

Propulsion with stiff polymers

Manoel Manghi (Toulouse)

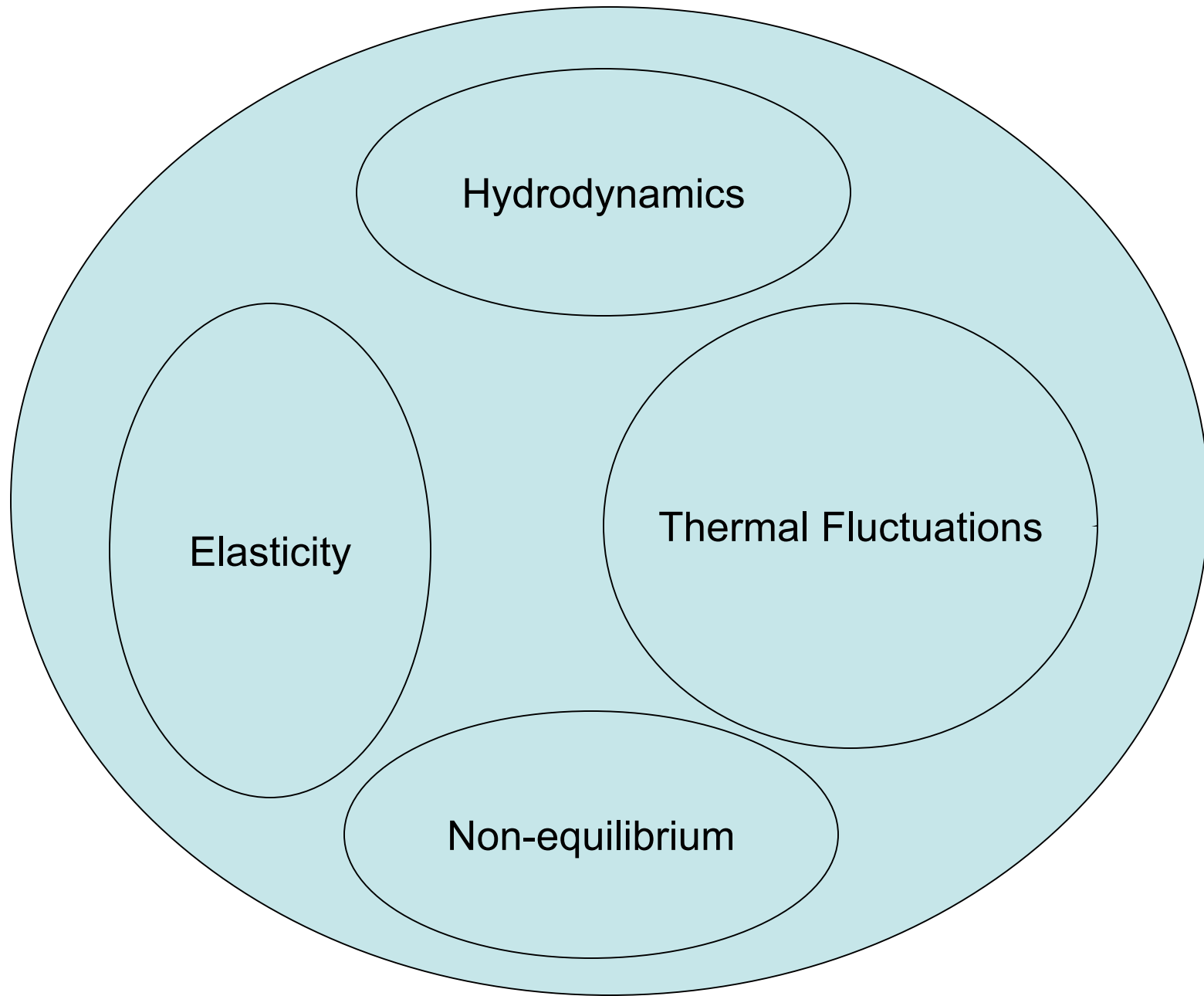
PRL 96, 068101 (2006)

Yong-Woon Kim (UCSB)

PRL in press (2006)

X. Schlagberger, Roland Netz (TUM)

- 1) sedimentation of polymers
- 2) electrophoresis of polymers
- 3) polymers in shear flow (unfolding of proteins in blood flow)
- 4) polymers at surfaces in shear or electric fields
(glycocalix deformation under shear)
- 5) driven stiff polymers -> propulsion



Hydrodynamics

Elasticity

Thermal Fluctuations

Non-equilibrium

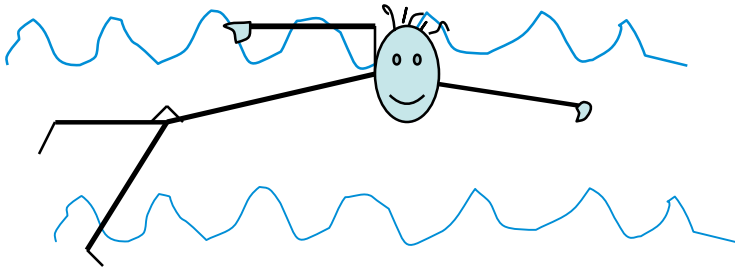
Hydrodynamics at low Reynolds numbers

Stationary Navier-Stokes equation $\eta \Delta \vec{v}(\vec{r}) - \nabla p = \rho(\vec{v} \cdot \nabla) \vec{v}$

If the Reynolds number $Re^{tr} = \frac{\rho}{\eta} l v \ll 1$, $Re^{rot} = \frac{\rho}{\eta} r^2 \omega \ll 1$,

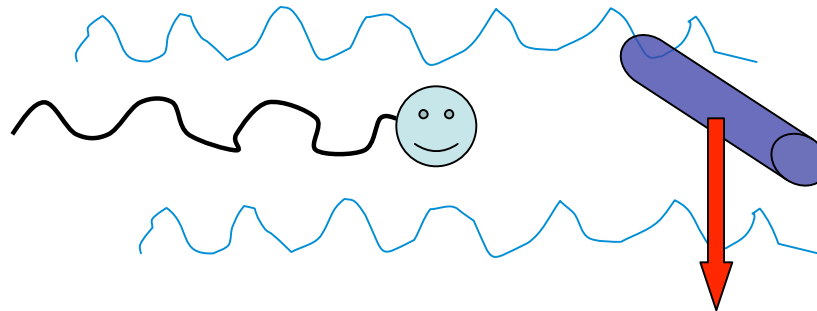
one obtains the **creeping flow equation**.

human



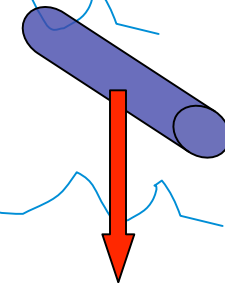
$$\left. \begin{array}{l} \text{H}_2\text{O}: \eta = 0.001 \text{ Pa s}; \\ \rho = 1000 \text{ kg/m}^3 \\ v = 1 \text{ m/s} \\ l = 1 \text{ m} \end{array} \right\} Re = 10^6$$

bacterium

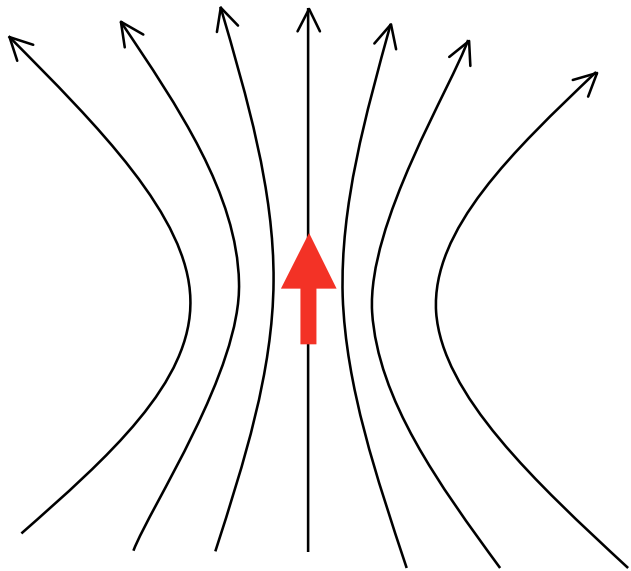


$$\begin{array}{l} v = 10^{-5} \text{ m/s} \\ l = 1 \mu \\ \rightarrow Re = 10^{-5} \end{array}$$

sinking cylinder



$$\begin{array}{l} v \sim 10^{-7} \text{ m/s} \\ l = 1 \mu \\ \rightarrow Re = 10^{-7} \end{array}$$



flow-field due to point-force at origin:

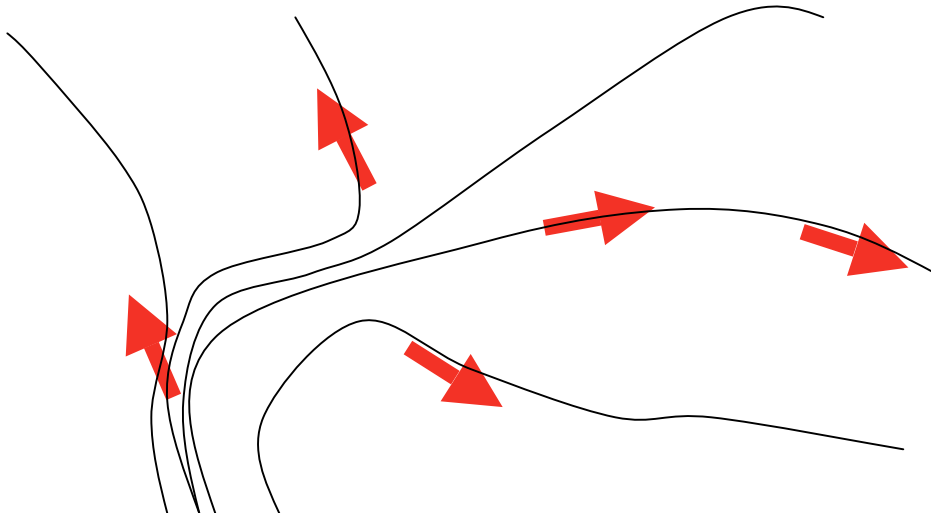
$$u^\alpha(r) = H^{\alpha\beta}(r) f^\beta \quad \alpha, \beta = 1, 2, 3$$

$$H^{\alpha\beta}(r) = \frac{1}{8\pi\eta r} \left[\delta_{\alpha\beta} + \hat{r}^\alpha \hat{r}^\beta \right]$$

(Oseen-Tensor)

for many particles the superposition principle is valid:

$$u^\alpha(r) = \sum_i H^{\alpha\beta}(r - r_i) f_i^\beta$$



invert to get forces for prescribed solvent velocity distribution !!

Next: add thermal noise

Theoretical Framework: Position Langevin Equation

Velocity of
i-th particle:

$$\cancel{m\ddot{r}_j(t)} \vec{\mu}_{ij} + \dot{r}_i(t) = \vec{\mu}_{ij} f_j(t) + \xi_i(t)$$

deterministic force $f_j(t) = -\partial U(t) / \partial r_j(t) + E$

Random force $\langle \xi_i(t) \xi_j(t') \rangle = 6 \vec{\mu}_{ij} k_B T \delta(t - t')$

Mobility matrix: $\vec{\mu}_{ij} = D_{ij} / k_B T = \mu_0 \delta_{ij} + \vec{H}(r_i, r_j)$

self mobility: $\mu_0 = (6\pi R \eta)^{-1}$ hydrodyn. interact.

equivalent to Smoluchowski equation for particle distribut. $W(r_j, t)$:

$$\frac{\partial W}{\partial t} = \sum_{i,j} \frac{\partial}{\partial r_i} \left[D_{ij} \frac{\partial W}{\partial r_j} - \mu_{ij} f_j W \right] \quad \text{with solution: } W \cong e^{-U/k_B T}$$

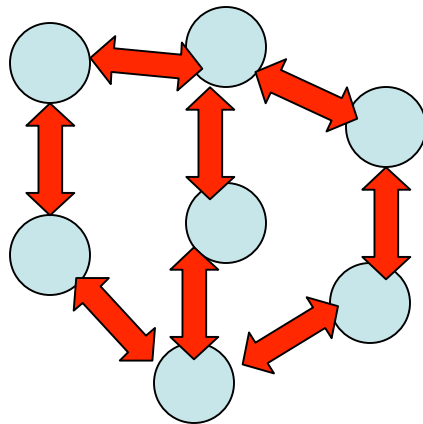
straightforward way to satisfy no-slip in multi-particle system

$$u_j^\alpha(r_j) = \sum_i H^{\alpha\beta}(r_j - r_i) f_i^\beta = \mu_0 f_j^\alpha + v_\infty(r_j) = v(r_j)$$

velocity of j-th
particle

solvent-velocity
due to other particles

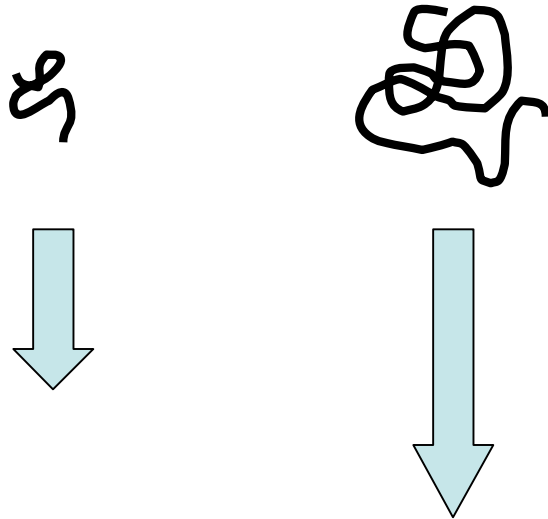
solvent-velocity
at particle position



cohesive/elastic forces in objects
automatically lead to solvent flow
stagnation

a few examples

Separation by sedimentation in the ultracentrifuge



G: force per monomer

N: monomer number

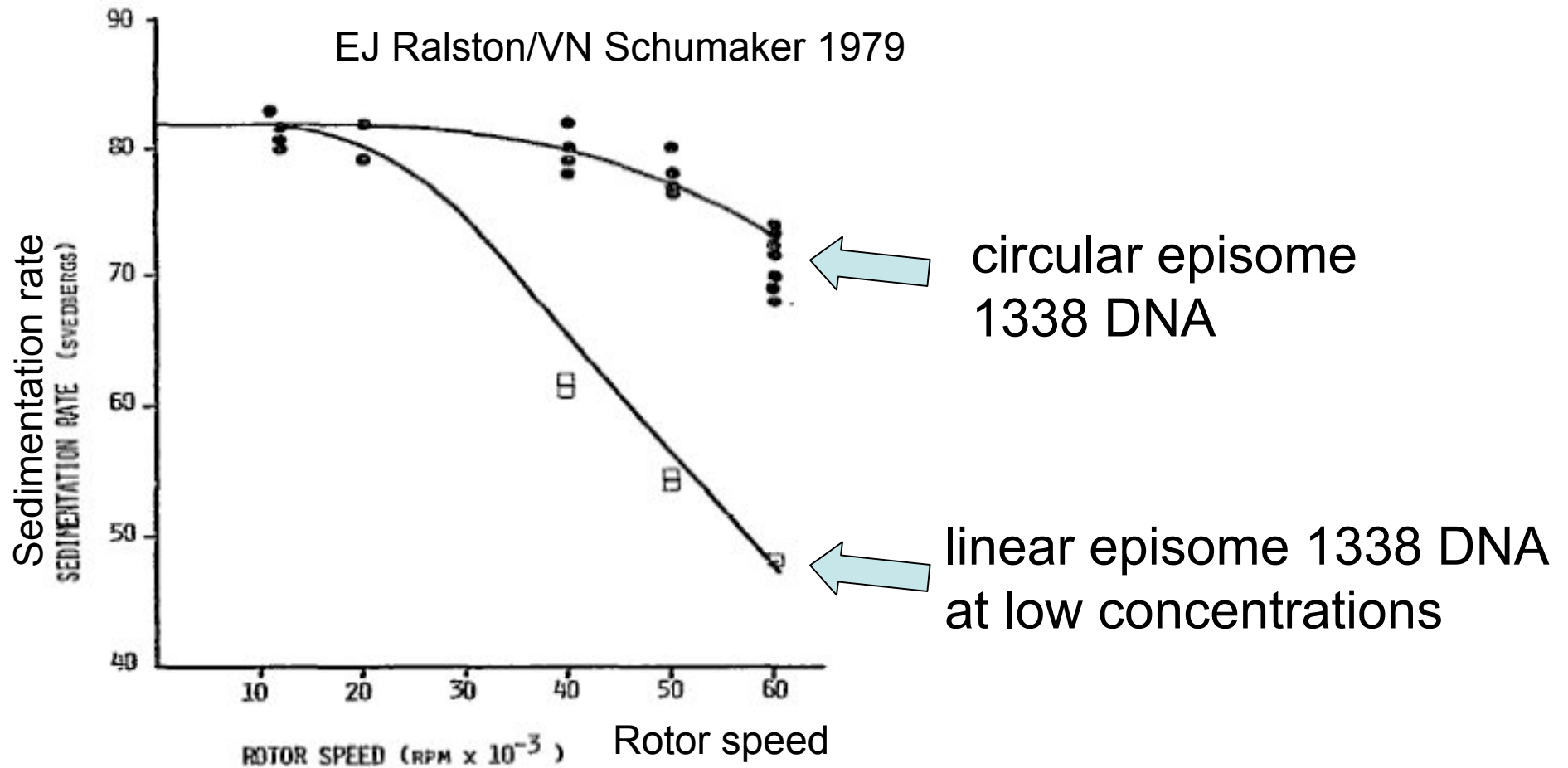
velocity $v = GN\mu$

mobility $\mu = 1/6\pi\eta R = 1/6\pi\eta N^{\nu}$

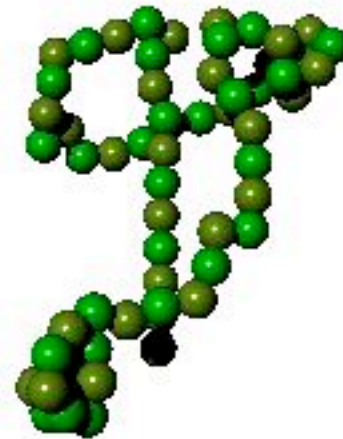
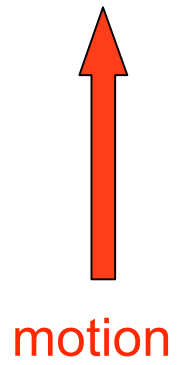
--> velocity $v \approx G N^{1-\nu}$

Why gel-electrophoresis is used for separating DNA (and not the ultracentrifuge)

- sedimentation rate of polymers goes down at high rotor speeds
- crossover is polymer-length dependent!



Crumpling of Flexible Chains (Xaver Schlagberger)



hydrodynamic simulations of sedimenting polymers

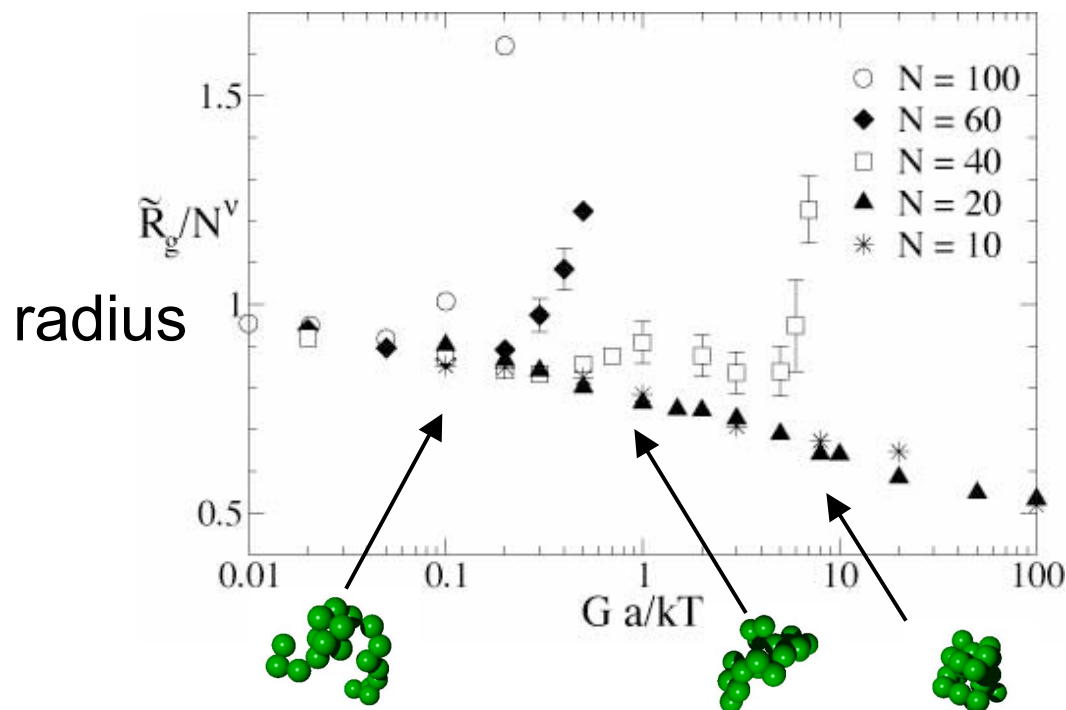
Xaver Schlagberger

hydrodynamic drag \rightarrow internal recirculation with velocity $v \approx GN/\eta R$

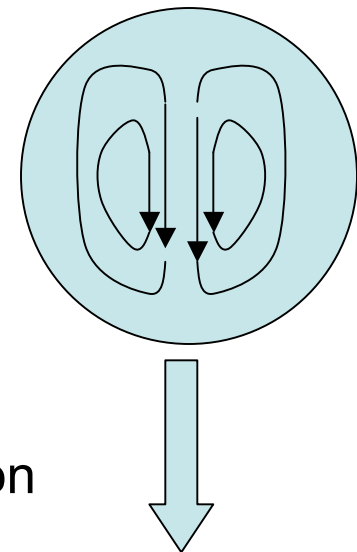
\rightarrow recirculation time scale $\tau_{flow} \approx R/v \approx \eta R^2 / GN$

compare with coil relaxation time $\tau_R \approx \eta R^3 / k_B T$

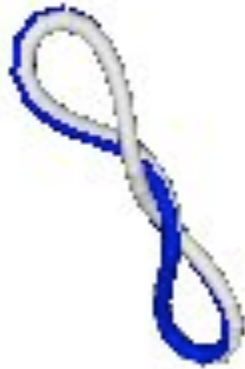
„scrambled/collapsed coil“ for $\tau_R > \tau_{flow}$ or $Ga/k_B T > N^{-2/3}$



sedimentation force



Sedimentation of twisted ring polymer
(conserved linking number) Hirofumi Wada

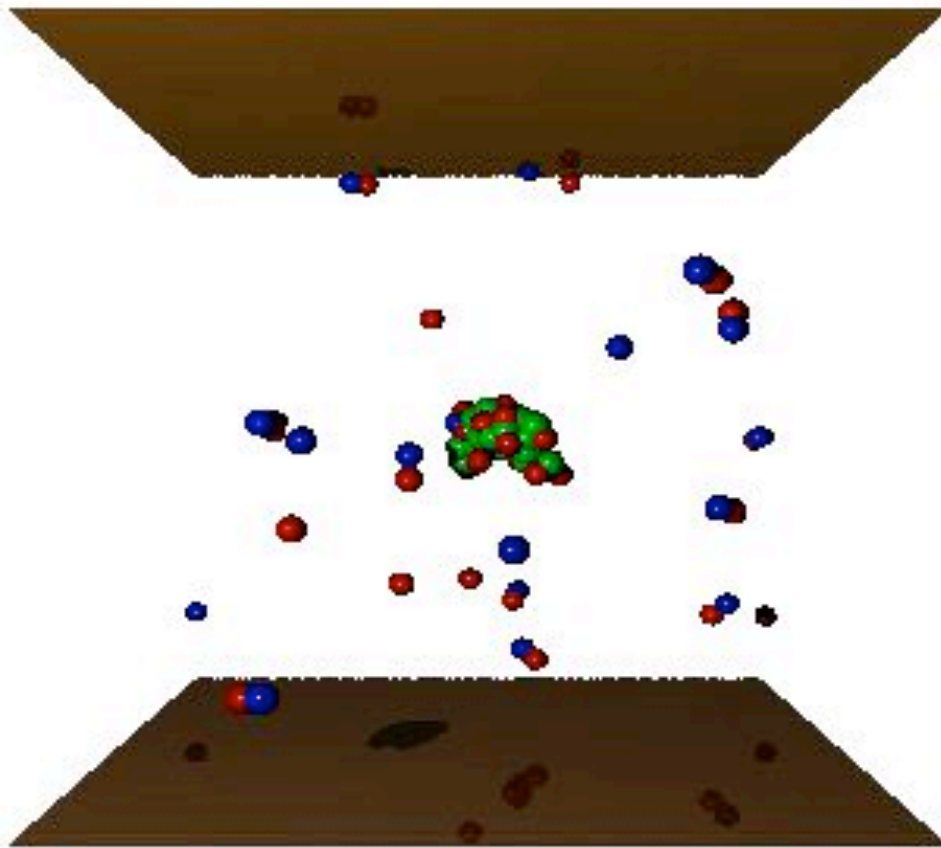


Dynamics of plectoneme formation in ring polymer

Hirofumi Wada



Electrophoresis of polyelectrolytes



20 monomers
40 counterions
20 coions



**electric field,
camera moves
with polymer !**

**minimal image BC
strong coupling $\Xi = 20$**

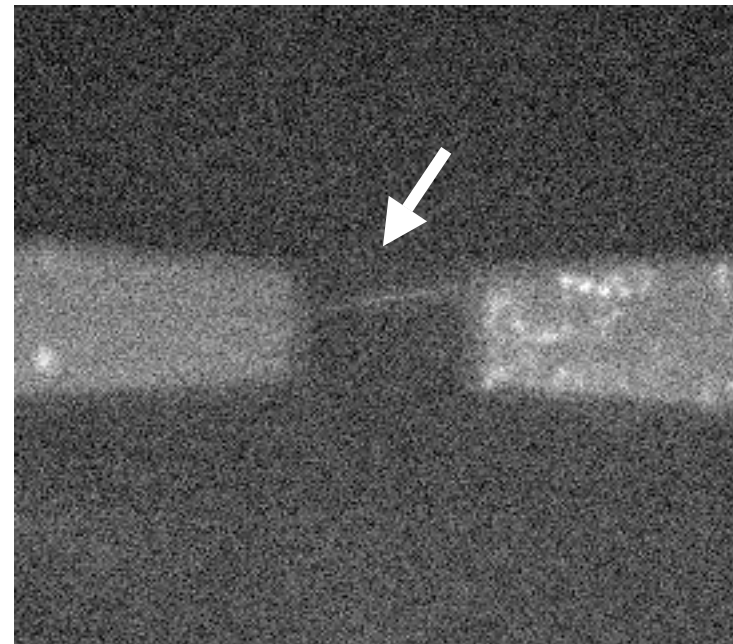
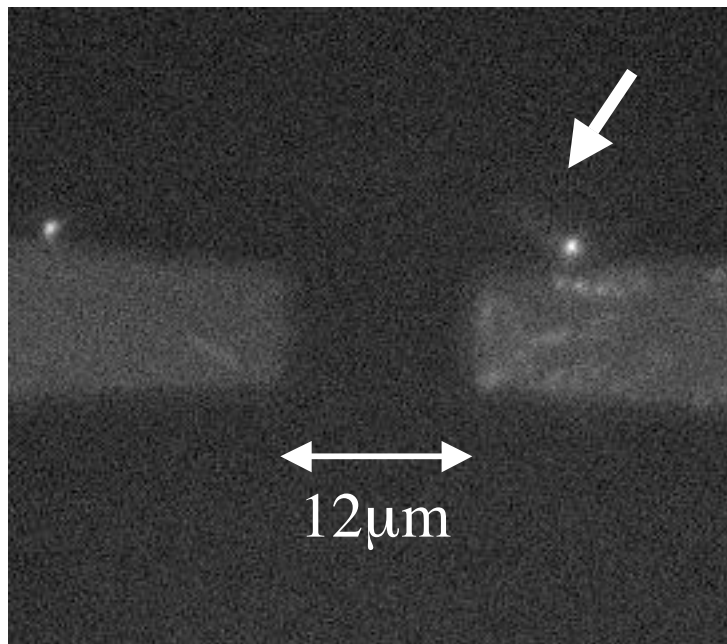
experimental observation of DNA stretching in fields

Beyer, Simmel (LMU)

4 Volts -> $E = 3 \times 10^5$ V/m

DNA length $17\mu\text{m}$

10mM HEPES, no added salt



DNA-molecules are typically stretched in free-solution electrophoresis experiments -> no length separation possible

Protein denaturation in shear flows

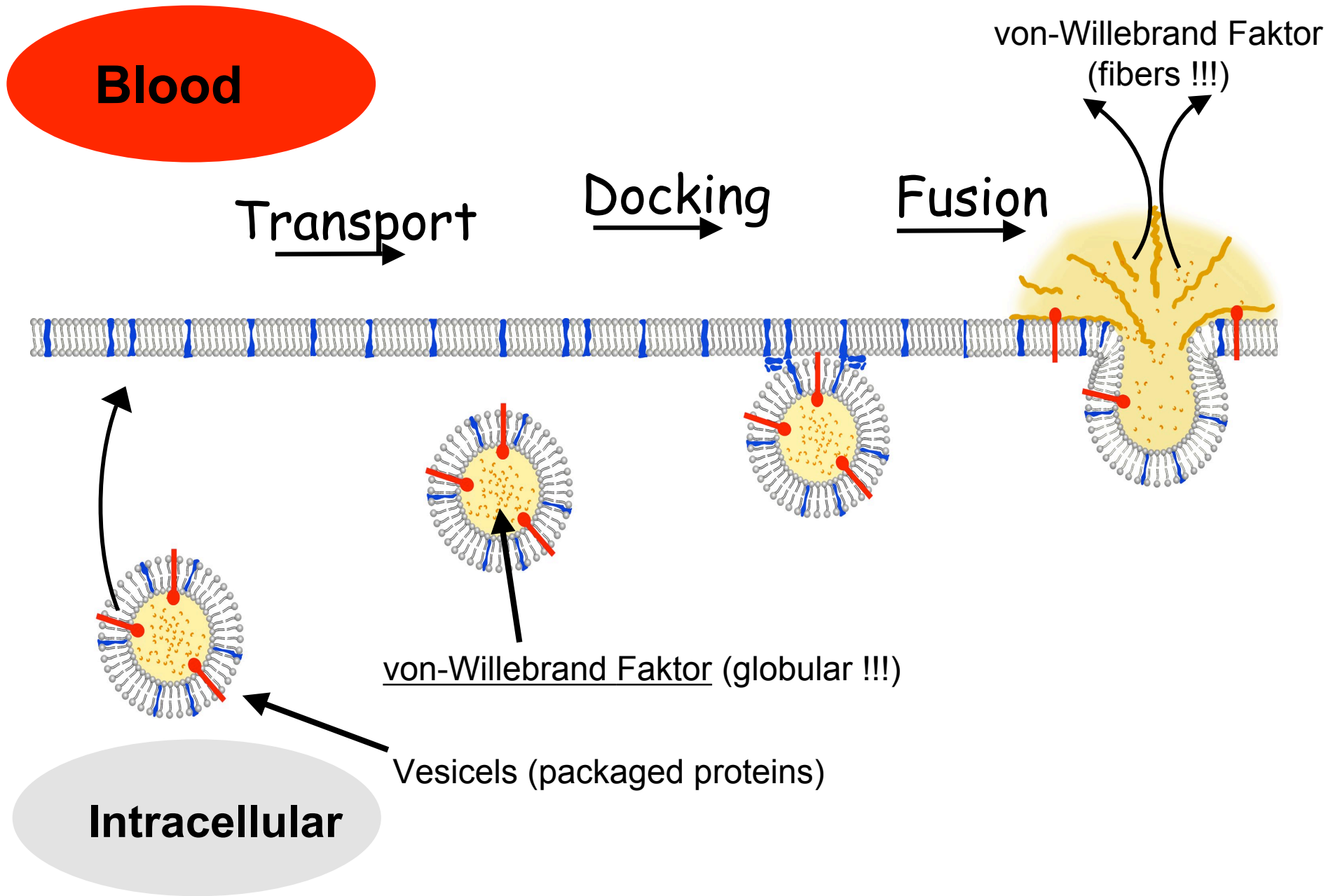
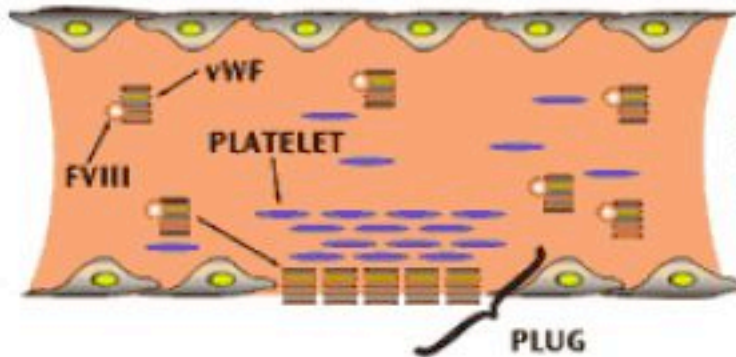


Figure 1. Normal Broken Blood Vessel



First, vWF proteins from the blood line up along the broken vessel wall and attract "sticky" platelets to form a plug.

Then the platelets attract strands of fibrin to strengthen the plug and form a clot. The clot helps stop the bleeding.

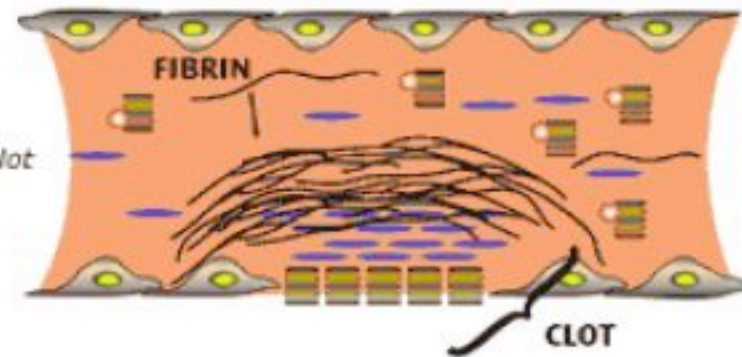
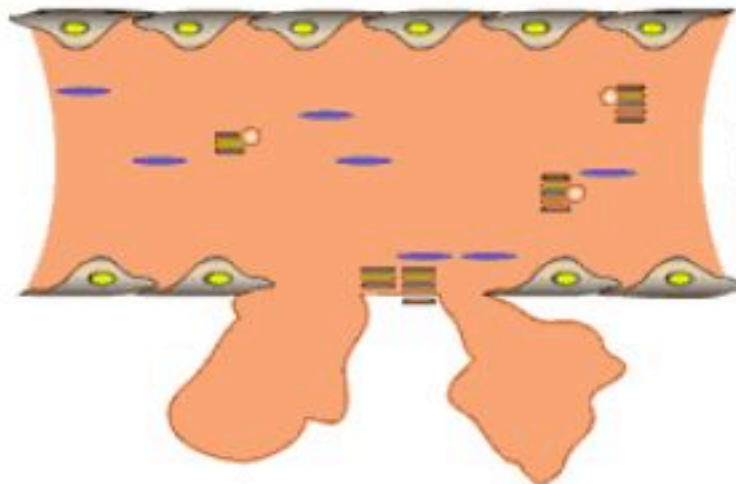


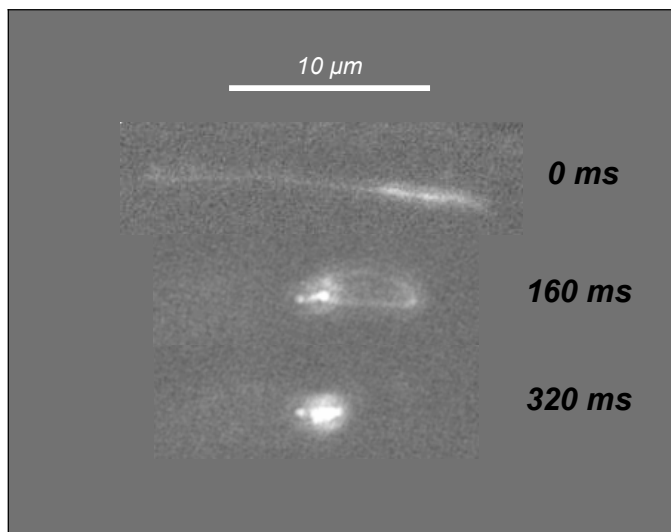
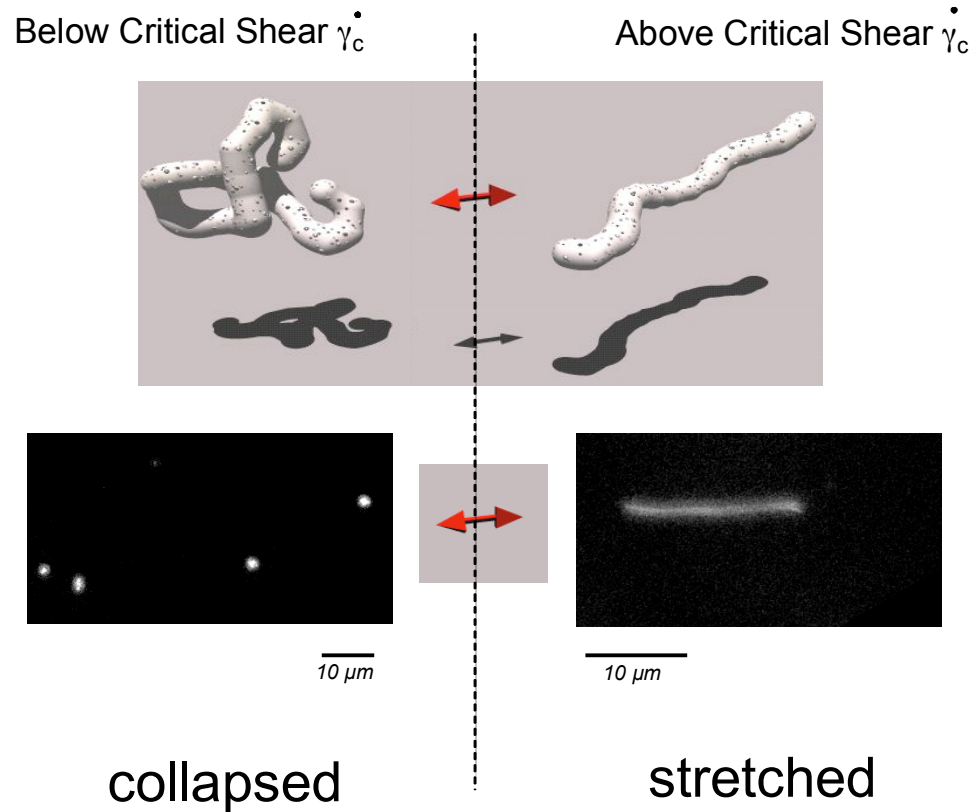
Figure 2. Broken Blood Vessel in vWD



When a person has vWD, there isn't enough vWF or the vWF is damaged. The clot may take longer to form or not form properly, and bleeding may take longer to stop.

unfolding occurs
also in bulk
(without collagen
substrate)

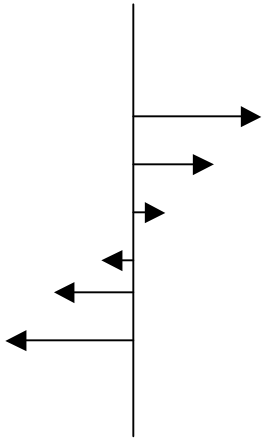
Schneider/Wixforth
(Augsburg)



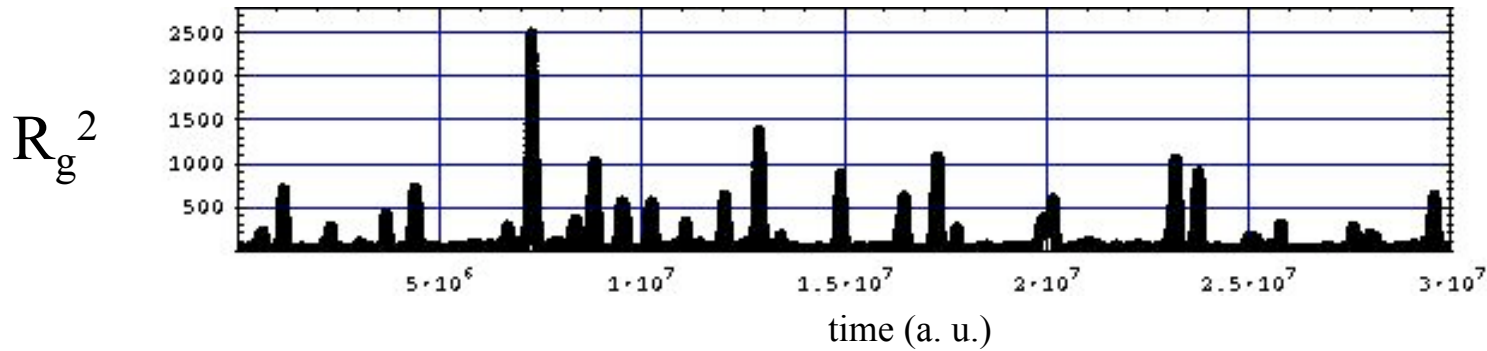
relaxation into globular state
once shear is turned off

in shear, $\varepsilon=2.5$, $\gamma=1.2$

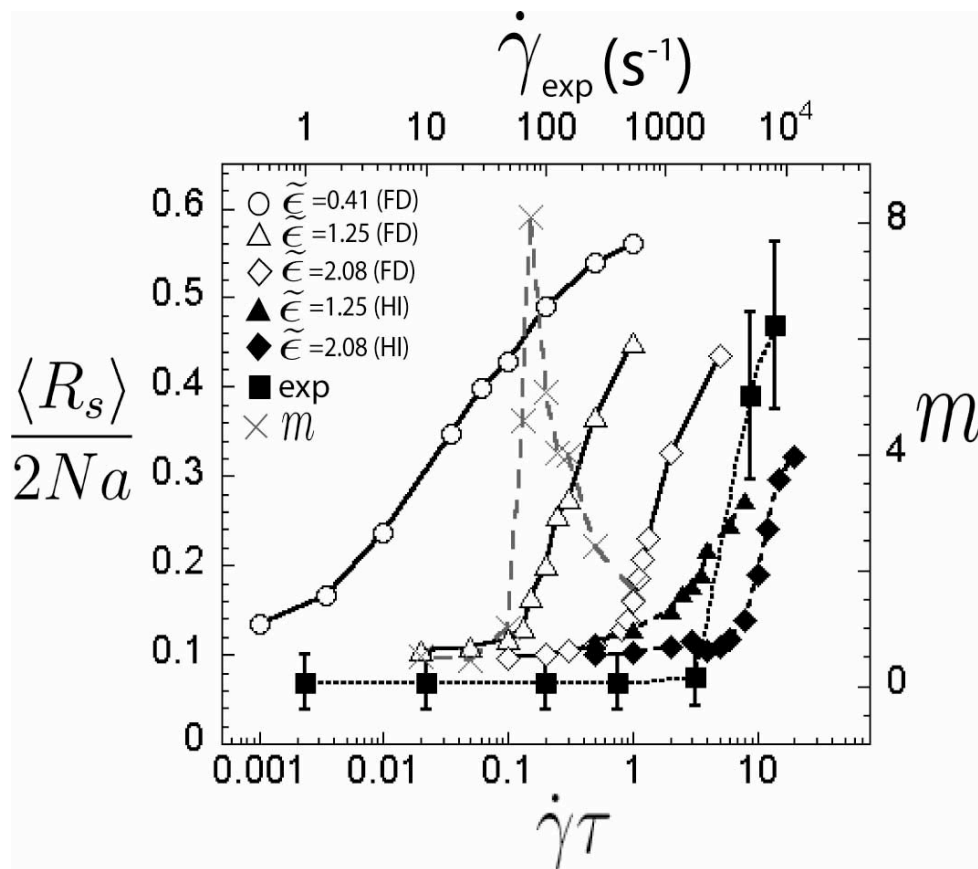
Alfredo Alexander-Katz



stretching dynamics



$$\gamma \sim \gamma^*$$



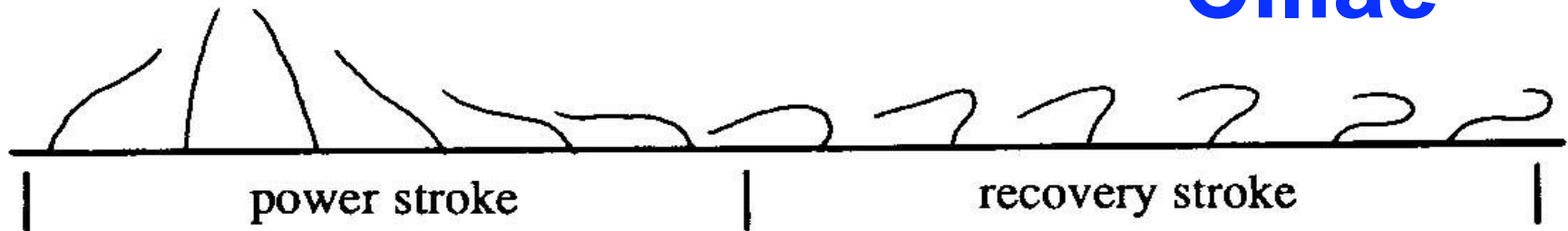
stretching response

unfolding becomes
abrupt for globular proteins
(in agreement with experiments)

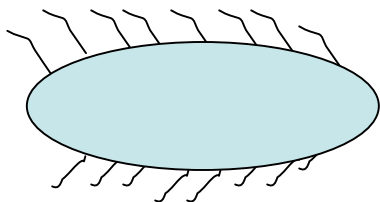
**Propulsion with
stiff polymers**

produce shear with beating polymers

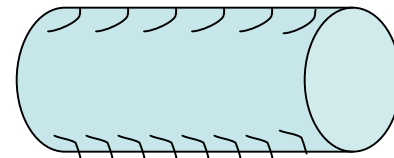
Ciliae



propulsion

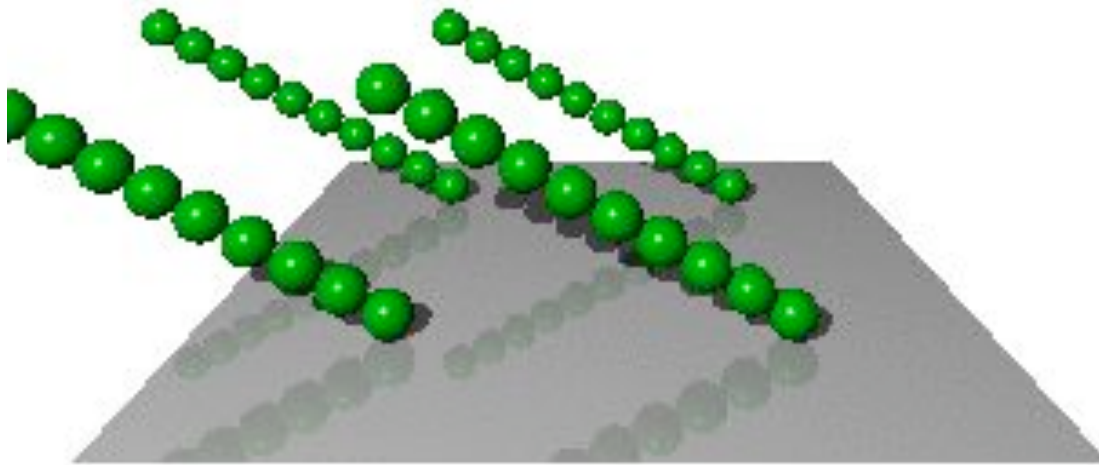


pumping



model polymer as isotropic elastic rod
apply asymmetric torque at the polymer base
-> measure net pumping velocity, efficiency, etc.

Yong-Woon Kim, RRN

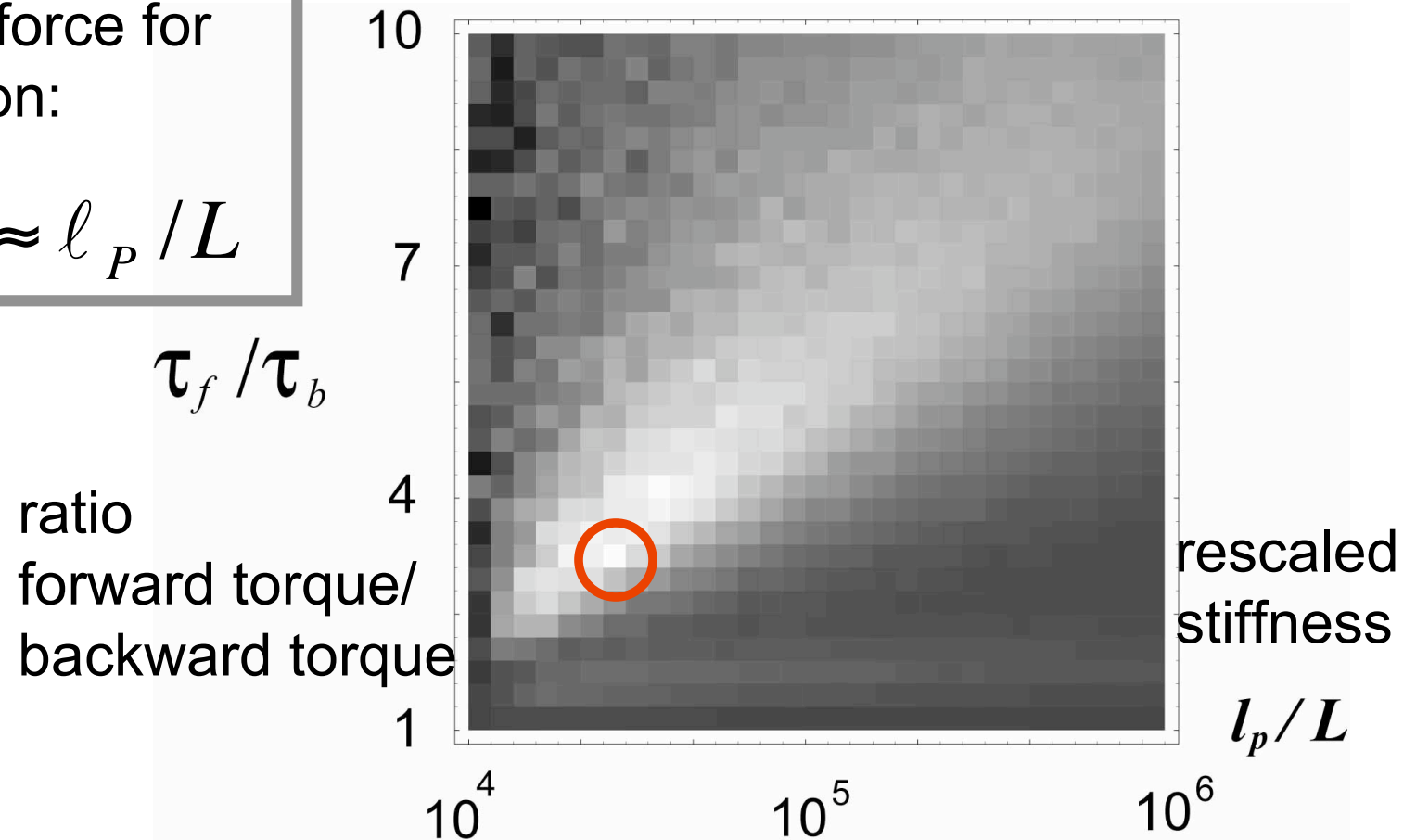


pumping efficiency = $\frac{\text{net solvent velocity}}{\text{power input}}$

1. condition:
threshold force for
deformation:

$$\tau_f / k_B T \approx \ell_p / L$$

2. Condition: asymmetry

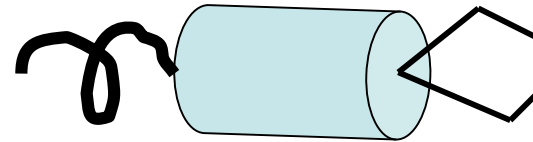


Symmetric motion: no net pumping

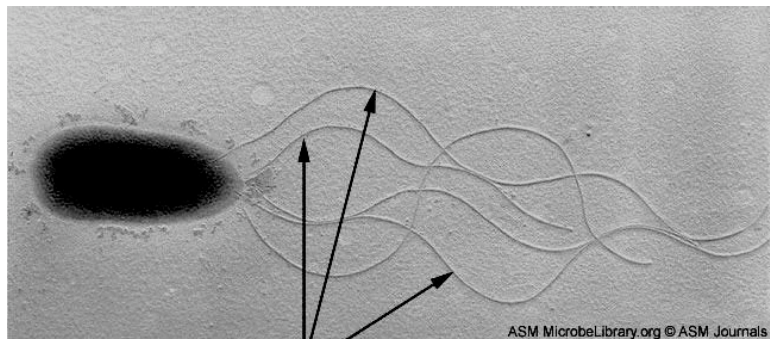
No elasticity: no net pumping (since reciprocal motion...)

propulsion with single rotating polymers

goal:
moving nanomachines



• Bacteria



Pseudomonas Putida



problem:

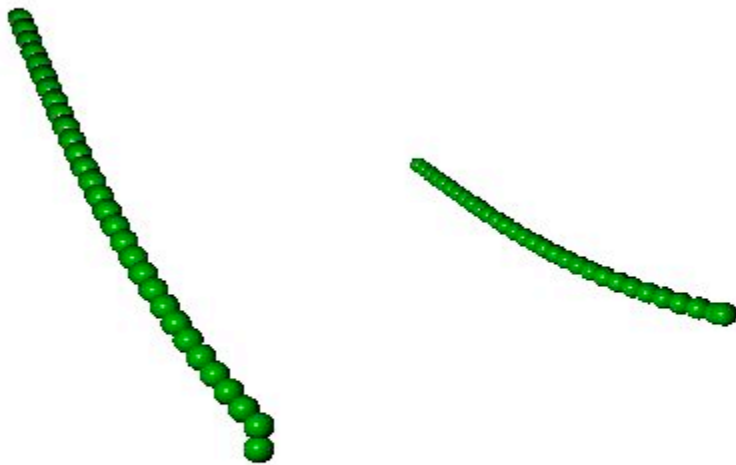
- need helical polymer,
- rotational sense determines thrust direction

question:

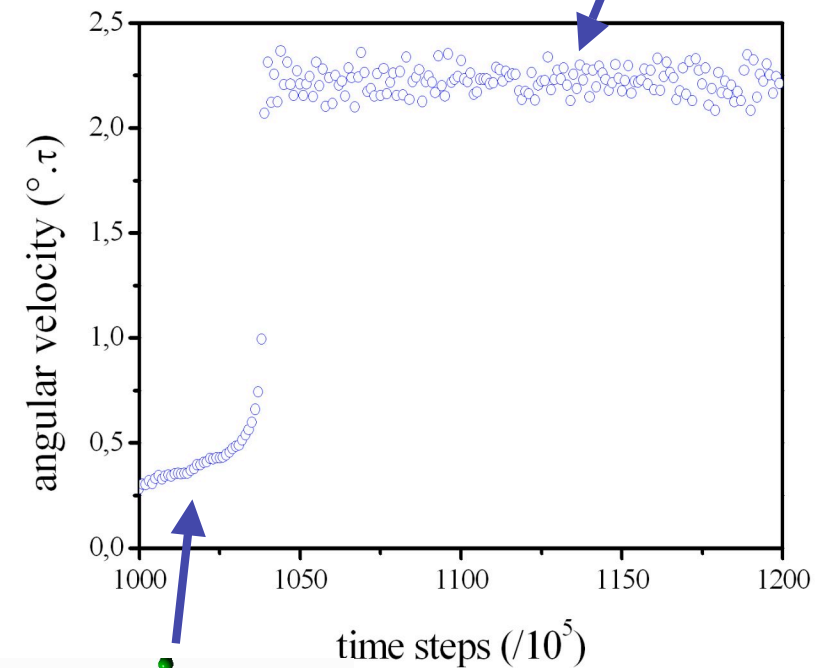
- propulsion possible with straight rotating polymers?

Dynamical transition for rotating straight flexible polymer

Manoel Manghi/
RRN



crankshafting motion



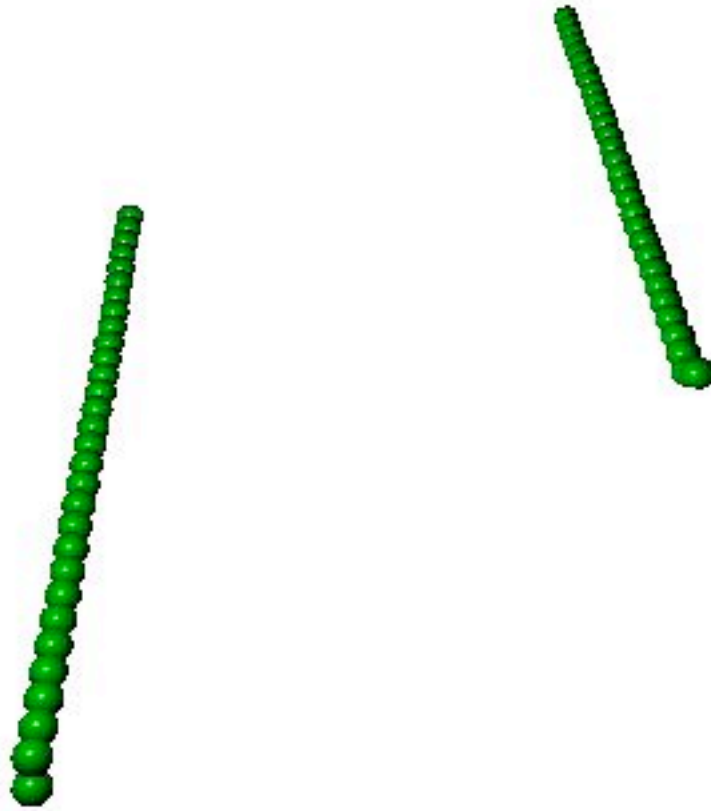
Torque = 4-5 kT/rad

$\ell_p = 6.7 L$

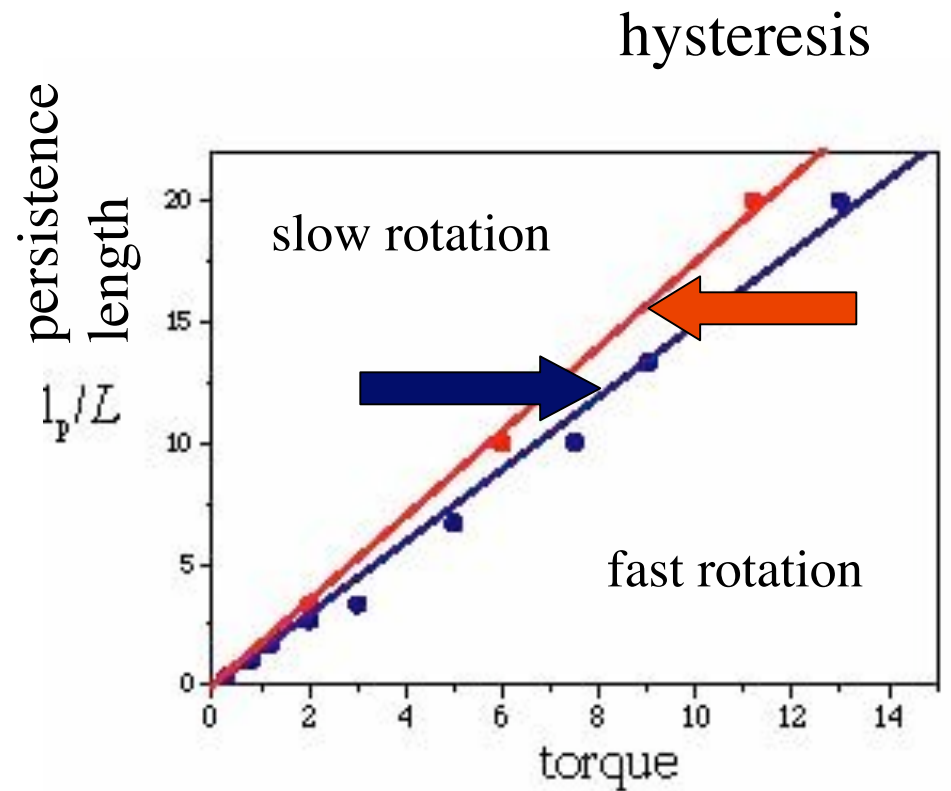
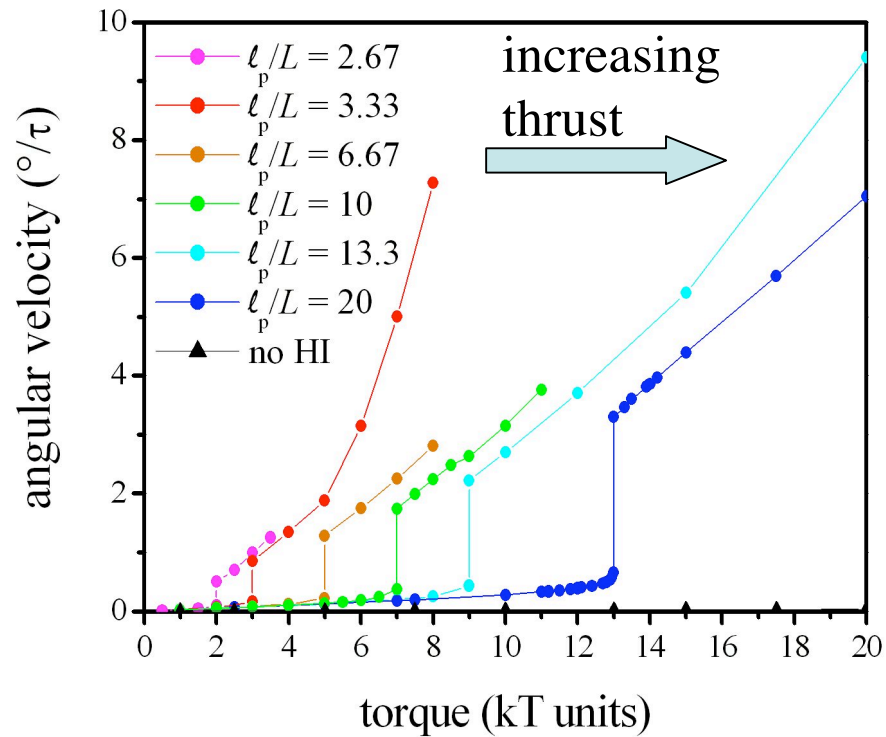
$N = 30$

axial rotation

flexible polymer , $l_p=L/3$
effects of thermal fluctuations?

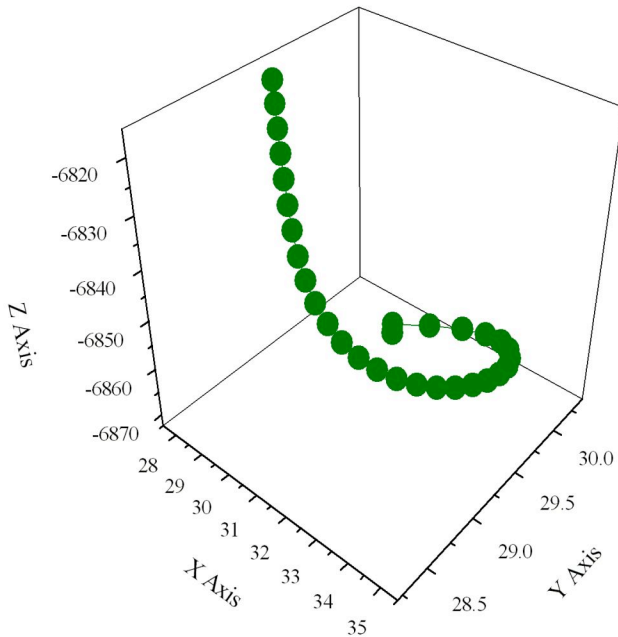


discontinuous non-eq. shape transition



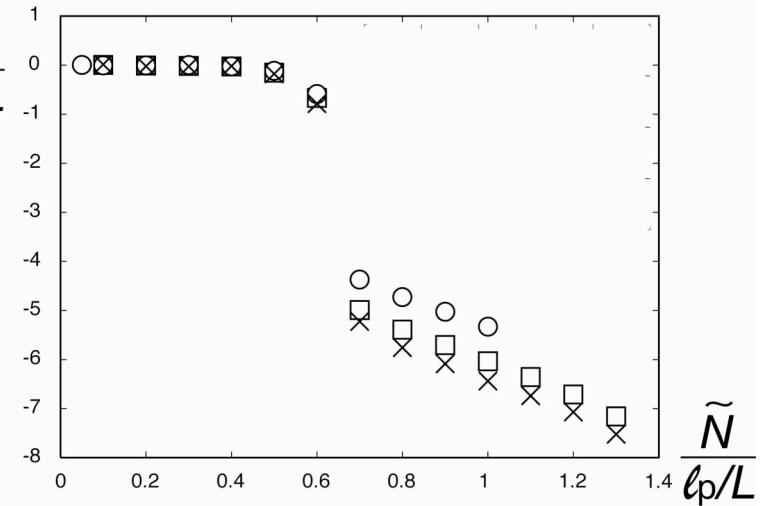
Propulsion due to breaking of time-reversal invariance of motion
 thrust direction independent of rotational sense !!

----> nano-force-rectifier



Propulsion velocity $10^3 \frac{\tilde{V}_z}{\ell_p/L}$

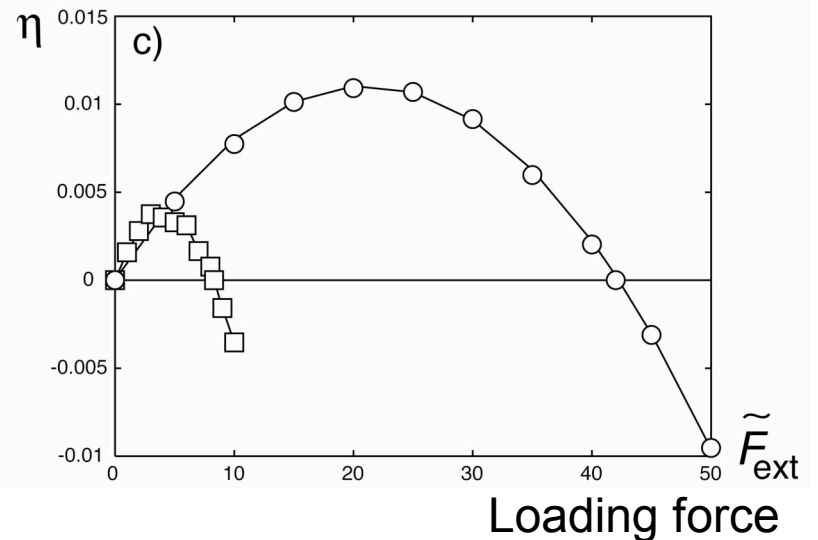
bending radius
 $1/R = N / (\ell k_B T)$
 transition at $R = L$



Propulsion efficiency η

$$\eta = \frac{\text{propulsion power}}{\text{rotation power}}$$

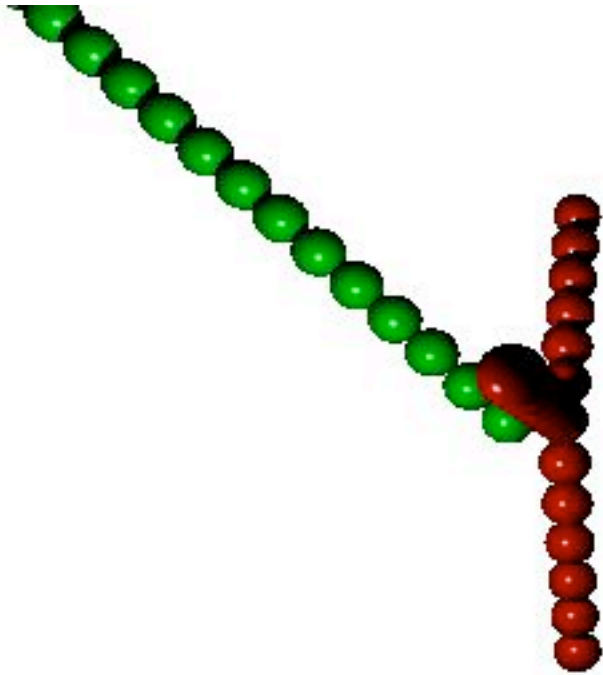
efficiency low (but not worse than bacterial flagellae)



Flagellum with a base: total force and torque must be zero

→ rotation leads to counterrotation

→ trajectories are complex



Hirofumi Wada

first form helix (difficult!):



then rotate, slowly



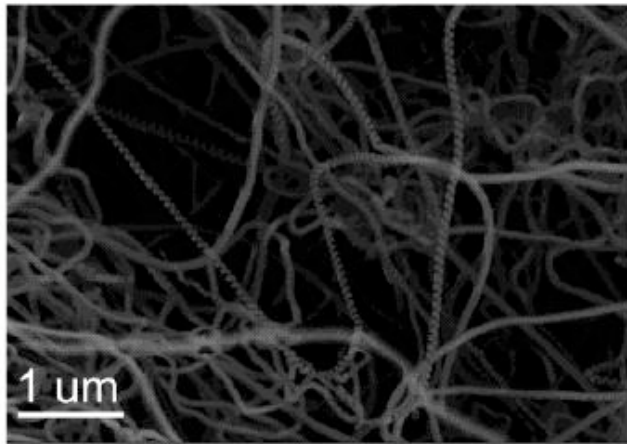
quickly
(periodic twist-stretch conversion)



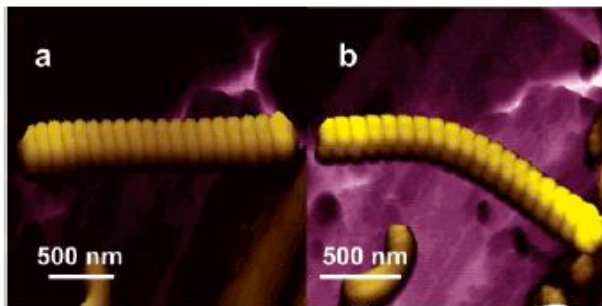
-shape (polymorphism??), bundling, efficiency.....

pulling on helical nanosprings

Hirofumi Wada (JSPS-fellow) & RRN



SEM characterization of as-synthesized silicon oxide nanowires

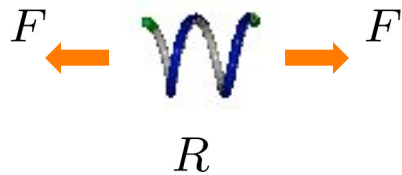


AFM manipulation of a helical silica nanospring

Hai-Feng Zhang *et al.* Nano Lett. **3** 577 (2003)



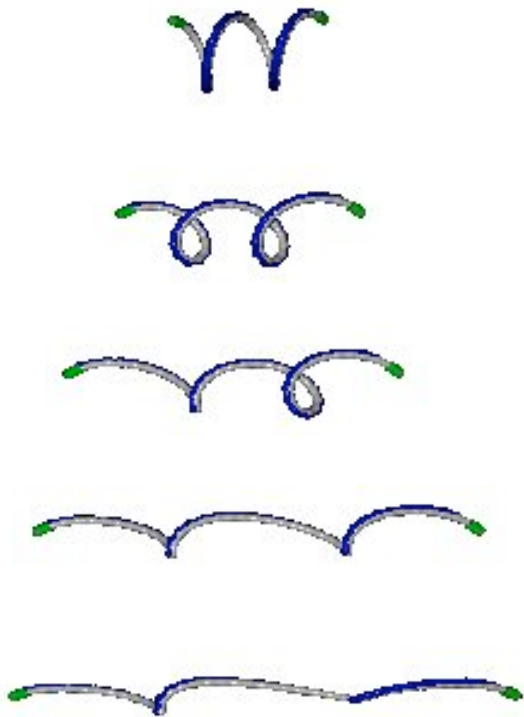
pulling on a nanospring



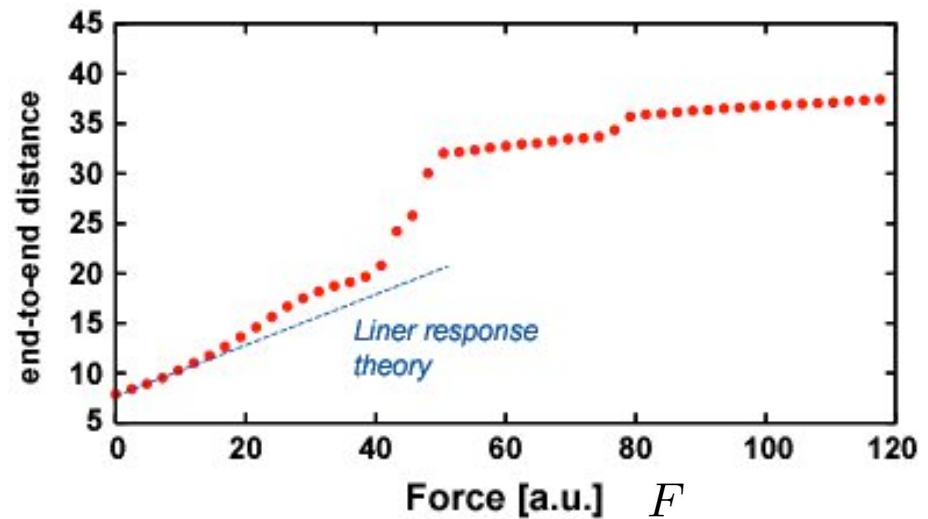
Linear response for small pulling force

$$F \approx K_{sp}(R - R_0)$$

$$K_{sp} = \frac{\Omega_0^2}{L} [4A\beta^2 + C(1 - \beta^2)^2]$$

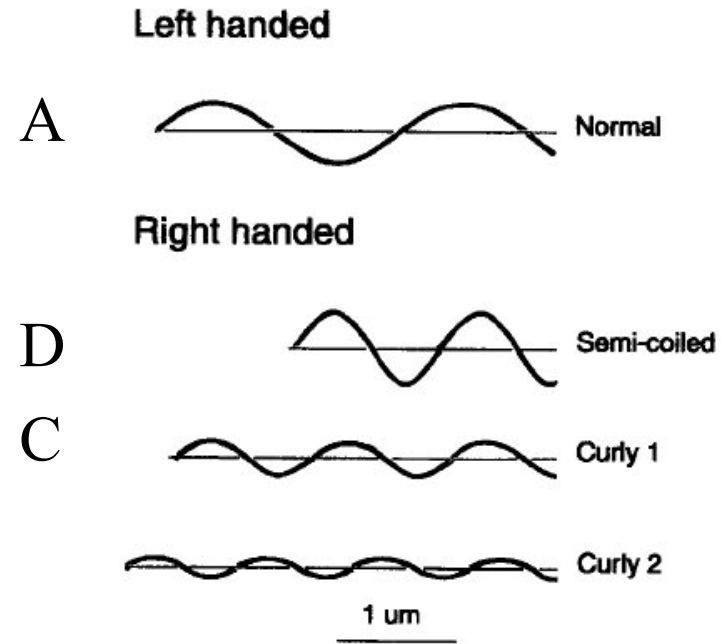
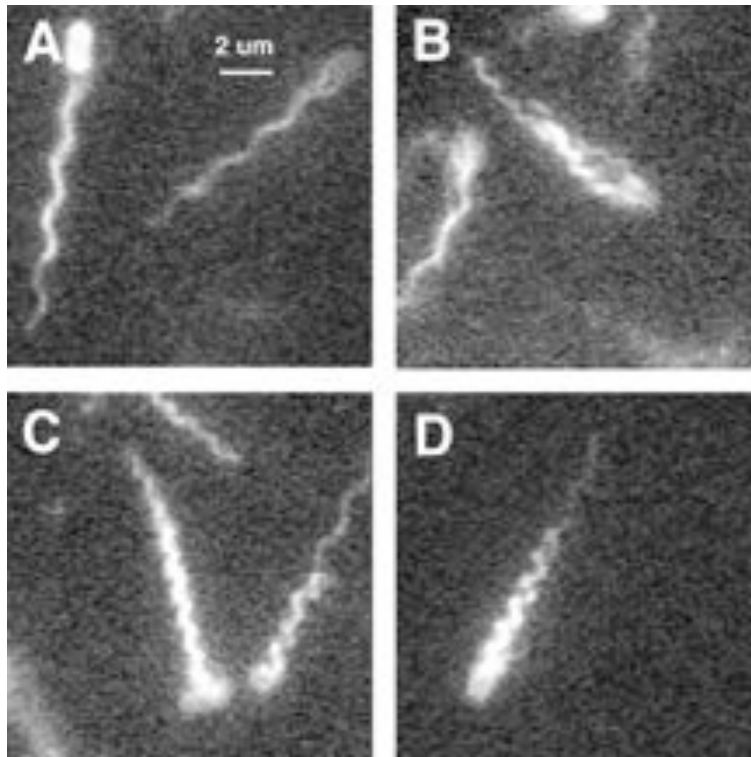


R



Force-extension curve reveals stretching instability of a helical nanospring.

Polymorphic transformations of flagellae as function of rotational sense and frequency



Dynamic conversion of bistable helix



design of simple propulsion devices
with stiff polymers
leads (naturally) to biomimetic structures