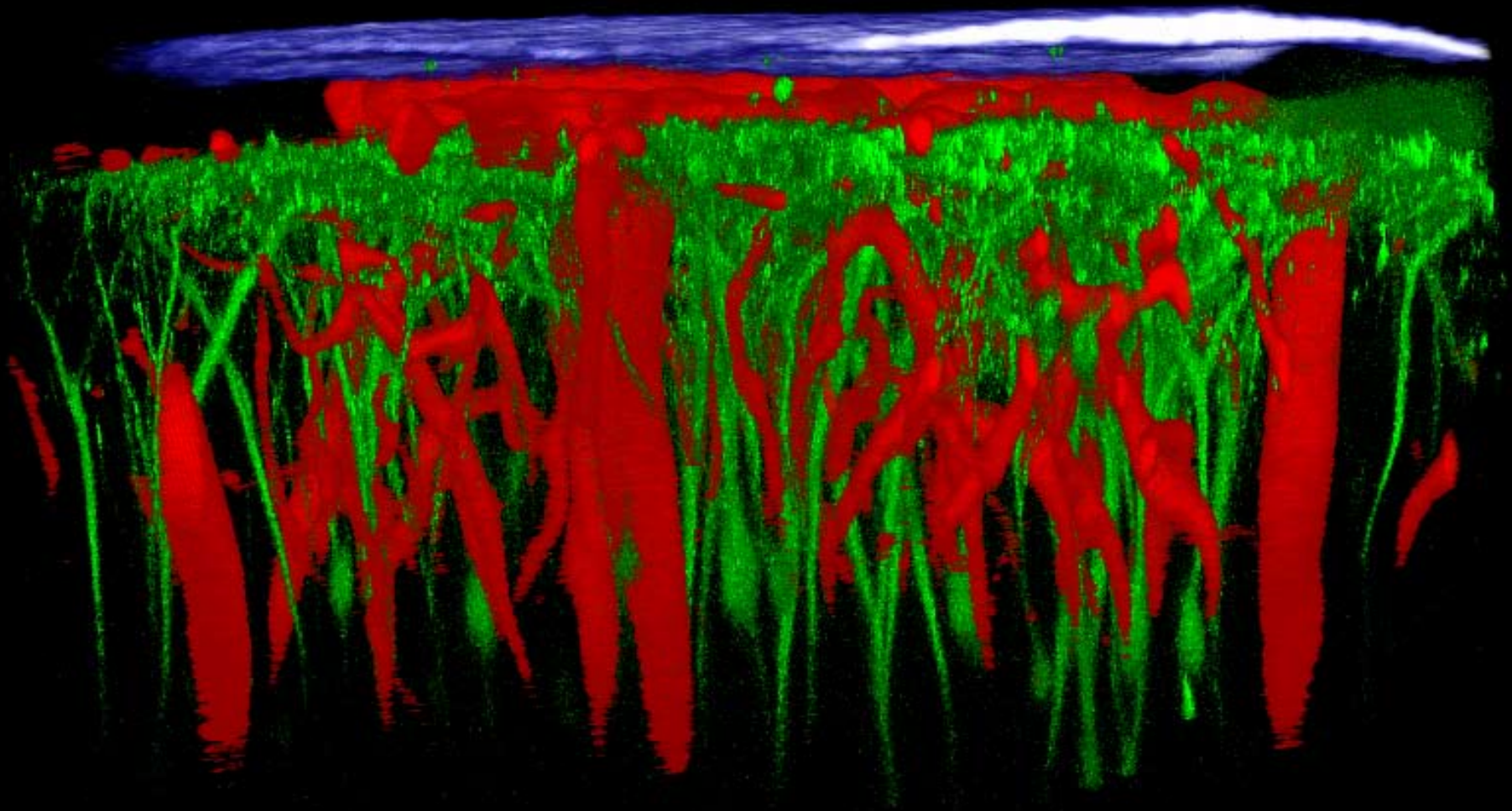


# Cortical angioarchitecture and vasodynamics

David Kleinfeld - UCSD

KITP program “Network architecture of brain structures and functions”



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# **Why should biologists be interested in cortical blood flow?**

## **Dynamic resource allocation**

**Blood is a limited resource, yet neural function depends on adequate flow**

## **Neuronal and vascular function are highly intertwined**

**Stroke and dysfunction of neurovascular control leads to neurological decline**

## **Correspondence of blood flow and oxygenation to neural activity**

**Functional magnetic resonant imaging (fMRI) and intrinsic optical imaging (IOS) are powerful noninvasive tools to study brain function. Yet fMRI and IOS rely on poorly understood relationships between blood dynamics and neural activity.**

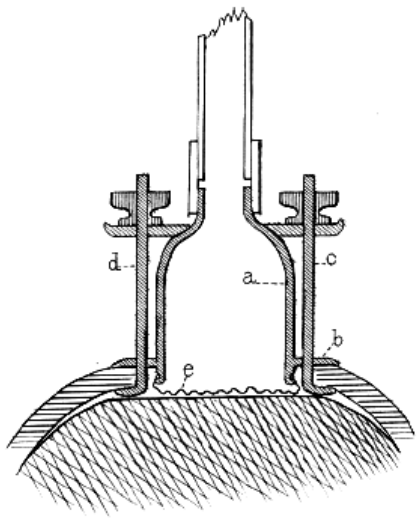
## **Our program**

**Connect network topology with the dynamics of flow**

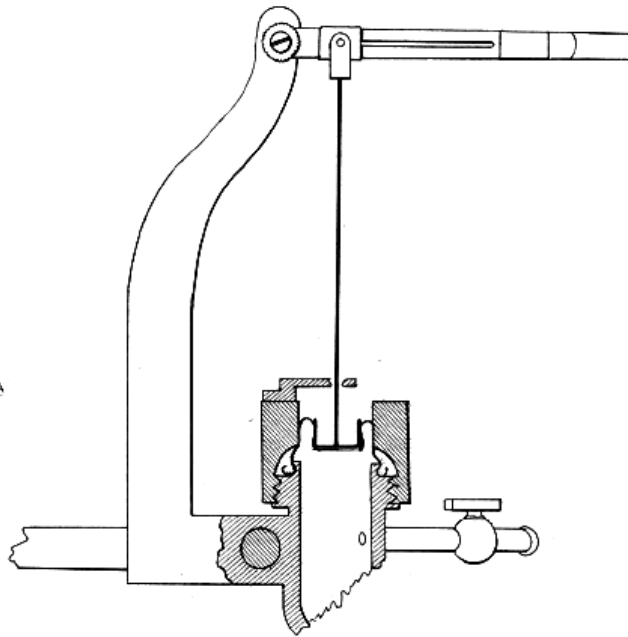
**Understand the consequences of perturbation to flow**

**Understand the mechanism of neuronal control of flow**

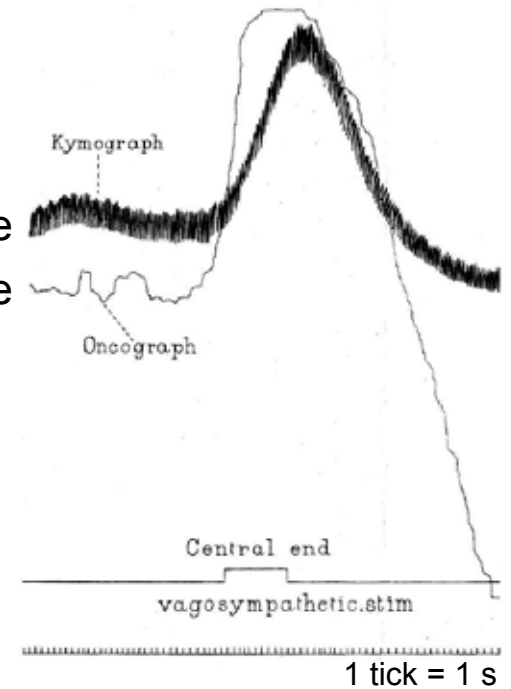
# Earliest publication on vasodynamics in the mammalian brain (Roy & Sherrington, J Physiol 1890)



Sealed, constant pressure chamber  
to record changes in brain volume



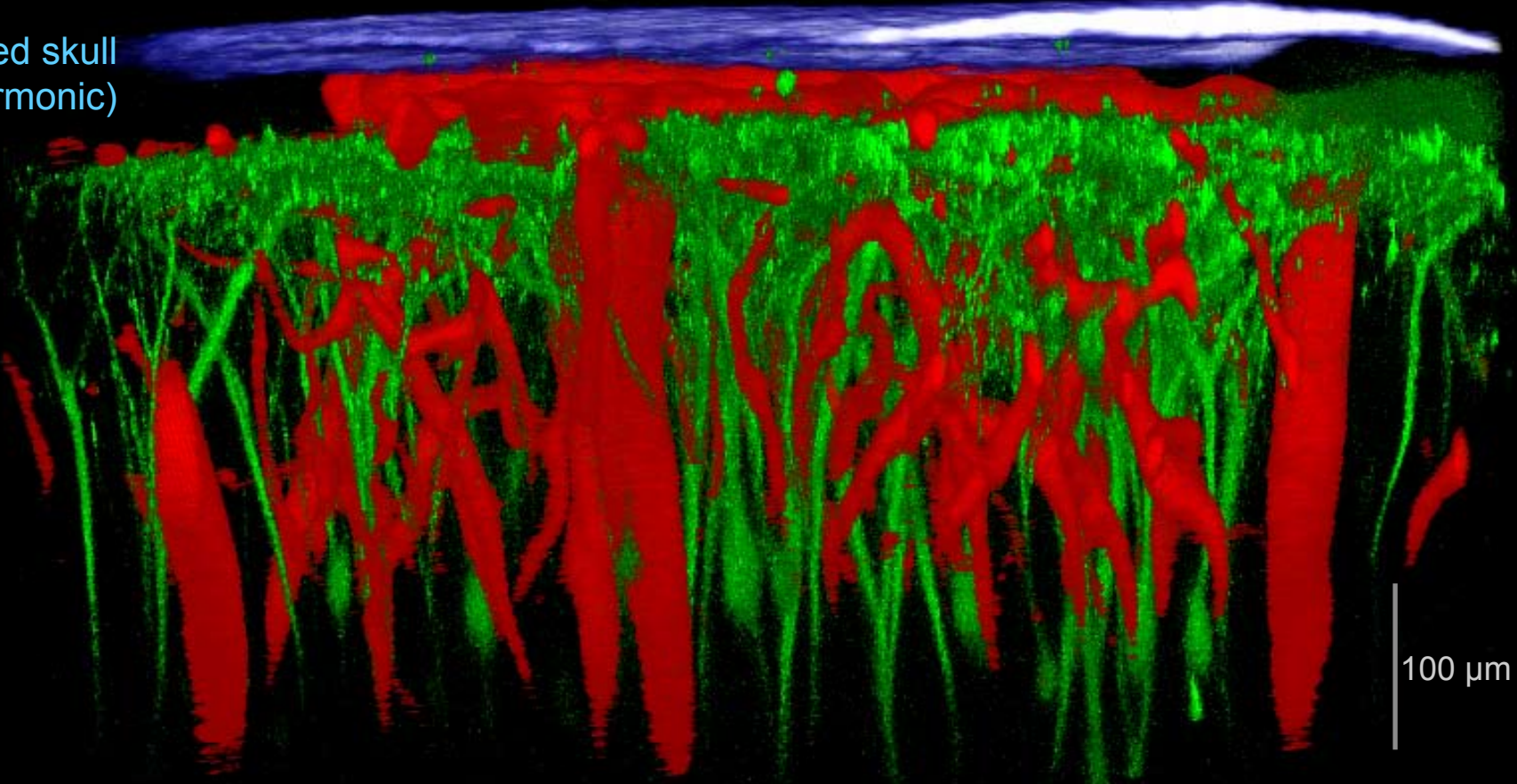
Systemic blood pressure  
Brain volume



Changes in brain volume in response to  
stimulation of the vagus (sympathetic nerve)

# Polished and reinforced skull for chronic in vivo two-photon imaging

Thinned skull  
(second harmonic)

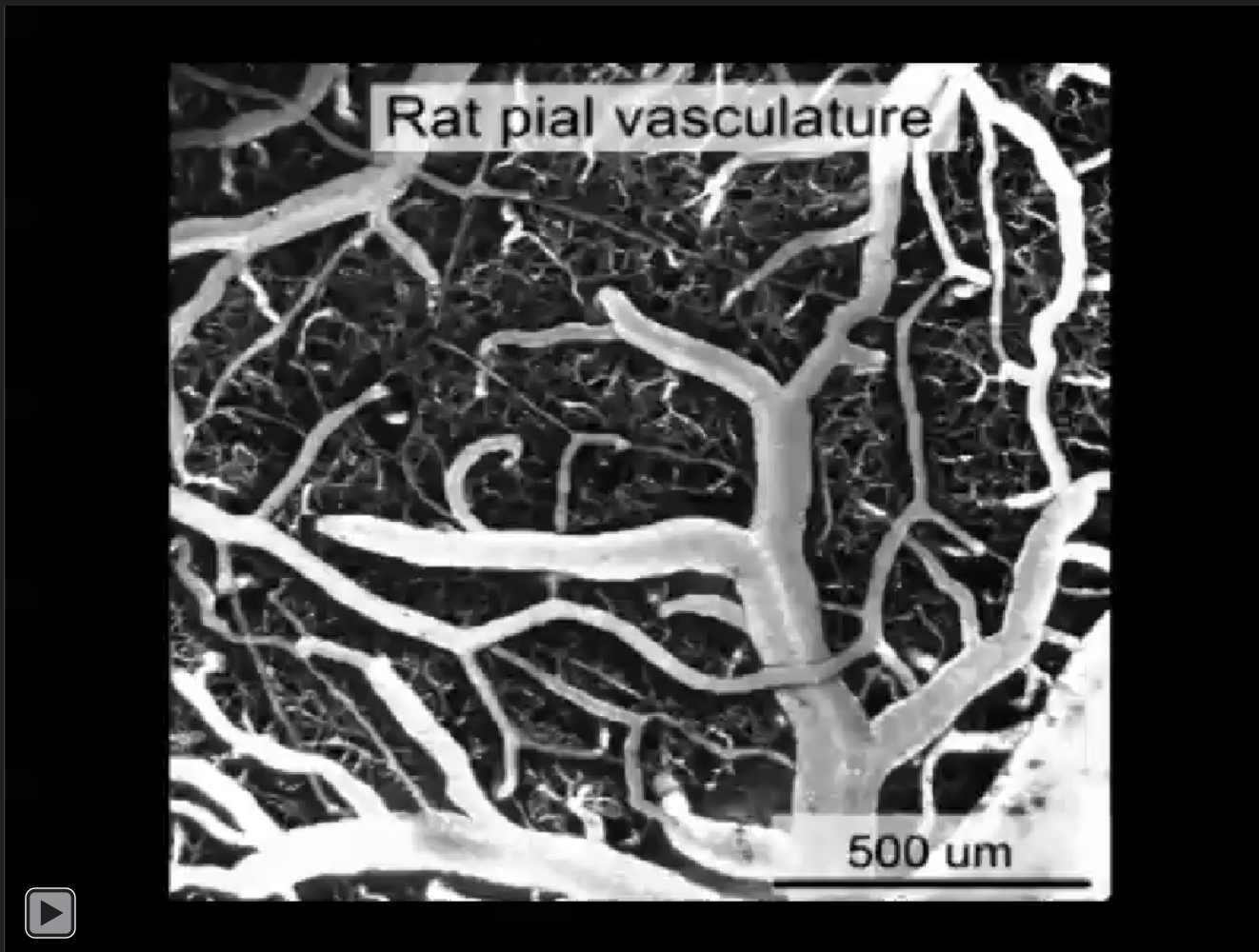


100  $\mu$ m

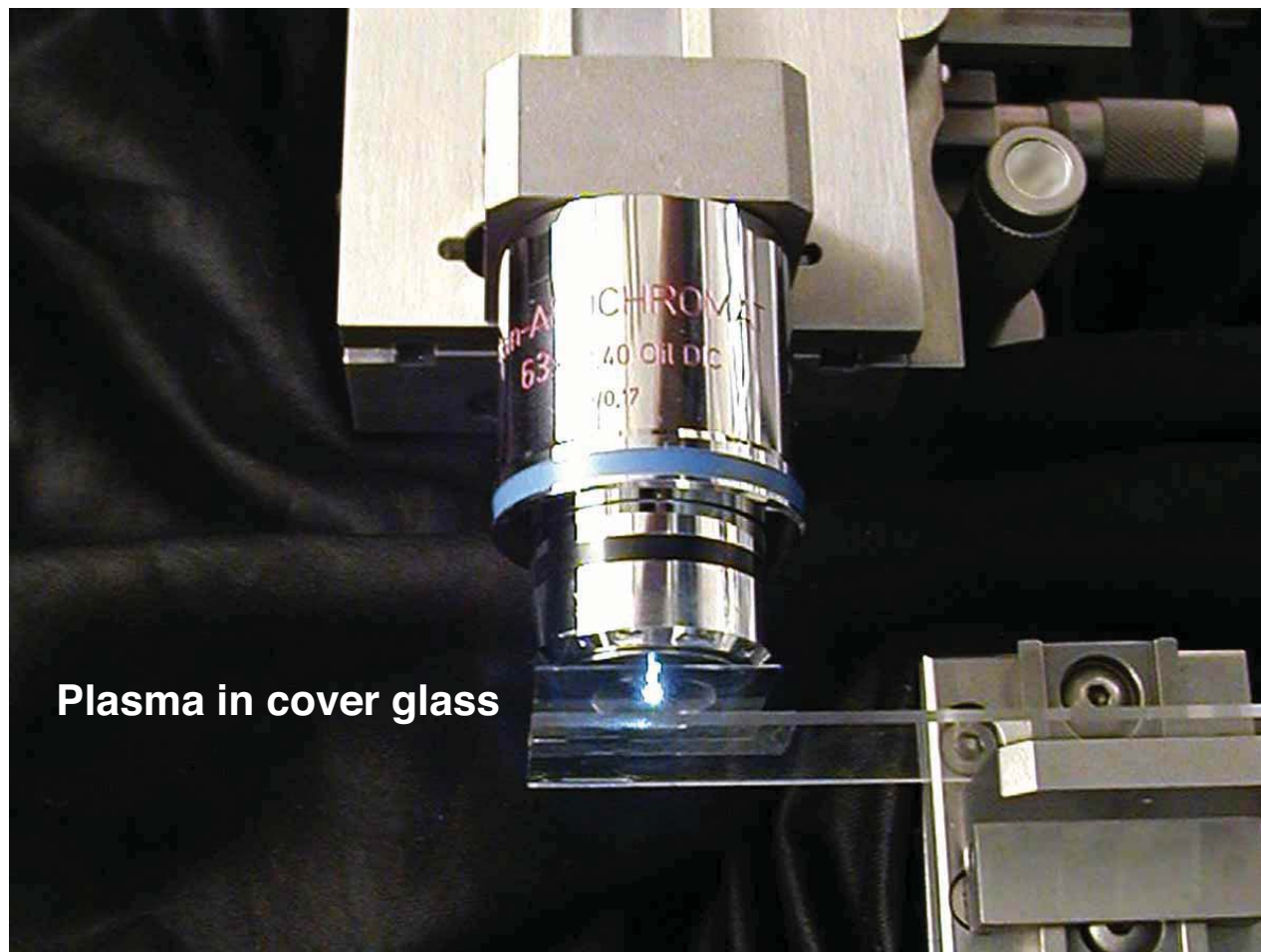
Blood vessels  
(fluorescein/dextran)

Neuronal dendrites  
(green fluorescent protein)

# Horizontal section of cortical vasculature via 2-photon microscopy



# Focused ultrashort laser pulses generate a plasma in materials



Schaffer, Brodeur, Garcia & Mazur (Optics Letters 2001)

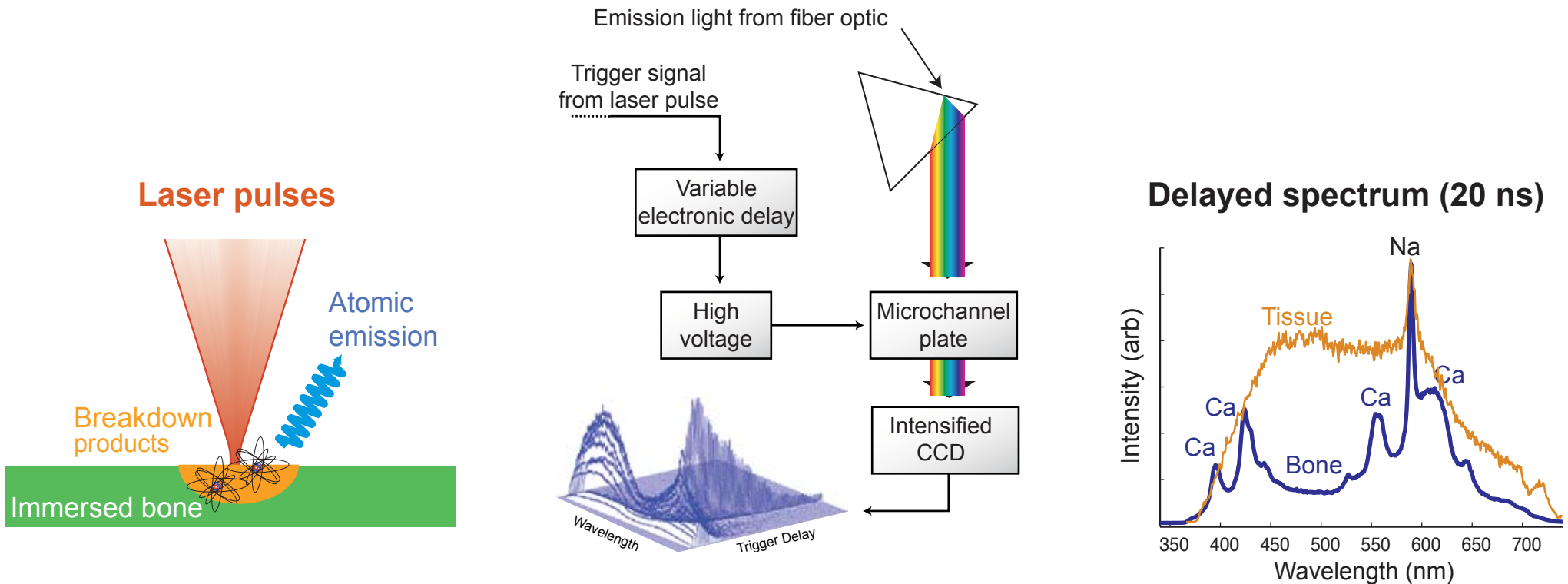
Loesel, Niemz, Nille & Juhasz (IEEE J Quantum Electronics 1996)

Stuart, Feit, Herman, Rubenchik, Shore & Perry (JOSA B 1996)

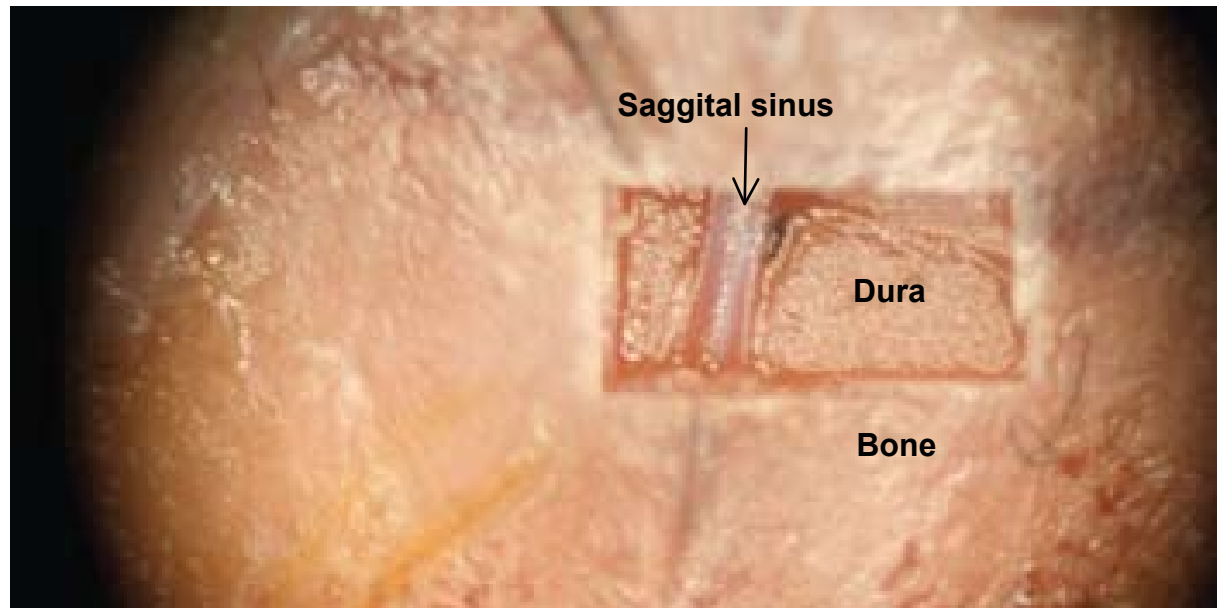
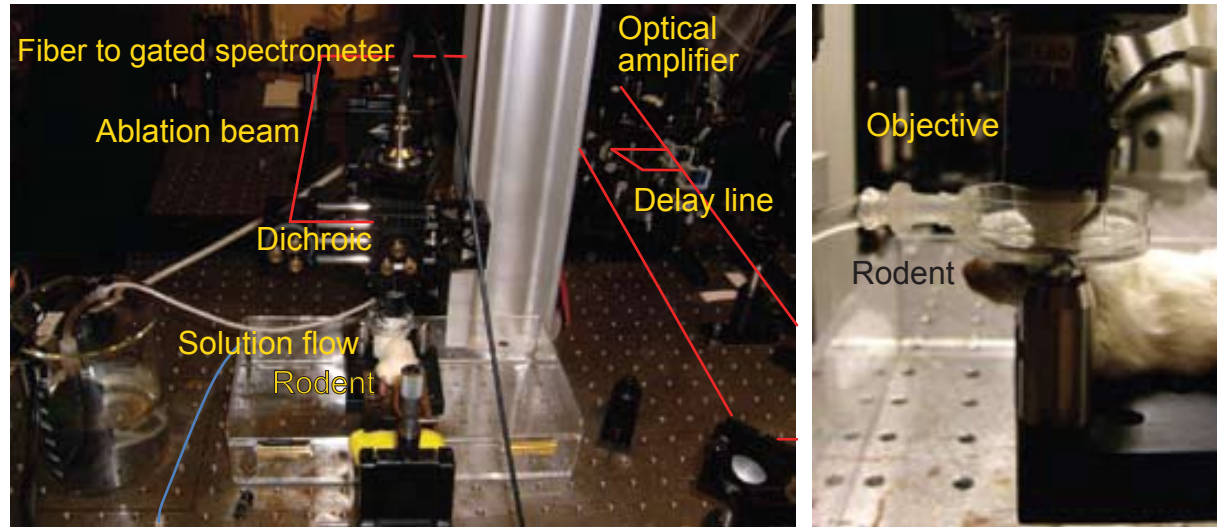


# Laser induced plasma ablation and spectroscopy with amplified ultrafast pulses for automated surgery with feedback control

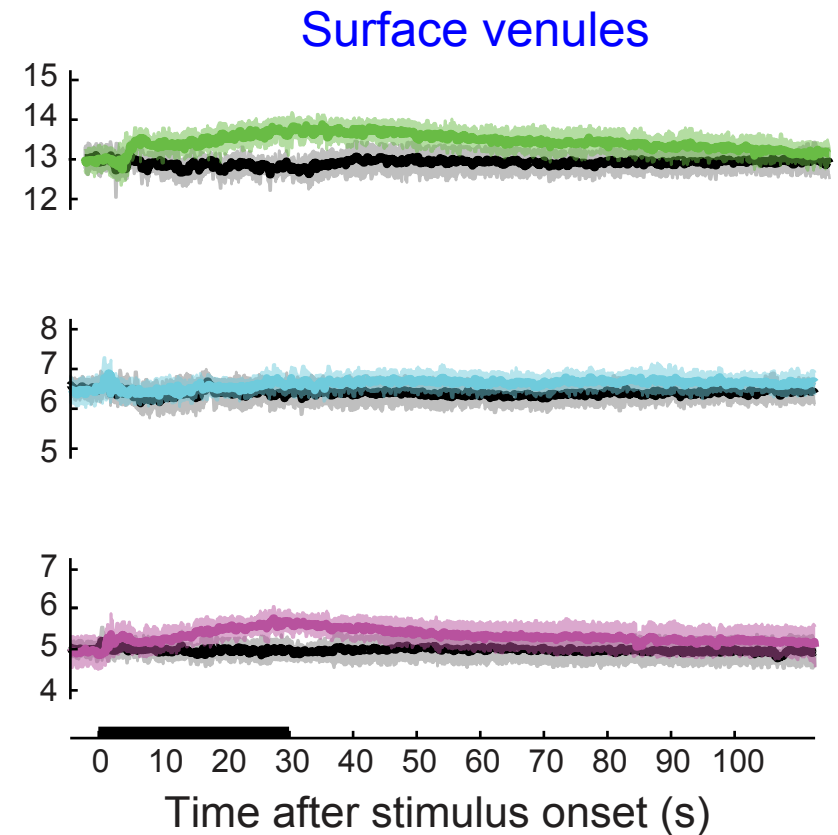
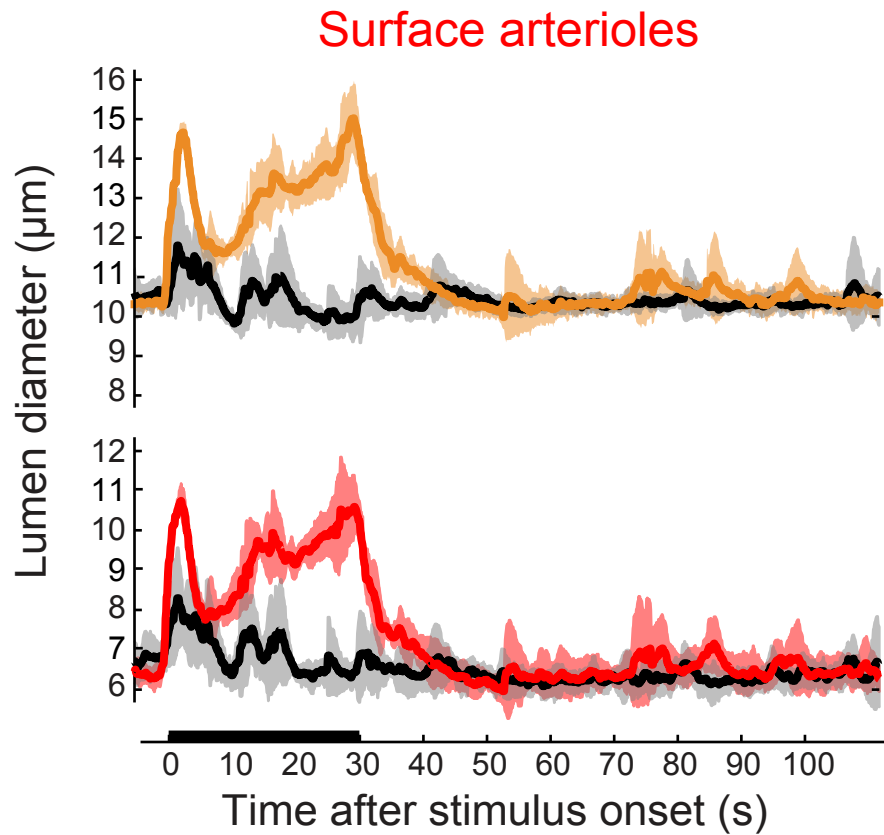
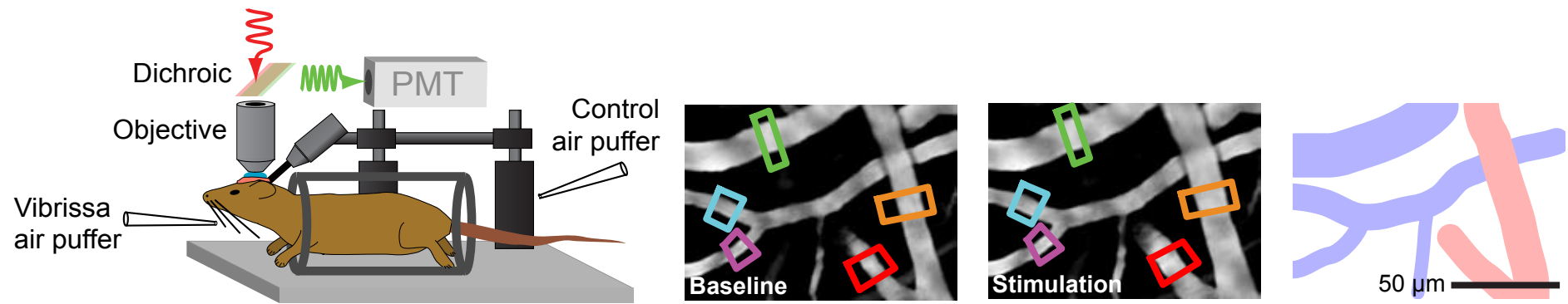
## Time-resolved spectrum



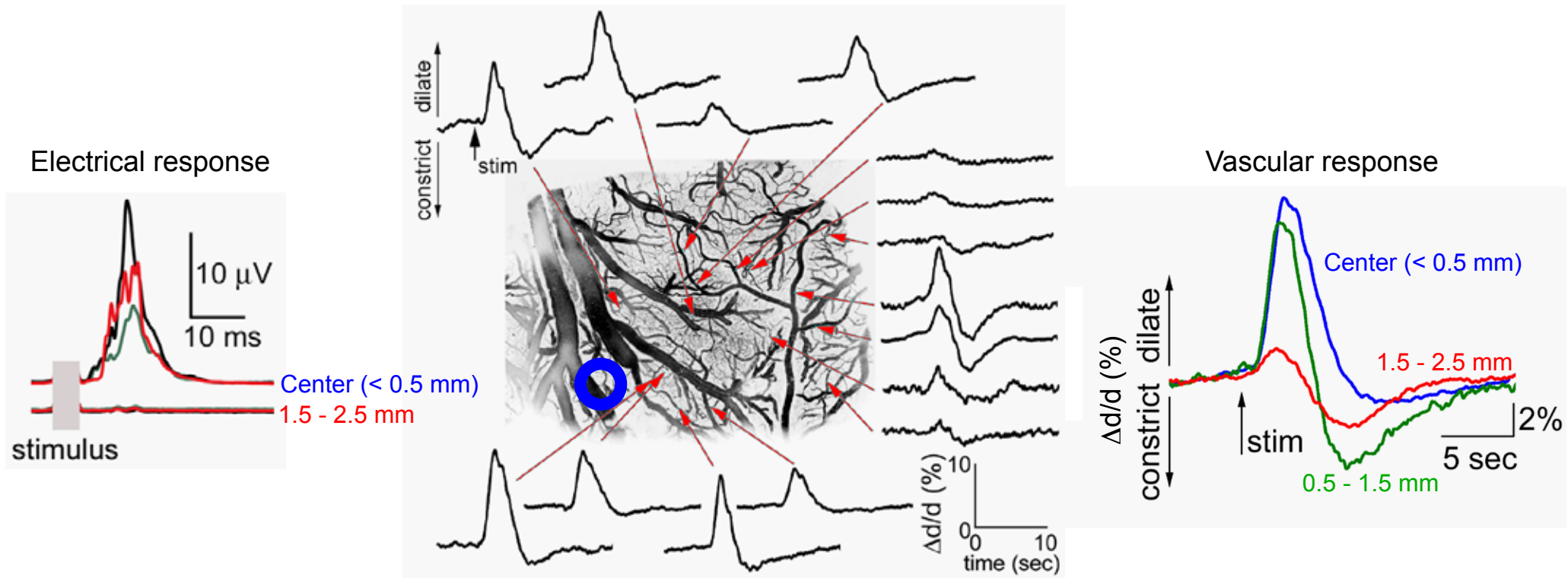
# All optical surgery with range finding and feedback (via LIPS and SHG)



# Vascular changes in response to stimulation are predominantly arteriole



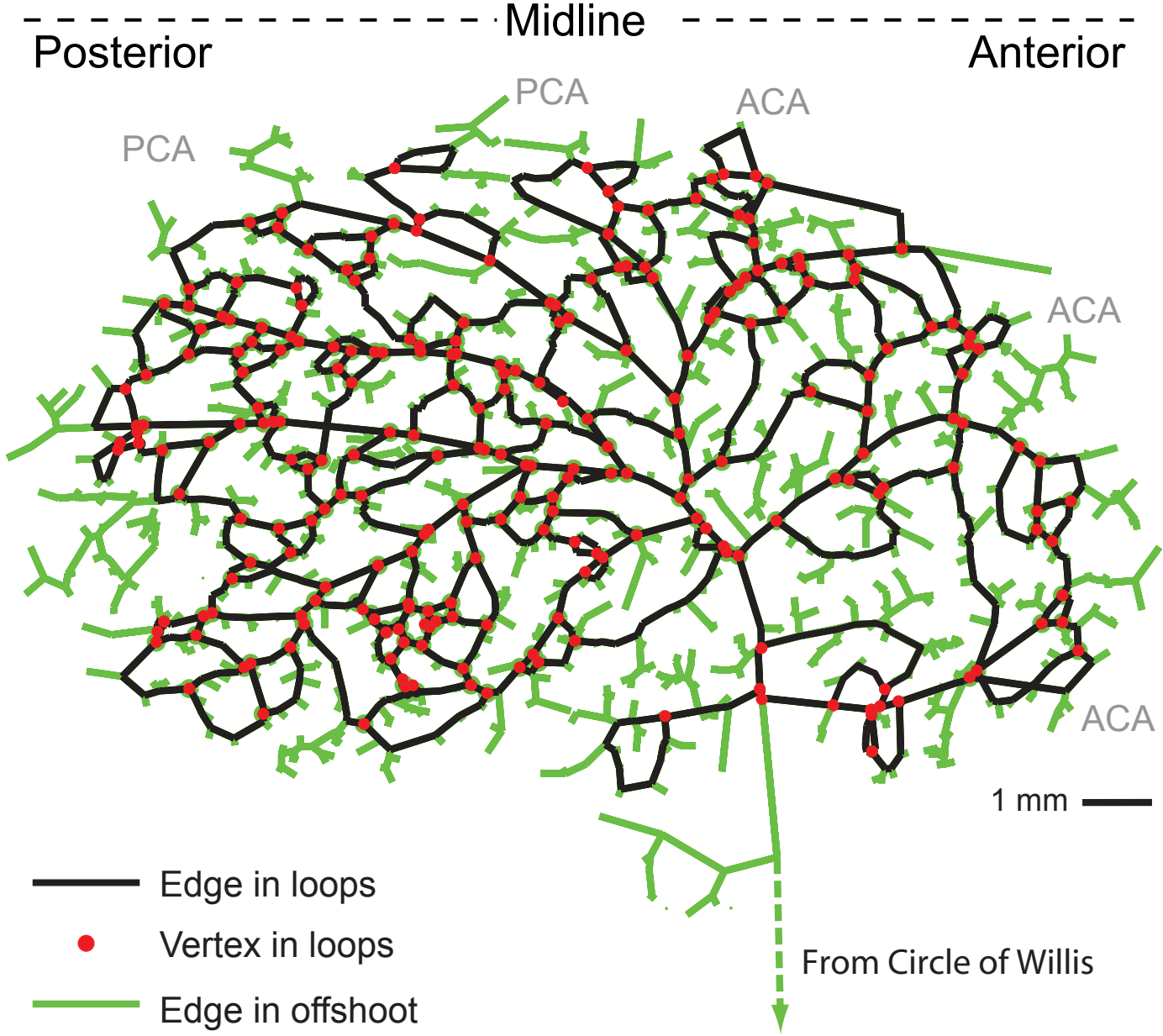
# Localized somatotopic stimulation leads to a center-surround pattern of vasodilation and vasoconstriction in contralateral surface arterioles



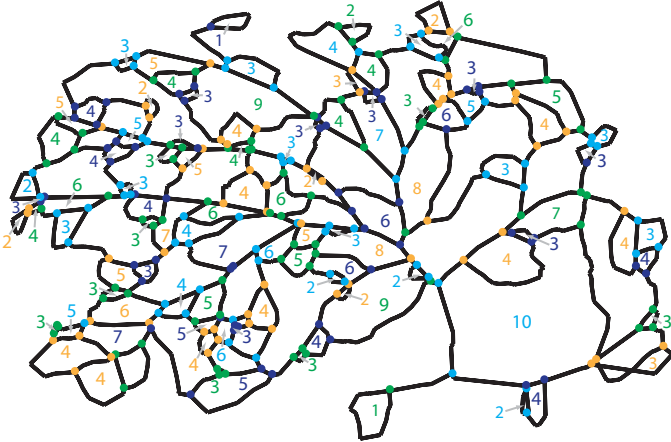
**Lesson: Surface arterioles can redistribute blood to areas with heightened activity.**

**Hypothesis: Redistribution results from a highly interconnected network rather than a tree structure.**

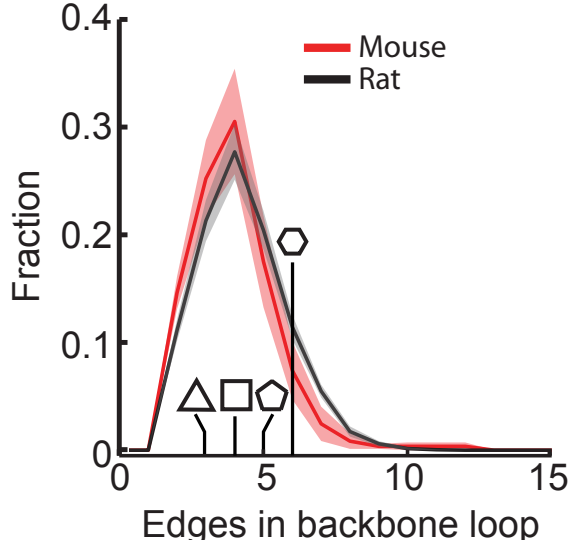
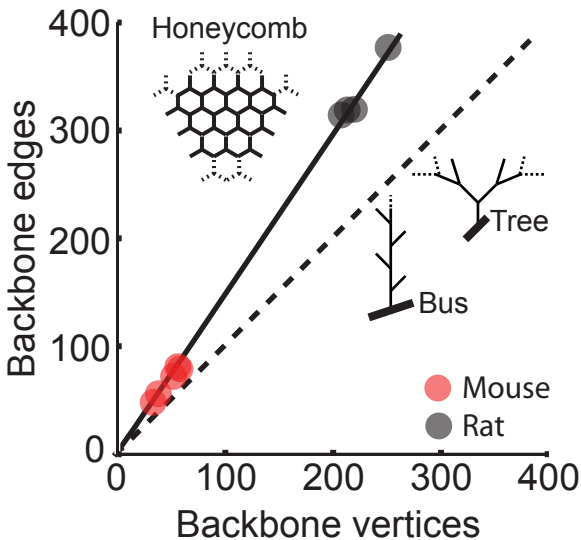
# Surface arteriole network consists primarily of interconnected loops



# Surface arteriole network consists primarily of interconnected loops



Kirchoff-like backbone loops



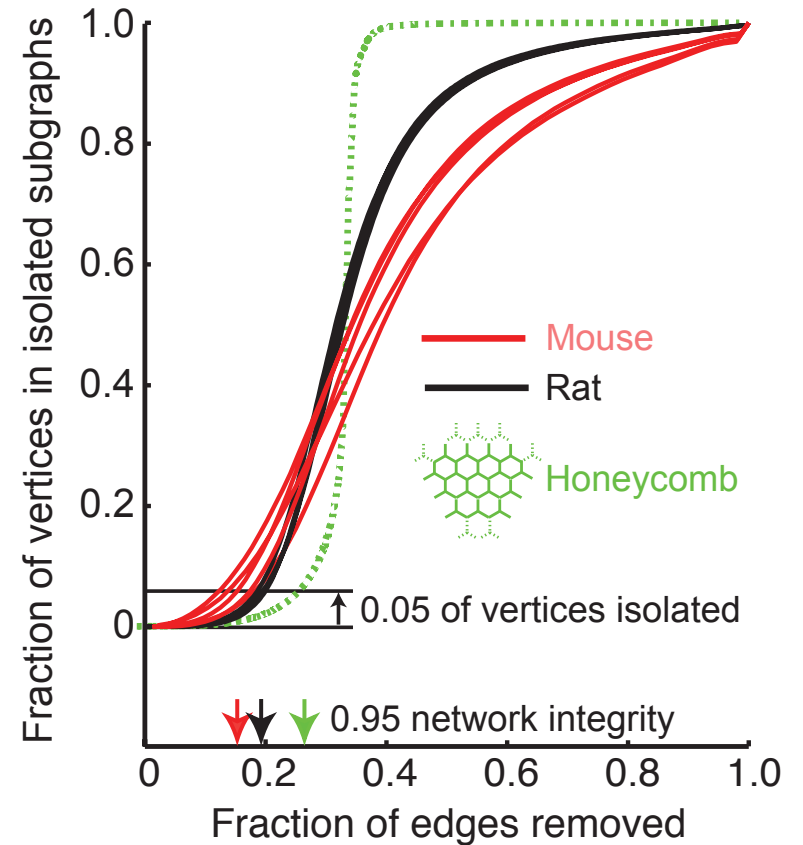
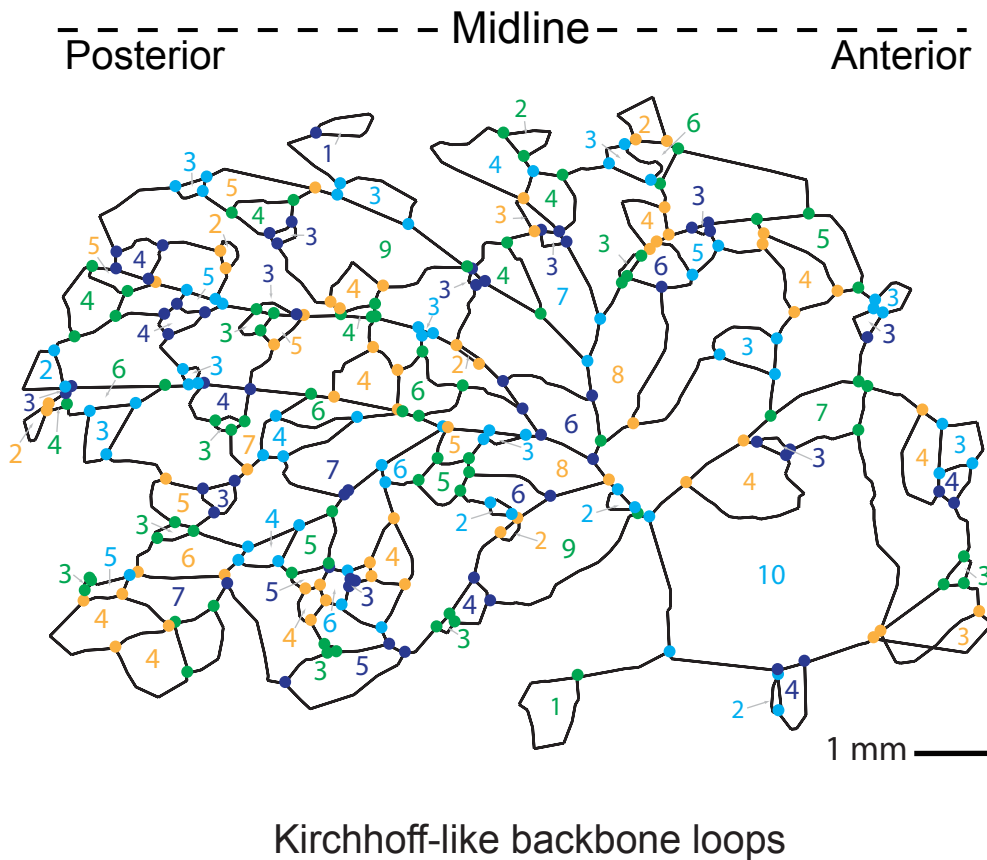
Coordination of 3 at every vertex - yet no simple lattice captures all features

**Lesson: Surface arterioles are arranged as a highly interconnected network.**

**Question: Does the surface angioarchitecture robustly circulate blood?**

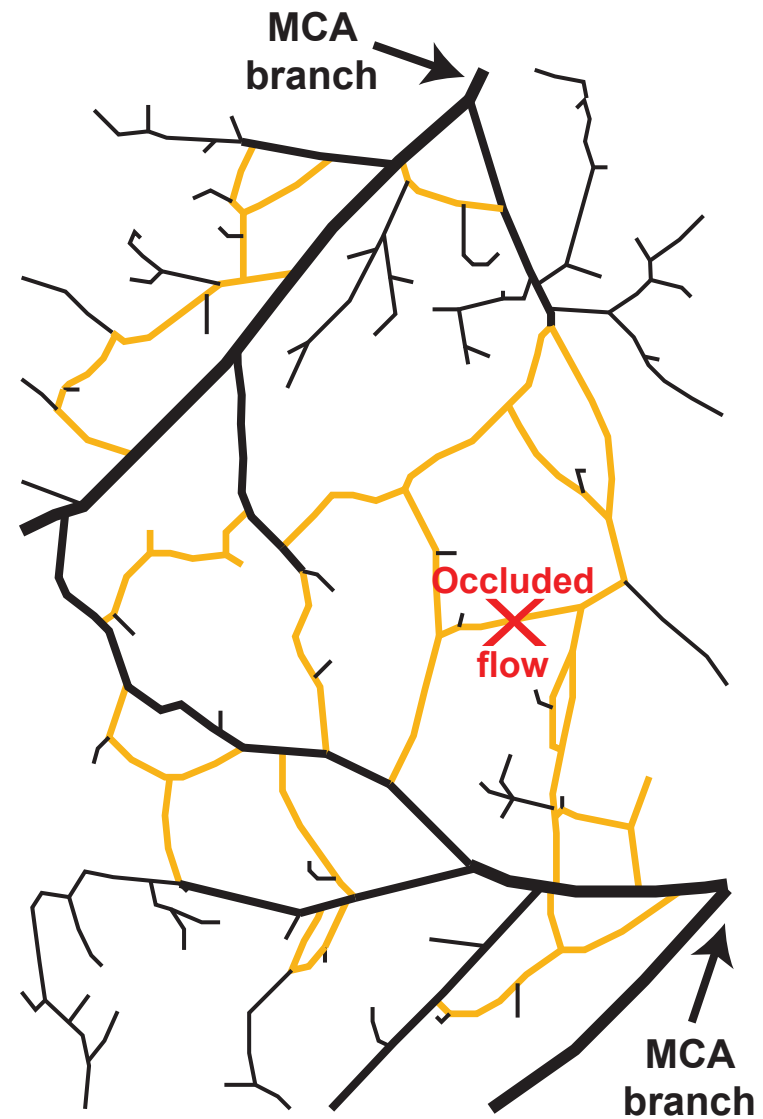
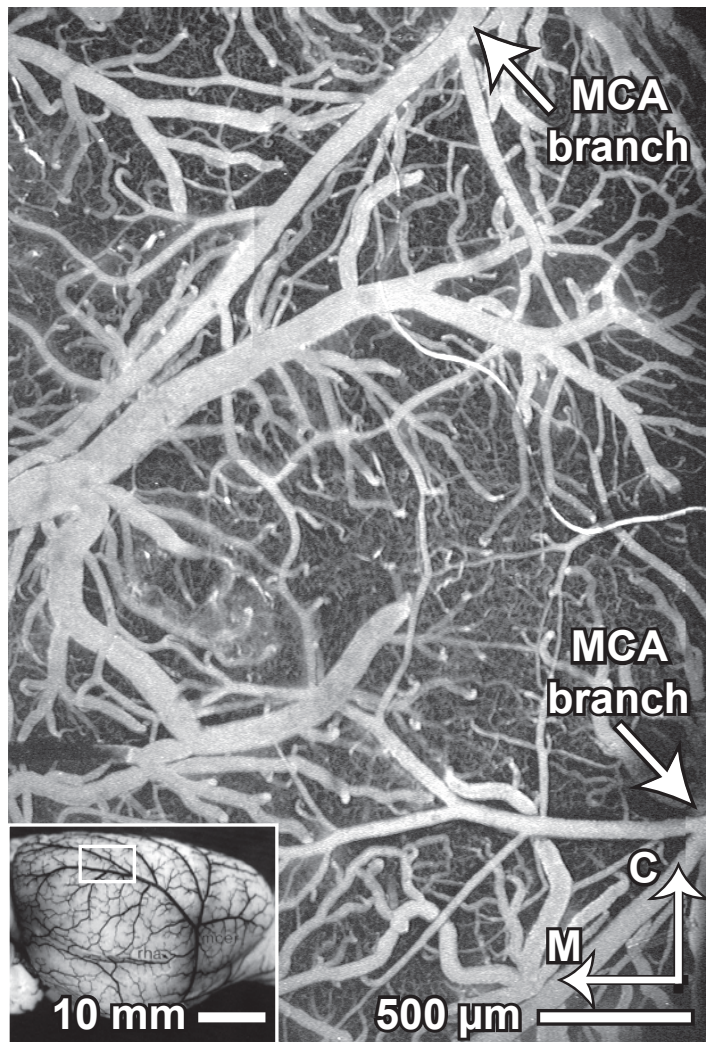


# Calculations support the robust nature of the surface network

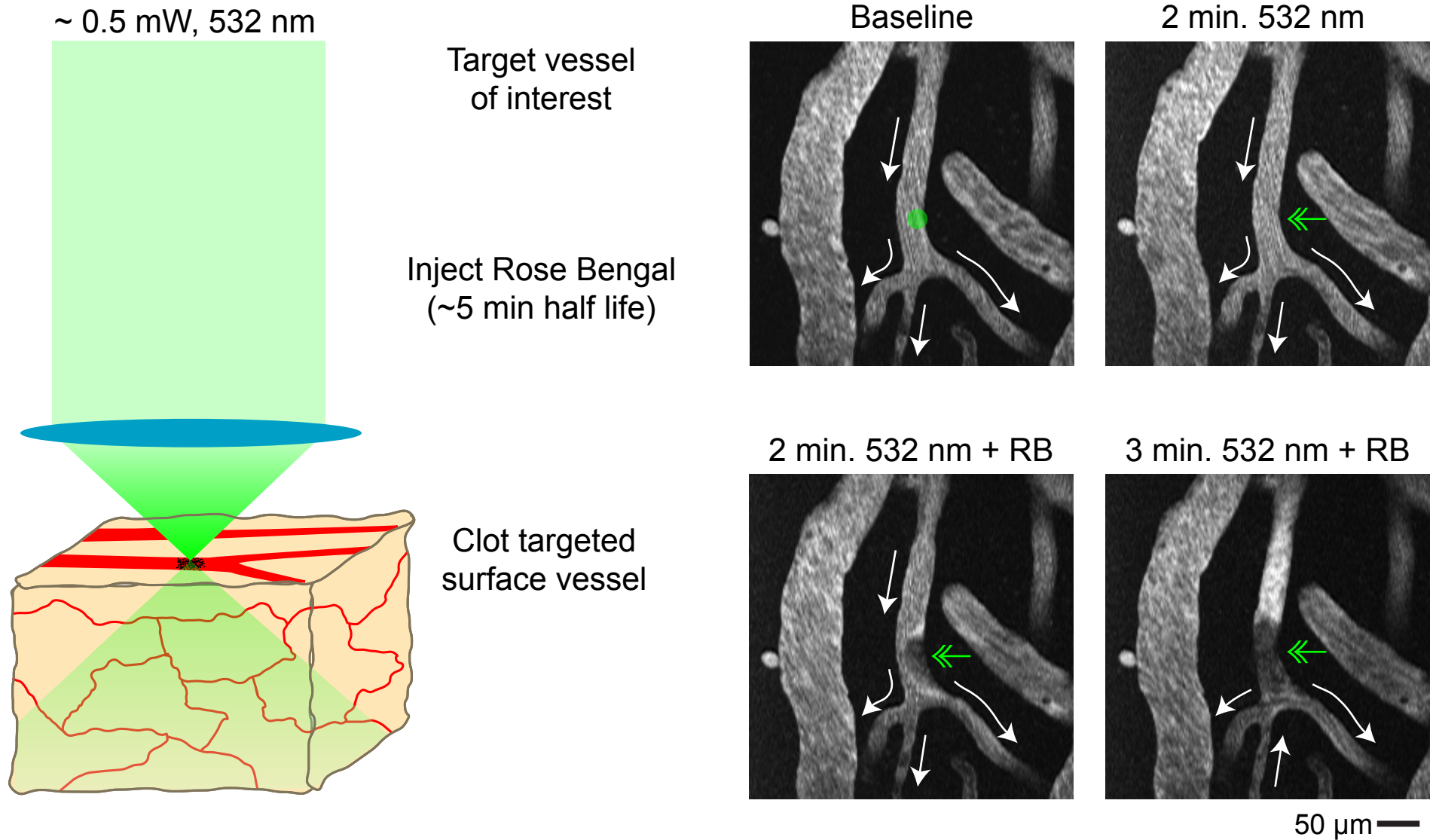


Prediction: Edges may be removed with minimal consequence to overall flow

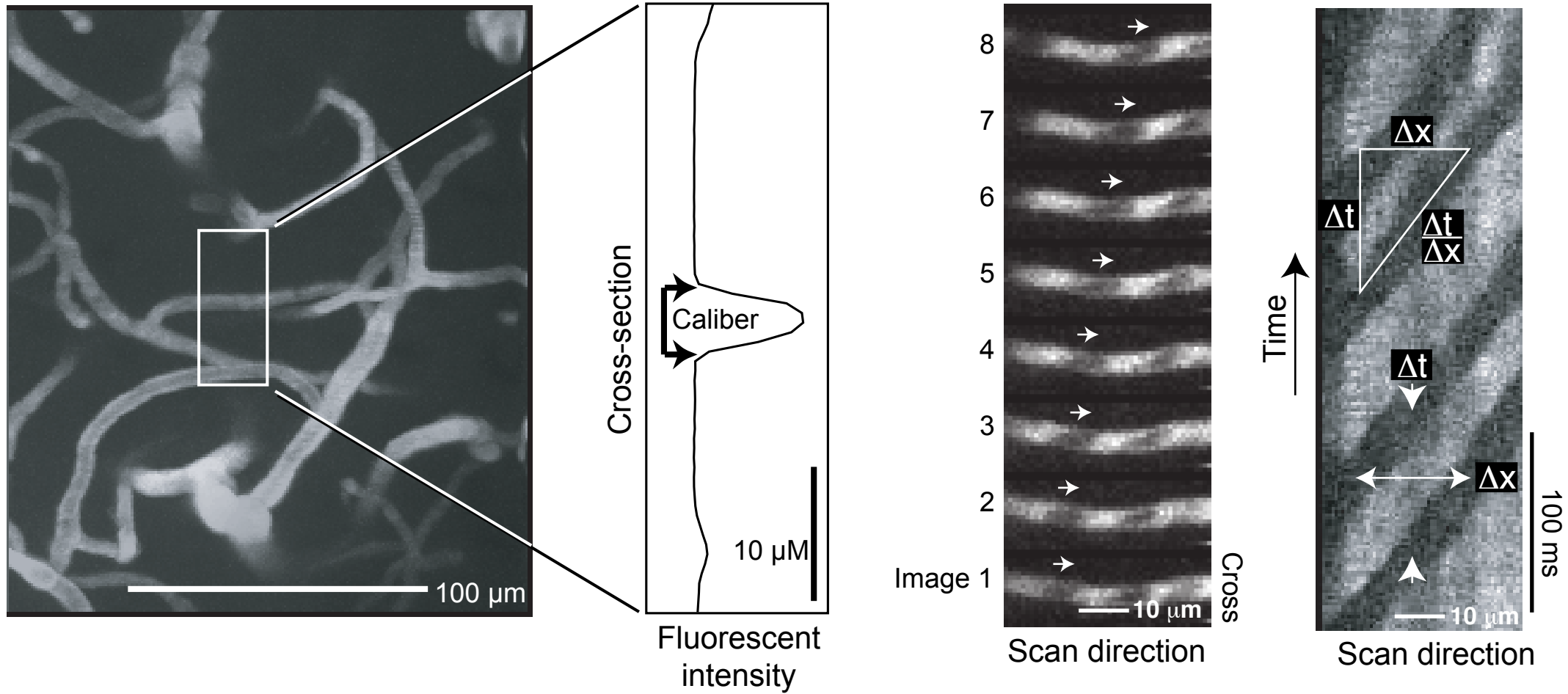
# Two-photon microscopy to image vasculature and target an occlusion



# Targeted photothrombotic occlusion to a selected surface vessel

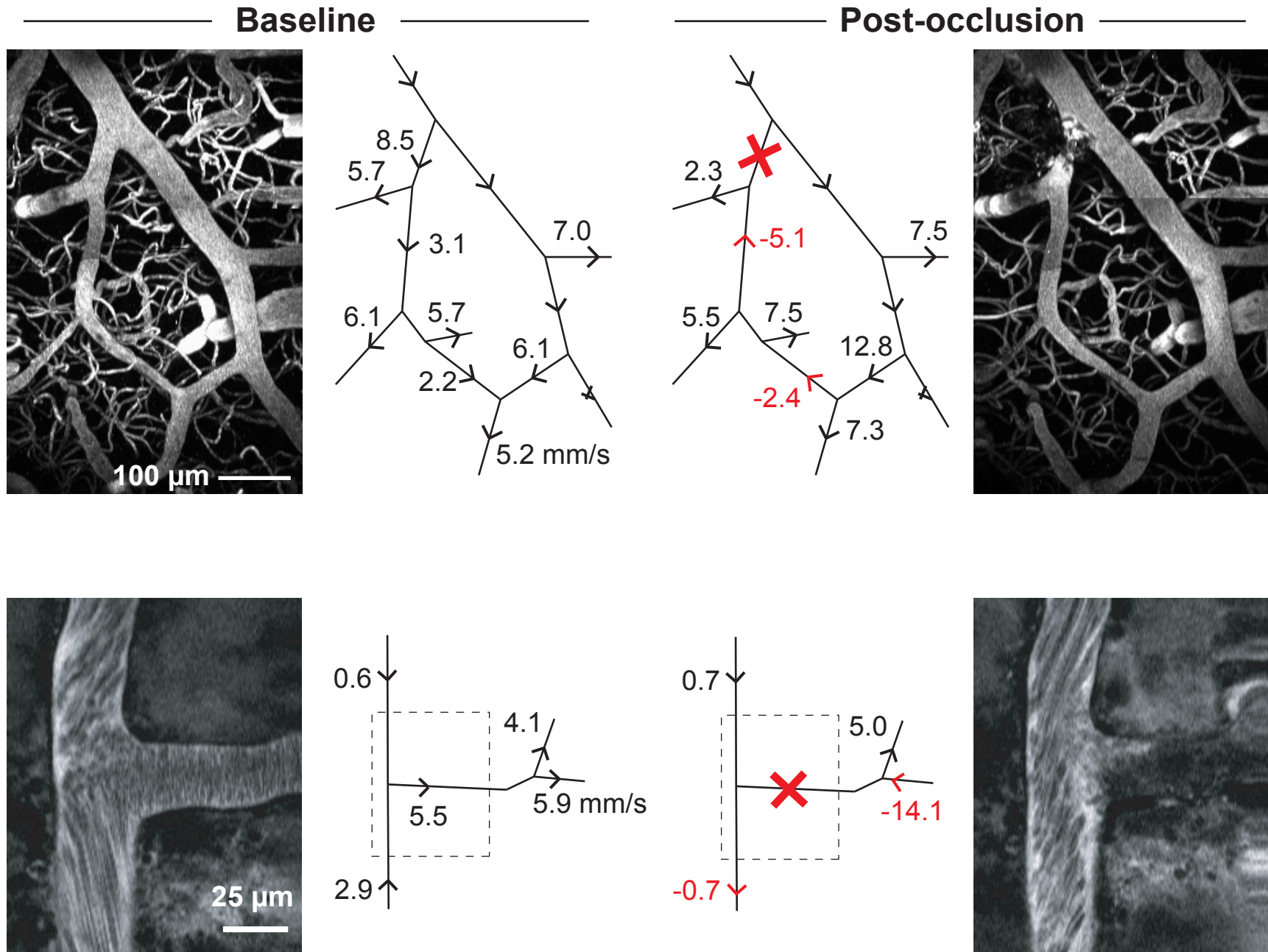


# *In vivo* two-photon microscopy for red blood cell tracking



Automatic calculation of speed  $v = (\Delta t / \Delta x)^{-1}$

# Reversal of RBC flow in downstream arterioles after targeted occlusion

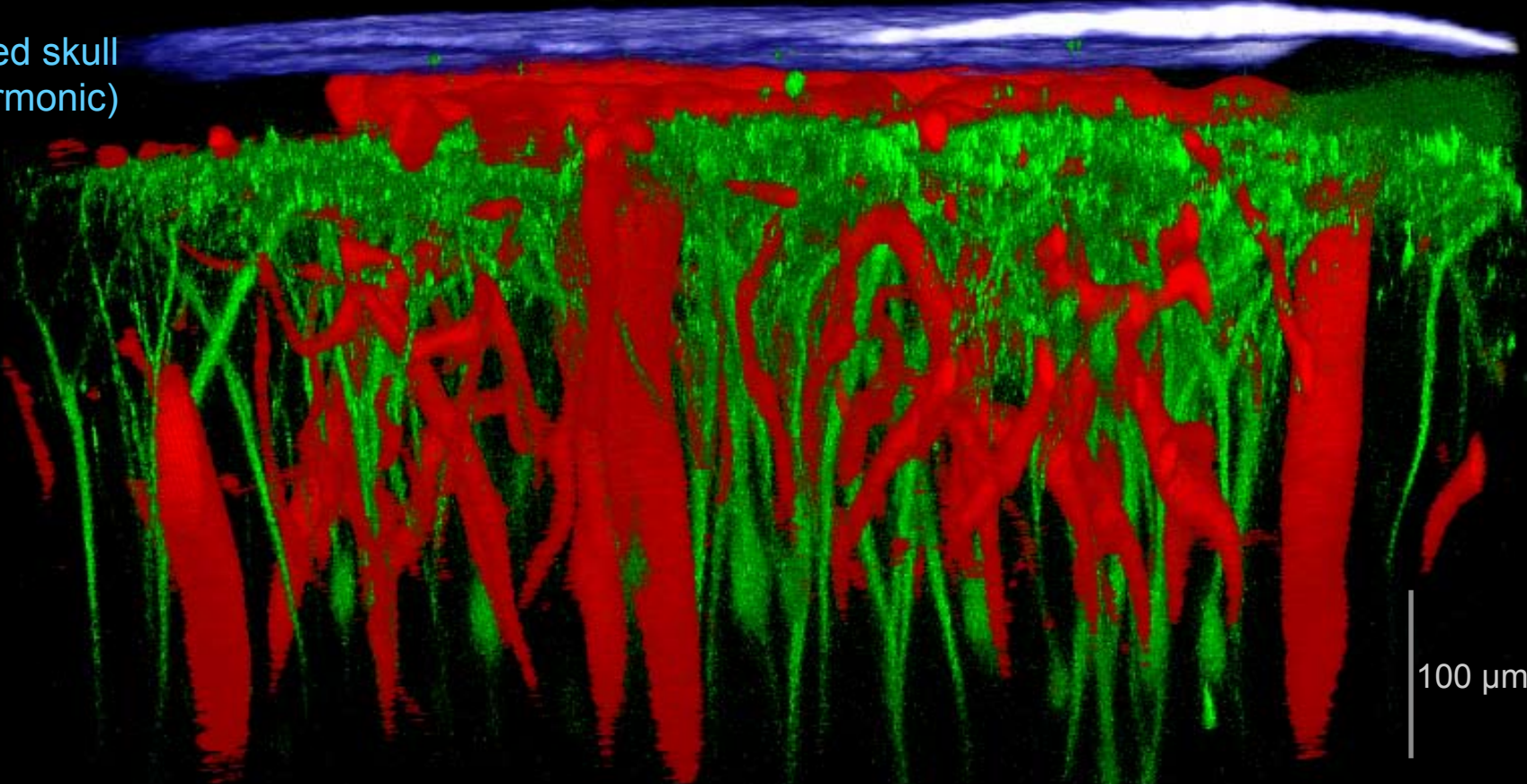


**Lesson: Blood flow in the surface arteriole network is robust.**

**We turn our attention to the routing of blood from the surface network to the subsurface microvasculature.**

# Polished and reinforced skull for chronic in vivo two-photon imaging

Thinned skull  
(second harmonic)



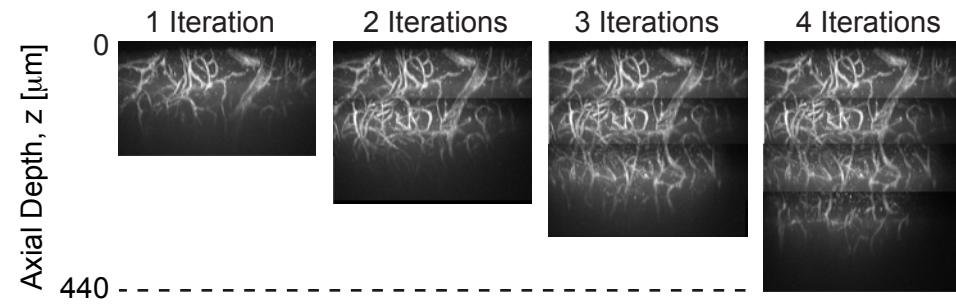
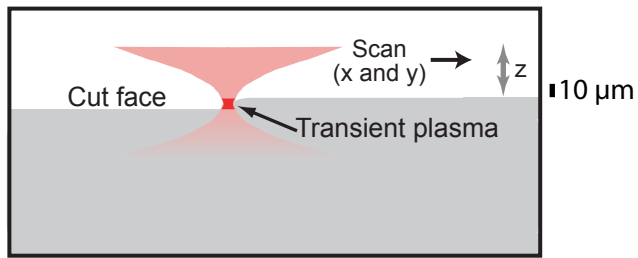
100  $\mu$ m

Blood vessels  
(fluorescein/dextran)

Neuronal dendrites  
(green fluorescent protein)

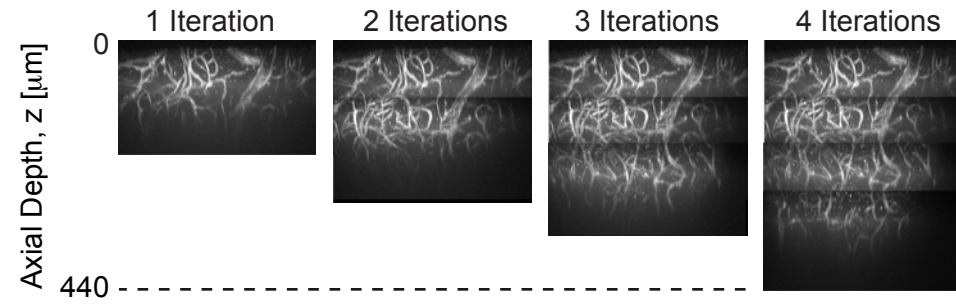
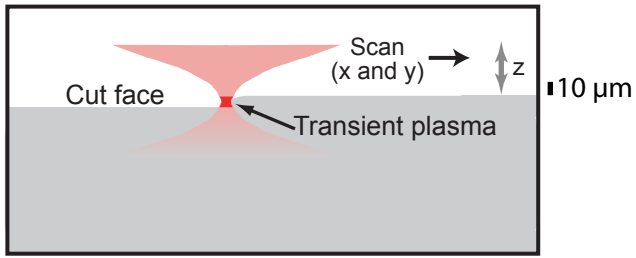


# All optical histology and automated tracing for vectorized anatomy

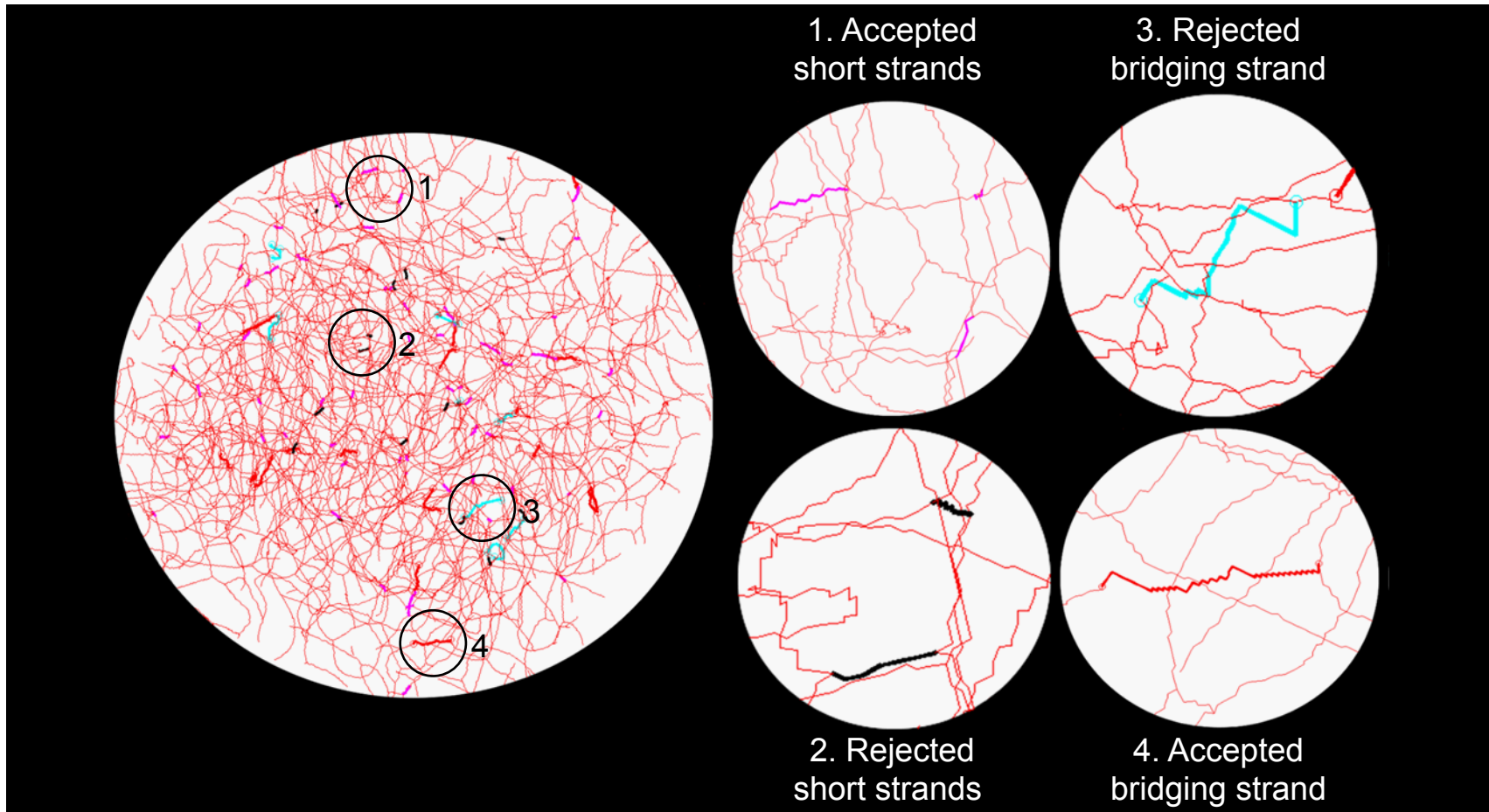


Tsai, Friedman, Ifarraguerri, Thompson, Lev-Ram, Schaffer, Xiong, Tsien, Squier and Kleinfeld (Neuron 2003)

# All optical histology and automated tracing for vectorized anatomy

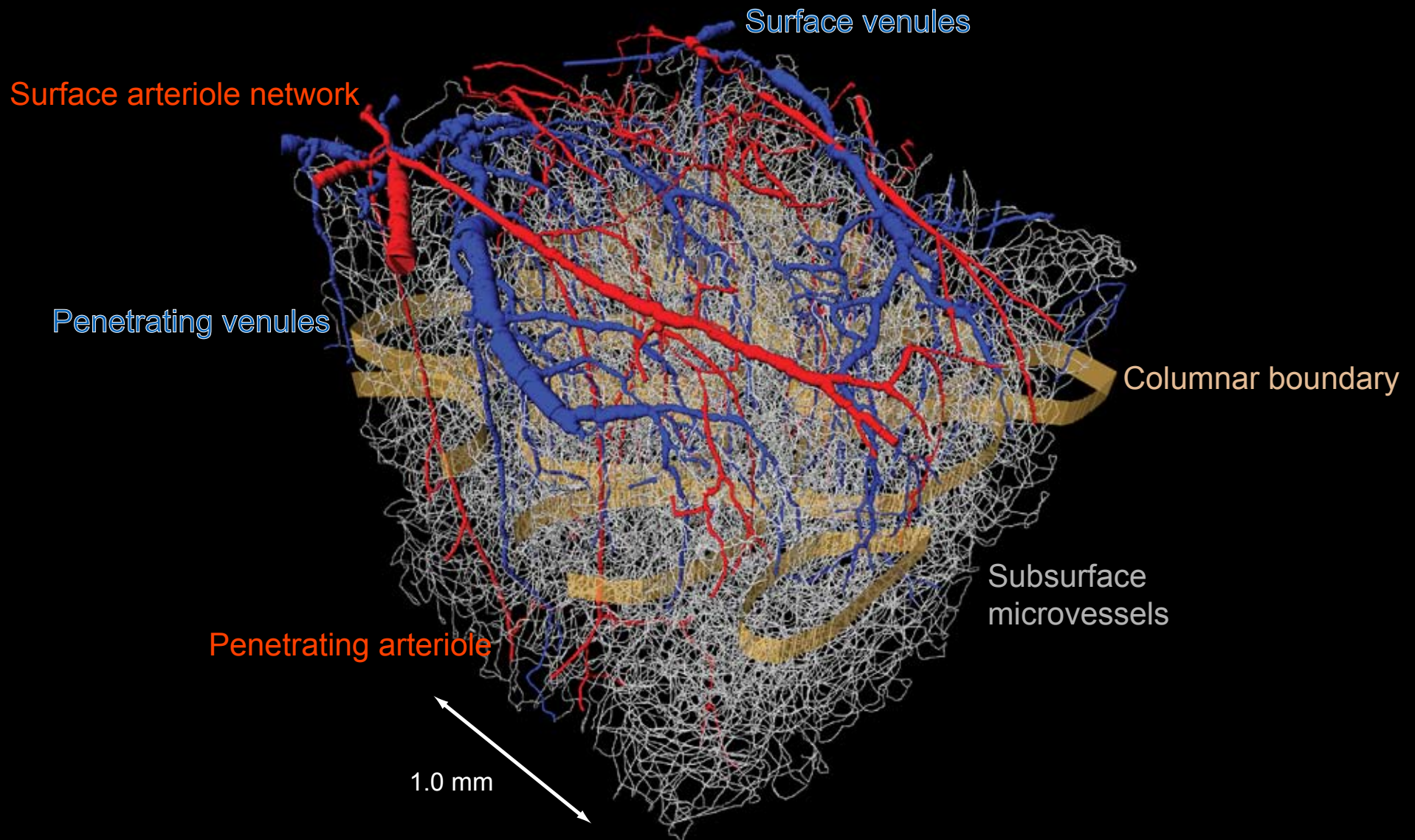


Tsai, Friedman, Ifarraguerri, Thompson, Lev-Ram, Schaffer, Xiong, Tsien, Squier and Kleinfeld (Neuron 2003)

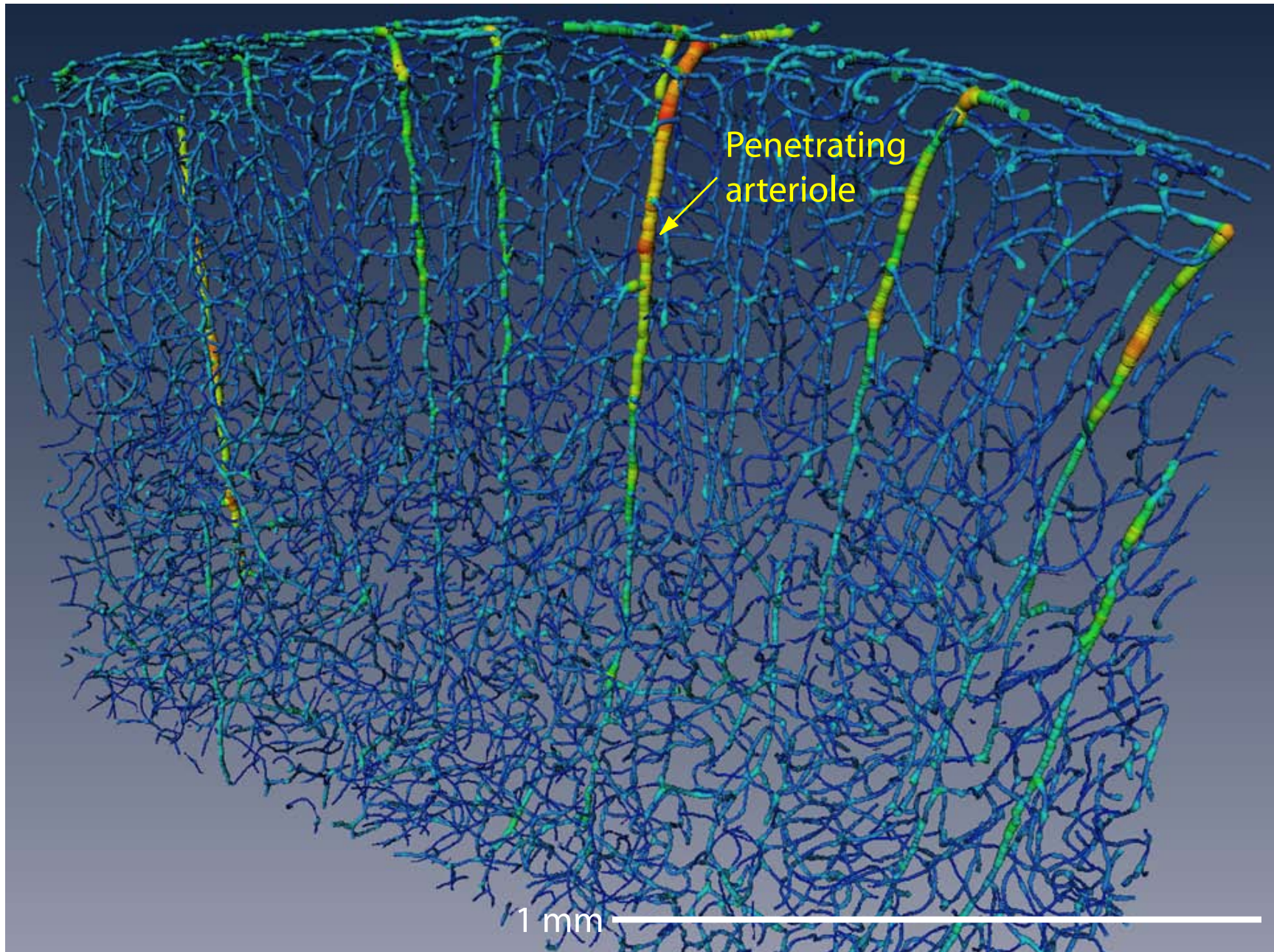


Kaufhold, Tsai, Blinder & Kleinfeld (MIAAB/MICCAI 2008; full paper submitted)

# Features of cortical vasculature from a 2 mm<sup>3</sup> angiotome

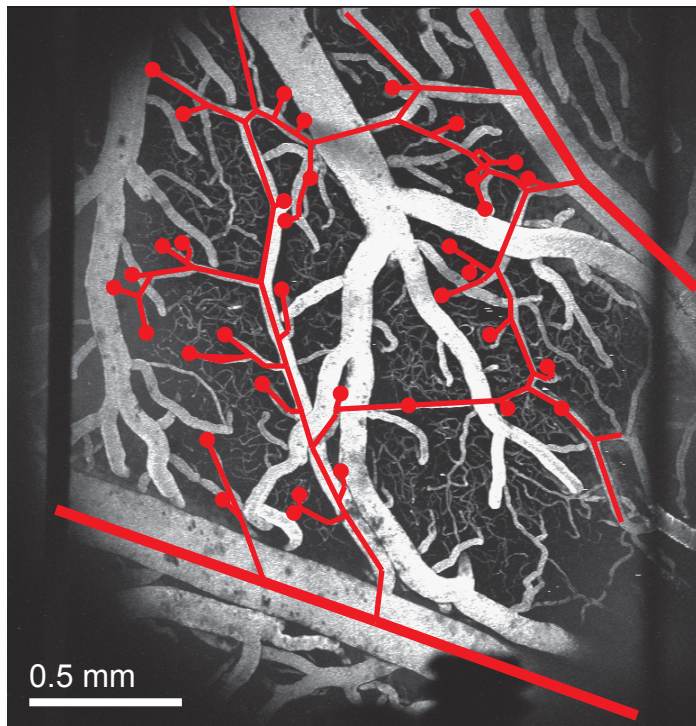


# Penetrating arterioles deliver blood to subsurface microvessels

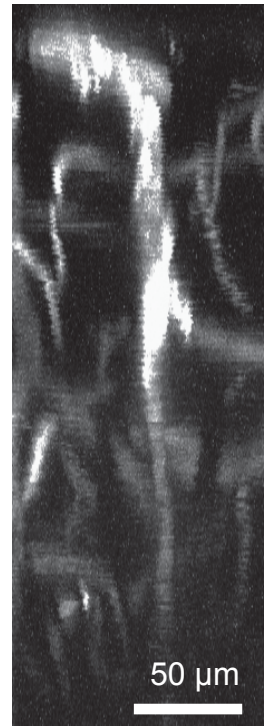
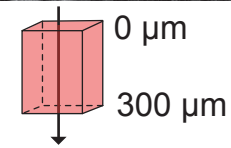
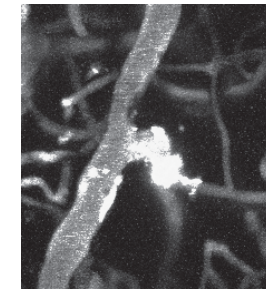
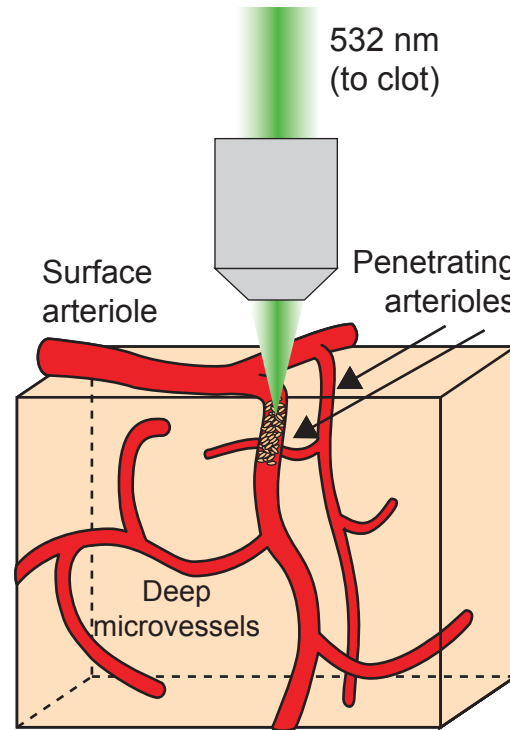


**Question: Is there collateral flow between neighboring penetrating arterioles?**

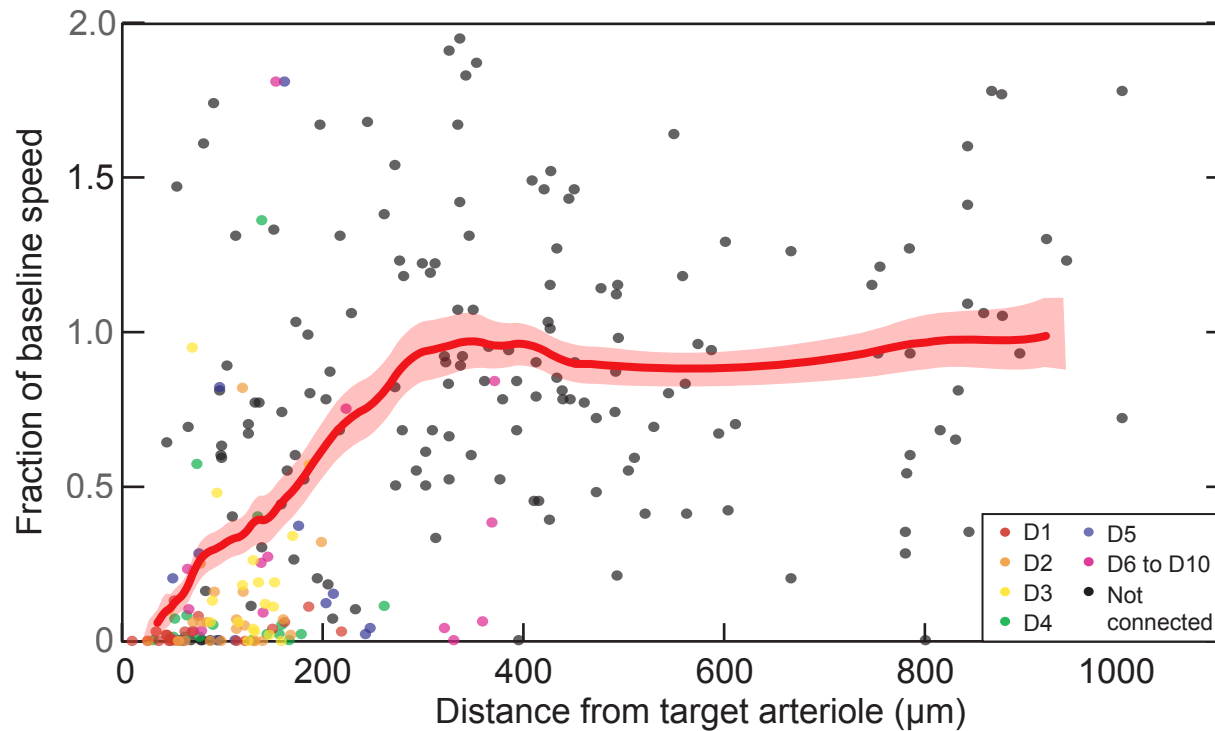
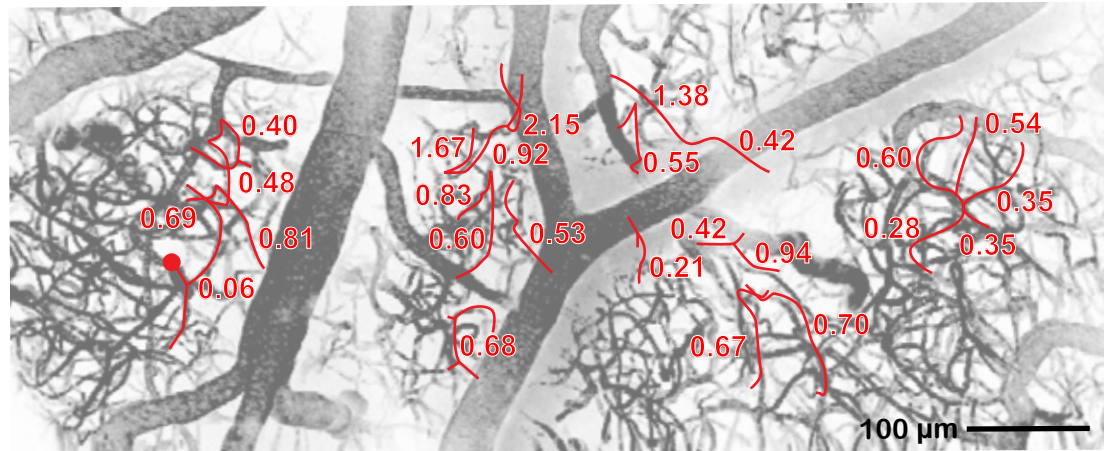
# TPLSM to target photothrombotic occlusion to a penetrating arteriole



- Penetrating arteriole



# Occlusion of a penetrating arteriole retards flow within a 400 to 600 $\mu\text{m}$ diameter “cylinder”



**Lesson: Penetrating arterioles are a bottleneck in the supply of blood to the subsurface microvasculature.**

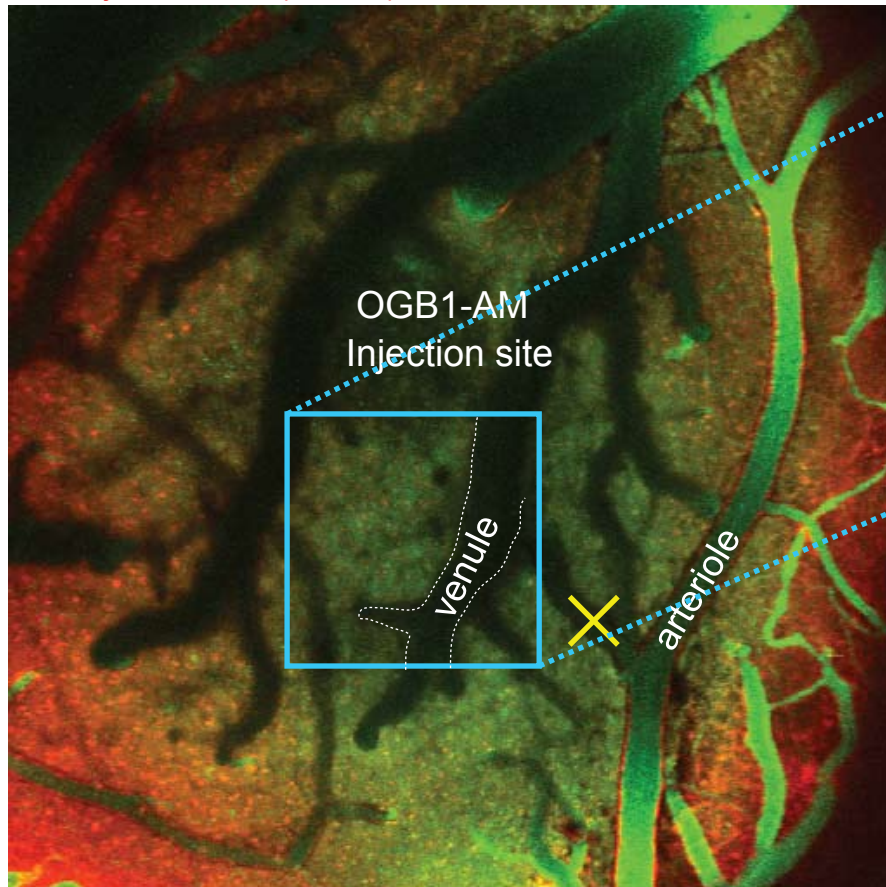
**Question: What is the chronic consequence of blockage of a single penetrating arteriole?**



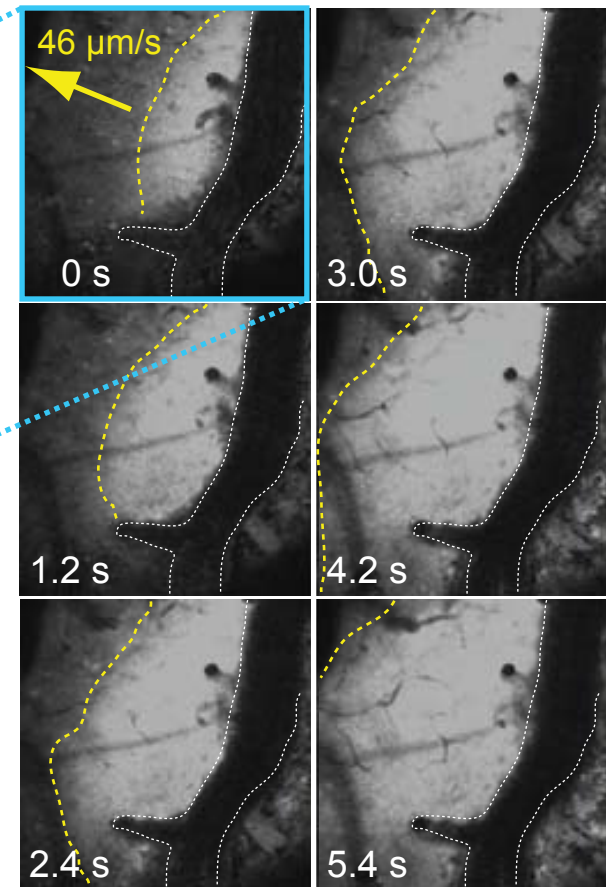
# Blockage of a single penetrating arteriole leads to waves of cortical spreading depression

## Pre-occlusion

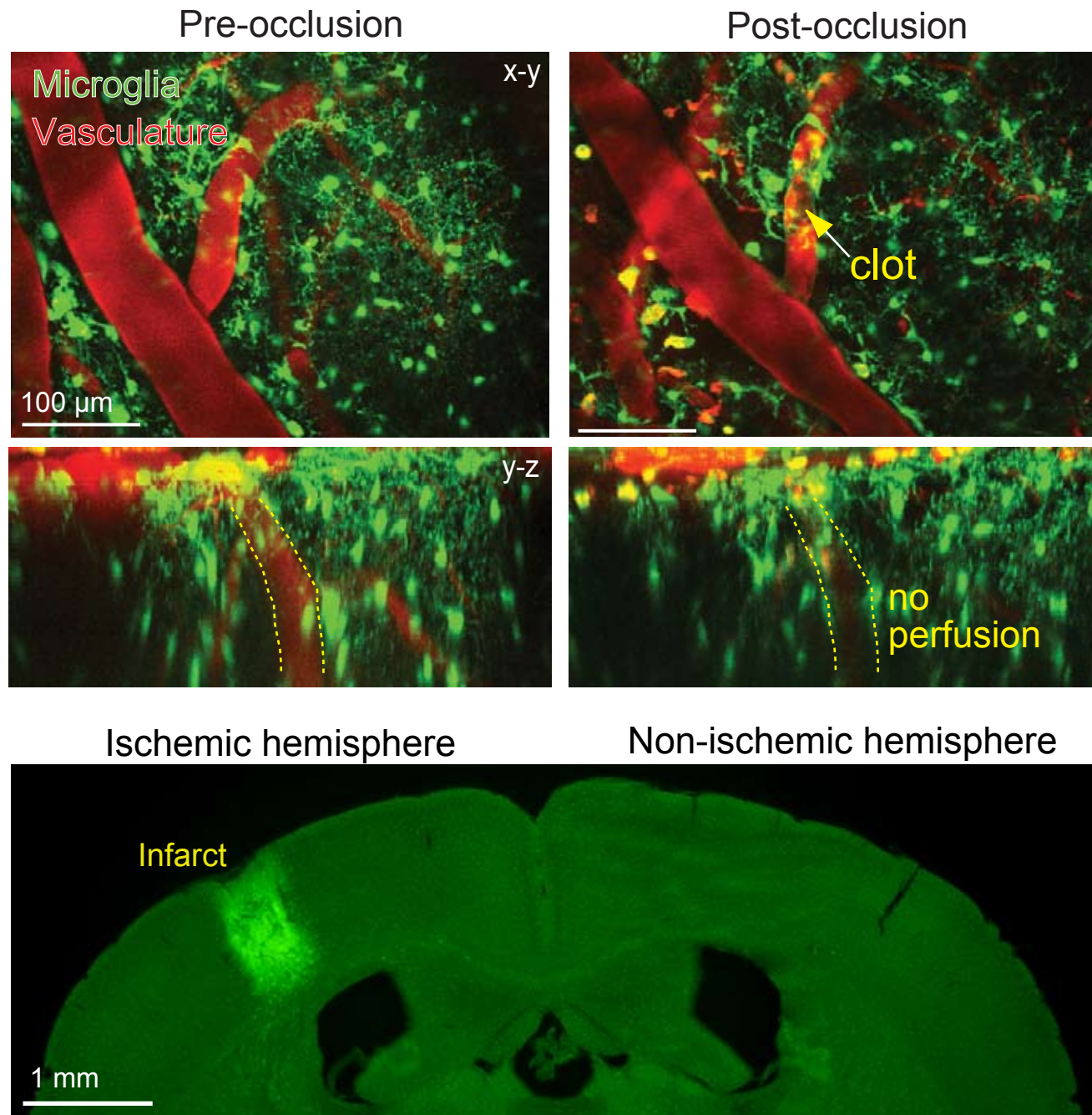
Cell calcium indicator (OGB1-AM or FITC)  
Astrocytes marker (SR101)



## Post-occlusion

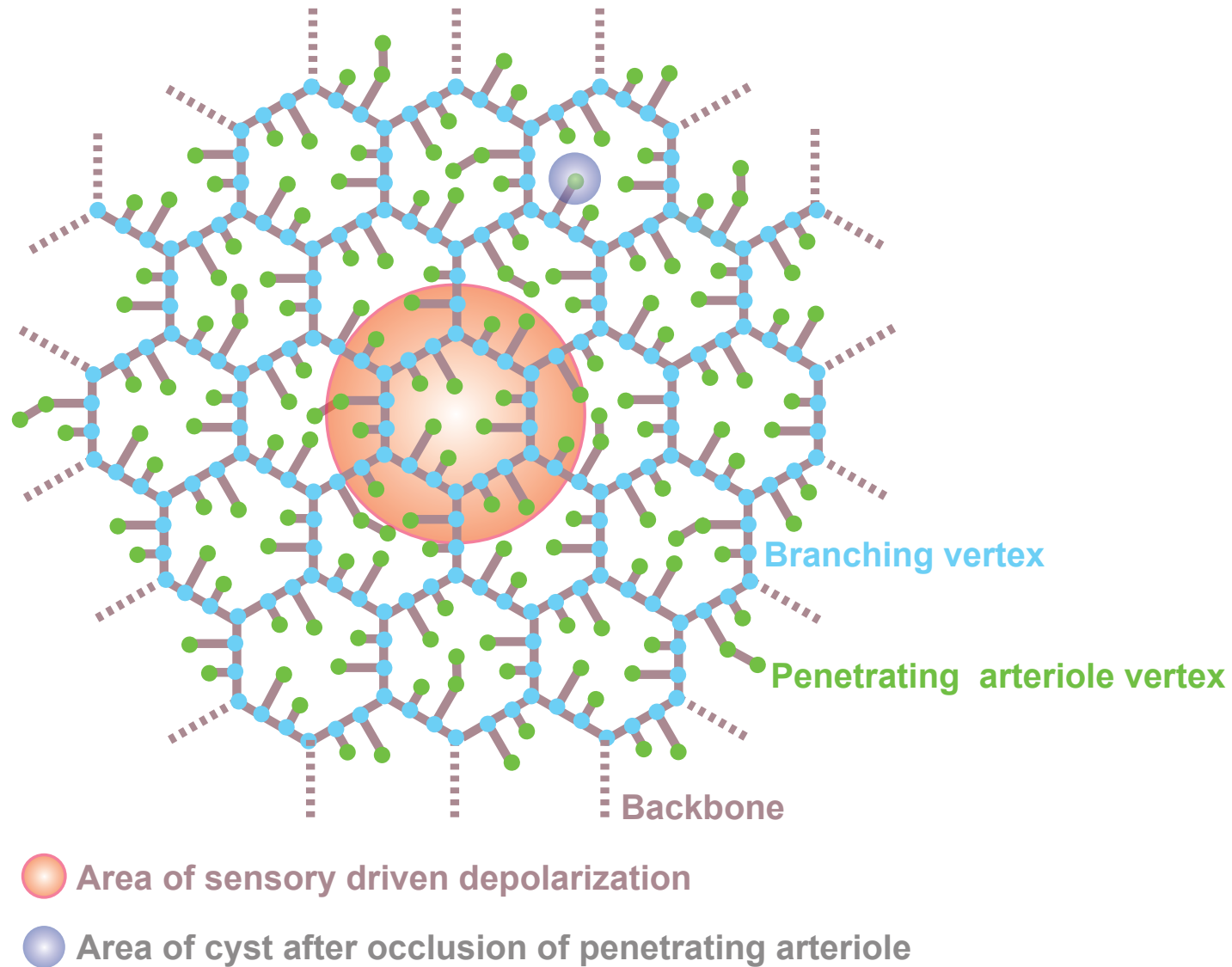


# Microglial invasion after occlusion of a single penetrating arteriole

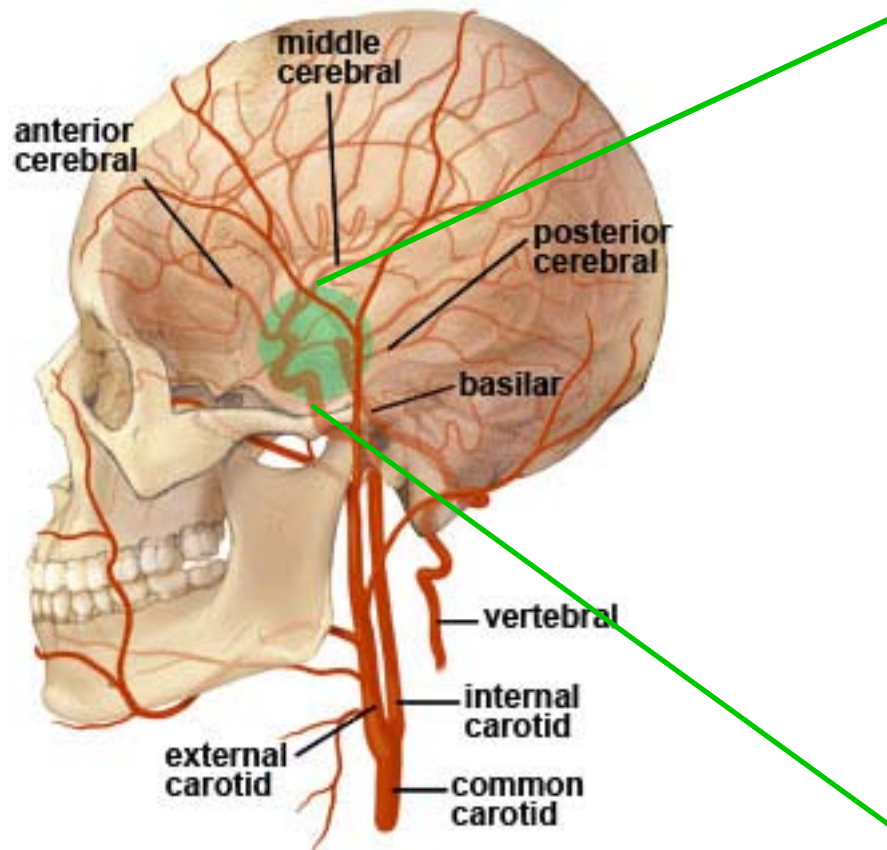


**Lesson: Blockage of a single penetrating arteriole leads to cell death throughout a region of cortex.**

# Summary of idealized topology and relative scales of the pial vasculature



# Surface loops add redundancy that is reminiscent of Circle of Willis

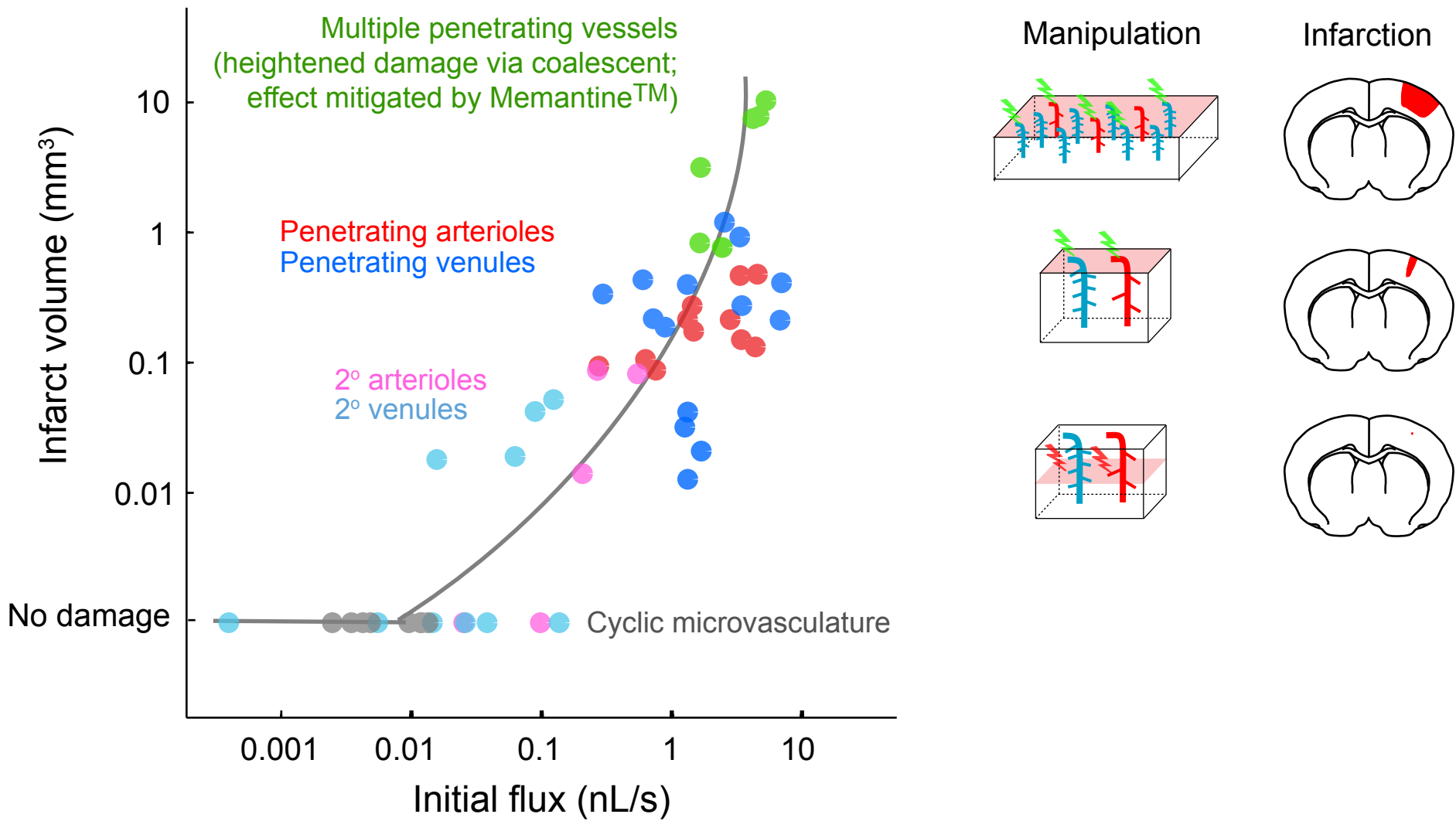


**These results open questions in biomedicine *and* biofluidics**

**Biomedicine: Are cortical micro-infarctions, seen in vascular dementia, a consequence of occluded penetrating vessels?**

**Biofluidics: Does each penetrating arteriole have an exclusive territory or does the subsurface microvasculature form a continuous network?**

# Hierarchy of microinfarcts from occlusions to 1-D penetrating vessels or their immediate downstream branches



Schaffer, Friedman, Nishimura, Schroeder, Tsai, Ebner, Lyden & Kleinfeld (PLoS Biology 2006); Shih, Friedman, Drew, Blinder, Lyden & Kleinfeld (JCBFM 2009); Shih, Tsai, Driscoll, Blinder, Drew, Friedman, Lyden & Kleinfeld (SfN abstract 2010)

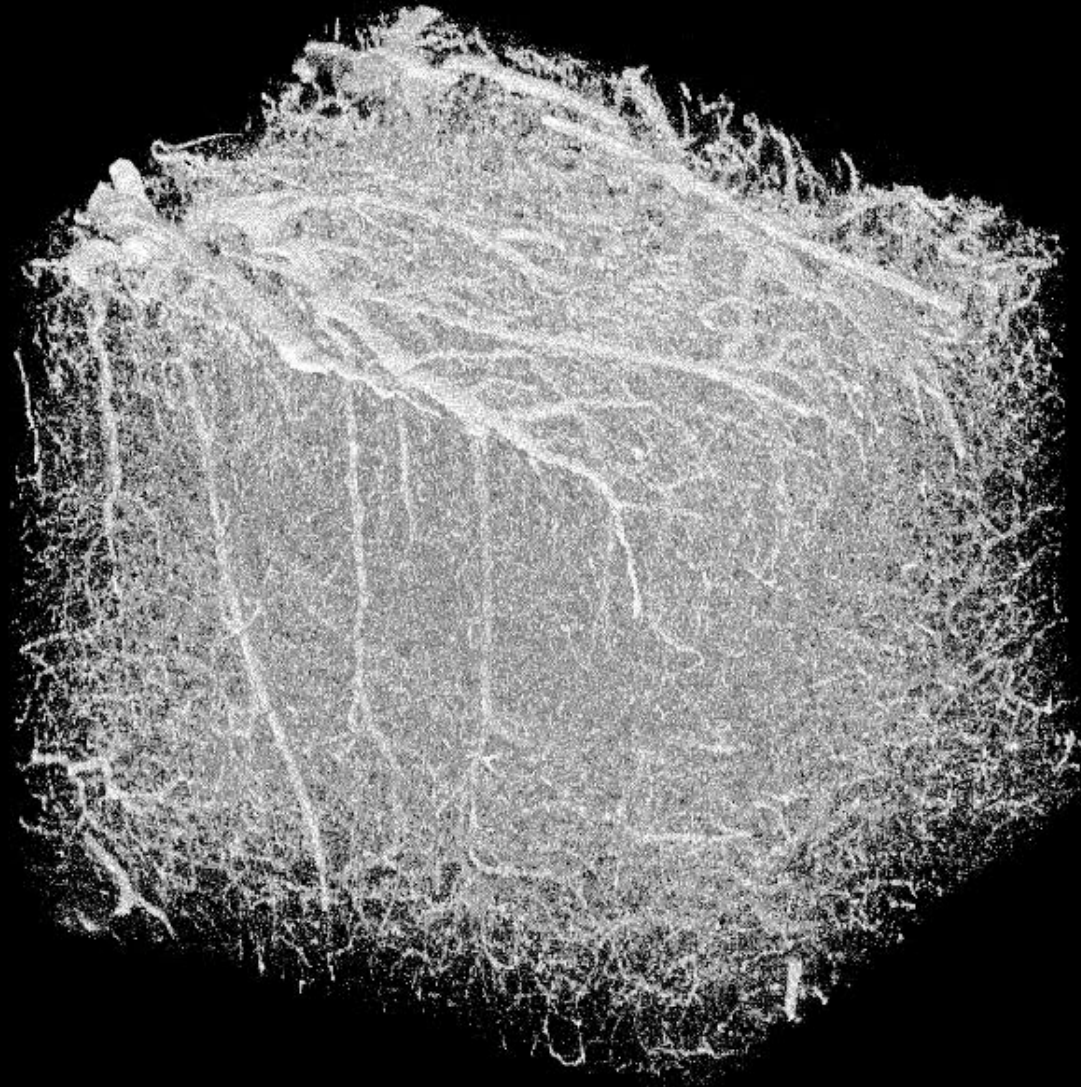
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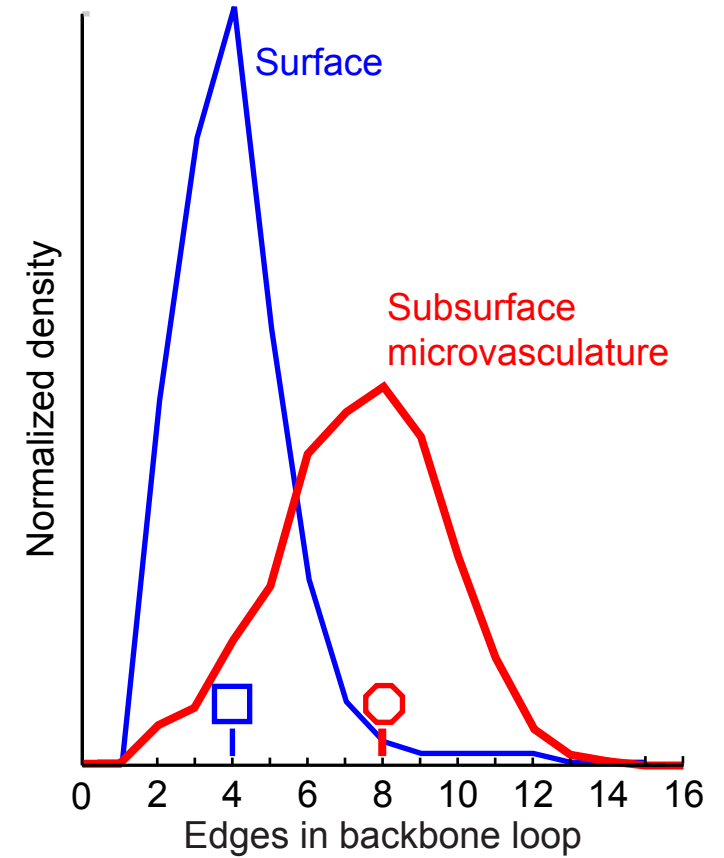
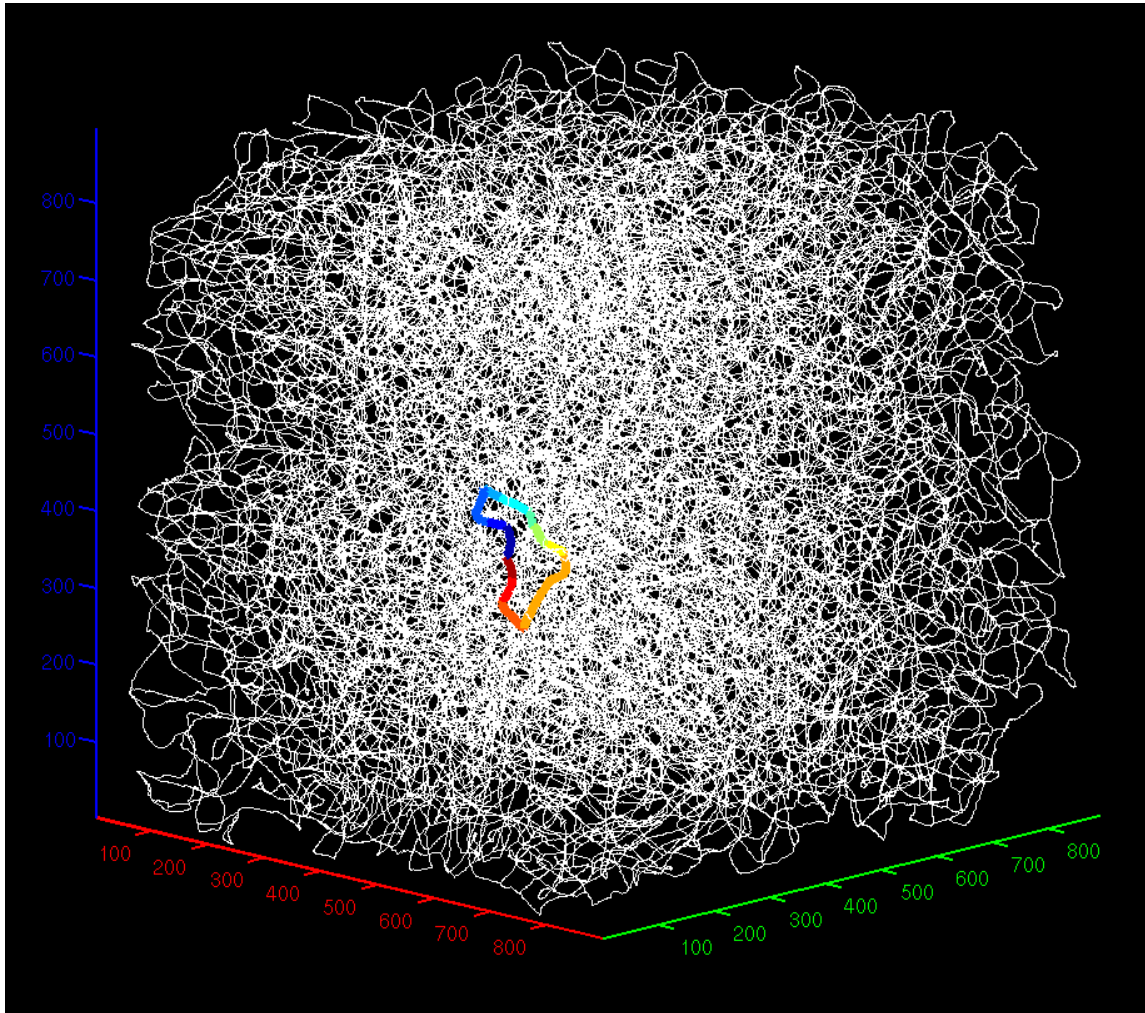
**Biofluidics: Does each penetrating arteriole have an exclusive territory or does the subsurface microvasculature form a continuous network?**



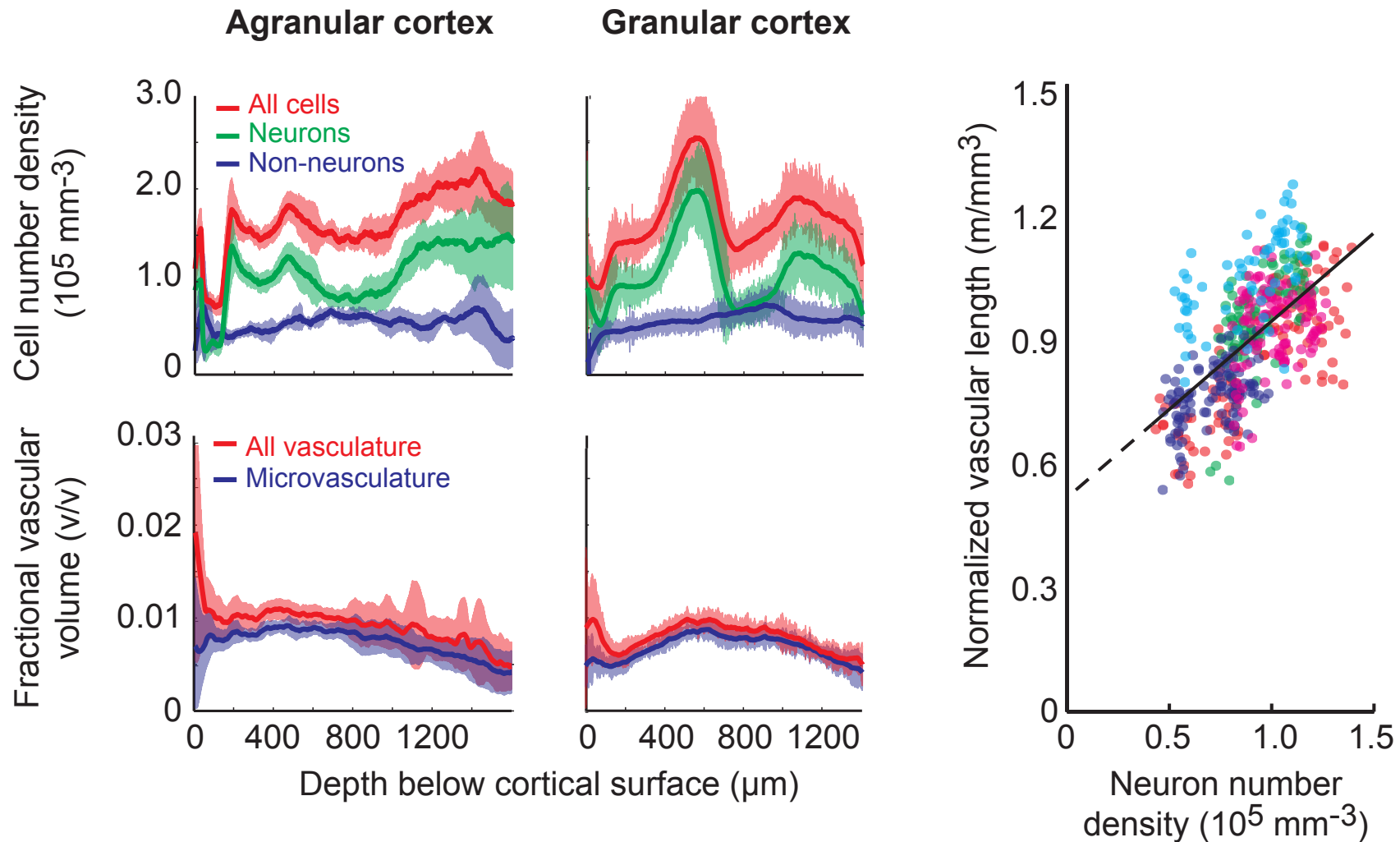
# Highlights of the rodent angiotome: A work in progress



# Subsurface microvessels form a highly interconnected network



# Statistics of vessels and cells in mouse neocortex from a complete inventory across 1 - 10 mm<sup>3</sup> slabs

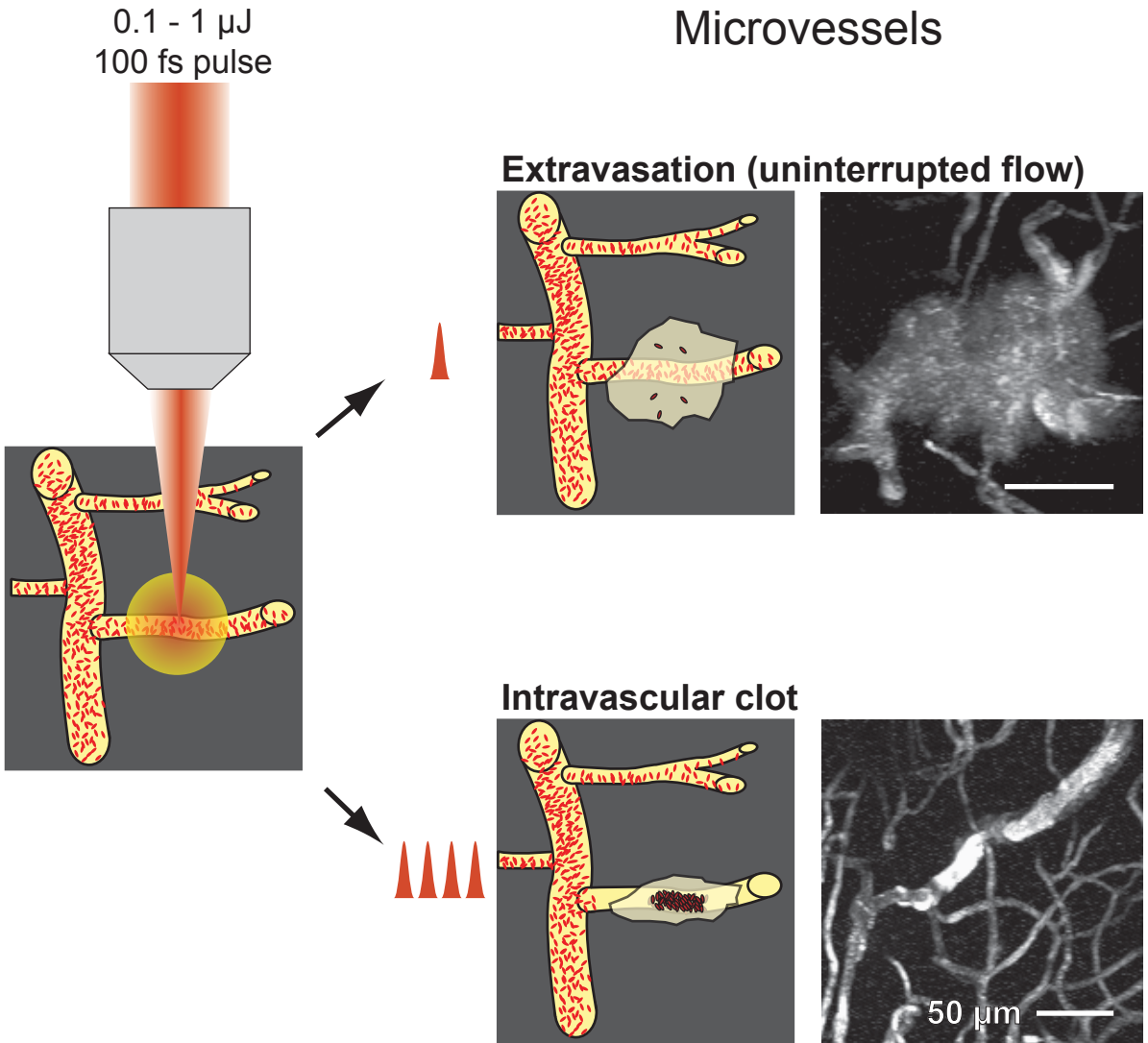


The set of shortest cycles in layer IV form a single connected graph



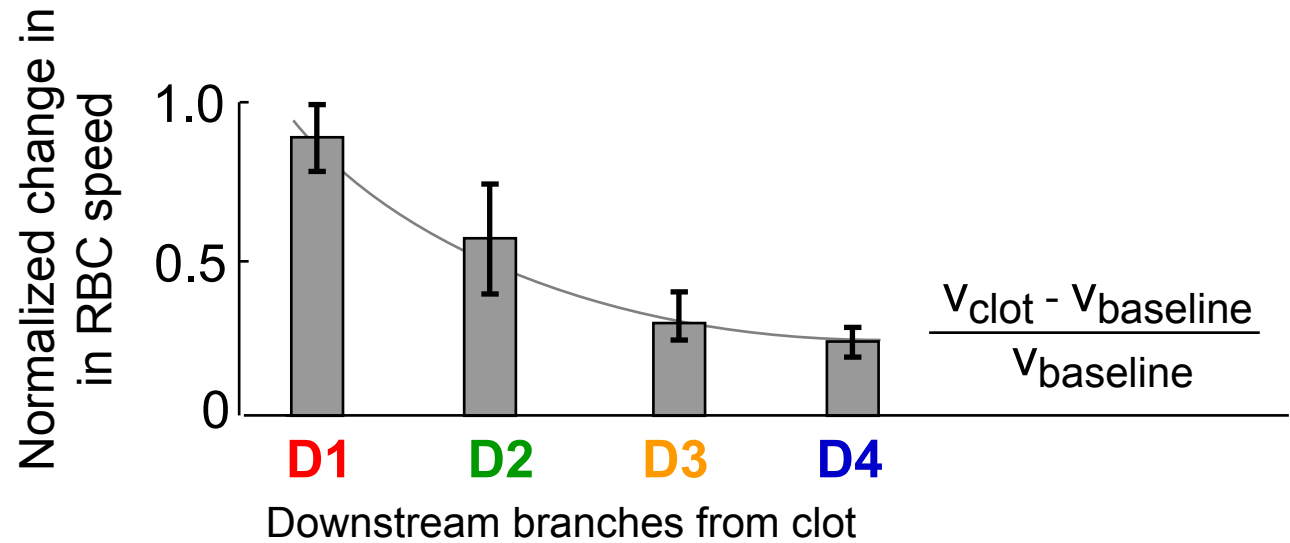
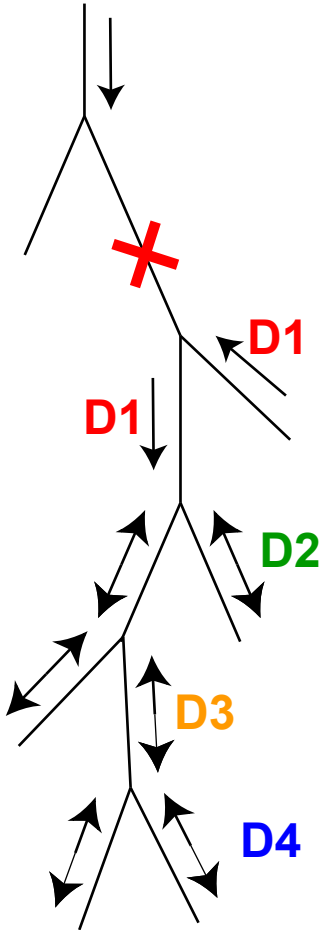
**Question: What is the consequence of blockage to a single microvessel, *i.e.*, do subsurface loops protect local flow?**

# Targeted disruption of subsurface microvessels by amplified laser pulses



Nishimura, Schaffer, Friedman, Tsai, Lyden & Kleinfeld (Nat Meth 2006)  
Tsai, Blinder, Migliori, Neev, Jin, Squier & Kleinfeld (Cur Opin Biotech 2009)

# Targeted occlusion to a subsurface microvessel blocks flow only in nearest, but not nearby, downstream branches



Consistent with long microvascular loops

**Lesson: Microvessels form closed loops that span multiple cortical columns – no evidence for exclusive territories.**

**Question: Why is flow spatially restricted even though we have complete lateral connectivity?**



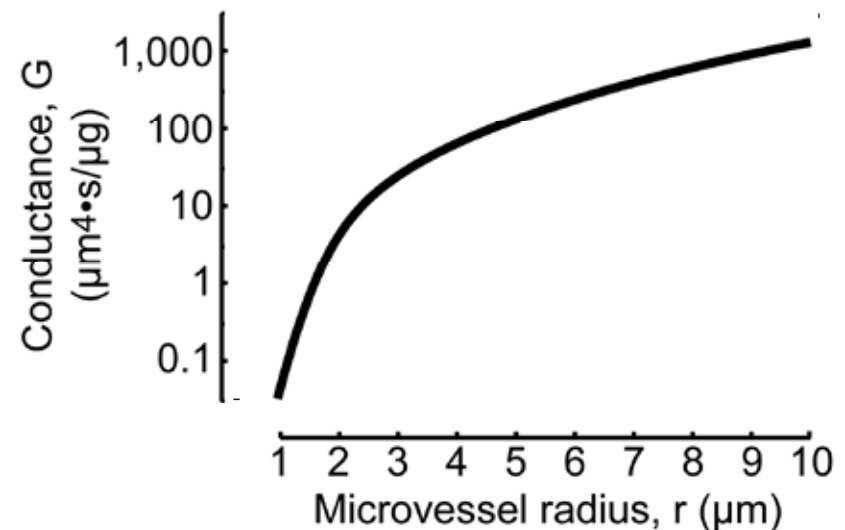
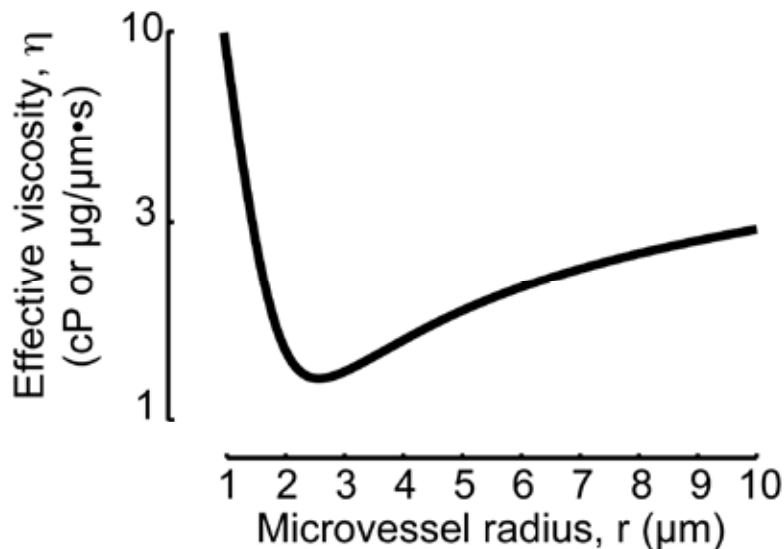
# Flow calculated between pairs of vertices using Kirchhoff's Law

Volume flux = [Conductance] (Pressure at vertex)

$$F = G P$$

For each pair of vertices (n,m), the measured vessel length  $l_{nm}$  and radius  $r_{nm}$  form the elements of  $\mathbf{G}$

$$G_{nm} = \frac{\pi}{8} \frac{r_{nm}^4}{\eta(r_{nm})} \frac{1}{l_{nm}} \quad \text{with} \quad G_{nn} = - \sum_{m \neq n}^{\text{all vessels}} G_{nm}$$



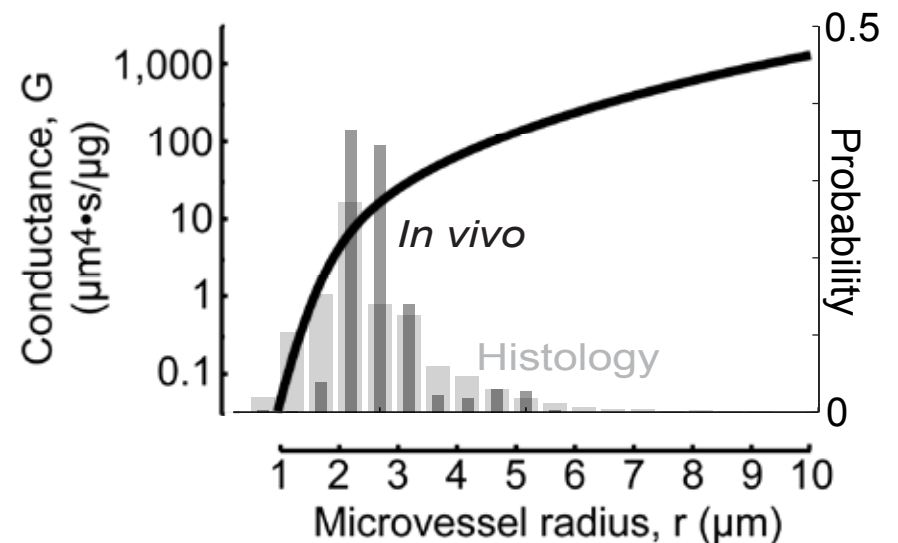
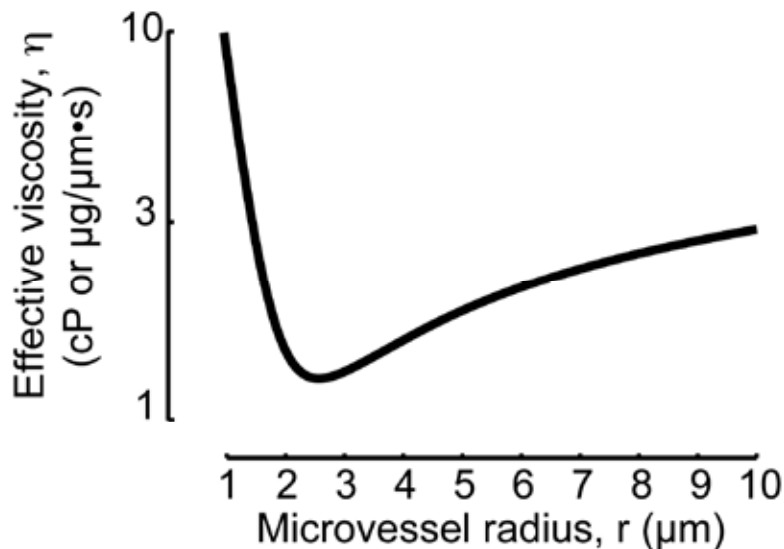
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$$G_{nm} = \frac{\pi}{8} \frac{r_{nm}^4}{\eta(r_{nm})} \frac{1}{l_{nm}} \quad \text{with} \quad G_{nn} = - \sum_{m \neq n}^{\text{all vessels}} G_{nm}$$

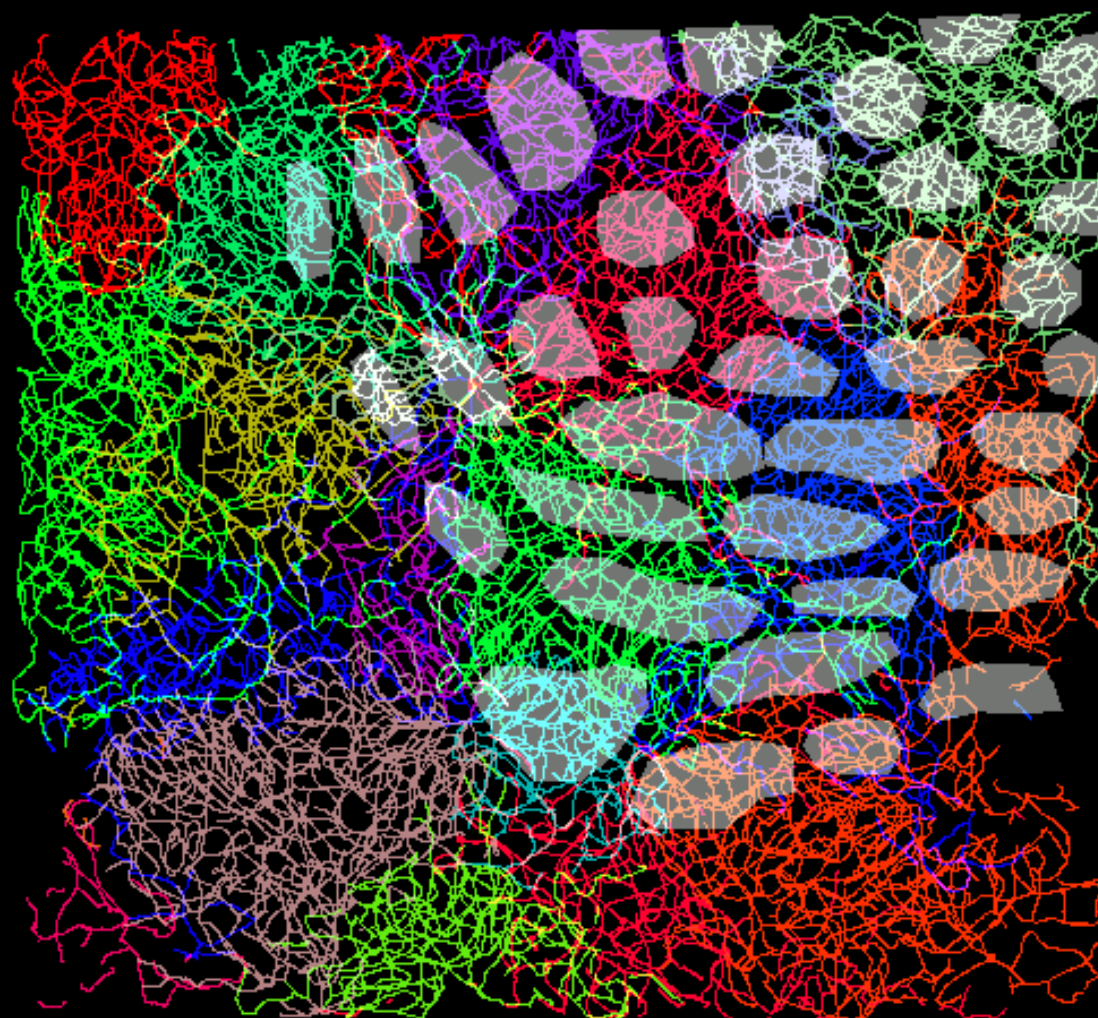
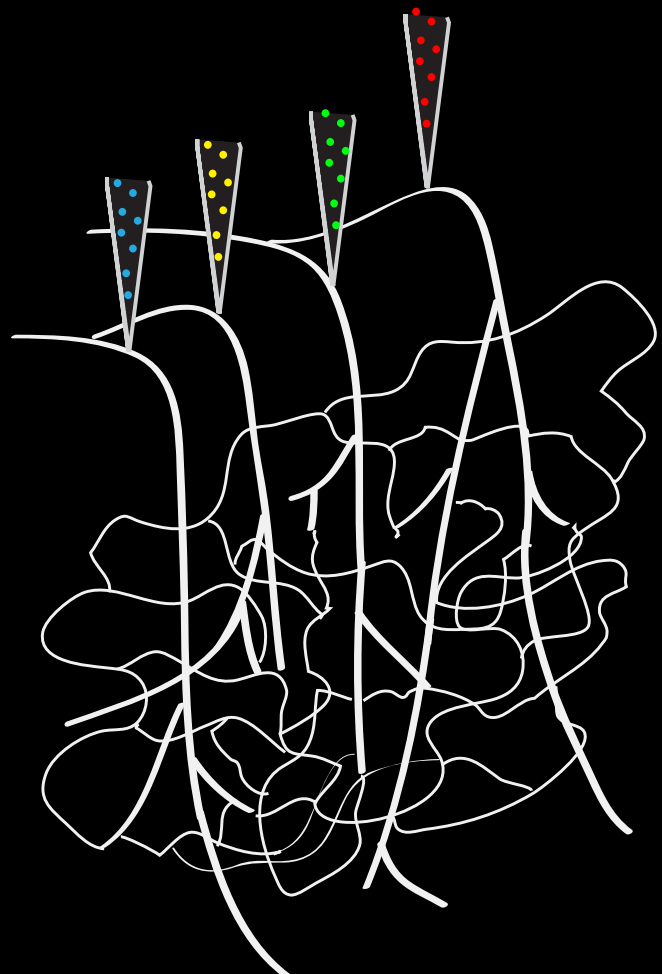



Pries & Secomb (American Journal of Physiology HCP 2005)

Tsai, Kaufhold, Blinder, Friedman, Drew, Karten, Lyden & Kleinfeld (J Neuroscience 2009)

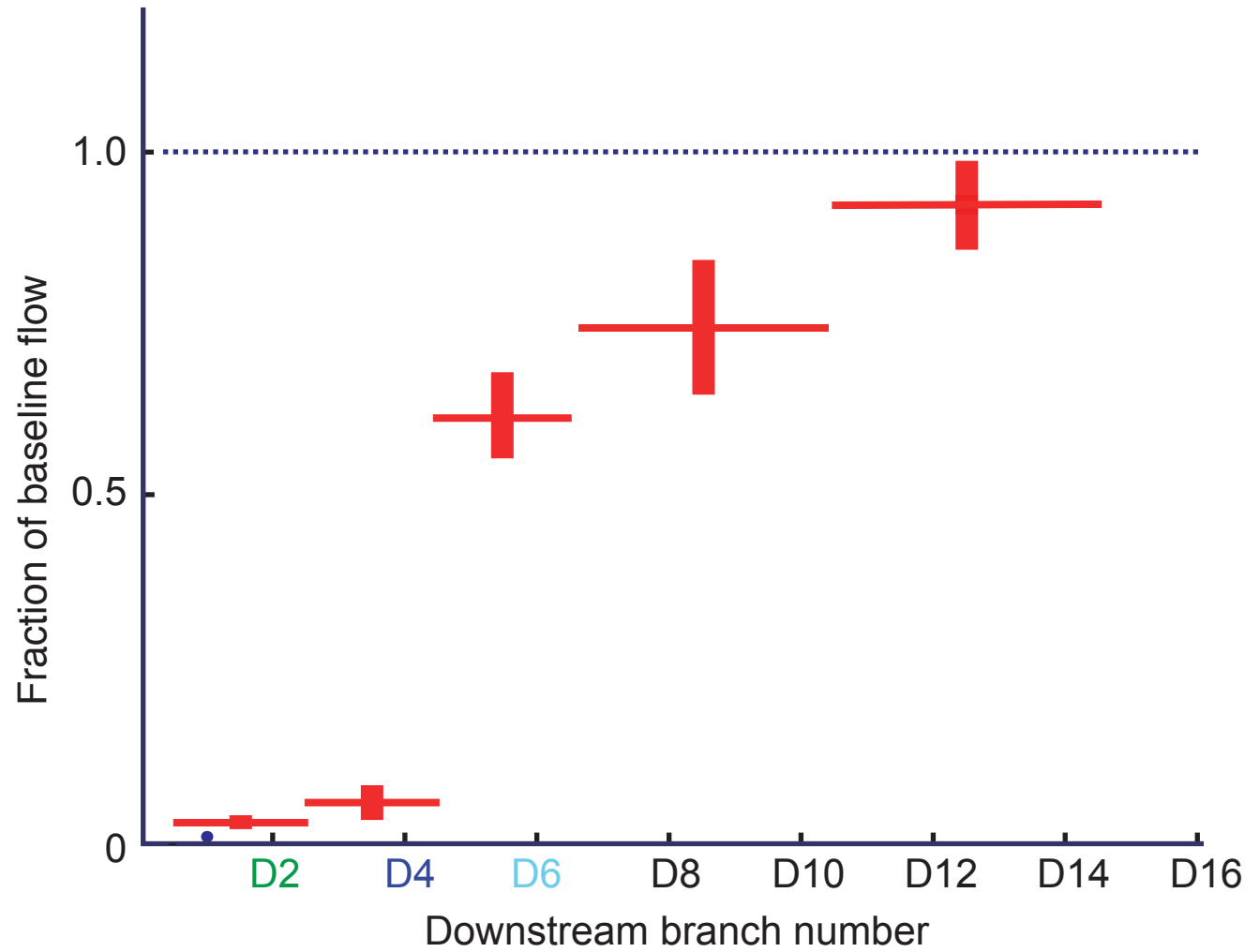
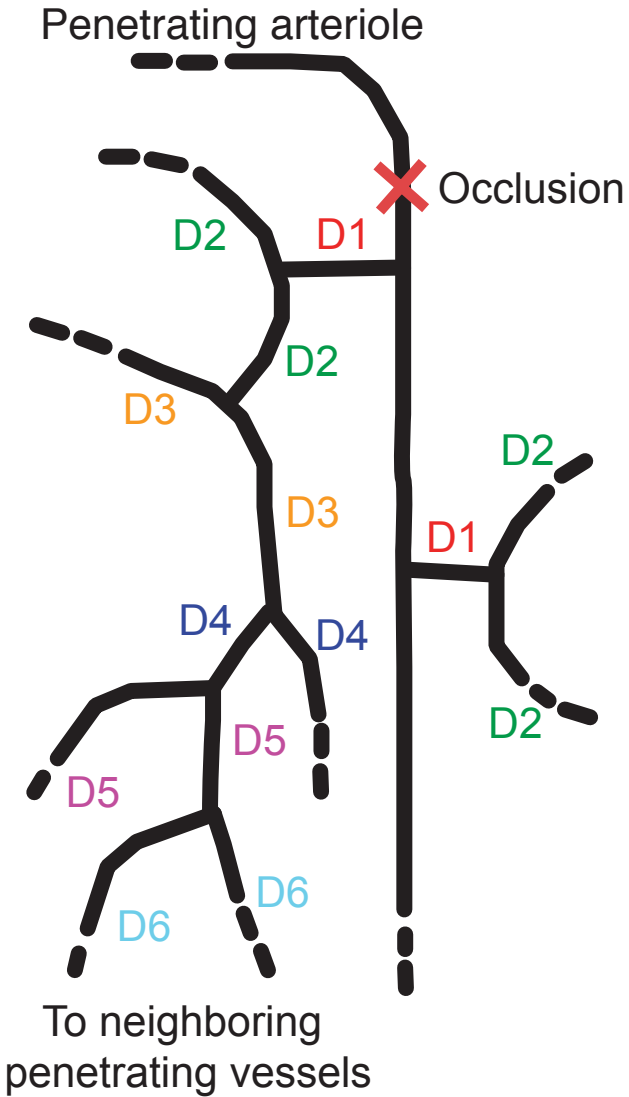
# Numerical analysis of “dye” injection to determine perfusion domains of individual penetrating arterioles


L4 (sensory input layer)



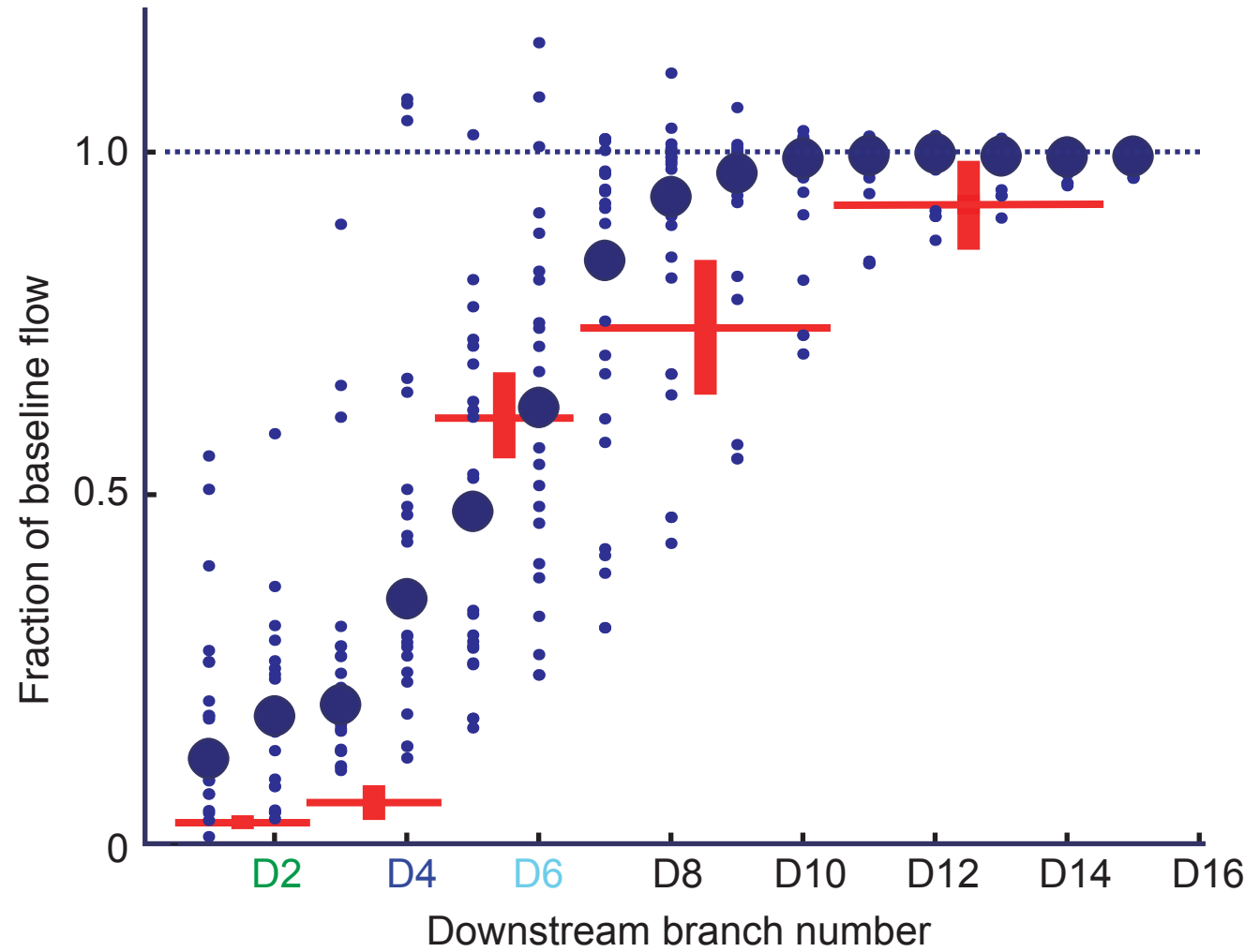
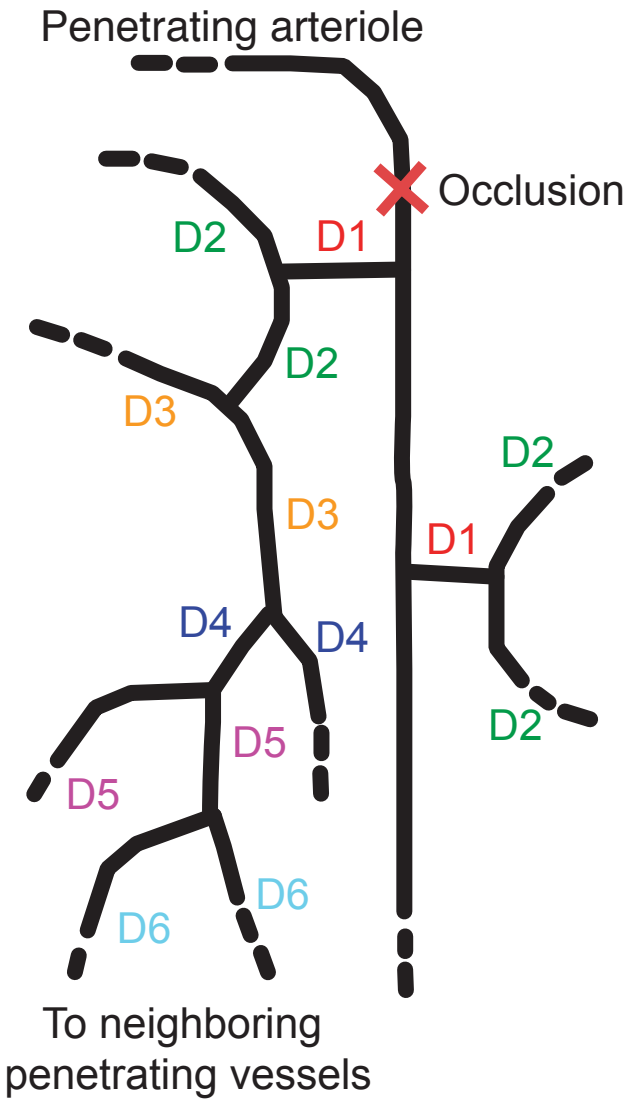
100  $\mu\text{m}$  



# Passive flow models the effect of an occlusion to a penetrating arteriole



 *In vivo* rat data of Nishimura, Rosidi, Iadecola & Schaffer (JCBFM 2010)

# Passive flow models the effect of an occlusion to a penetrating arteriole



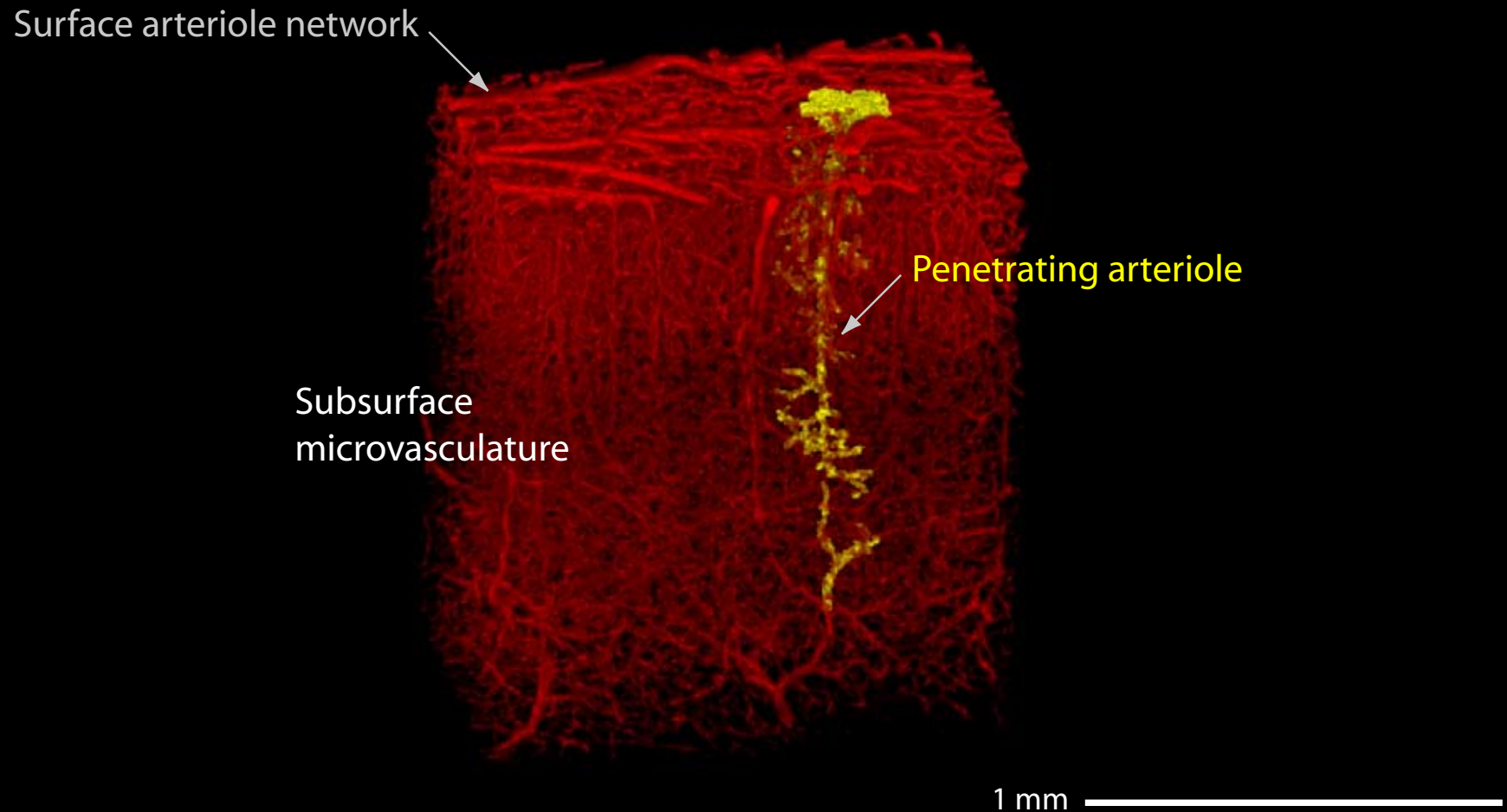
-  *In vivo* rat data of Nishimura, Rosidi, Iadecola & Schaffer (JCBFM 2010)
-  Flow analysis using mouse vascular reconstruction and Secomb's  $G(r, l)$ .

**Lesson: Lateral connectivity of the subsurface vasculature does not guarantee the perfusion of brain tissue.**

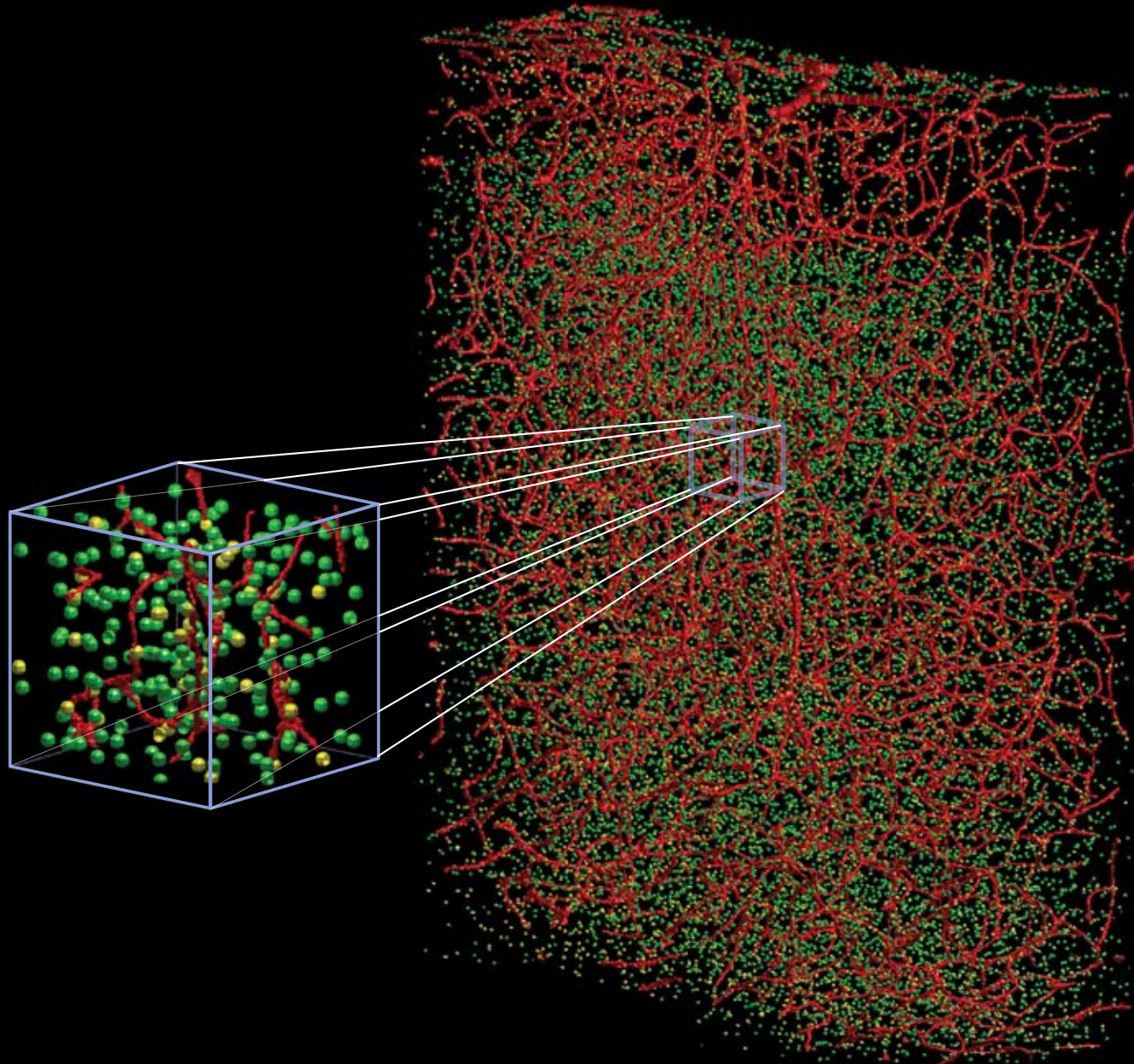
# Summary on angioarchitecture

**Redundancy:** Surface vascular network as a 2-D distribution grid  
Subsurface microvascular network as a 3-D grid

**Fragility:** Penetrating vessels as 1-D conduits from surface to microvessels  
Subsurface microvascular dominated by isotropic resistance to flow



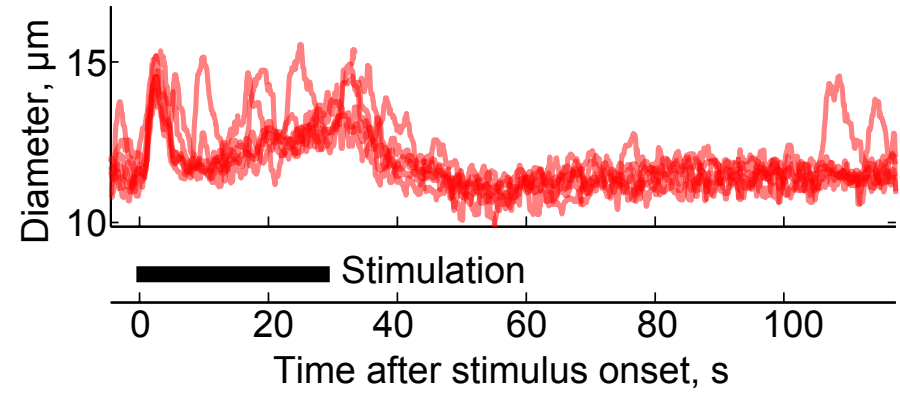
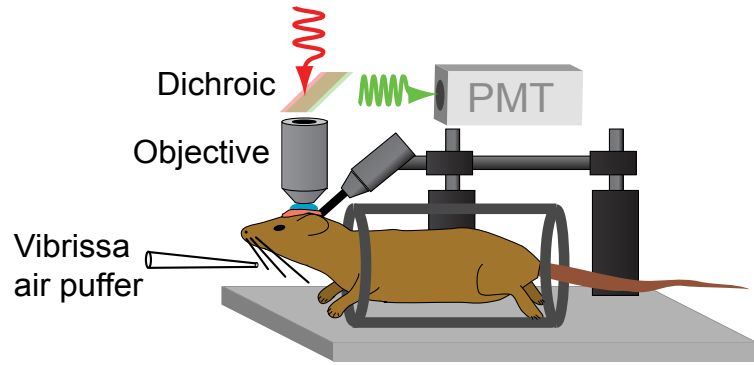
# Next steps: Delimiting changes in blood flow by patterns of neuronal activity



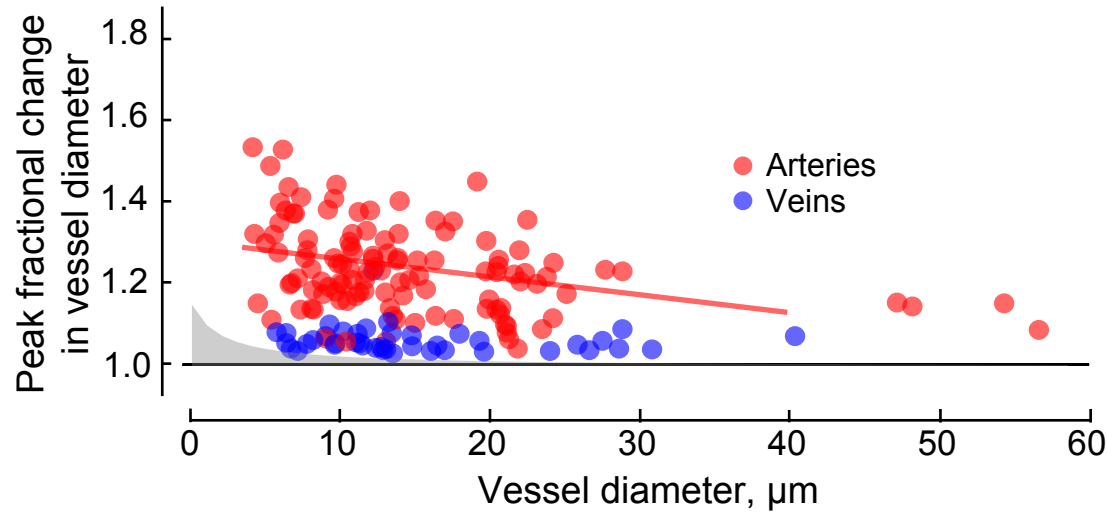


**Question: How much does cortical blood flow fluctuate in the absence of sensory input?**

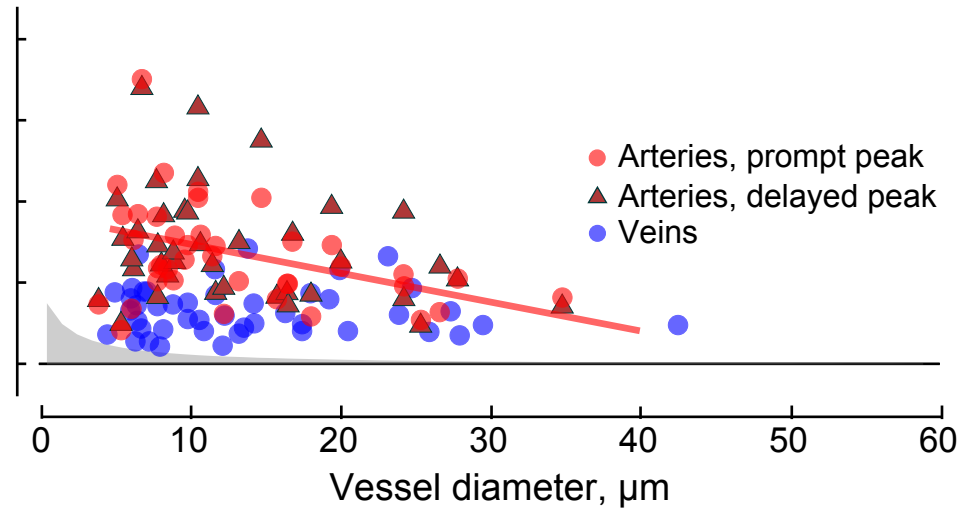
# Changes in arteriole diameter in response to stimulation are the same magnitude as the fluctuations in diameter (vasomotion)



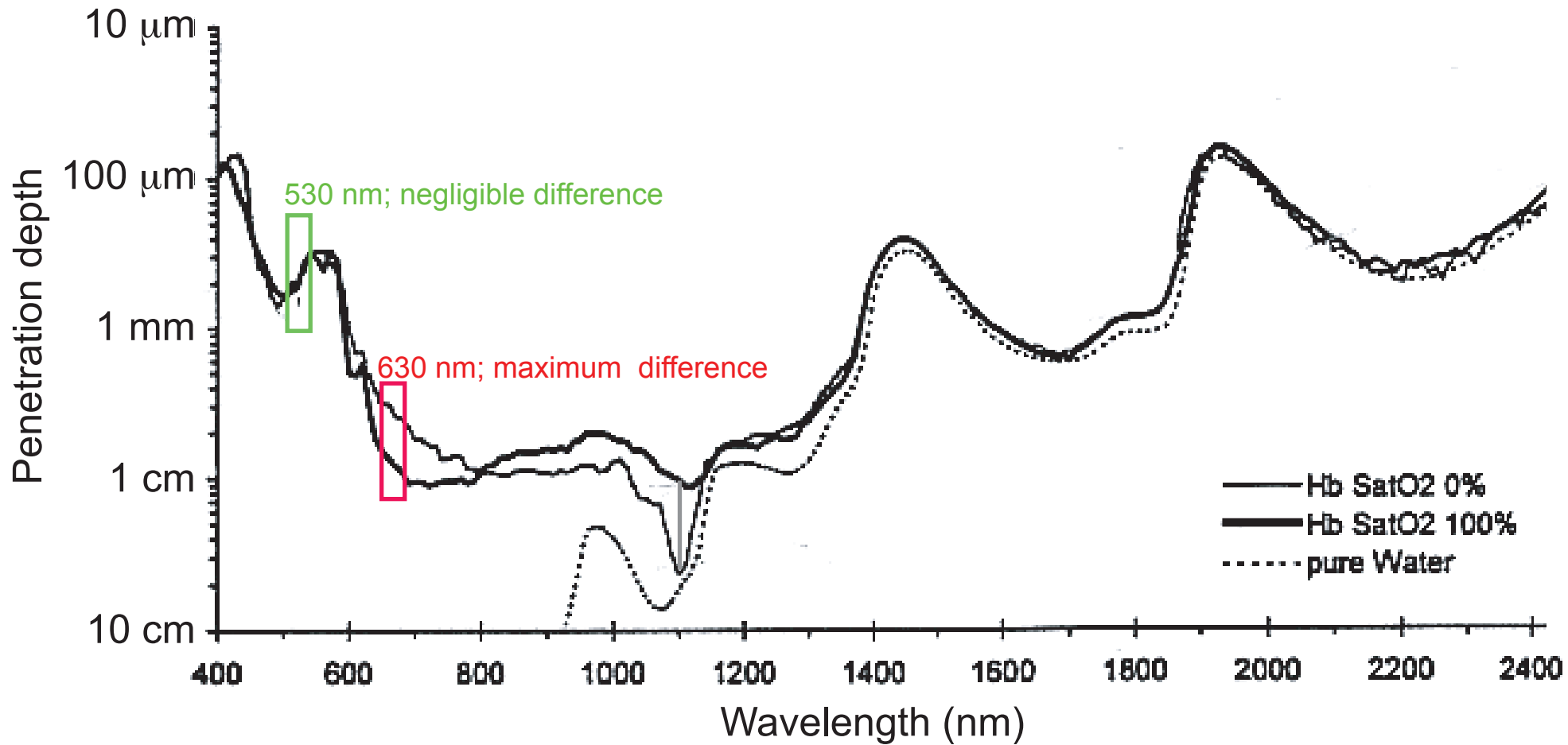
Spontaneous



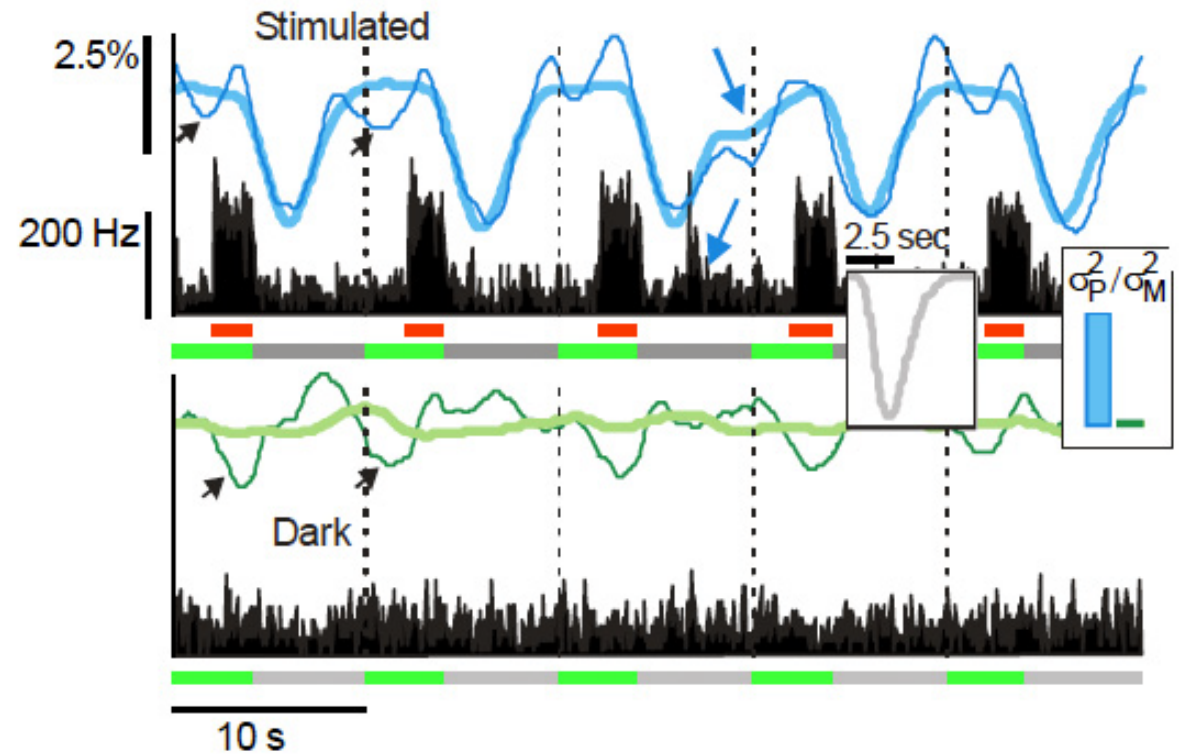
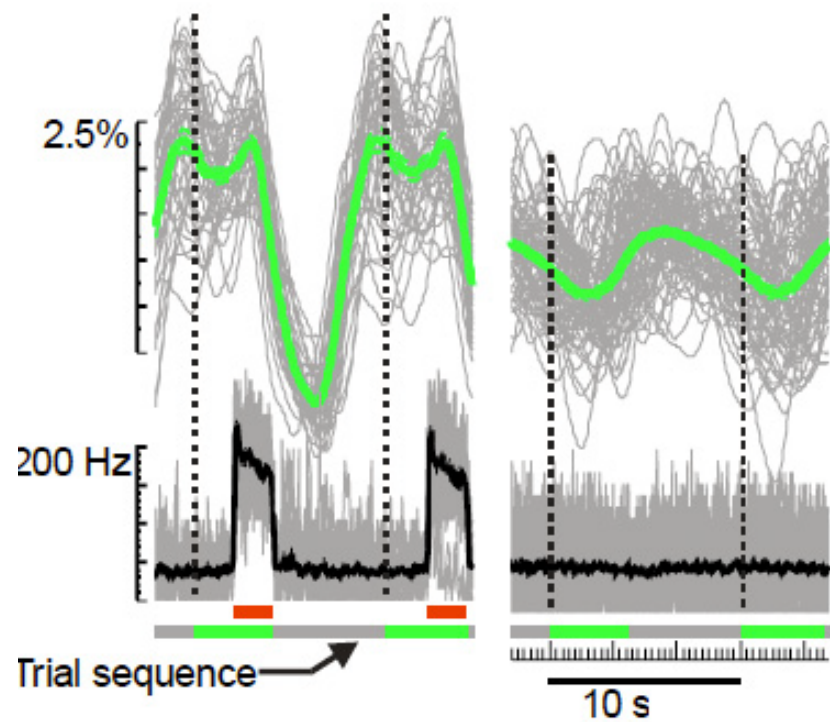
Average vibrissa stimulation



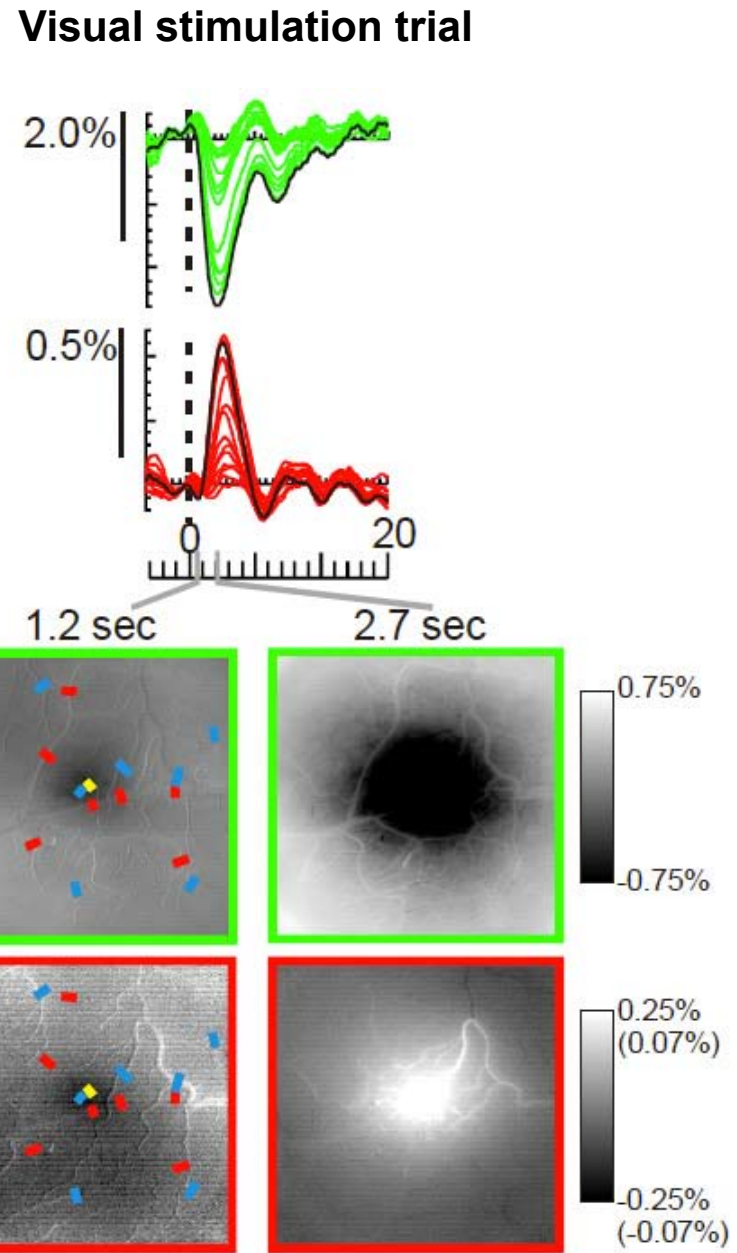
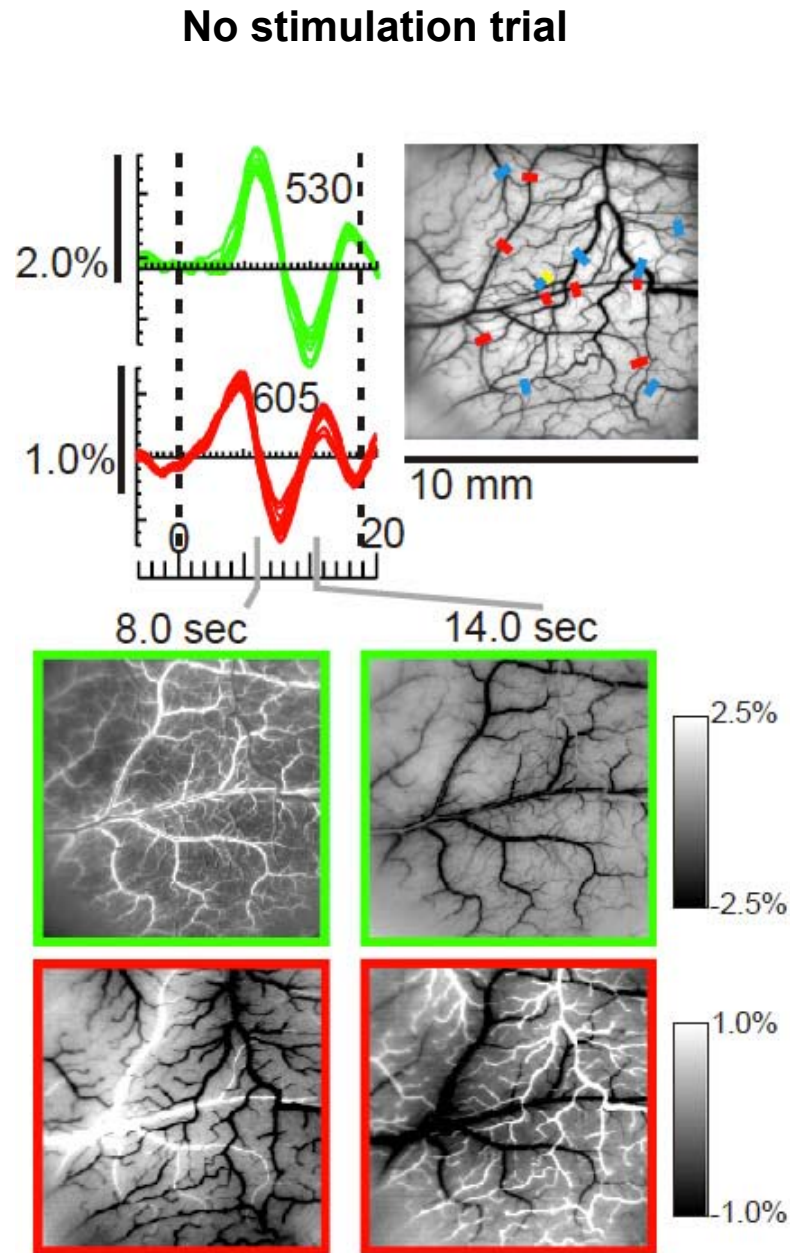
# Absorption by oxyhemeoglobin versus deoxyhemoglobin: Origin and BOLD fMRI and IOS signals



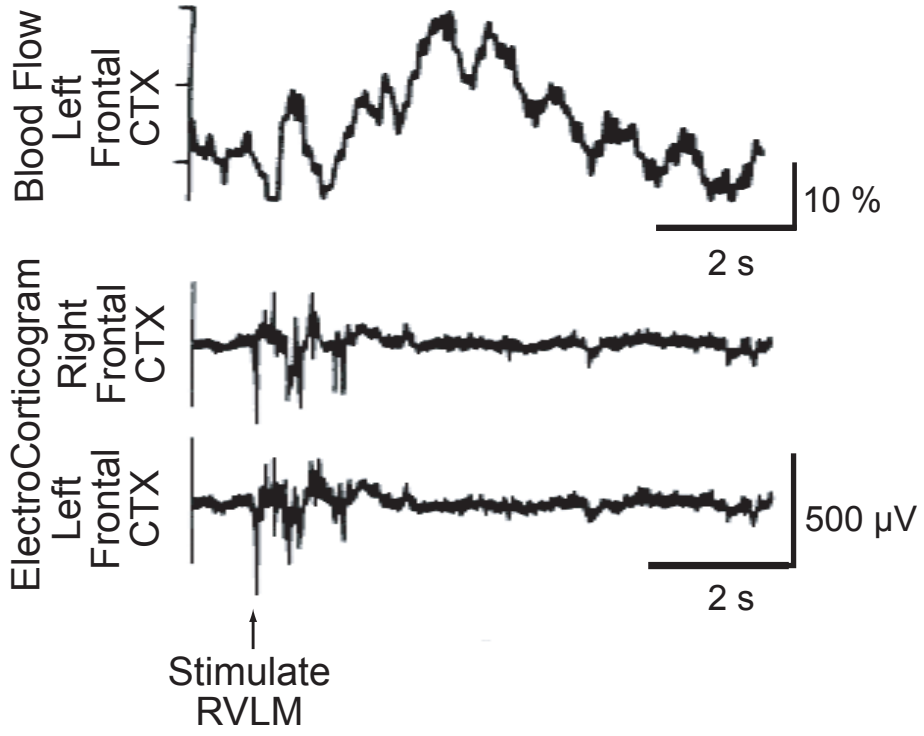
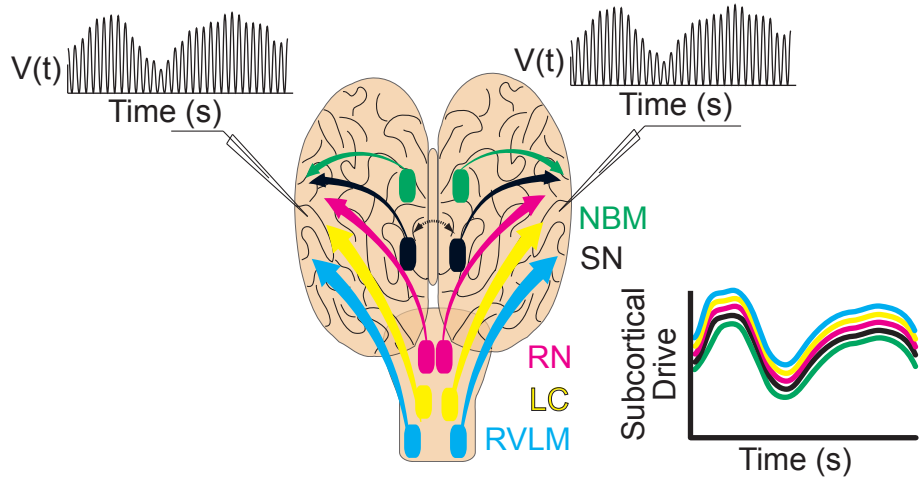
# Changes in blood oxygenation synchronized to trial onset and not stimulation



# Changes in blood oxygenation synchronized to trial onset and not stimulation



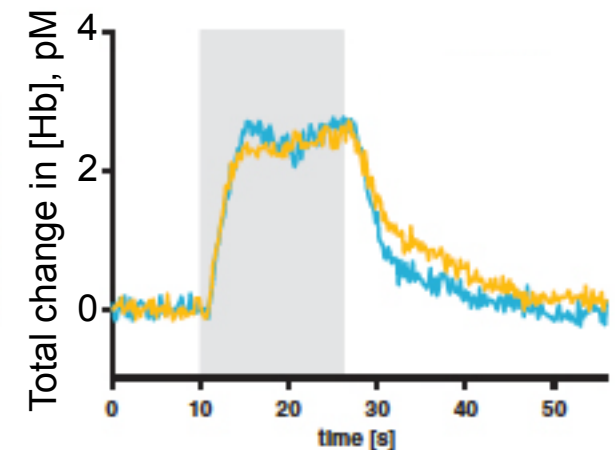
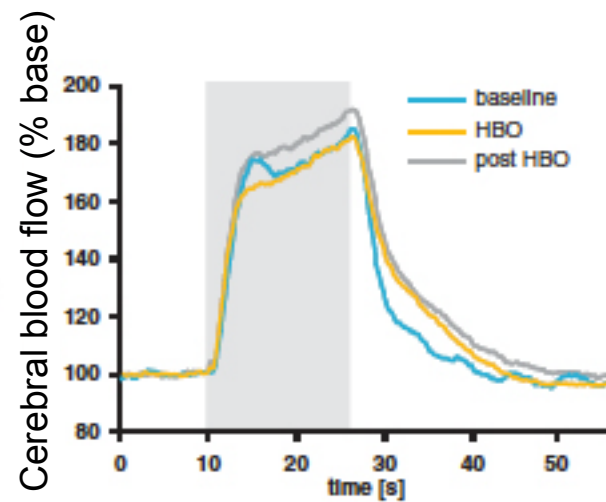
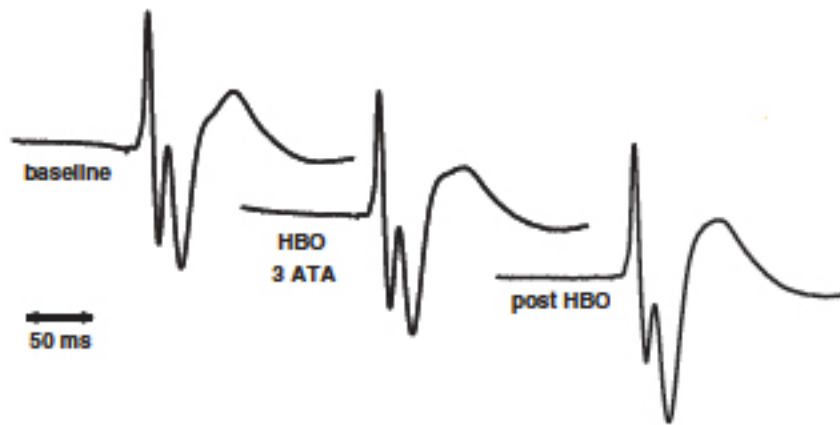
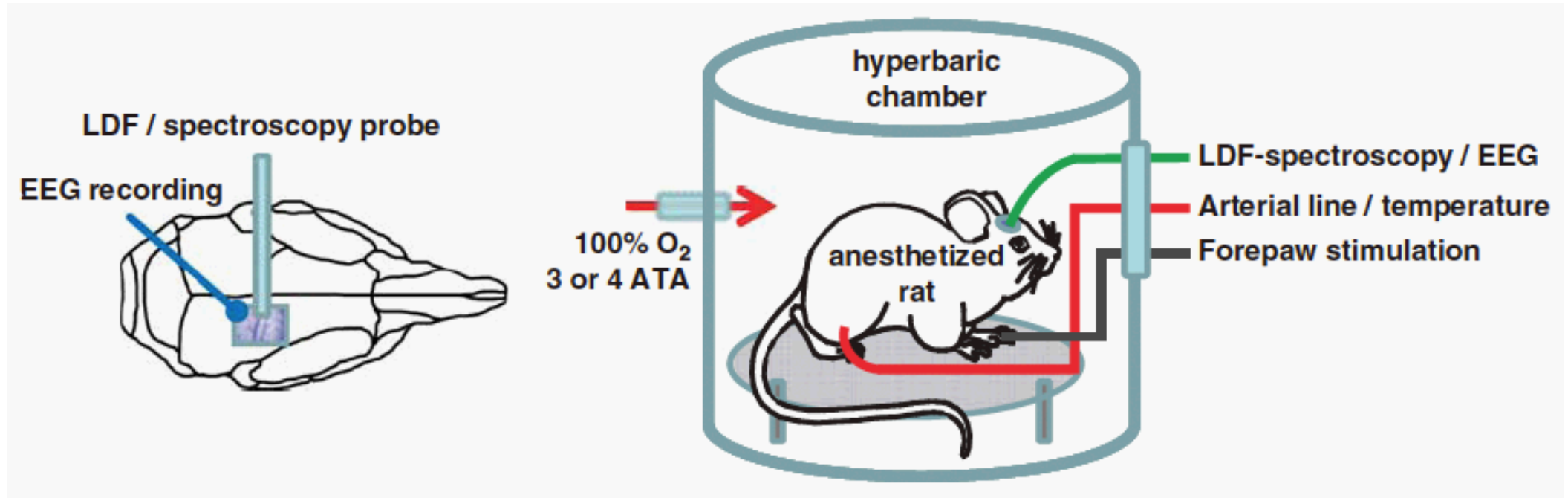
# Subcortical substrates for control of cortical blood flow and LFP power



**Lesson: Fluctuations in flow and potential subcortical variations to flow are as large as stimulus induced changes!**

**Question: Is the need for O<sub>2</sub>, as occurs with stimulus driven neuronal activity, enough to trigger increased blood flow?**

# Hyperoxygenation (3 atmospheres) does not affect cerebral blood flow

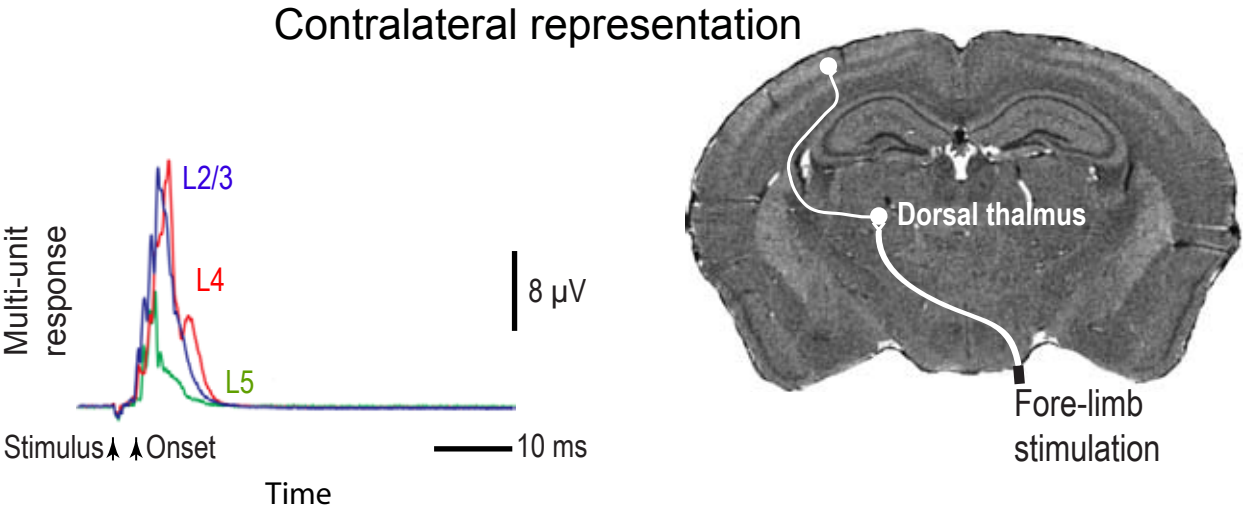




**Question: Metabolites aside, does an increase in neuronal activity necessarily imply an increase in blood flow?**

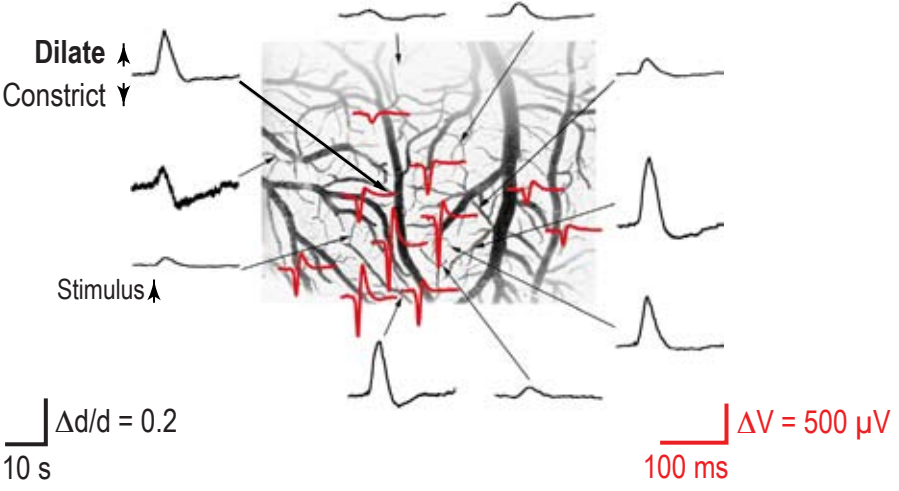
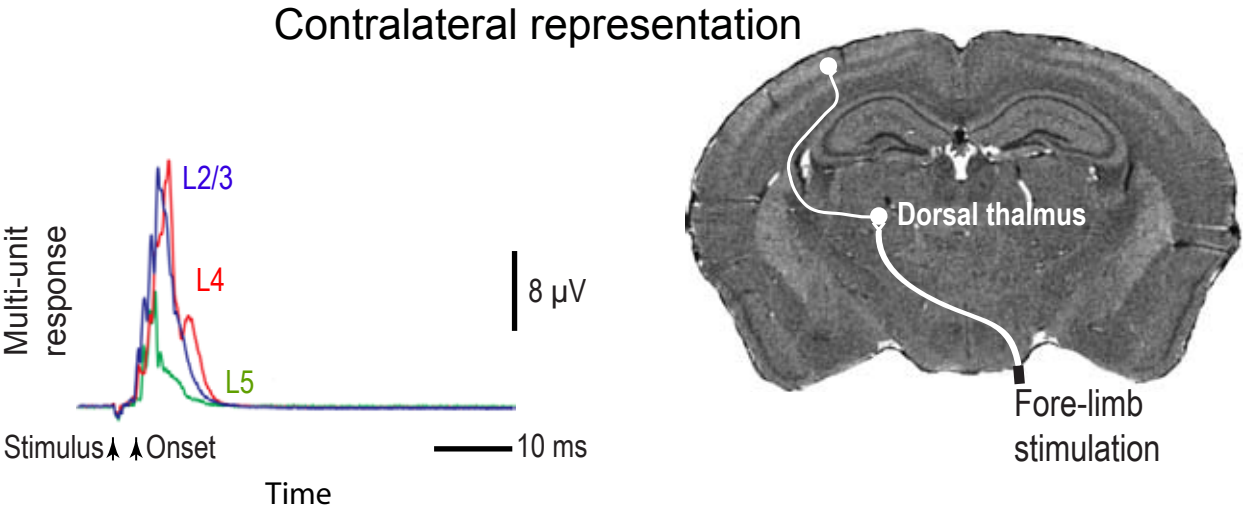
# Neuronal activity changes blood flow - but logic remains to be deciphered

Example of changes in the lumen of surface arterioles for the two cortical hemispheres



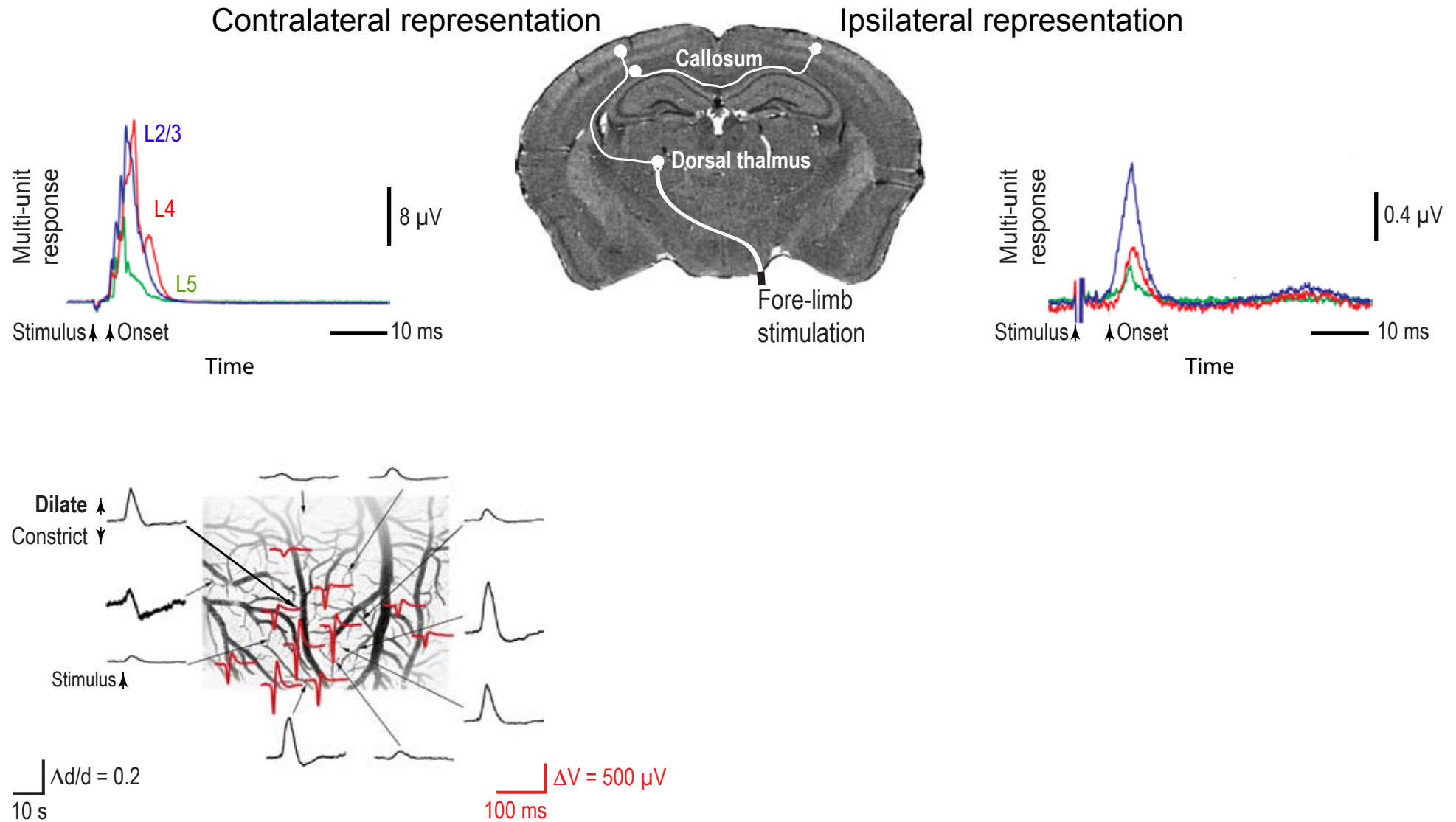
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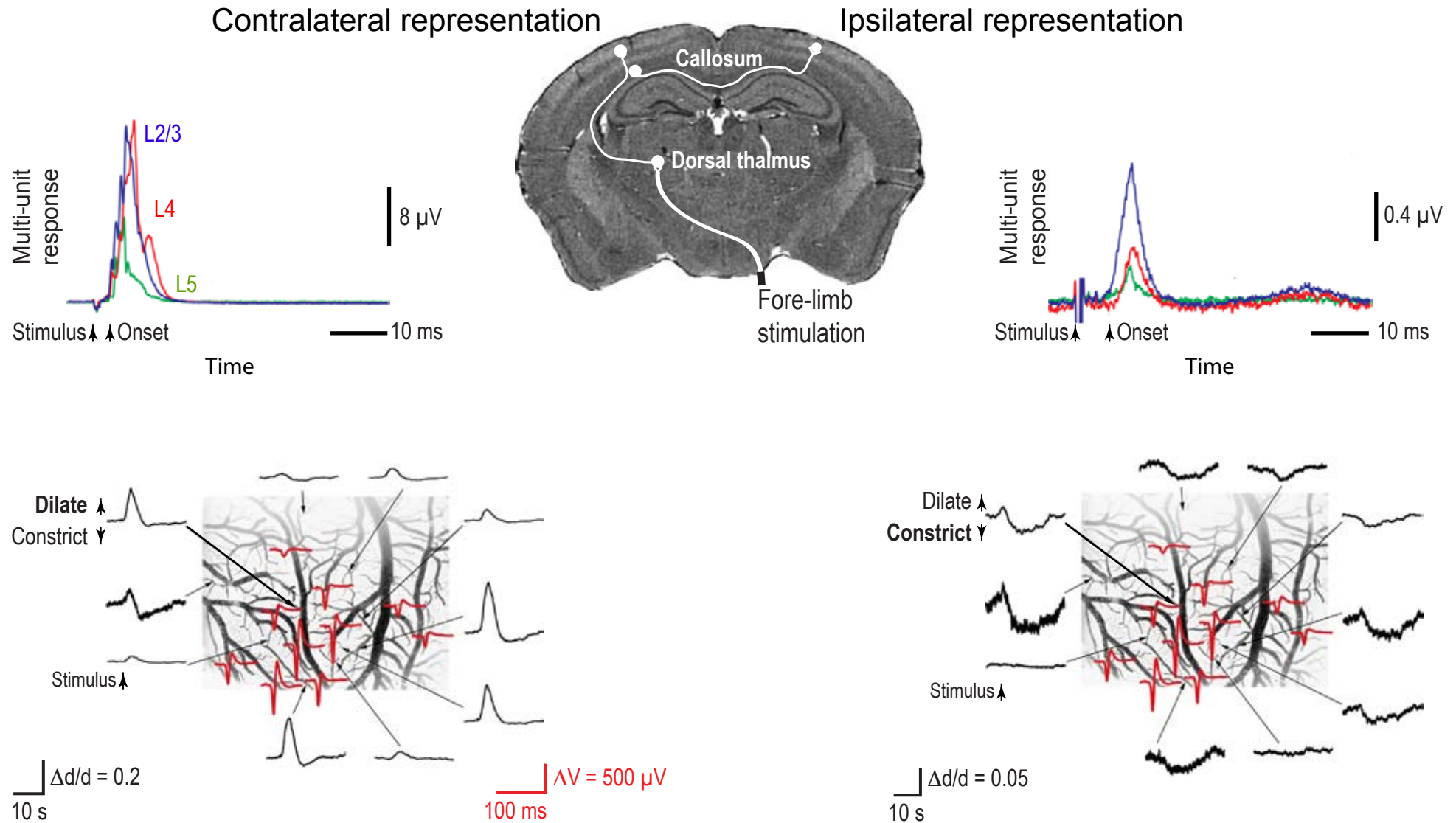
# Neuronal activity changes blood flow - but logic remains to be deciphered

Example of changes in the lumen of surface arterioles for the two cortical hemispheres



# Neuronal activity changes blood flow - but logic remains to be deciphered

Example of changes in the lumen of surface arterioles for the two cortical hemispheres



# Neural activity leads to competition between dilation and constriction

- Pyramidal activation of astrocytes leads to release of vaso modulators (e.g., Harder and McVicar laboratories)
- Interneuron release of neuropeptides (e.g., Hamel and Rossier laboratories)
- Extrinsic feedback from subcortical inputs, particularly from cholinergic and serotonergic nuclei

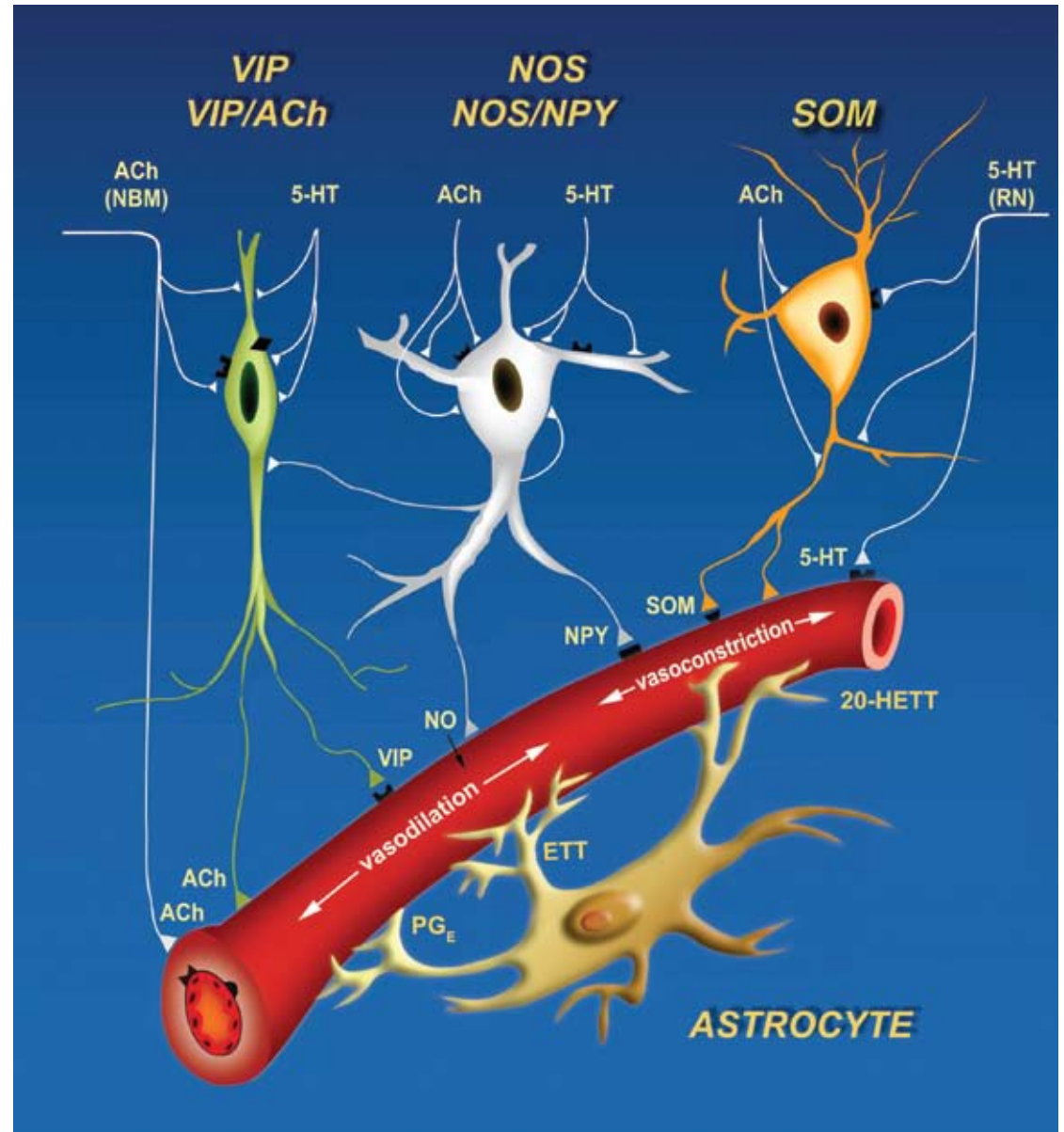
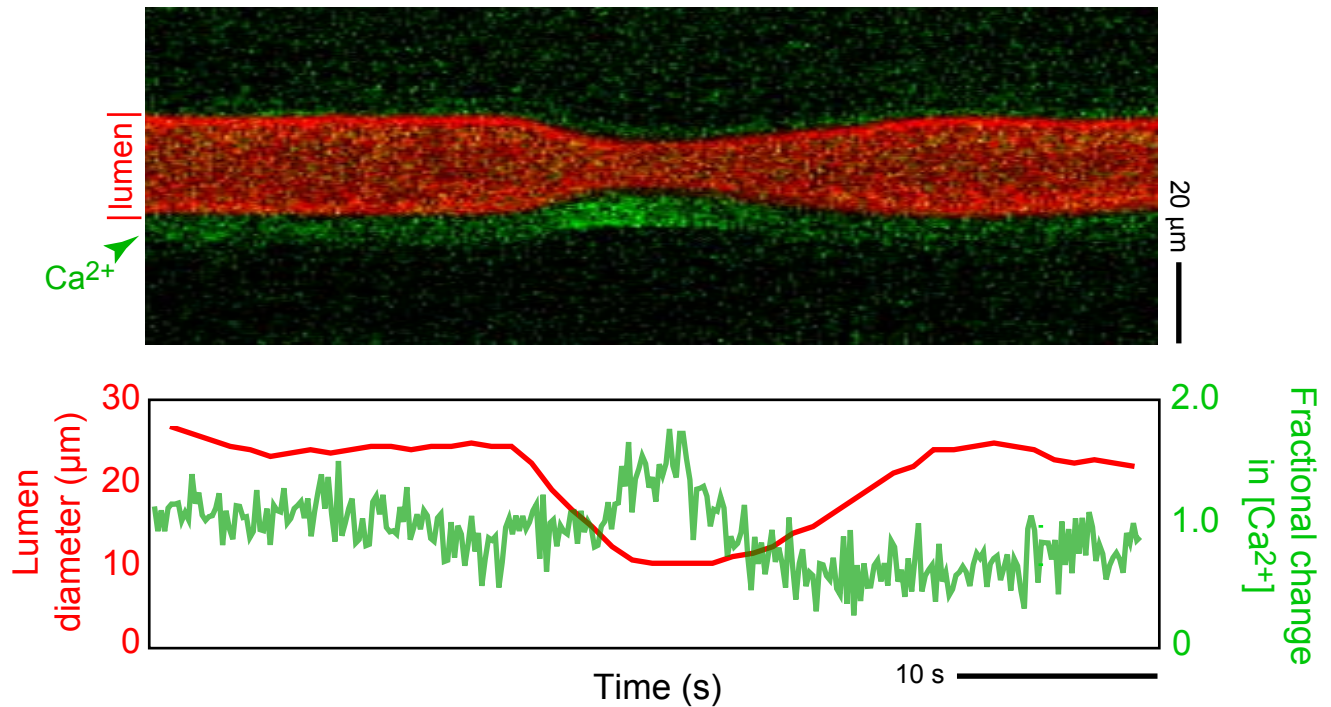
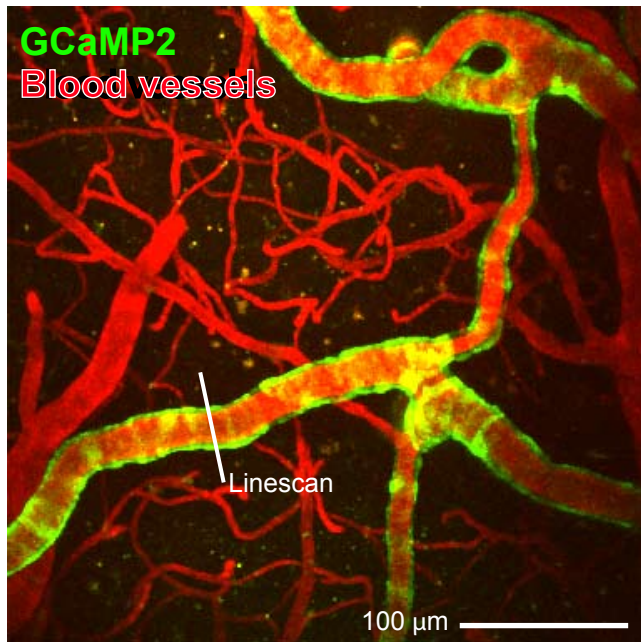


Figure adapted from Cauli, Tong, Rancillac, Serluca, Lambolez, Rossier & Hamel (J Neurosci 2004) and Iadecola & Nedergard (Nat Neurosci 2007)

# Smooth muscle activation may be imaged concurrent with blood flow

Example of measurements of muscle  $[Ca^{2+}]$  and lumen diameter in  $\alpha$ -actin-BAC-GCaMP2 mice\*



\*Ji, Feldman, Deng, Greene, Wilson, Lee, Johnston, Rishniw, Tallini, Zhang, Wier, Blaustein, Xin, Nakai & Kotlikoff (JBC 2004)  
Kleinfeld, Blinder, Drew, Driscoll, Muller, Tsai & Shih, (Front Neuroenergetics 2011)

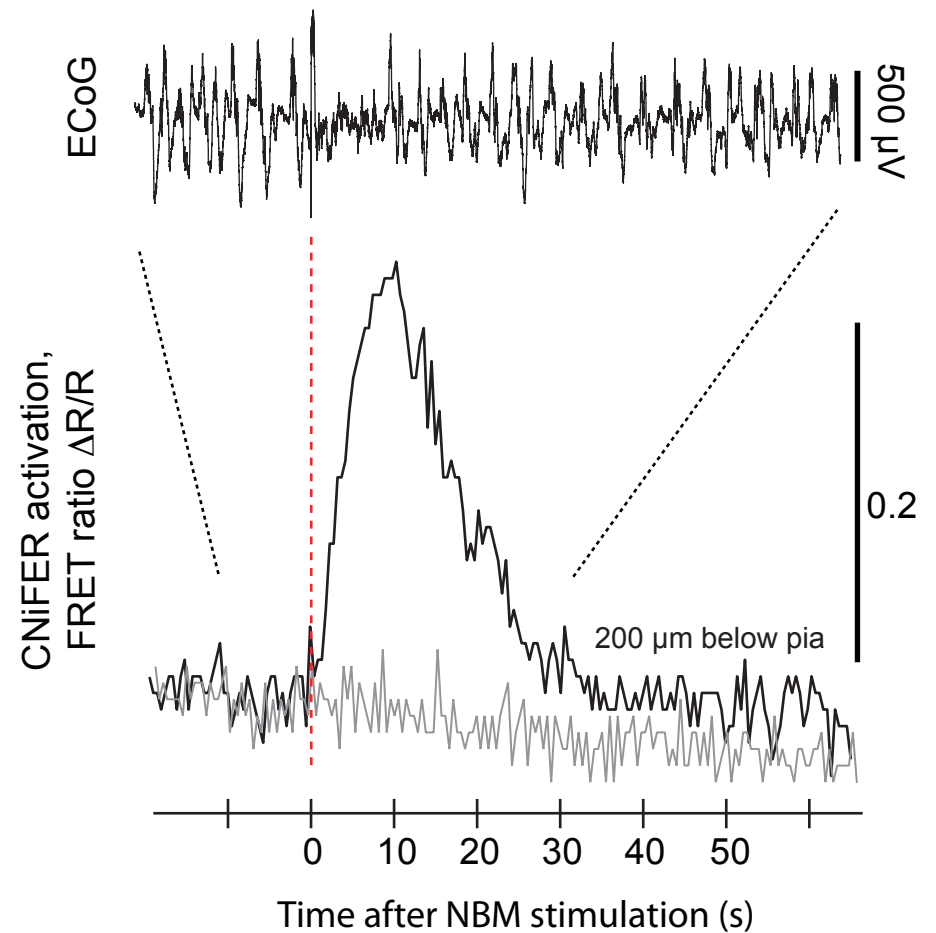
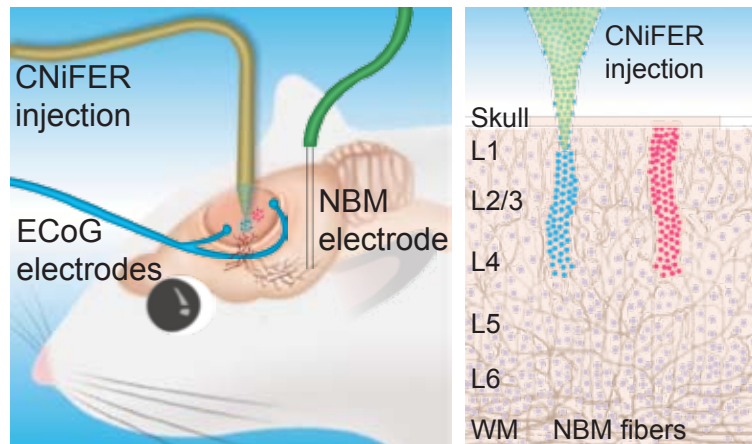
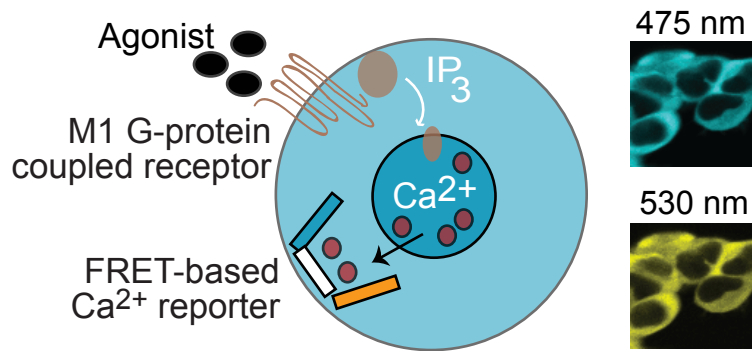
## CNiFERs for *in vivo* detection of vasoactive signaling molecules

| Vascular receptor      | G-protein        |                   | Neuronal agent |         | Astrocyte agent |         |
|------------------------|------------------|-------------------|----------------|---------|-----------------|---------|
|                        | Pathway*         | Action            | Constrictor    | Dilator | Constrictor     | Dilator |
| Neuropeptide Y         | G <sub>i/o</sub> | cAMP ↓            | ■              |         |                 |         |
| Somatostatin           | G <sub>i/o</sub> | cAMP ↓            | ■              |         |                 |         |
| Acetylcholine (M5)     | G <sub>q</sub>   | IP <sub>3</sub> ↑ |                | ■       |                 |         |
| Serotonin              | G <sub>q</sub>   | IP <sub>3</sub> ↑ |                | ■       |                 |         |
| Vasointestinal peptide | G <sub>s</sub>   | cAMP ↑            |                | ■       |                 |         |
| Endothelin (B)         | G <sub>i/o</sub> | cAMP ↓            |                |         | ■               |         |
| Adenosine (A2A)        | G <sub>s</sub>   | cAMP ↑            |                |         |                 | ■       |
| Prostaglandin (E2)     | G <sub>q</sub>   | IP <sub>3</sub> ↑ |                |         |                 | ■       |

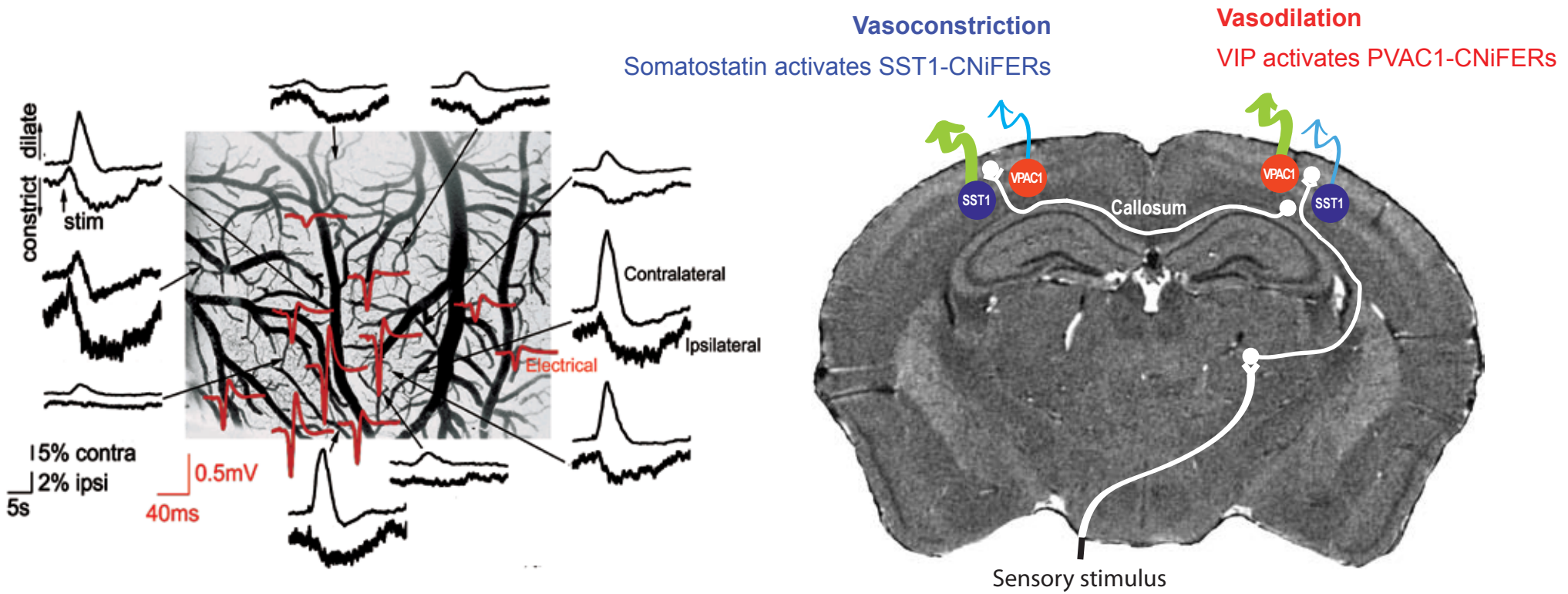


# *In situ* neurotransmitter receptor activation monitored by cell-based neurotransmitter fluorescent engineered reporters (CNiFERs)

Example of extracellular ACh detection by M1-CNiFERs chronically implanted in rat



# Example of a proposed experiment: SOM- and VIP-CNiFERs to detect change in balance of constriction versus dilation



# Goal: Quantitative model to explain cortical blood flow dynamics

$$\text{Vascular output} \equiv f \left( \begin{array}{c} \text{Smooth muscle tension} \\ \vdots \\ \text{Vascular architecture} \end{array} \right)$$

$$\text{Smooth muscle tension} \equiv f \left( \begin{array}{c} \text{SOM+ inhibitory interneuron activation} \\ \text{VIP+ inhibitory interneuron activation} \\ \vdots \\ \text{Astrocyte activation} = f(\text{excitatory cell activation}) \\ \text{Extracortical and endothelial contributions} \\ [\text{O}_2]_{\text{tissue}} \end{array} \right)$$

# Cortical angioarchitecture and vasodynamics

Thank you for your attention!

