The result of their investigations was a description of the cell cycle as a biochemical machine that oscillated between two states, mitosis and interphase, and whose oscillations were independent of the completion of many of the cell cycle events.

Cell cycle progression in the EARLY EMBRYO:

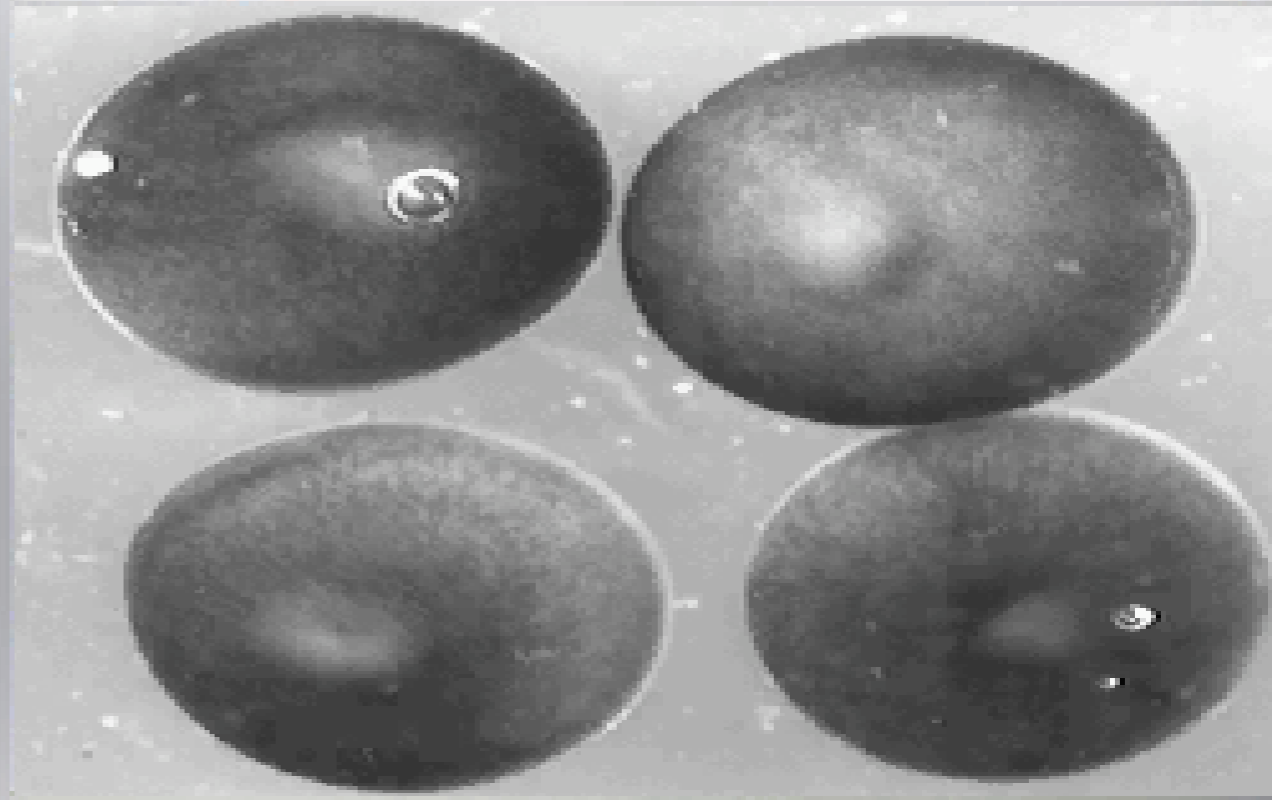
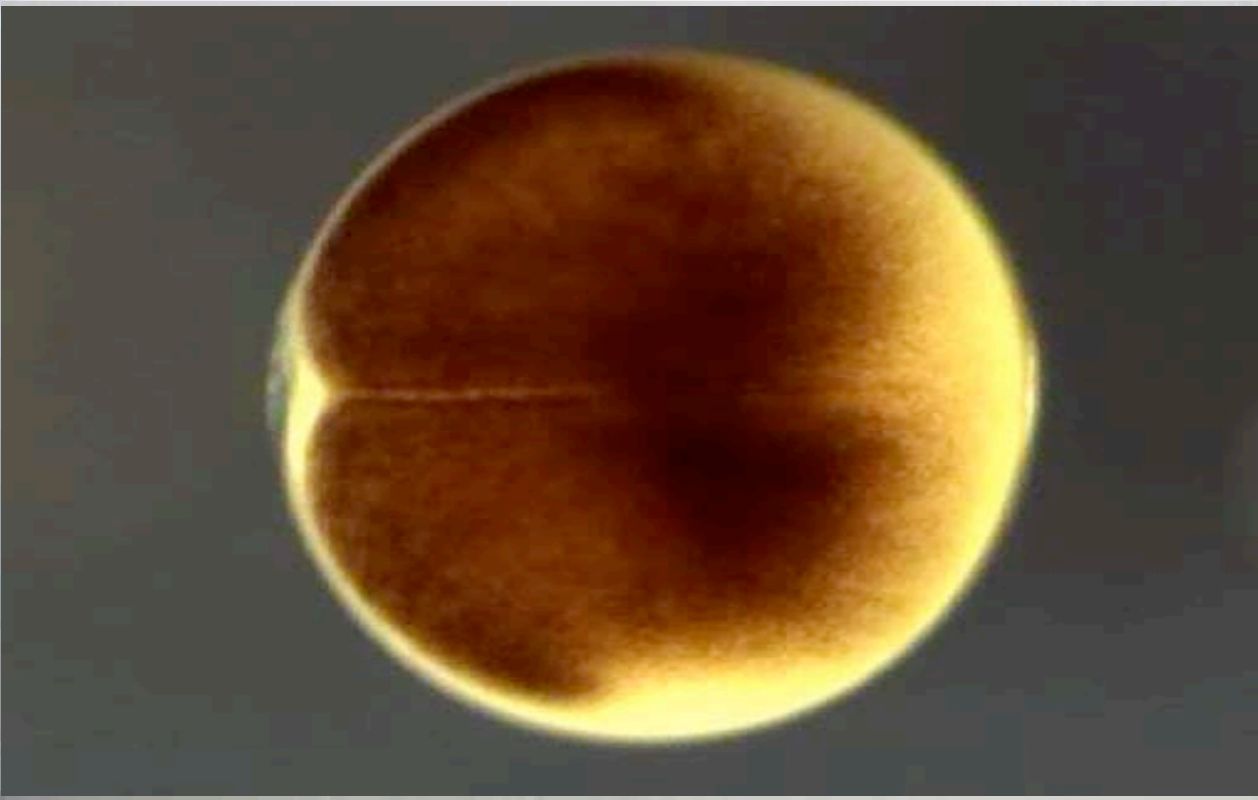
An autonomous oscillator, running just like a CLOCK - "Oscillations of M-phase-promoting activity independent of the completion of many of the cell cycle events."

Cell cycle progression in SOMATIC CELLS:

Sets of dependent reactions, just like DOMINOES - "Any fusion between two interphase cells at different stages of the cell cycle, the advanced nucleus (in G2) waits for the completion of events in the retarded nucleus (in G1) before progressing in the cell cycle."

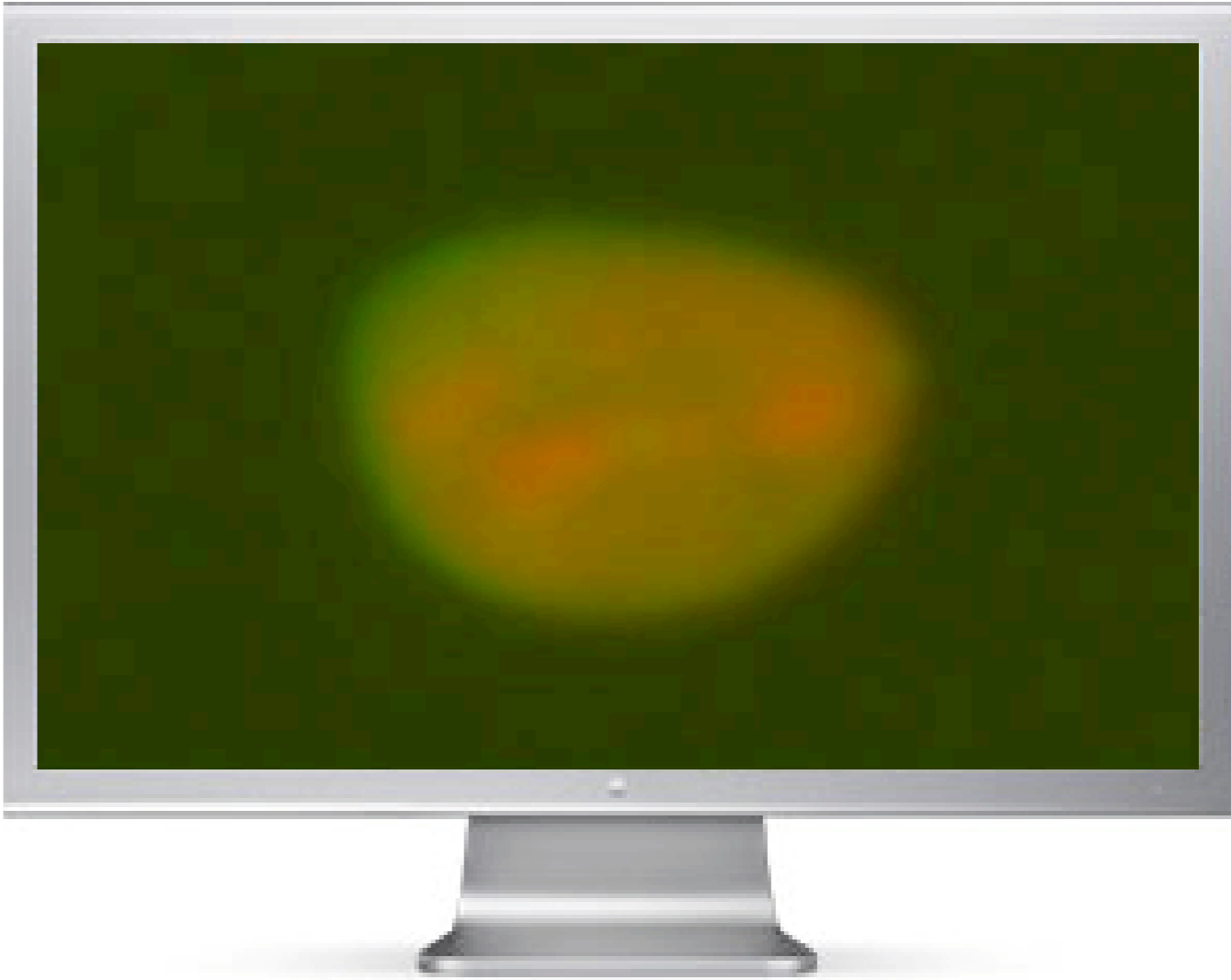
**no growth or checkpoints until after
division 12 (MBT), approximately 6 hpf**

nocodazole-treated embryos

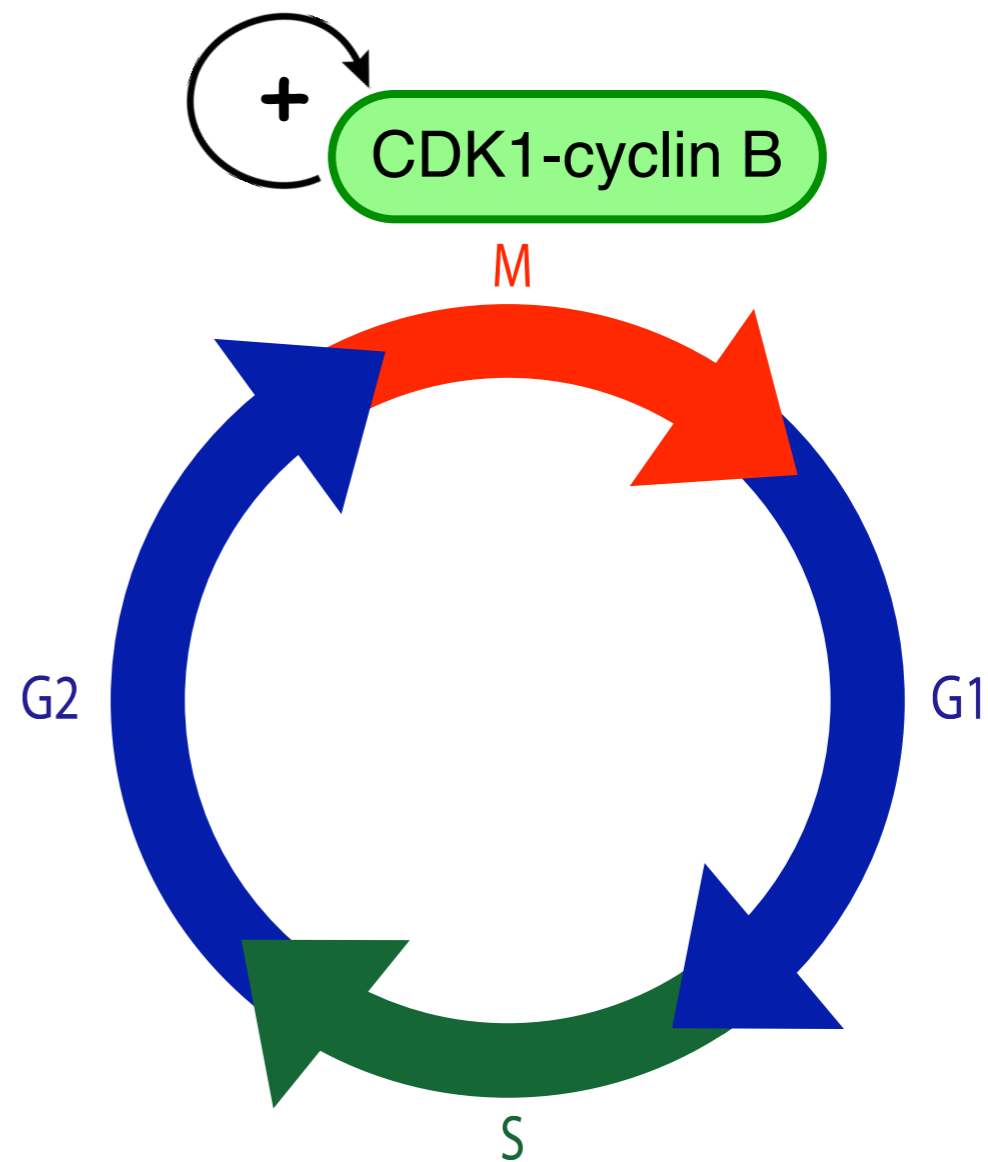
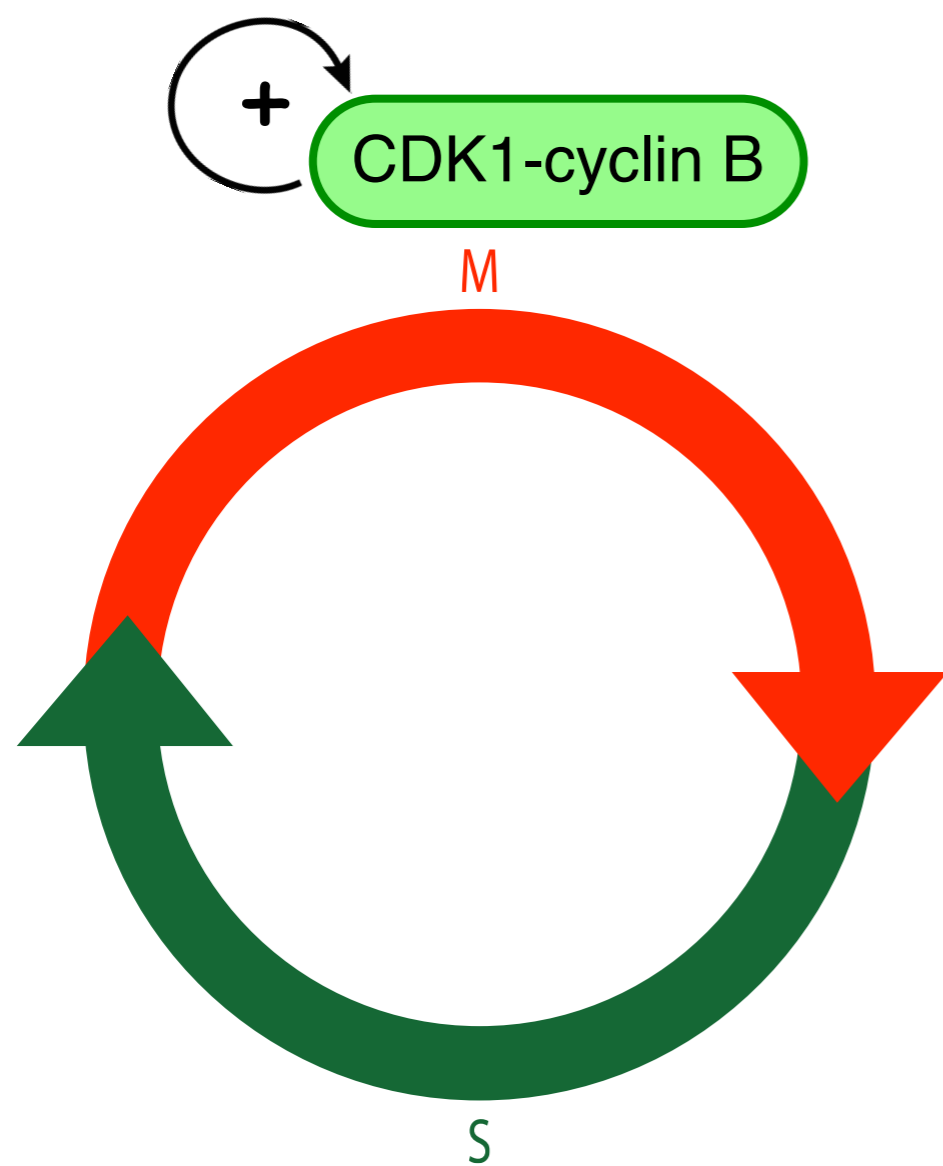


**no growth or checkpoints until after
division 12 (MBT), approximately 6 hpf**

**growth periods & checkpoints enabled;
quiescence, differentiation, senescence**



Do the design principles underlying the M-phase circuit differ between...

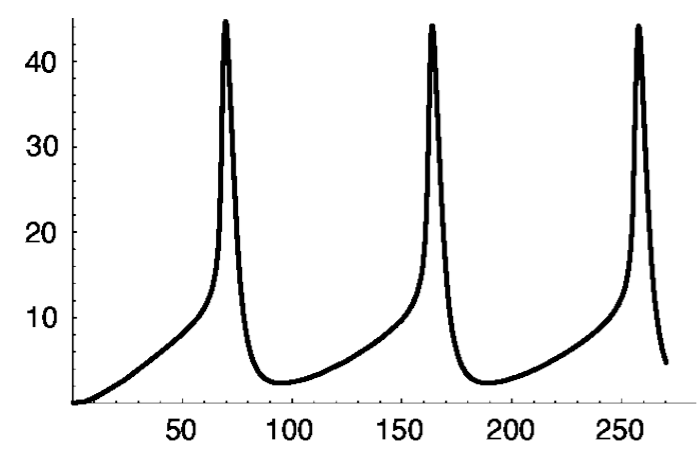


...and of barriers to oncogenesis?

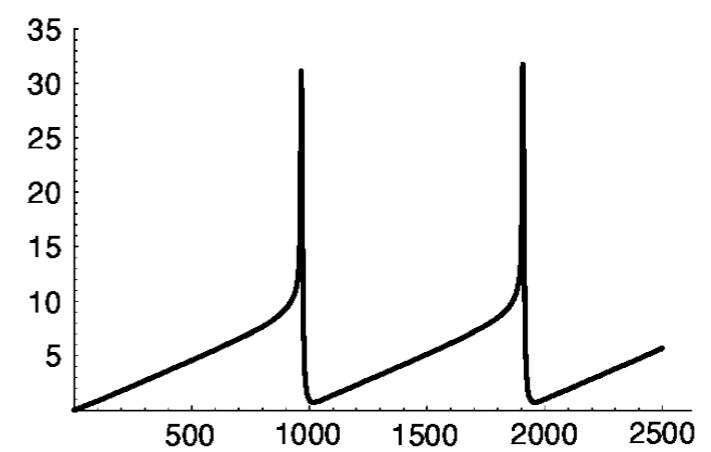
Somatic Controls = f(Transcriptional regulation? Checkpoints? Positive Feedback?)

Is positive feedback a biochemical necessity for proper M-phase control, from embryo to adult?

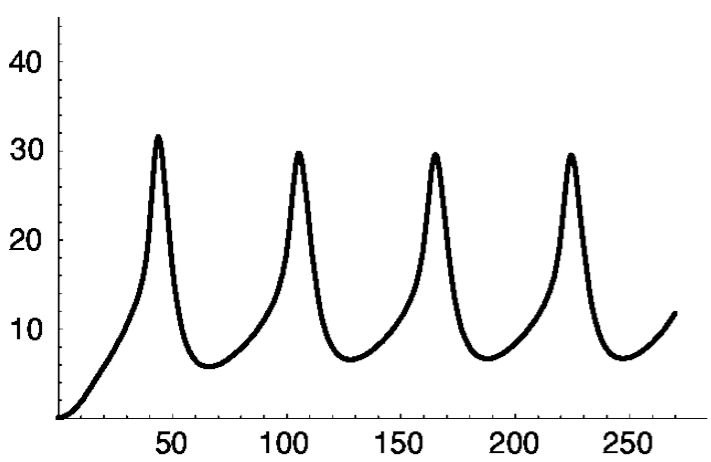
If this IS the case, then the somatic system may very well depend on positive feedback...
Simplest Case: Consider the Rate of Cyclin Synthesis & [CDK1AF]



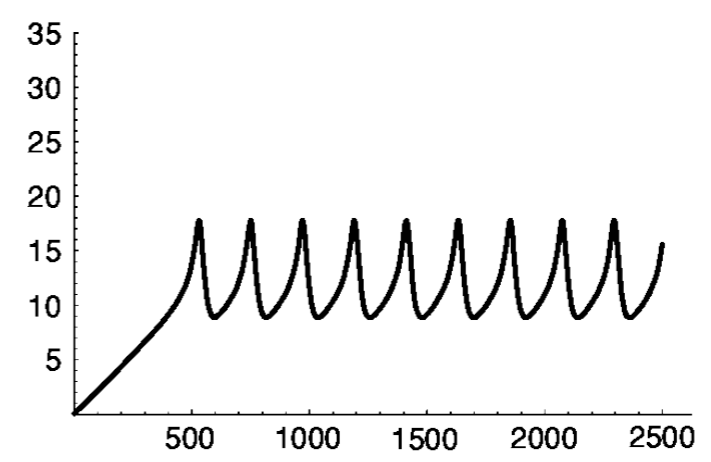
2 nM/min, 0 nM AF



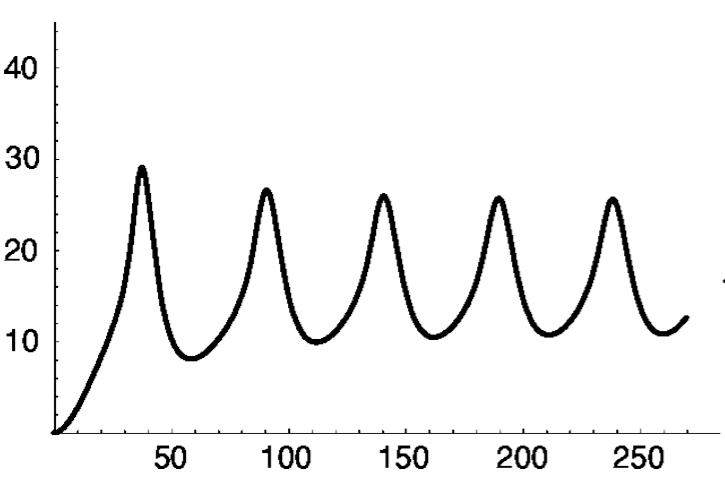
0.08 nM/min, 0 nM AF



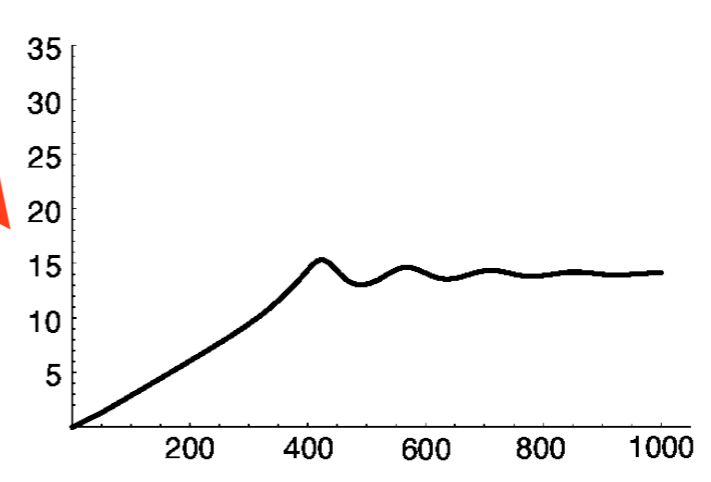
2 nM/min, 50 nM AF



0.08 nM/min, 50 nM AF



2 nM/min, 100 nM AF



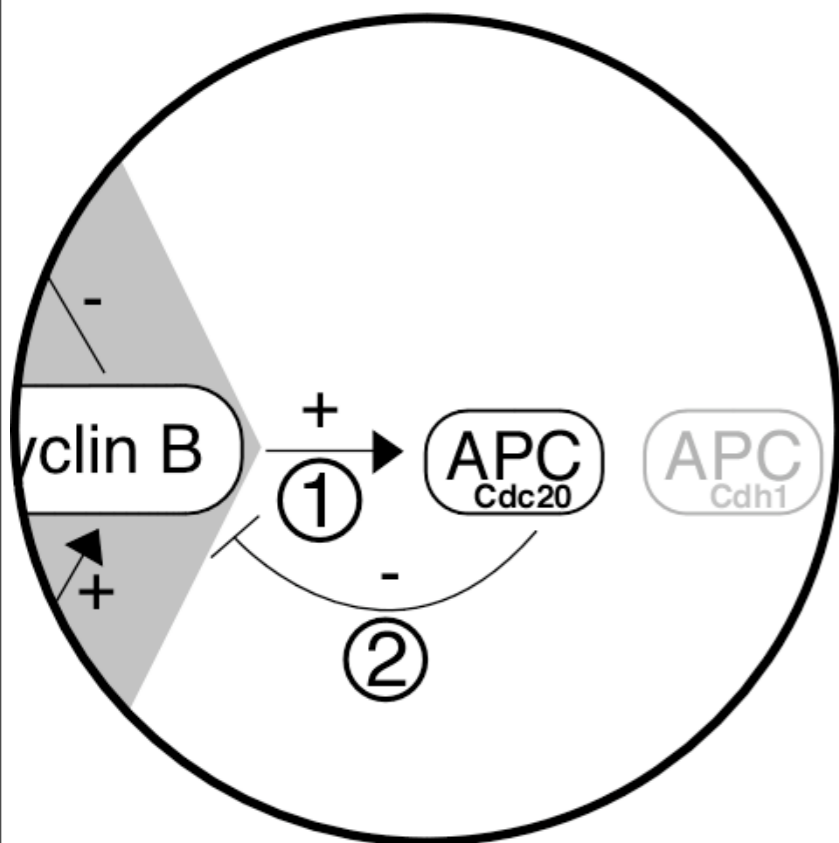
0.08 nM/min, 100 nM AF

EARLY EMBRYO example

SOMATIC CELL example

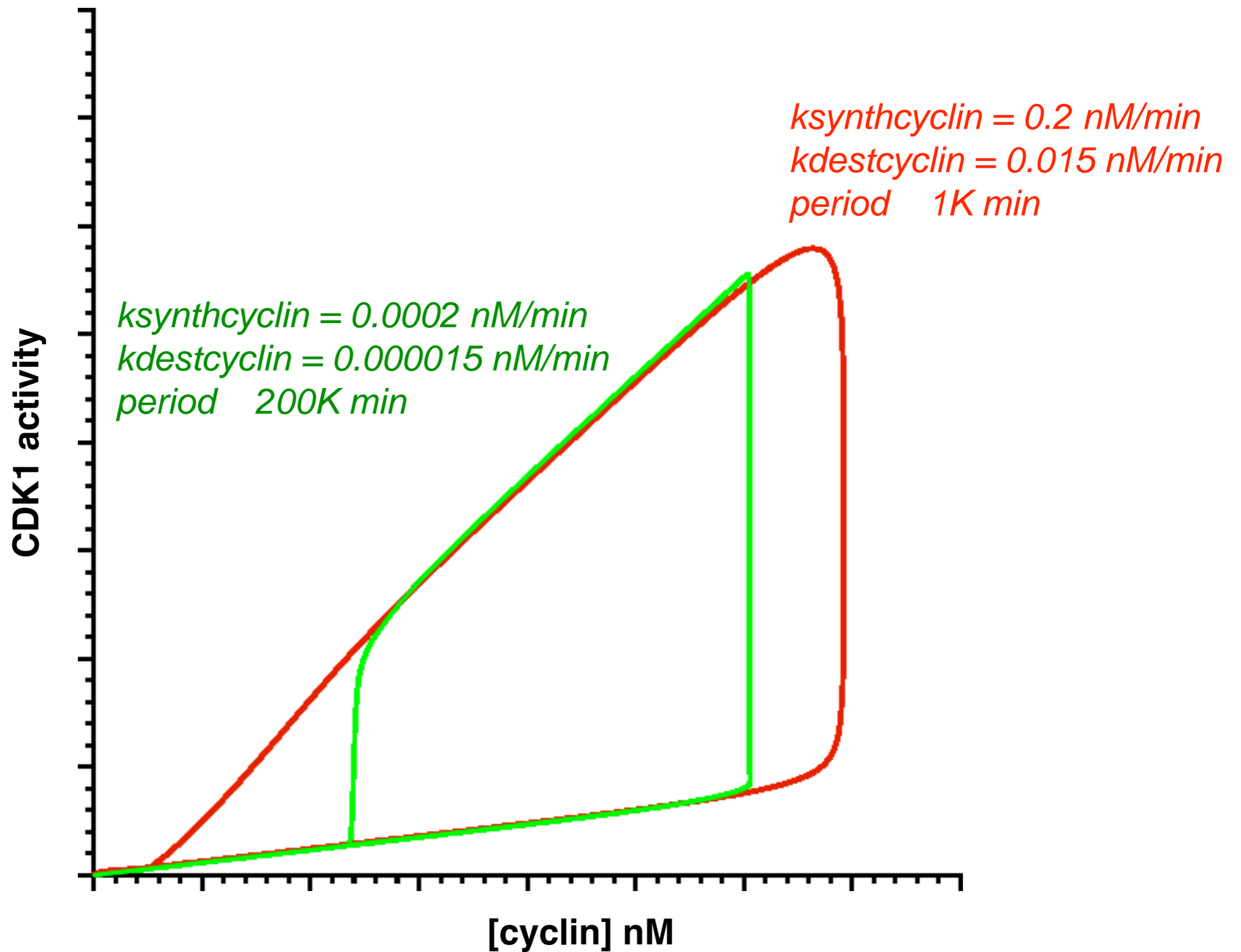
Negative-Feedback Loops and the Cell Cycle: From Early Embryo to Adult

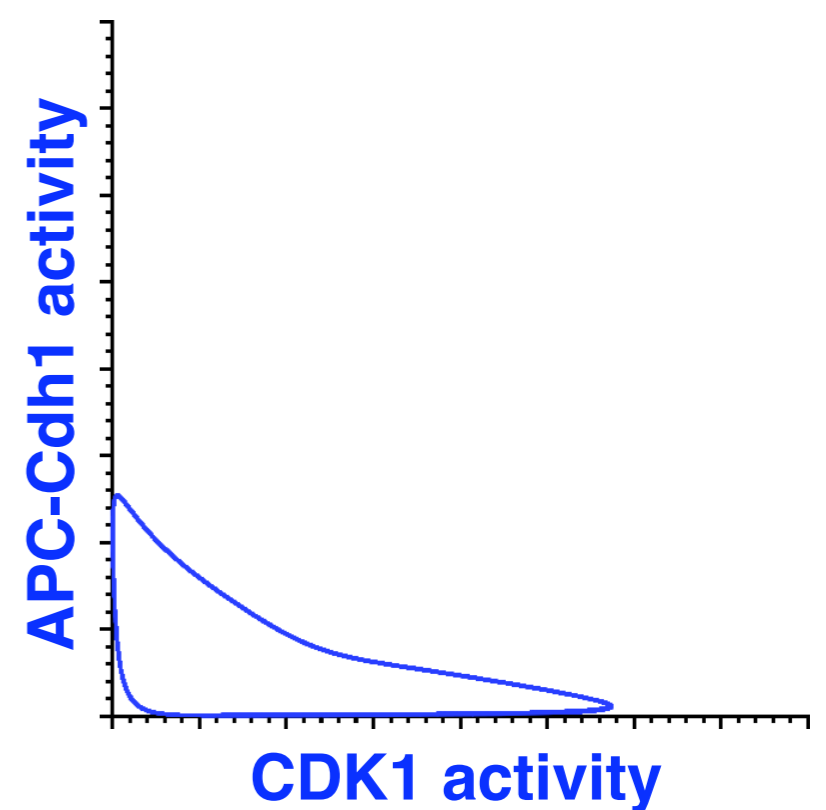
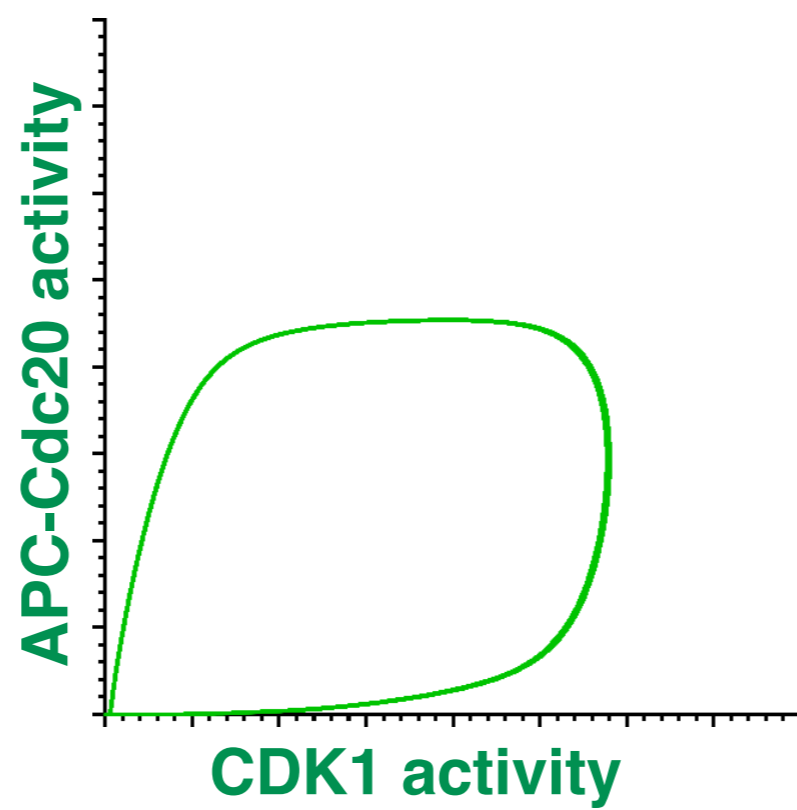
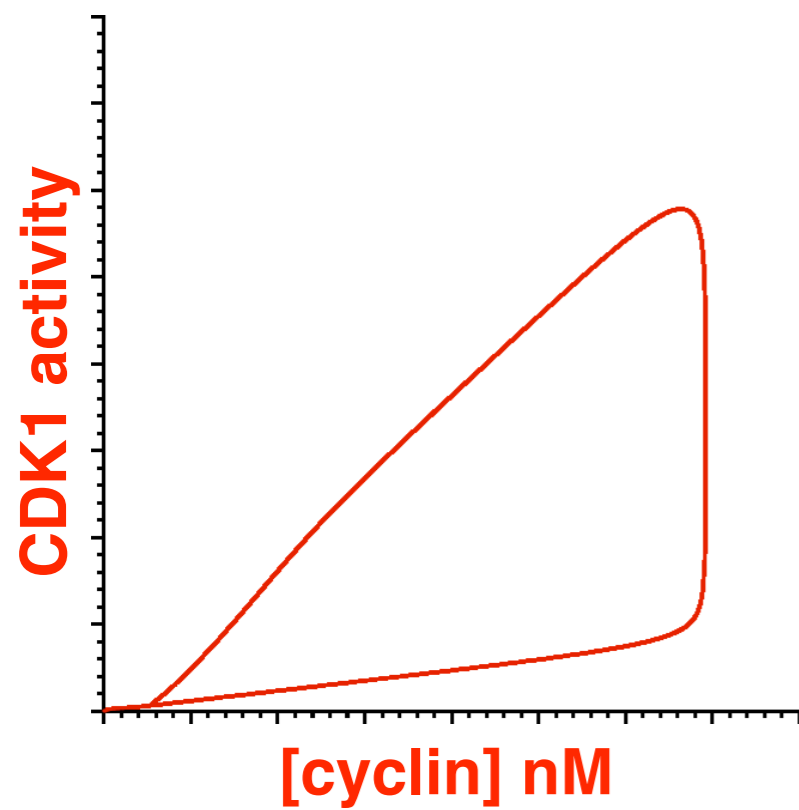
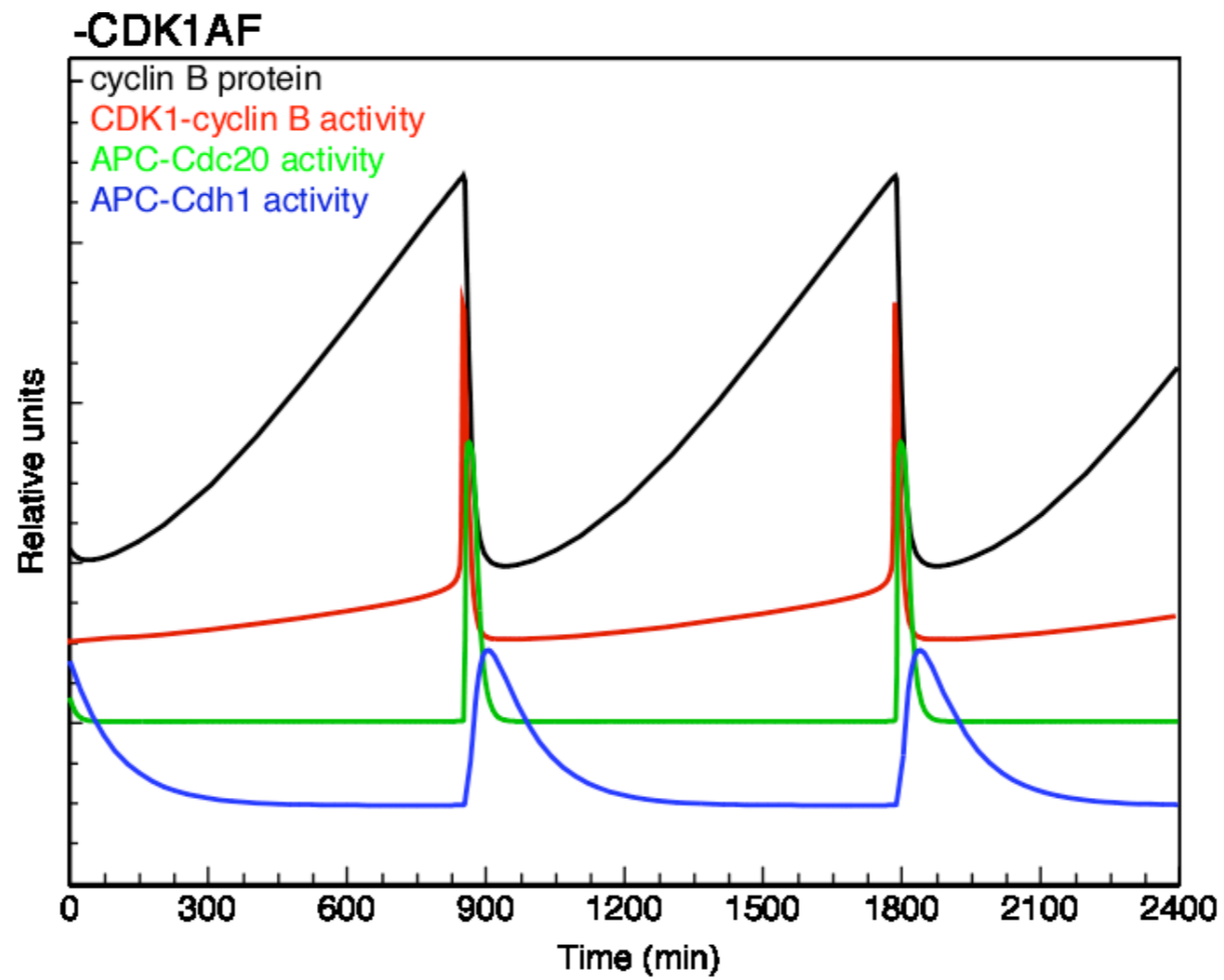
early embryonic cell



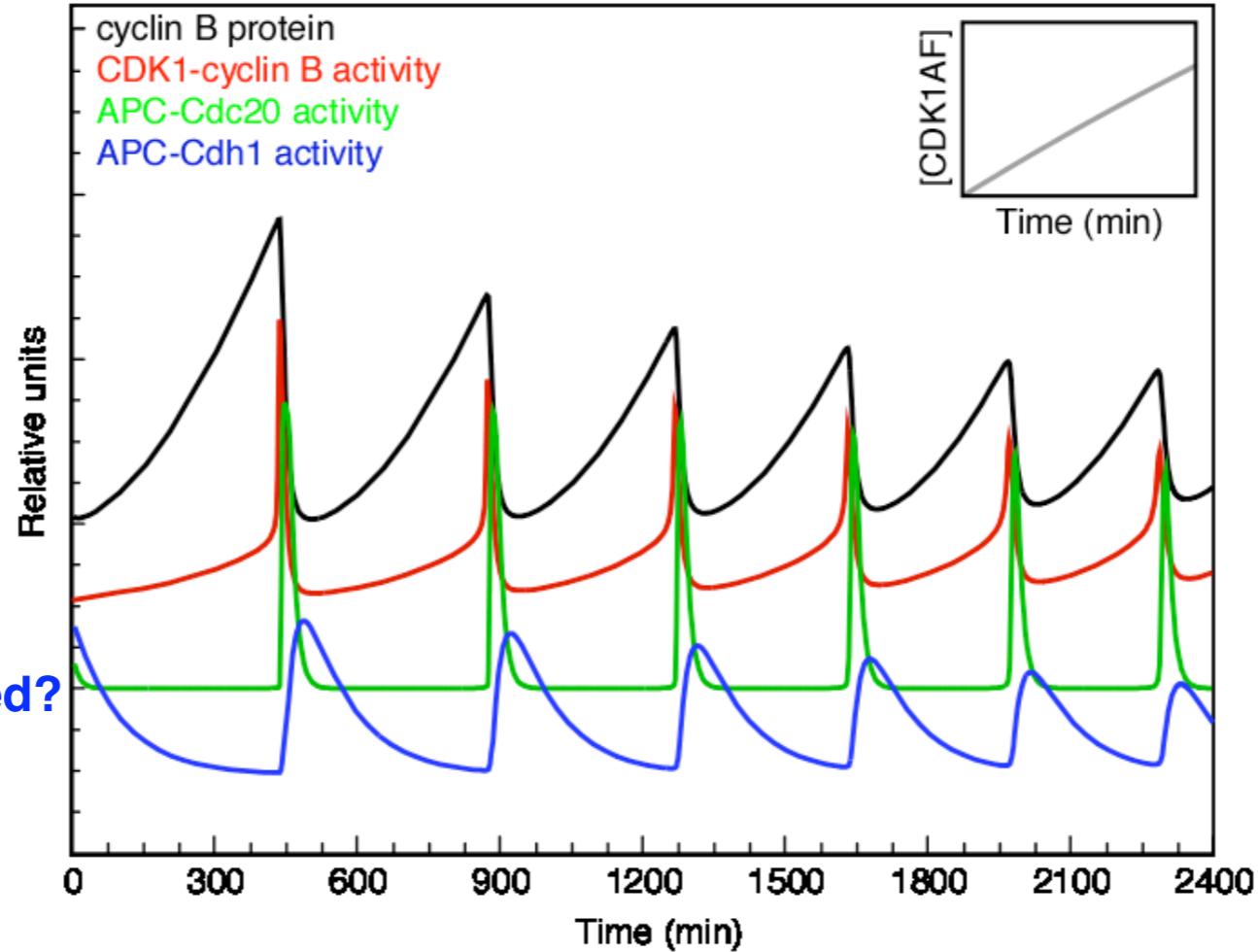
- 16 coupled ordinary differential equations
 - ksynth (cyclin)* - “stimulus parameter”
 - kdest (APC-Cdc20 & APC-Cdh1)* - “destruction parameter”
- 44 parameters total
- scripted in *Mathematica*

The "Somatic" Model & its Overshoot of the Hysteresis Loop





+CDK1AF



1) Early mitosis?

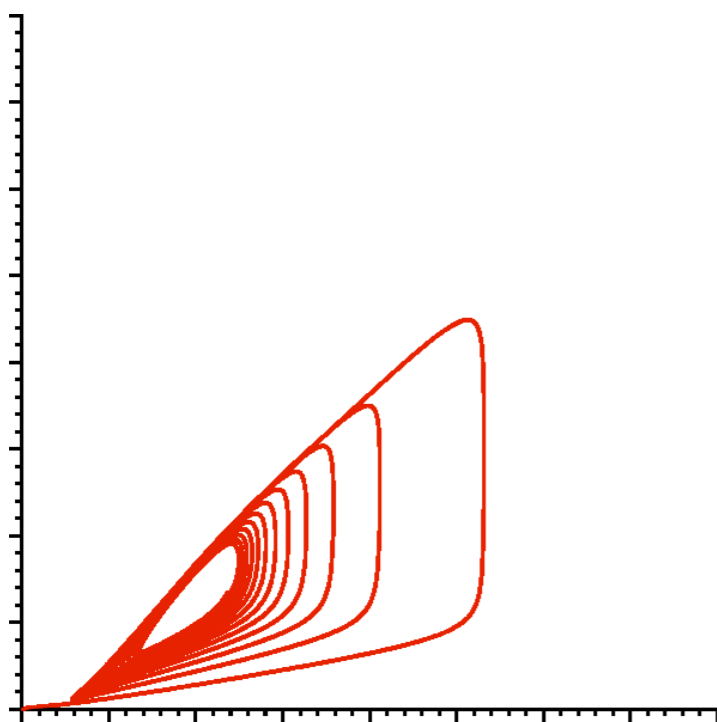
2) Increased frequency?

3) Some damping?

4) Cdc20 activity maintained?

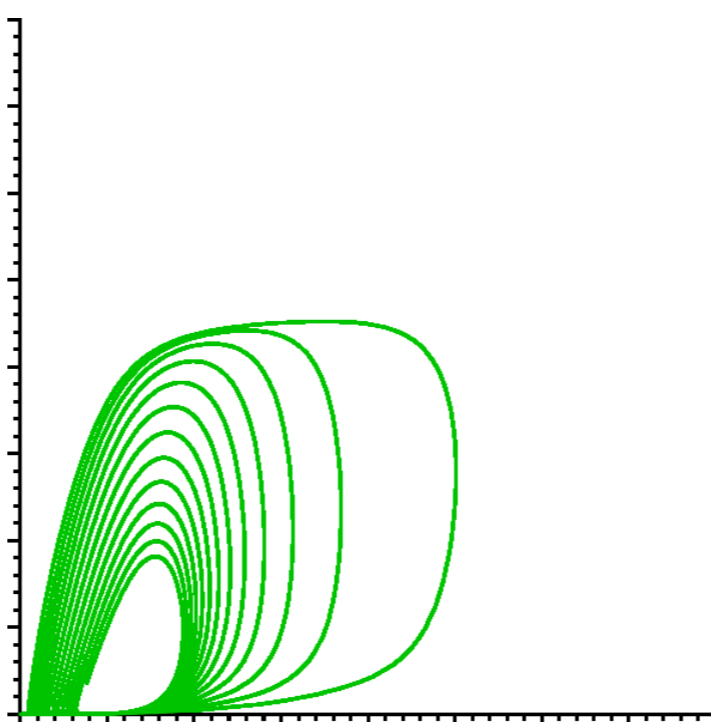
5) Cdh1 activity period reduced?

CDK1 activity



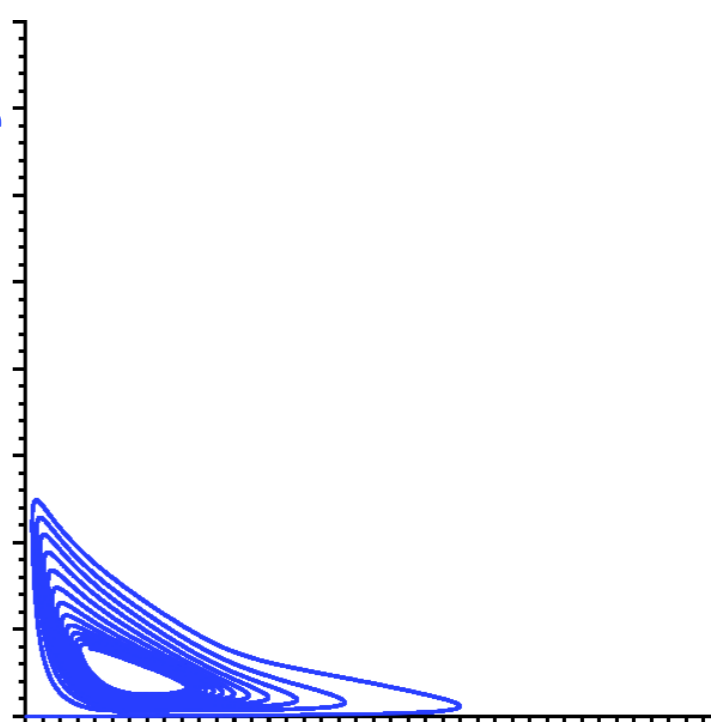
[cyclin] nM

APC-Cdc20 activity



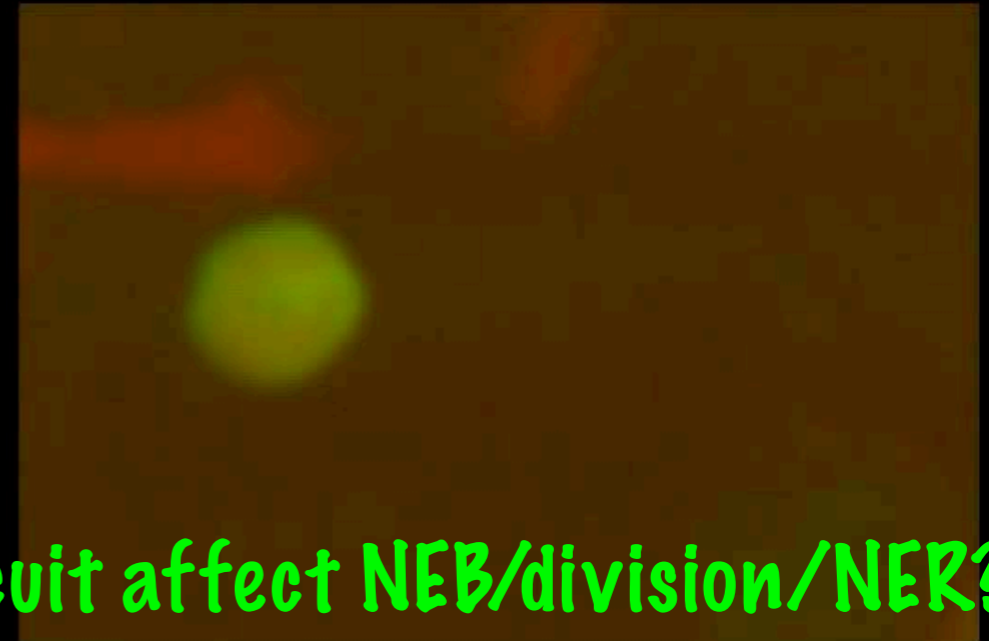
CDK1 activity

APC-Cdh1 activity



CDK1 activity

YFP-Lamin As A Nuclear Biosensor



How will the positive-feedback short-circuit affect NEB/division/NER?

YFP-Lamin A1

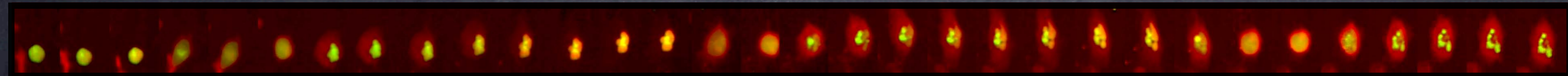
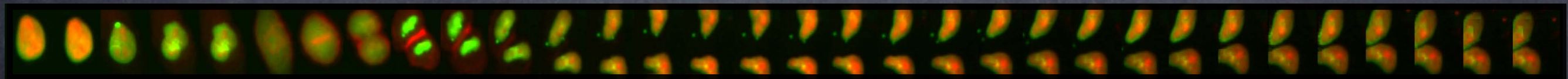
MBS-Red

CFP-CDKIWT (not shown)

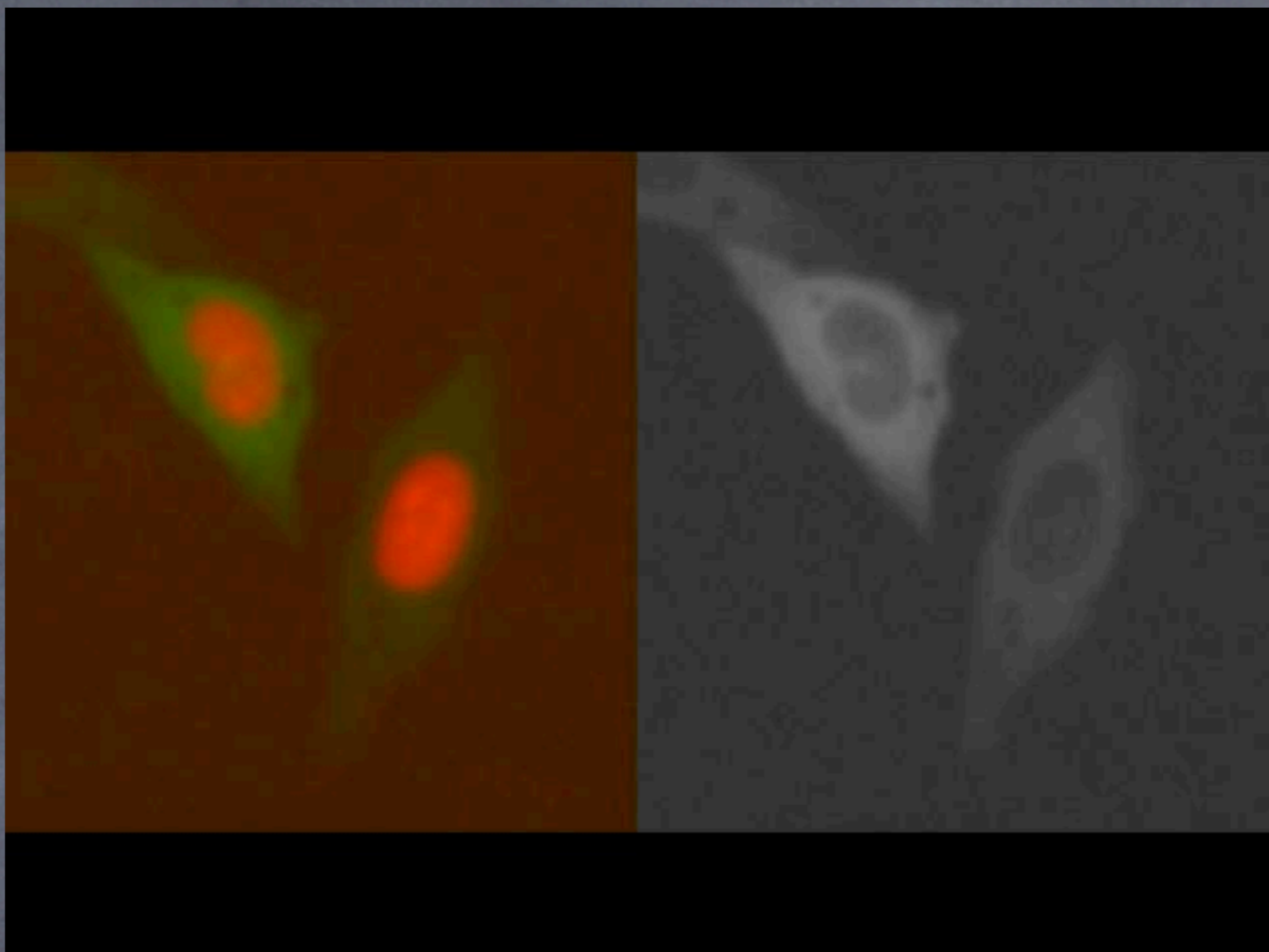
YFP-Lamin A1

MBS-Red

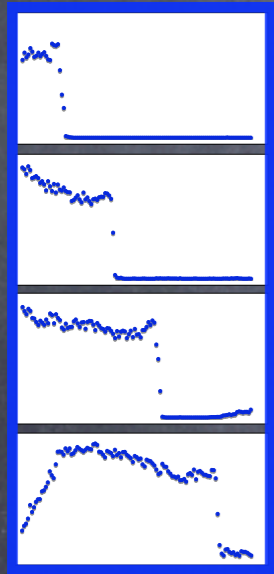
CFP-CDK1AF (not shown)



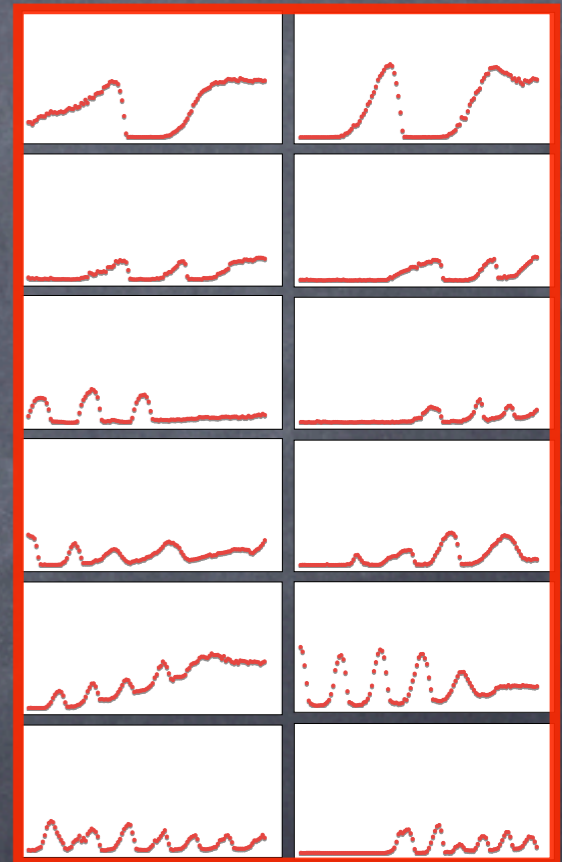
Cyclin B1-YFP As A Negative-Feedback & CDK1 Sensor



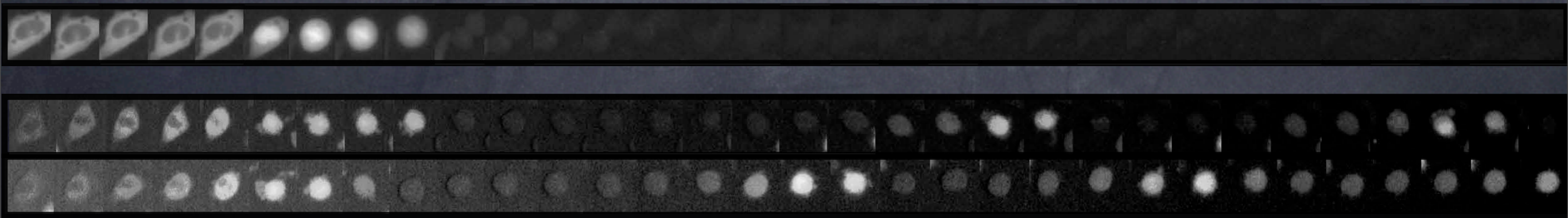
CDKIWT



CDK1AF



Cyclin B1-YFP
MBS-Red
GFP-CDK1AF (in cell shown)

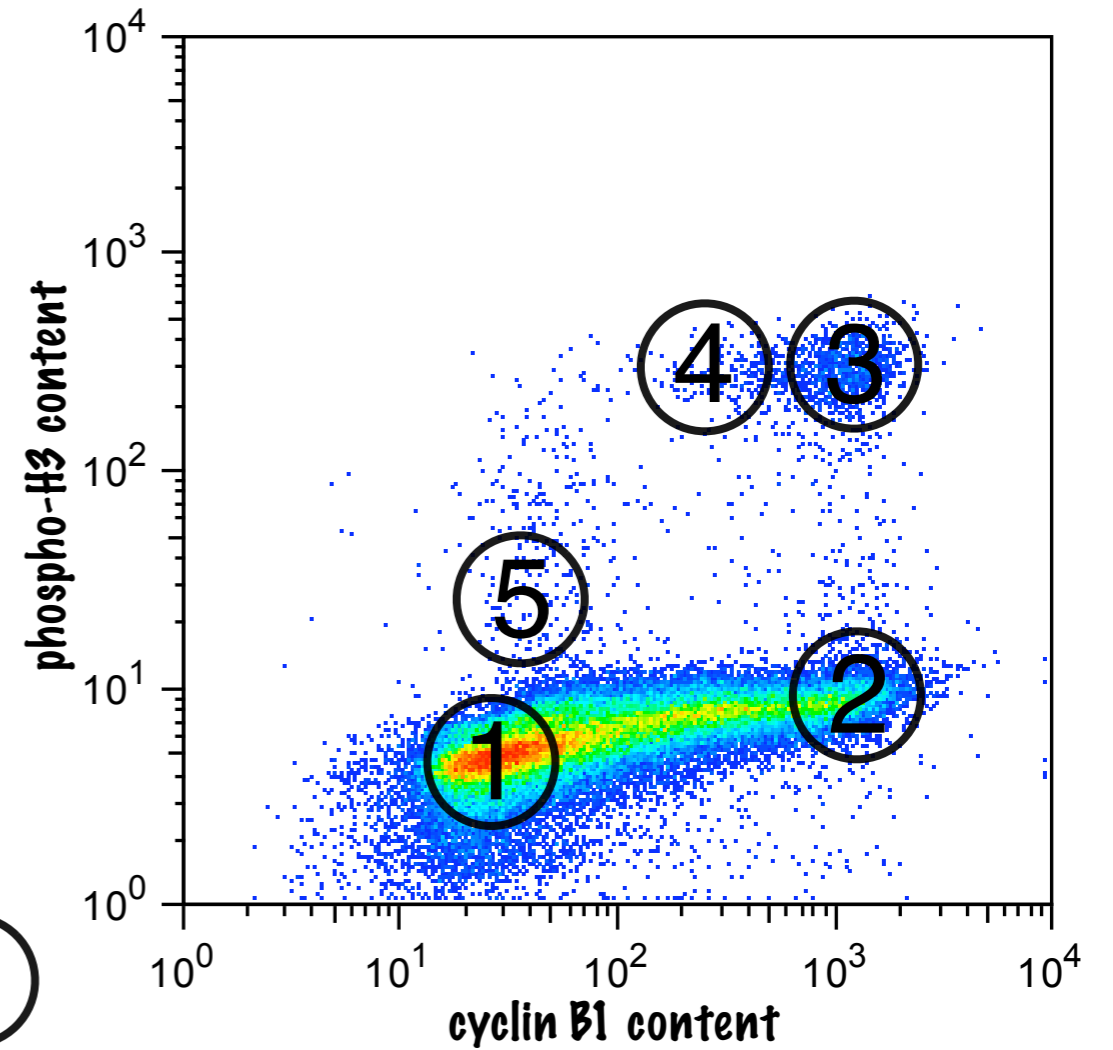
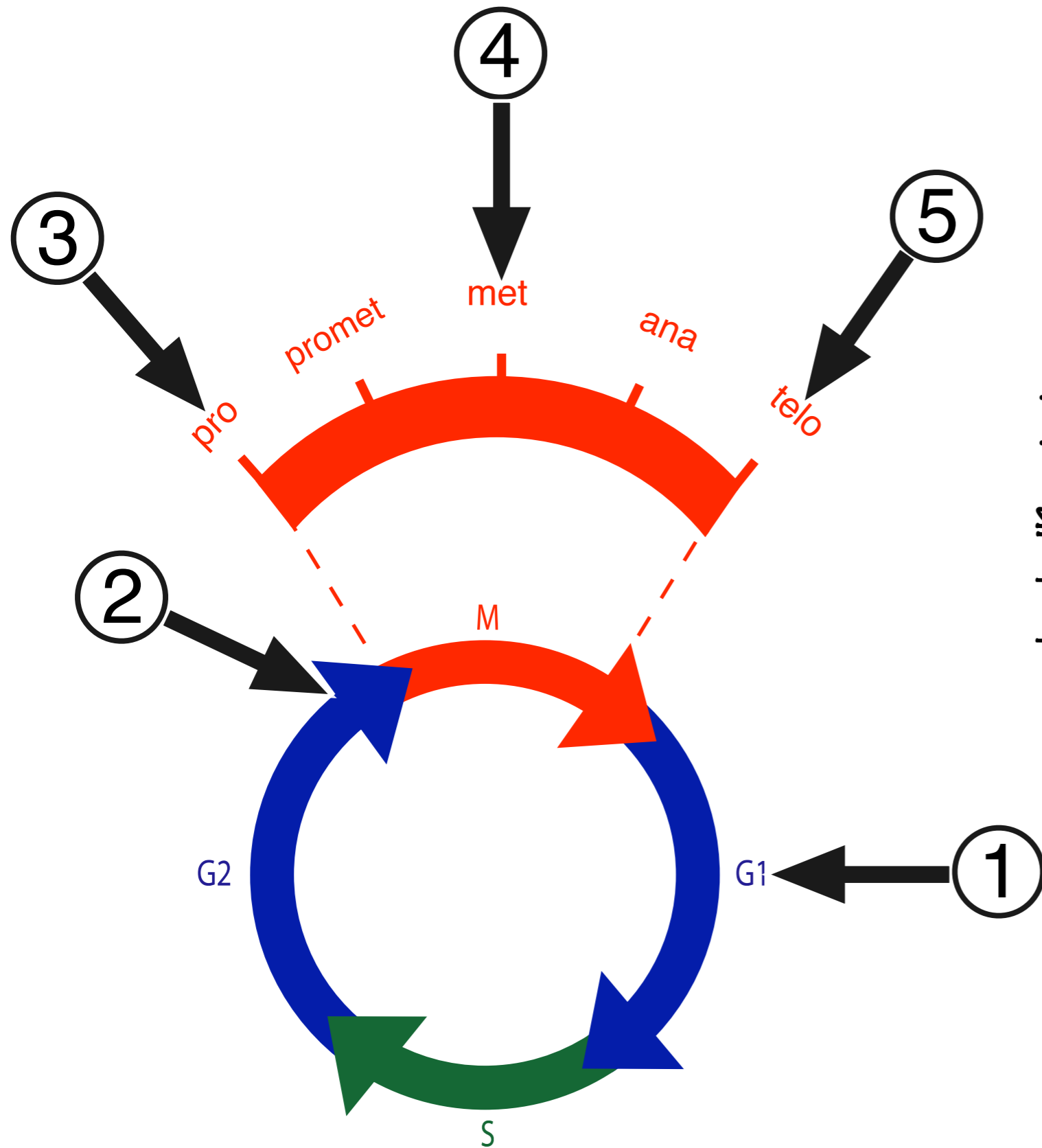


Which mitotic cyclins are involved in the aberrant M-phase-like/interphase-like oscillations that were observed in single cells during the live-cell imaging?

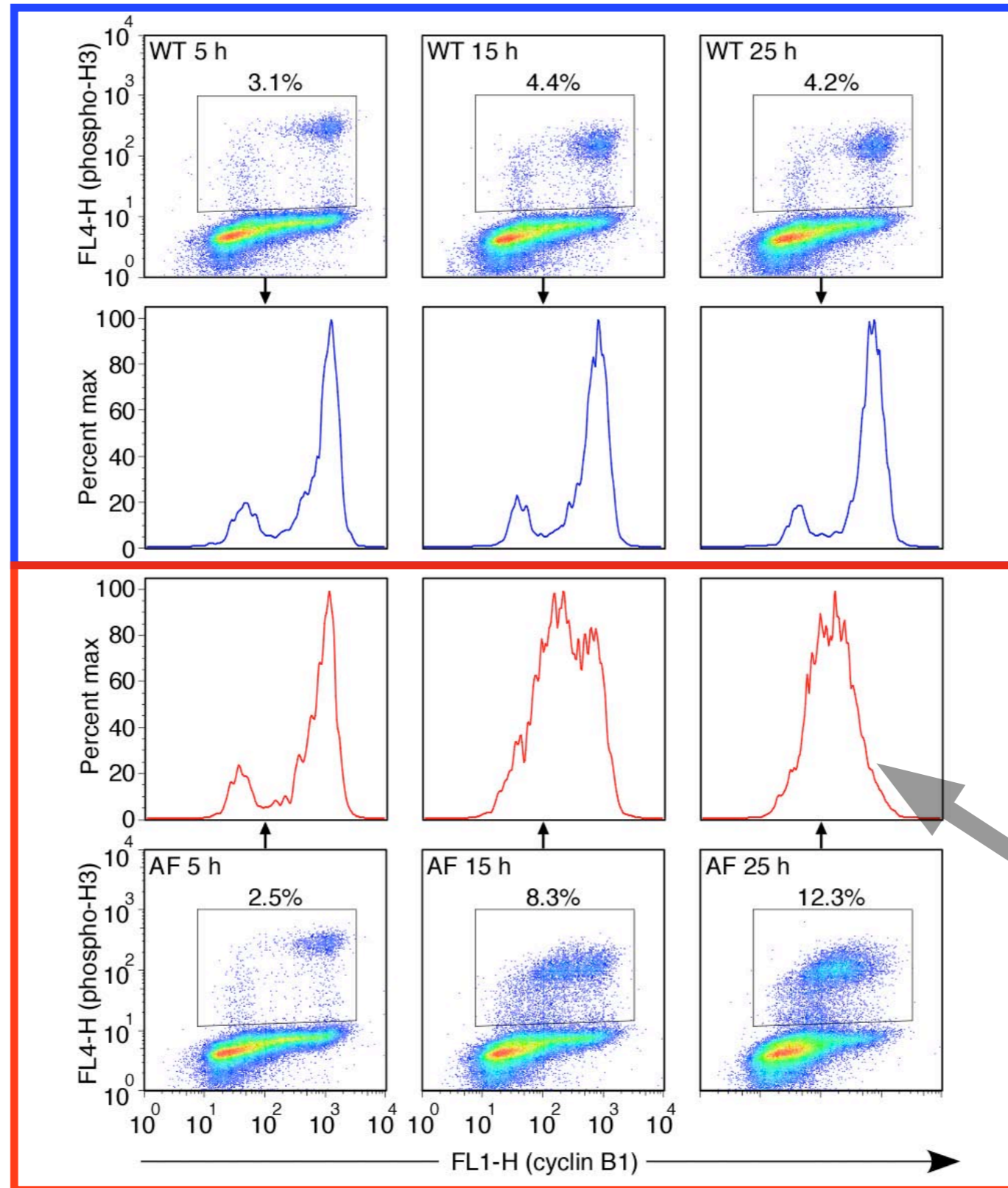
How can we dissect what is going on in large a population of cells **WITHOUT extreme perturbation (even asynchronous), but **WITH** molecular sensitivity?**

The BANE of CELL BIOLOGISTS: If not working in egg extracts, or “imaging” live individual cells with limited sets of sensors, we are working with cell populations.

Single-Cell Analysis of Cyclin B1 Content

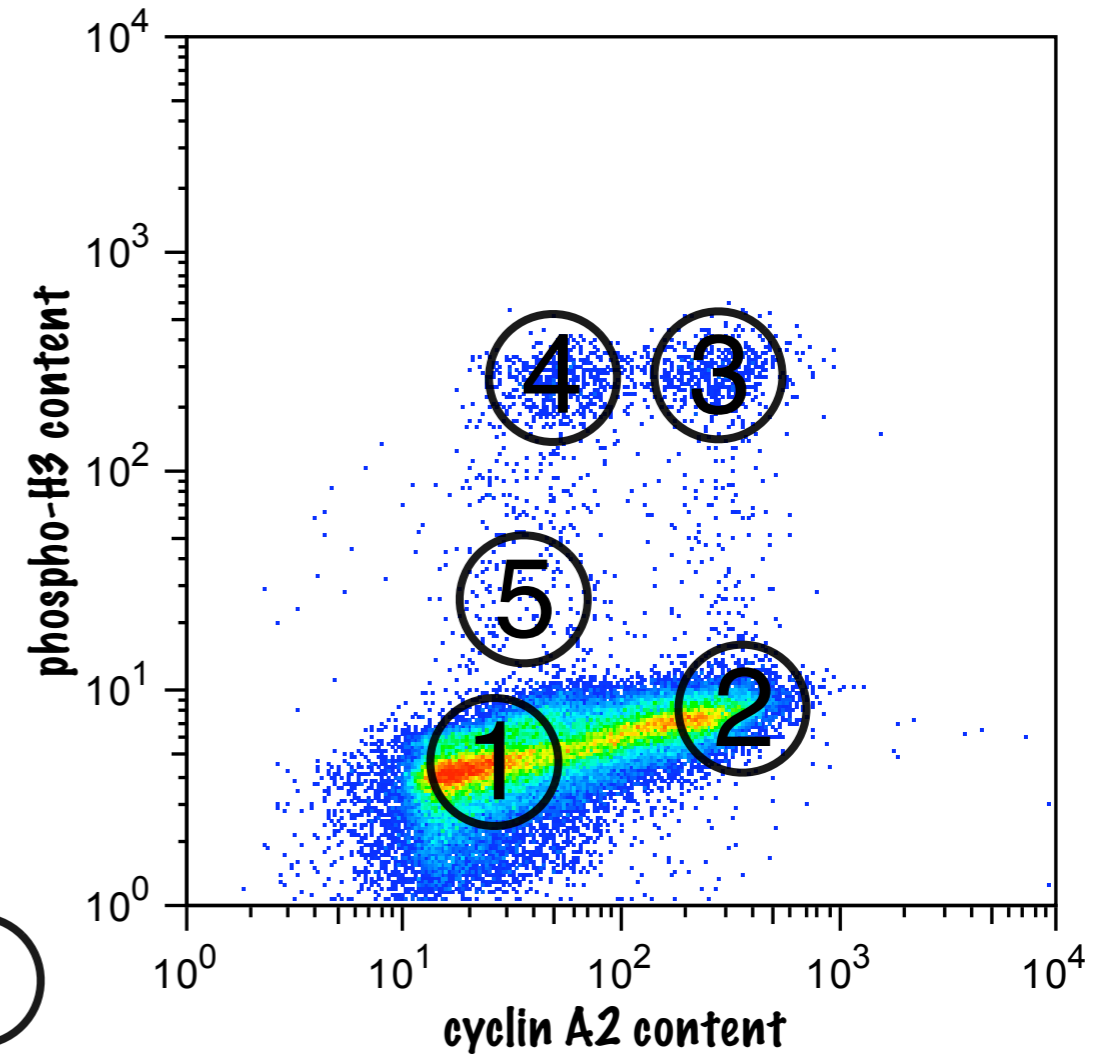
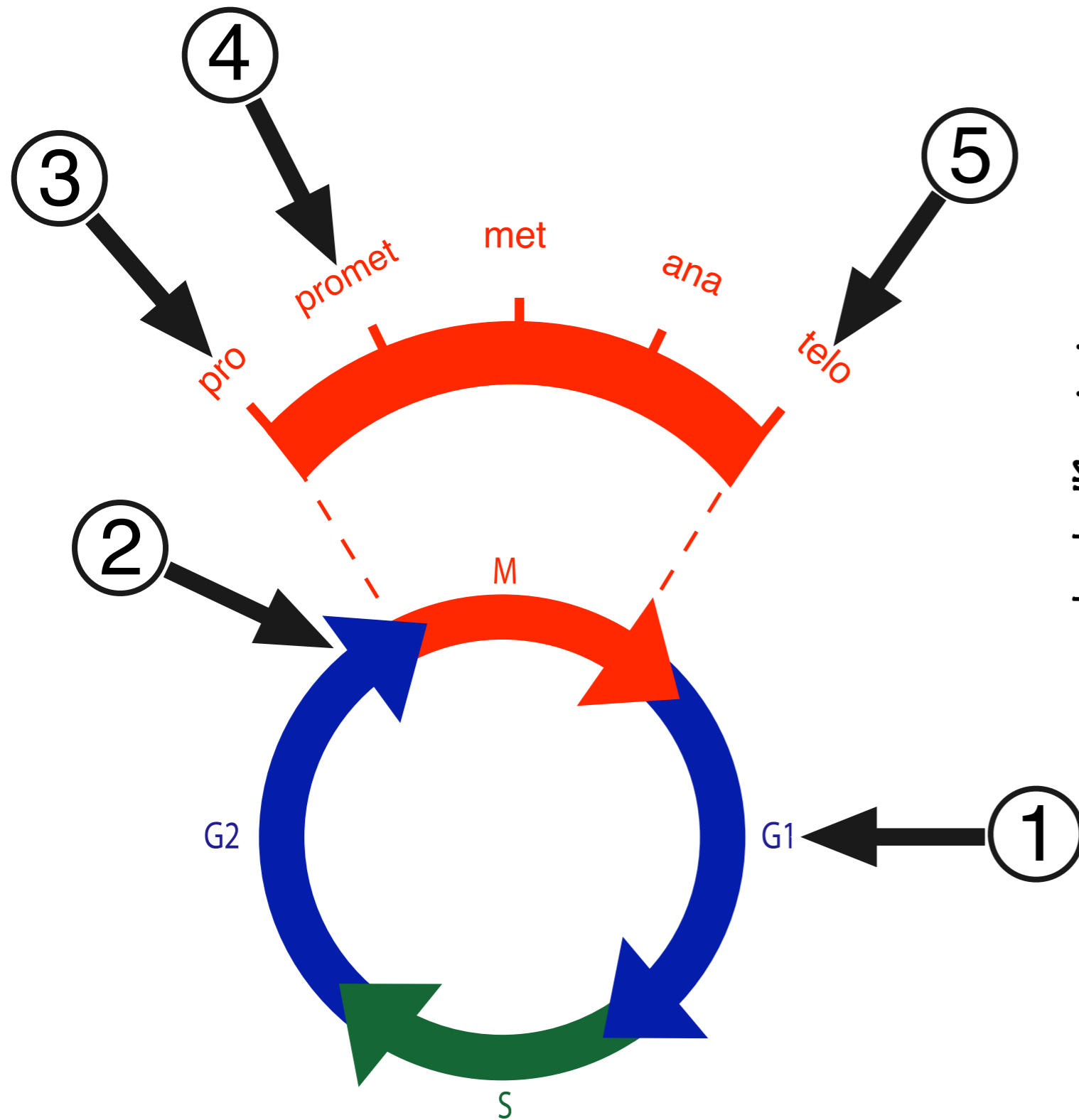


Single-Cell Analysis of Cyclin B1 Content

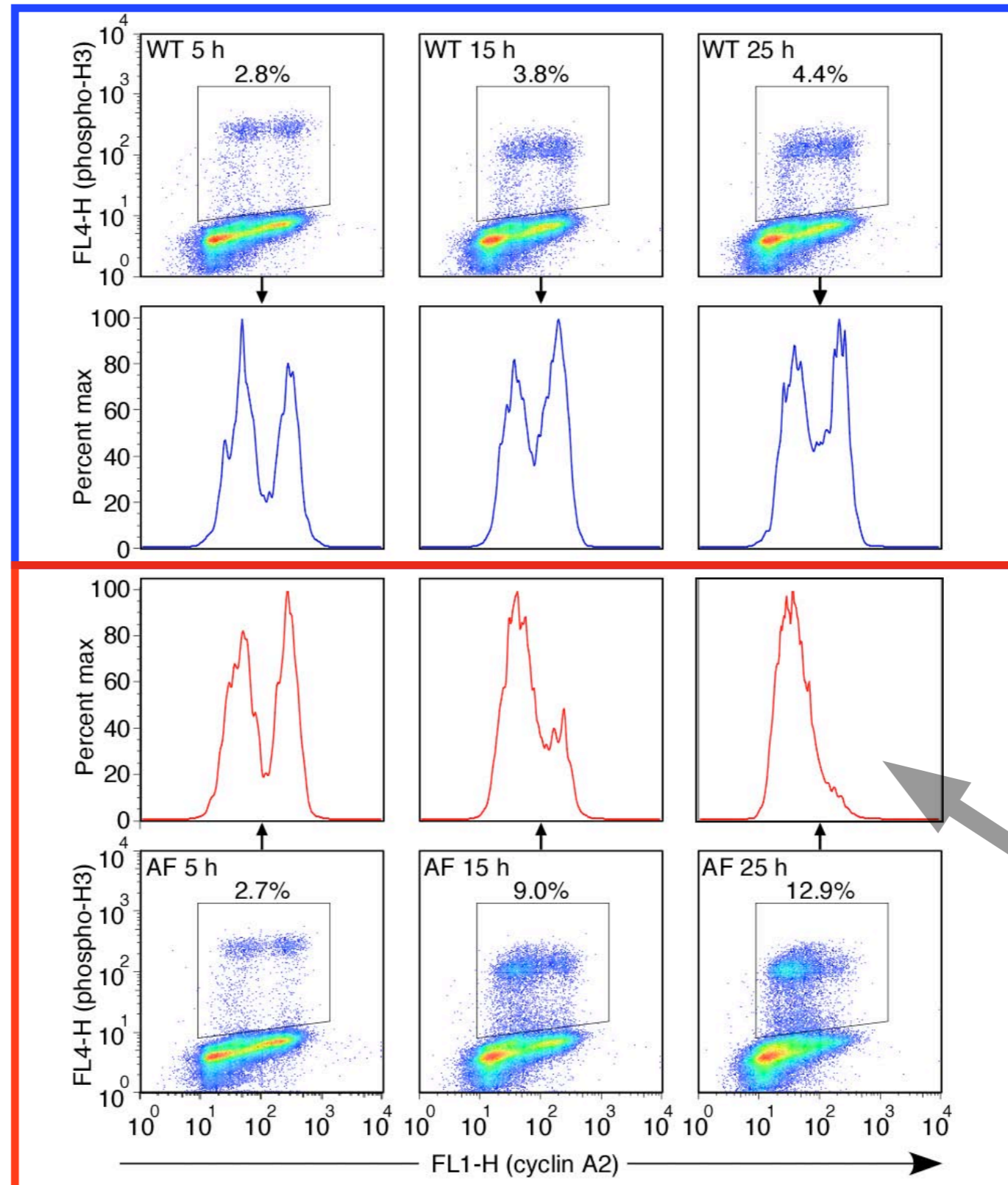


CDK1AF causes MORE cells to enter an M-phase-like state, though they are expressing INTERMEDIATE levels of cyclin B1

Single-Cell Analysis of Cyclin A2 Content



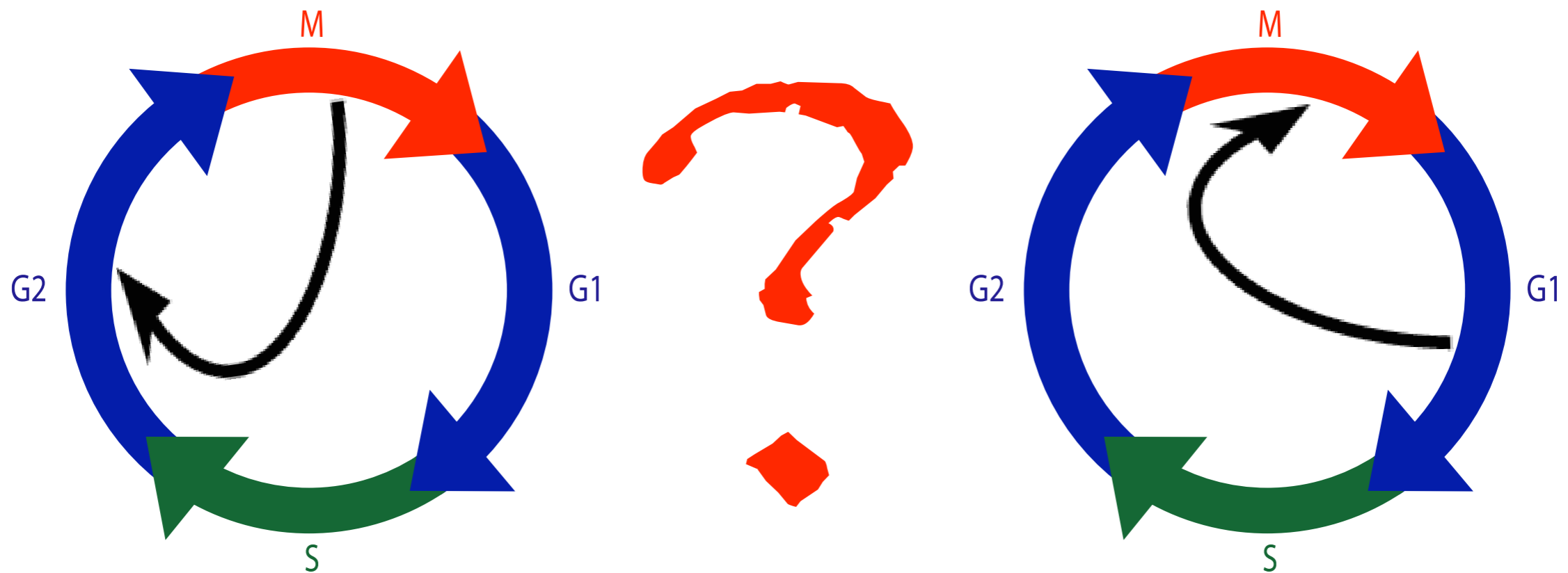
Single-Cell Analysis of Cyclin A2 Content



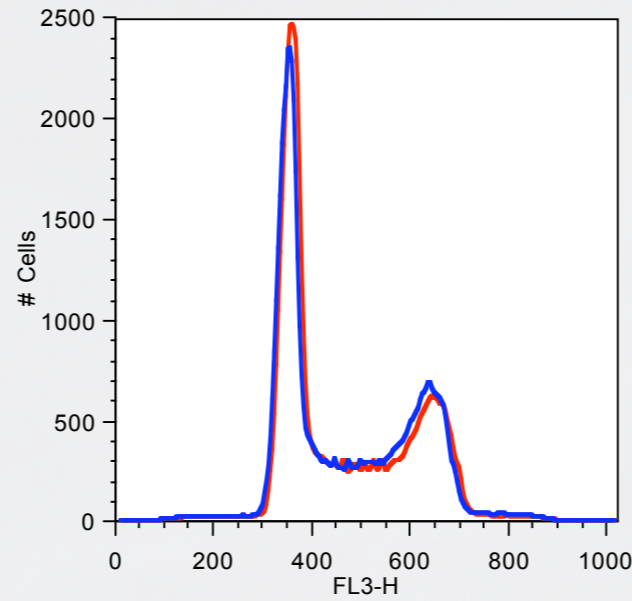
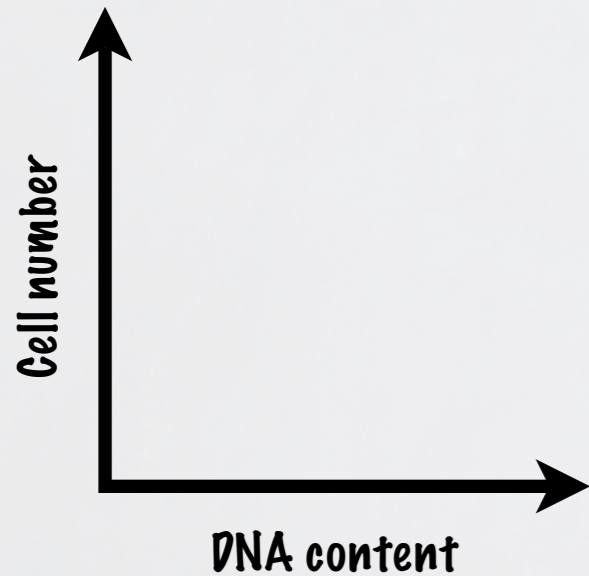
CDK1AF causes an accumulation of M-phase-like cells that are devoid of cyclin A2

With cyclin A2 absent from the perturbed, M-phase-like populations, cyclin B1 is the sole mitotic cyclin driving the abnormal CDK1AF-induced oscillations.

Are these cells experiencing a G2-M defect, or are they daughter cells with an M exit-G1 initiation defect.



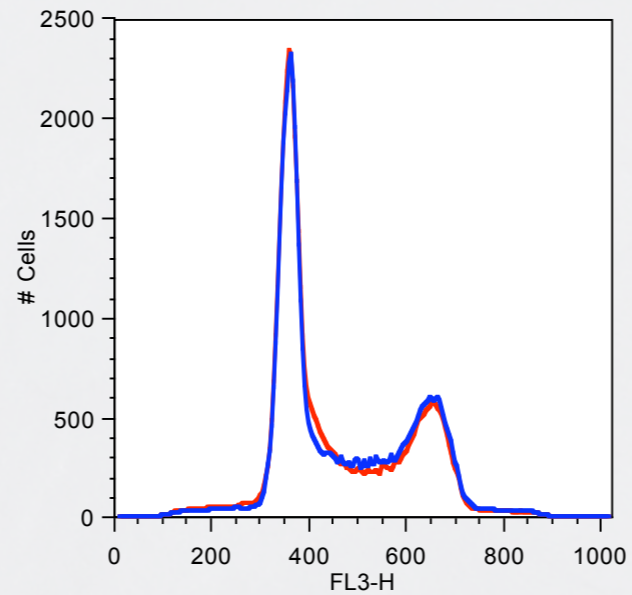
No, but they do begin to differentiate between repeated DNA content cells. Cells seem to be rapidly moving from G1 to S phase.



WT_5 DNA GATE 60000
AF_5 DNA GATE 60000

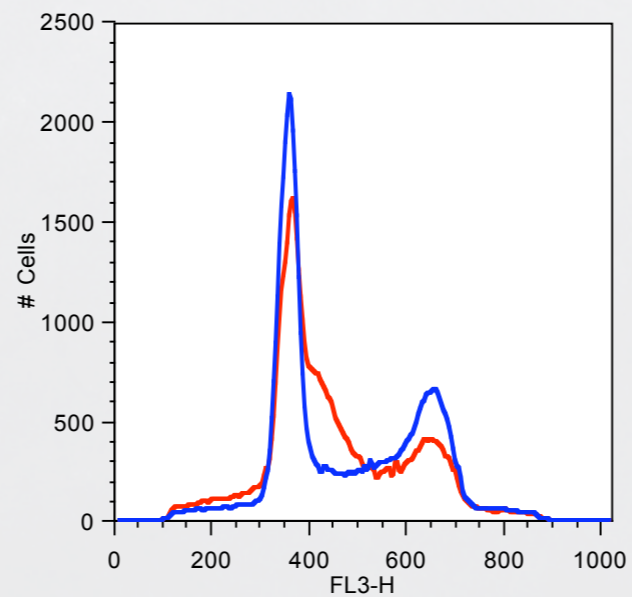
Post-transfection (h)

5



WT_15 DNA GATE 60000
AF_15 DNA GATE 60000

15



WT_25 DNA GATE 60000
AF_25 DNA GATE 60000

25

Live-Cell Analyses of G1-, S-, and M-phase Progression

**CDKI^{WT}-transfected
w/MBS & PCNA-YFP**

S end/G2 start (8:32:34 PM)

G2 duration ≈ 1.75 h

M (10:12:12 PM)

M duration ≈ 0.67 h

cytokinesis/G1 start (10:53:20 PM)

G1 duration ≈ 8.5 h

S start (7:25:51, 7:31:48 AM)

**CDKI^{AF}-transfected
w/MBS & PCNA-YFP**

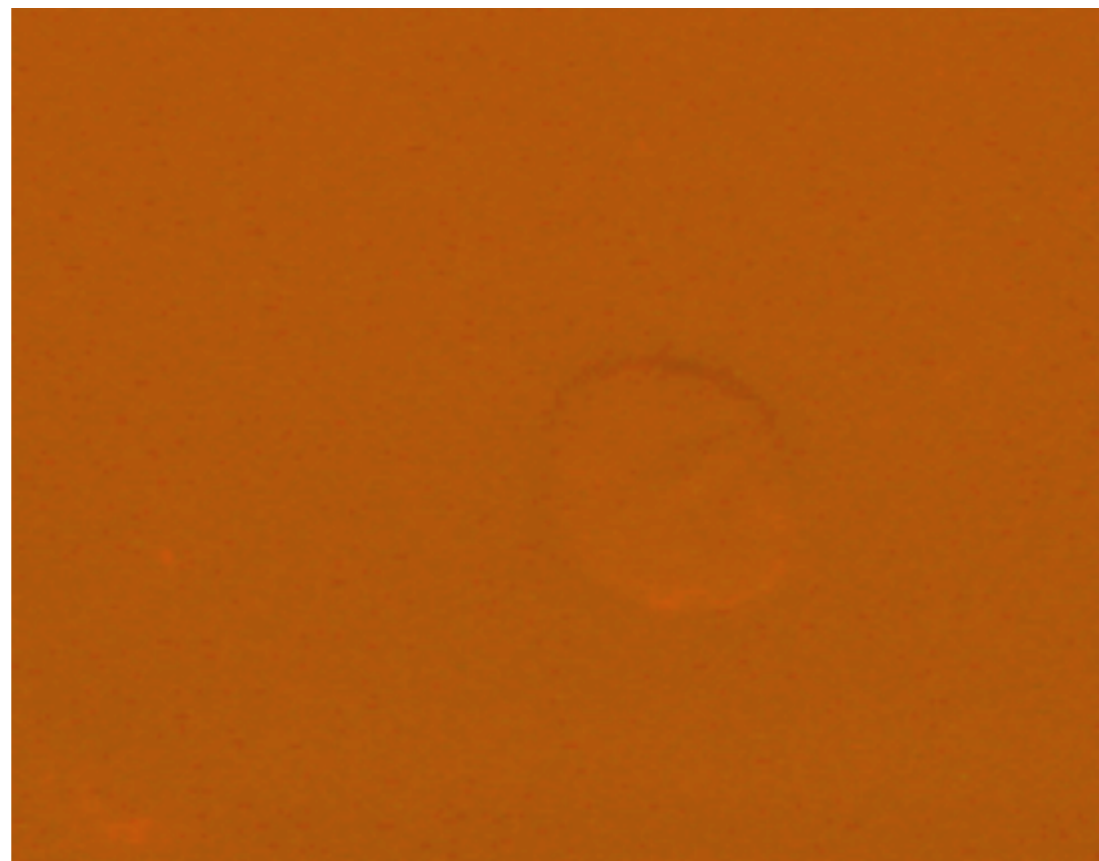
cytokinesis/G1 start (11:02:07 PM)

G1 duration ≈ 3.5 h

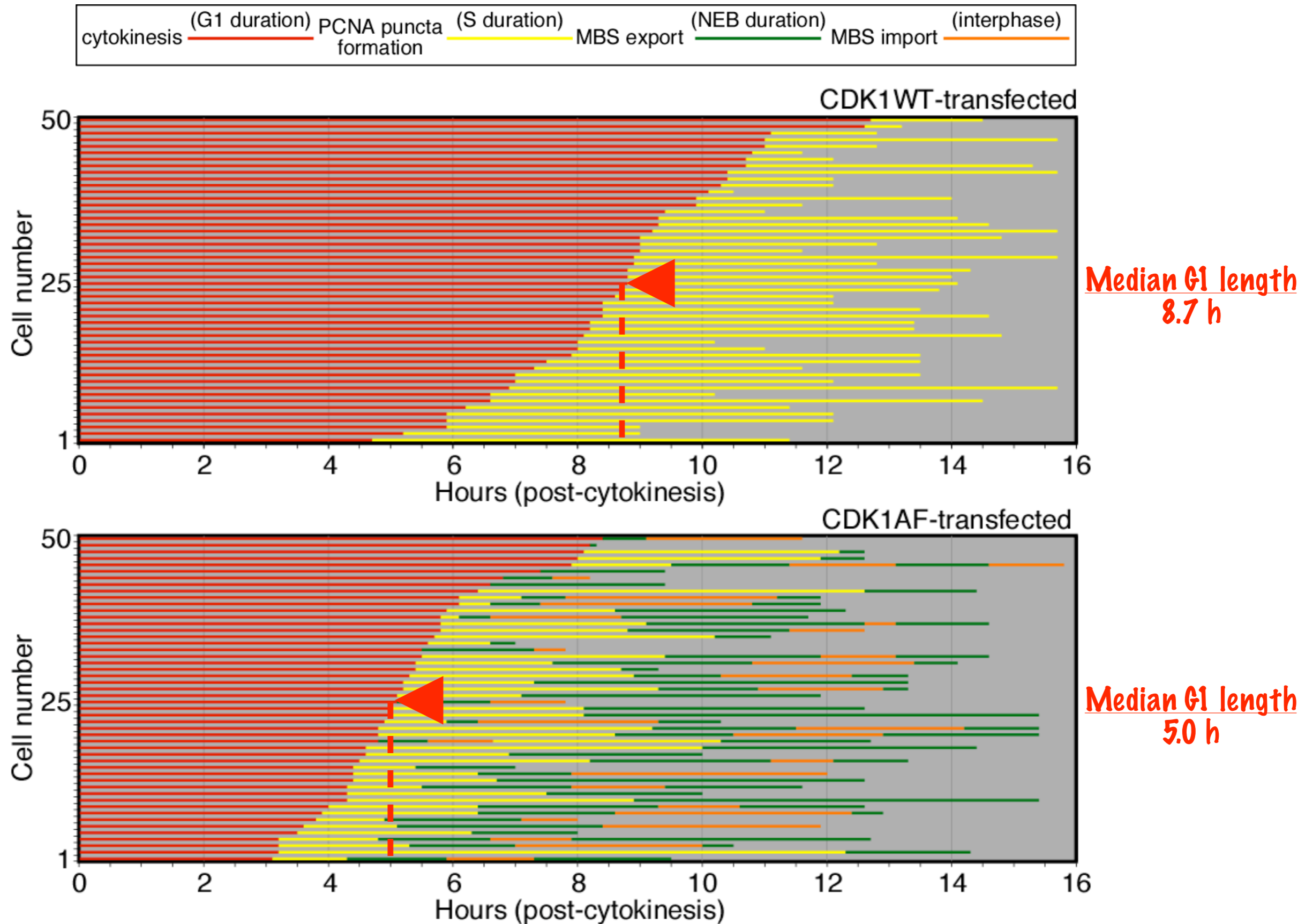
S start (2:30:31 AM)

S duration ≈ 2.0 h

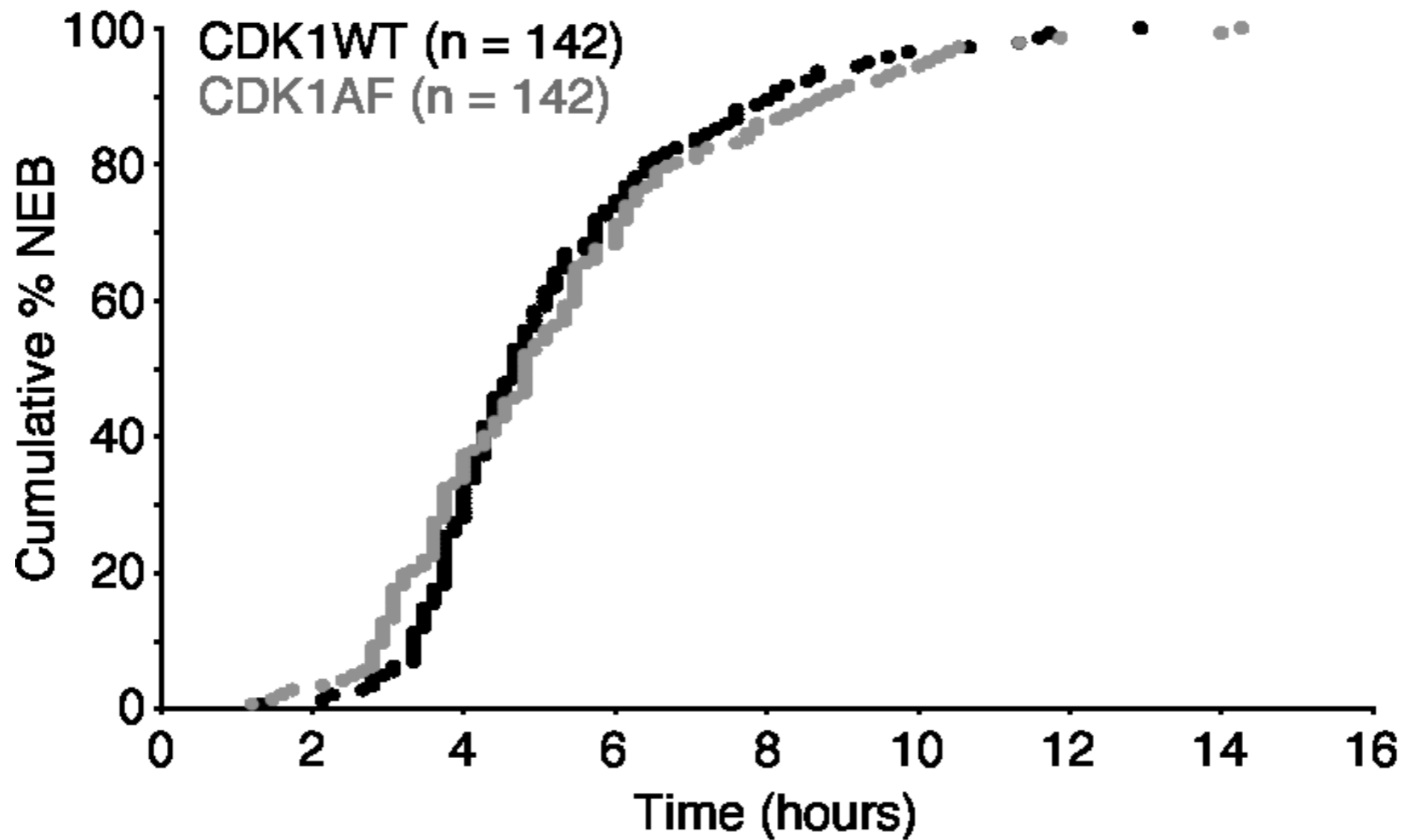
NEB (3:57:57, 4:27:42 PM)



Live-Cell Analyses of G1-, S-, and M-phase Progression



Does CDK1AF cause early mitosis in a cell population released from thymidine?



Cyclin A2 Regulates Nuclear-Envelope Breakdown and the Nuclear Accumulation of Cyclin B1

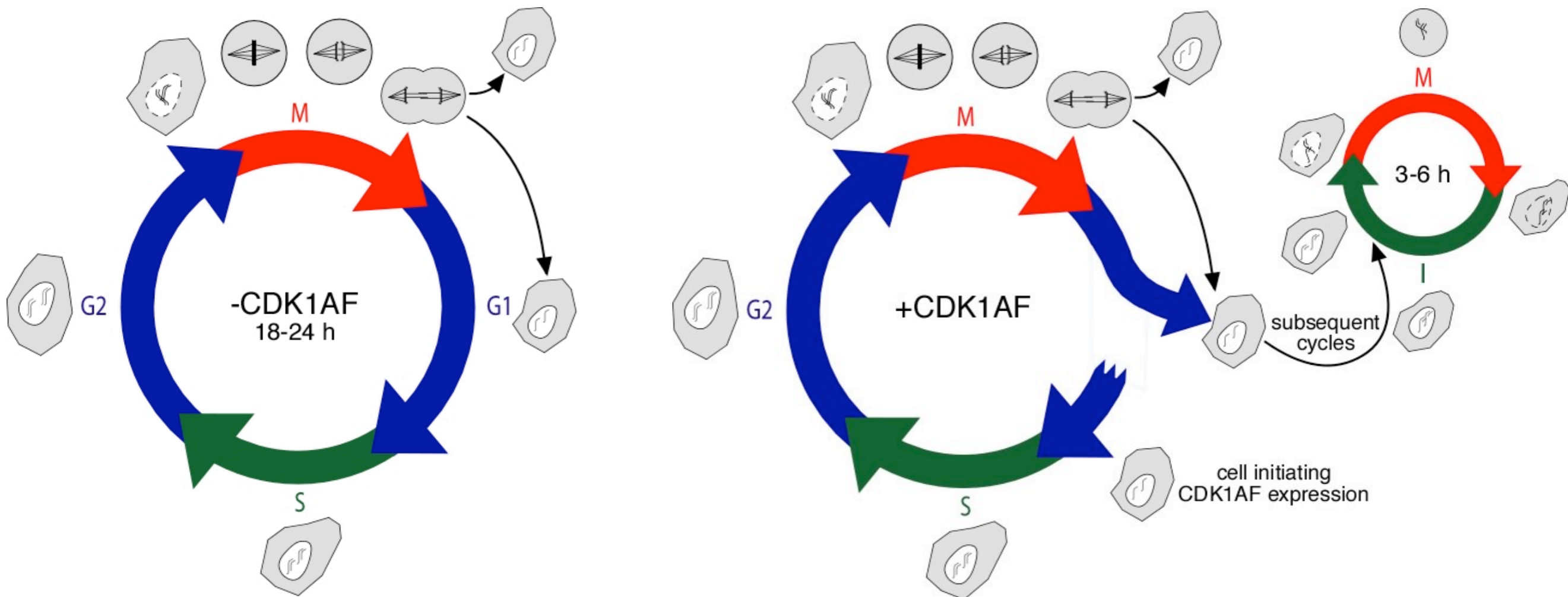
Delquin Gong,^{1,5,*} Joseph R. Pomerening,¹ Jason W. Myers,¹ Christer Gustavsson,² Joshua T. Jones,² Angela T. Hahn,^{1,3} Tobias Meyer,² and James E. Ferrell, Jr.^{1,4,*}

¹Department of Chemical and Systems Biology, CCSR, MC 5174

²Department of Chemical and Systems Biology,

pathways [8]. We synchronized cells by double thymidine block and transfected them with the cyclin d-siRNAs singly or in combination during the first thymidine wash-out. 24 hr after transfection, cells were released from the second thymidine block. We assessed cyclin levels by immunoblotting (Figure 1A) and followed mitotic progression by automated epifluorescence microscopy, and quantified the duration of mitotic biosen-

- 1) Early mitosis? X
- 2) Increased frequency? ✓
- 3) Some damping? ✓
- 4) Cdc20 activity maintained? ✓
- 5) Cdh1 activity period reduced? ✓



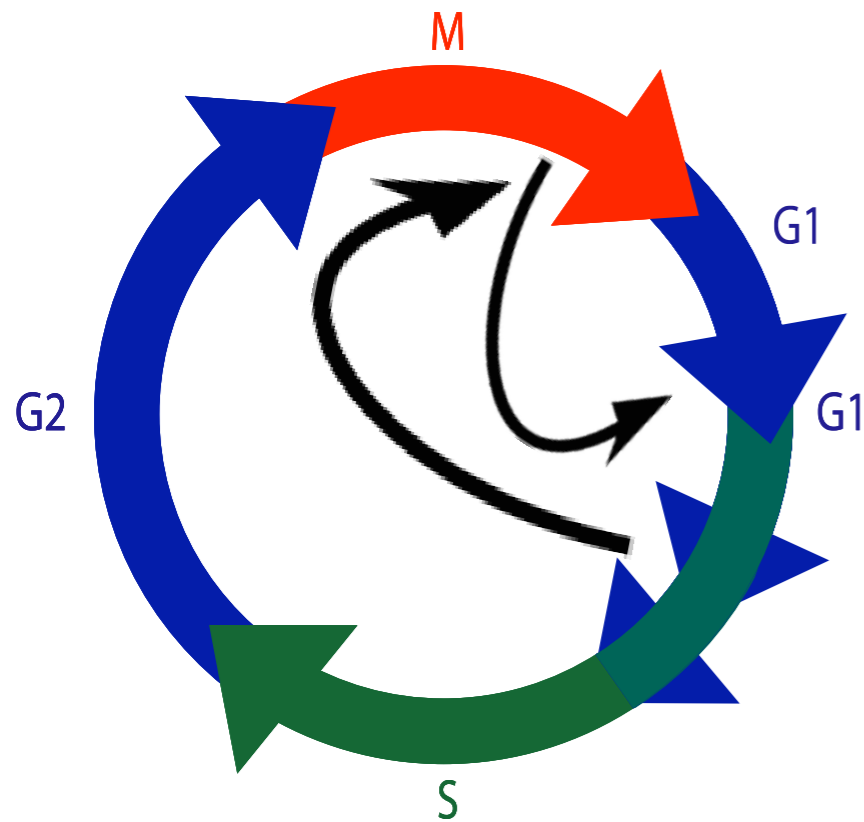
Summary-Part III

Bypassing the positive feedback loops responsible for CDK1 activation with CDK1AF causes multiple cellular defects within a SOMATIC CELL type, including:

exchanges of M-phase-like and interphase-like periods with intermediate M-phase phosphorylation states;

periods of cyclin B stability and instability, when there should only be ONE per cell cycle;

a rapid oscillatory behavior that emerges from cells that divide, and either enter an M-phase-like state from G1 (16% of the time), or cells shorten G1 (by approx. 4 h), induce a precocious S phase, then stimulate an M-phase-like state (84% of the time).



Positive feedback in CDK1 activation provides discrete M- and S-phases in the embryonic context (to oscillate properly).

Positive feedback in CDK1 activation ensures a sufficient G1 period, and thus, a properly delayed S phase onset in the somatic context (no oscillation).



Thank You.

Ferrell Lab

-Prof. Jim Ferrell

-Sun Young Kim

-Jason Myers

-Delquin Gong

Meyer Lab

-Prof. Tobias Meyer

-Josh Jones

-Angie Hahn

Tom Wehrman

Mark Hammer

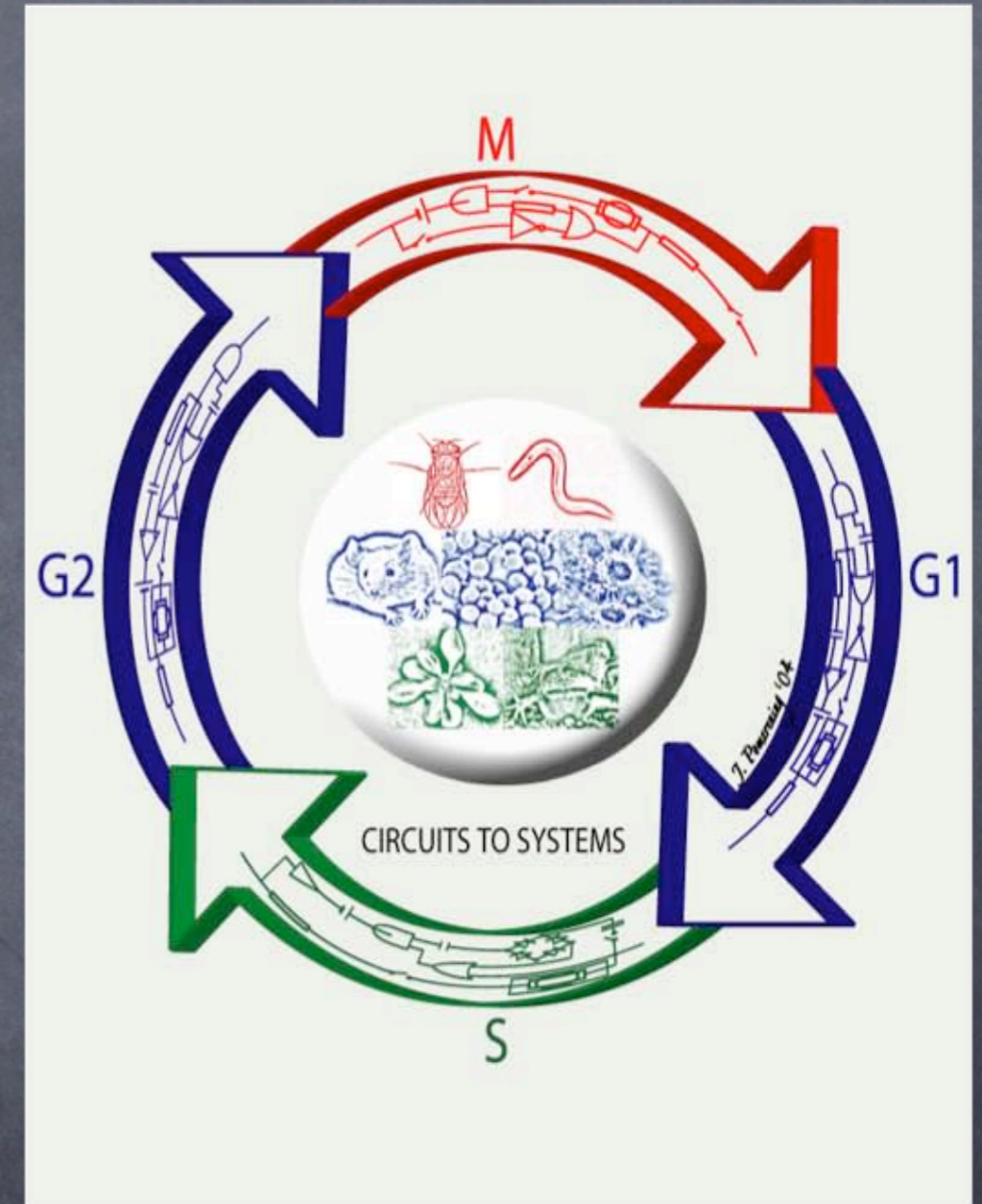
Stanford HTBC Facility

M. Cristina Cardoso

Max Delbrück, Berlin

Quantitative Chemical Biology Program

Quantitative Chemical Biology



Kavli Institute for Theoretical Physics - University of California-Santa Barbara