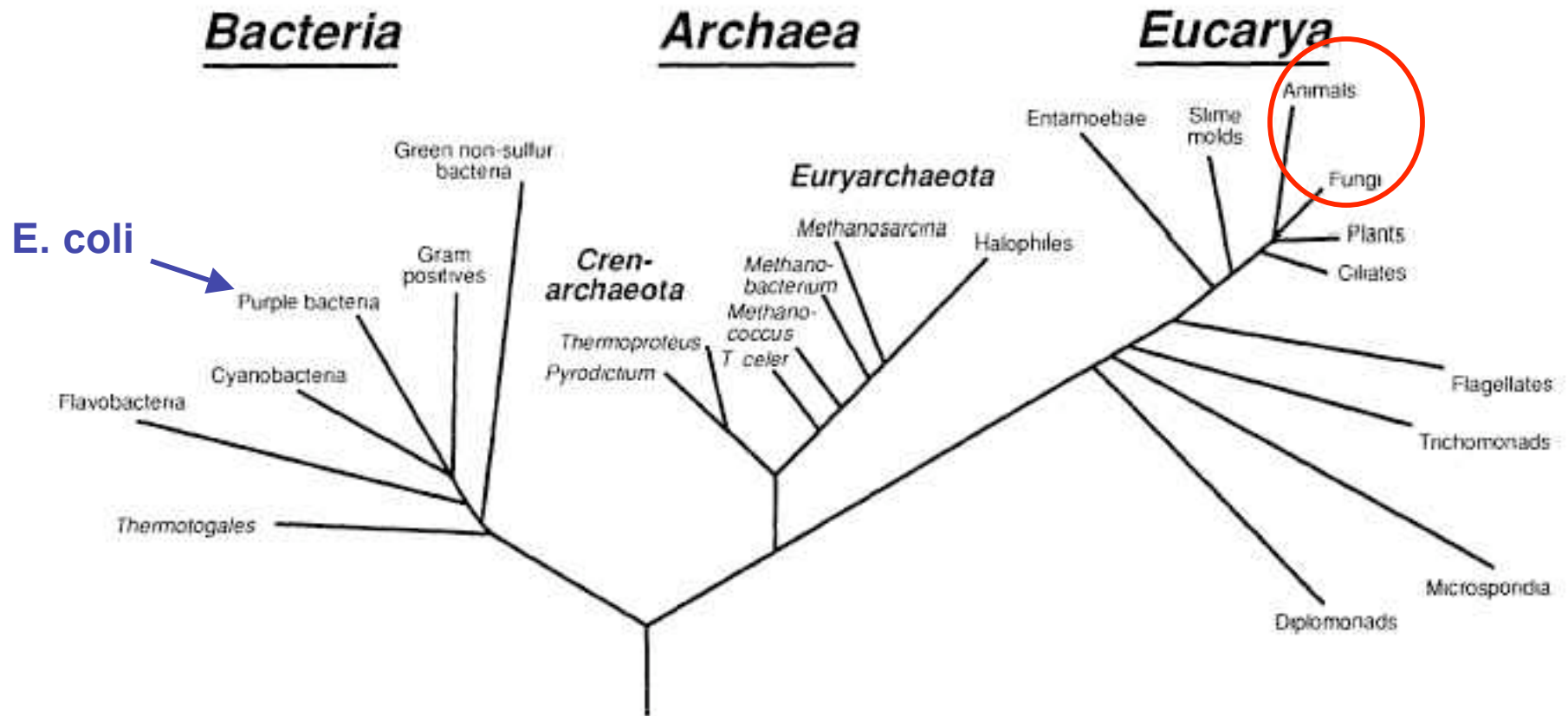




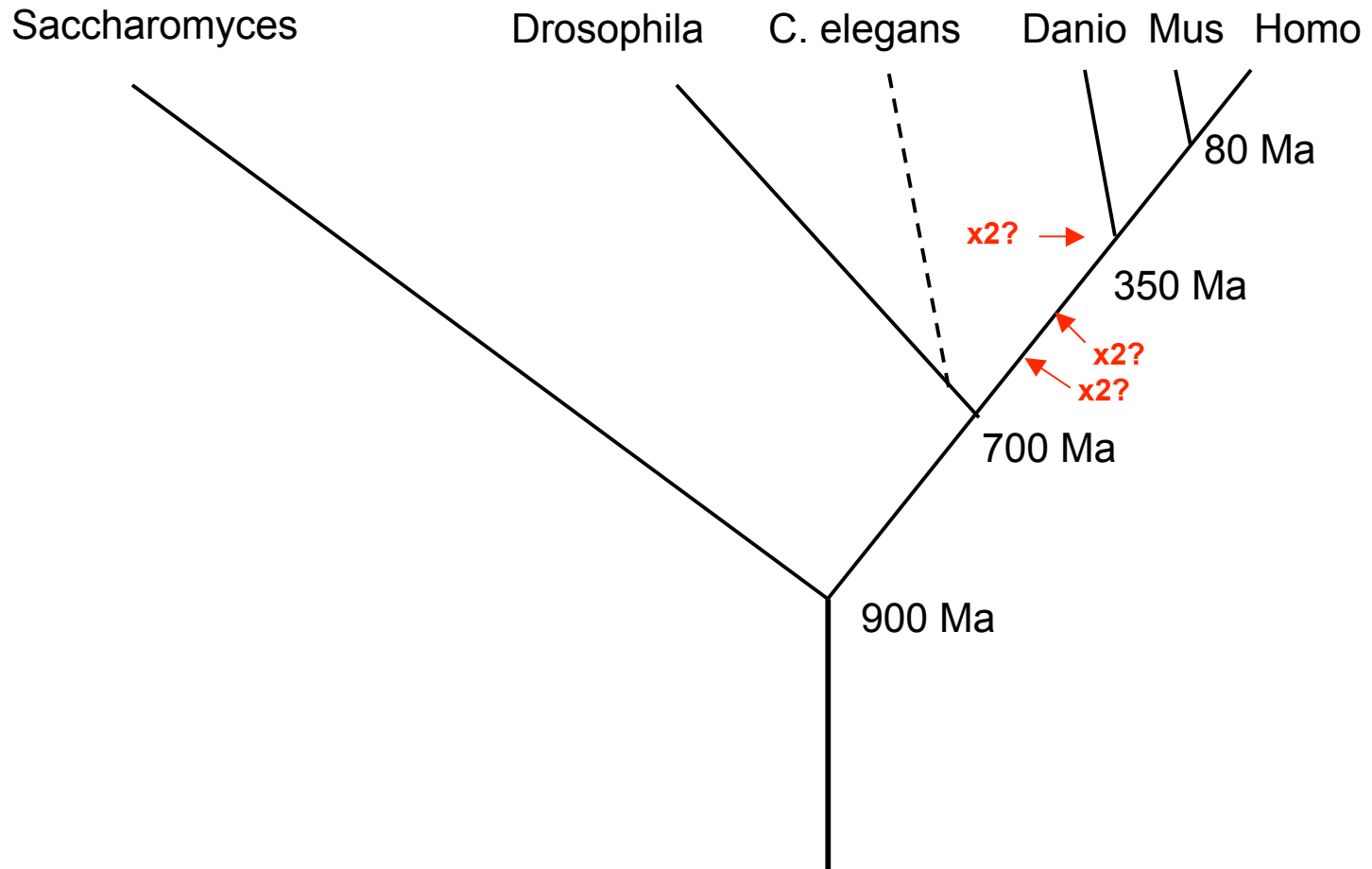
Genetic Architecture of the Visual System

Baier lab
UCSF

The Universal rRNA Tree

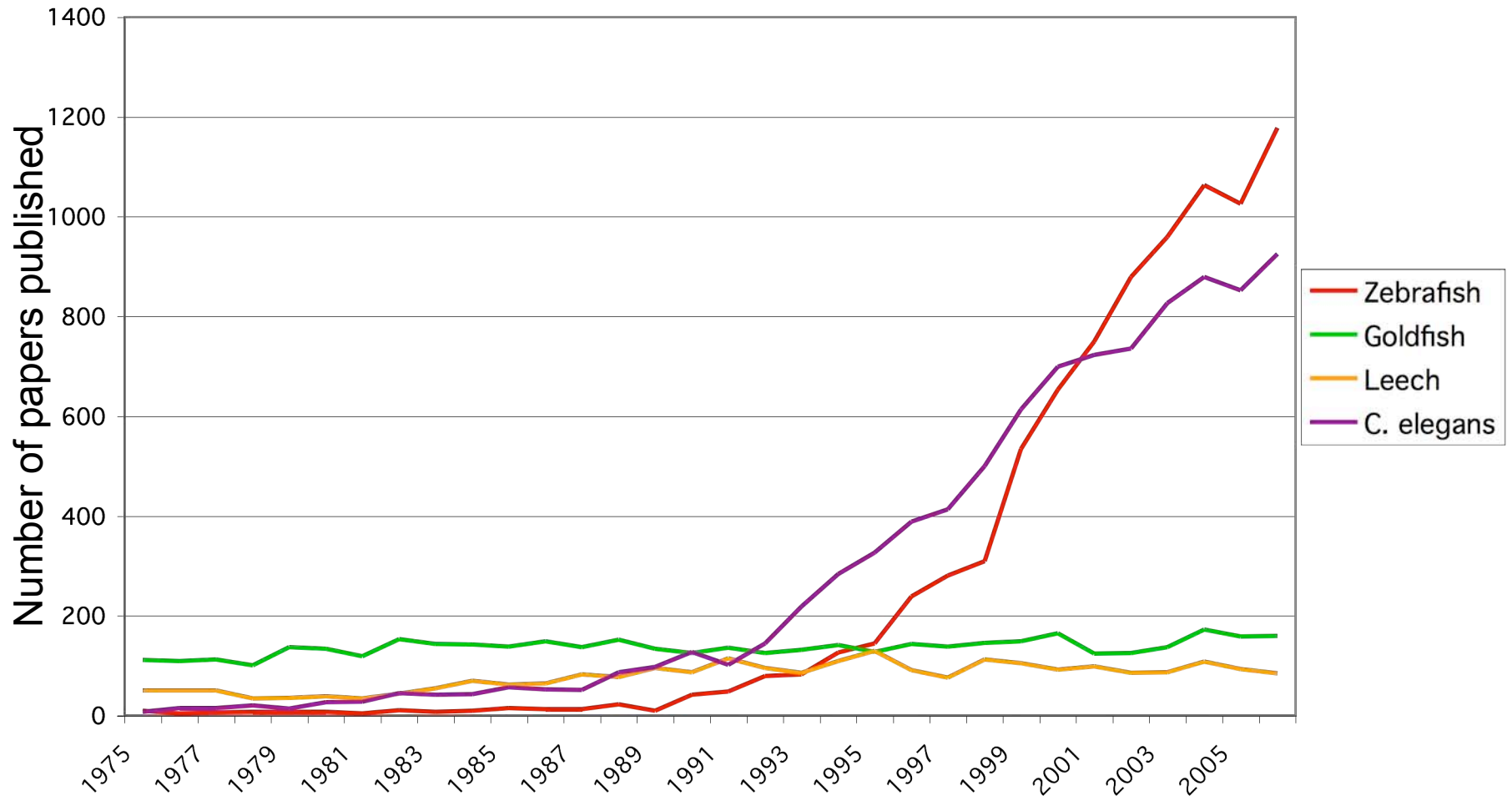


Phylogeny of genetic model organisms



(Numbers are based on fossil evidence, molecular clocks, or best guesses.)

Explosion of research literature on genetic models



↑ Brenner (1974)
↑ Streisinger (1981)

2007: 4,737 publications
on "zebrafish"

Experimental advantages of zebrafish for neuroscience research

- 1) Genetics
- 2) Optics
- 3) Behavior
- 4) Chemistry

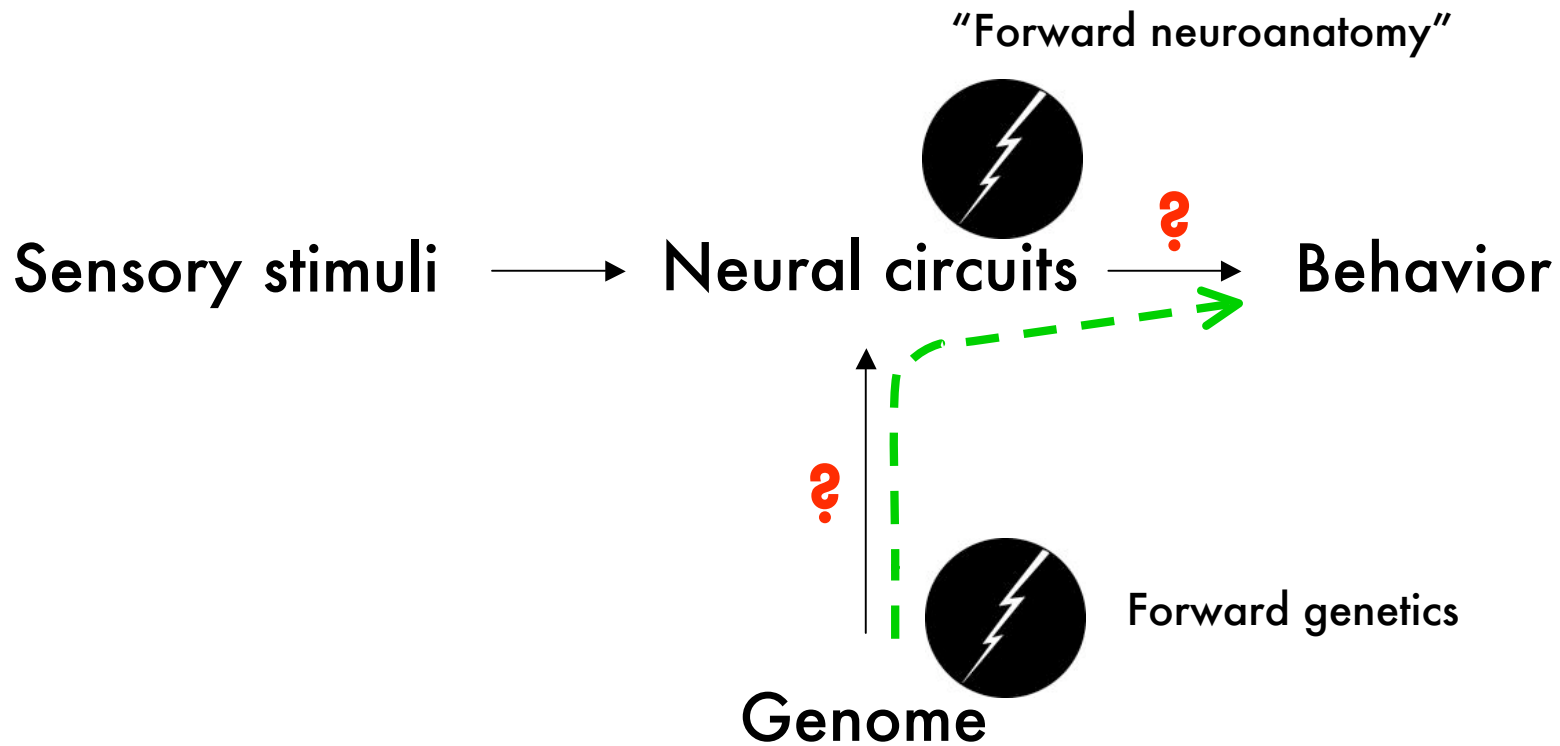
Outline of the talk

- 1) Genetic approaches to neural circuits & behavior
- 2) Assembly and function of retinotectal connections
(*two little stories*)
- 3) New tools for systematic functional dissection of circuits

How can we untangle the brain?

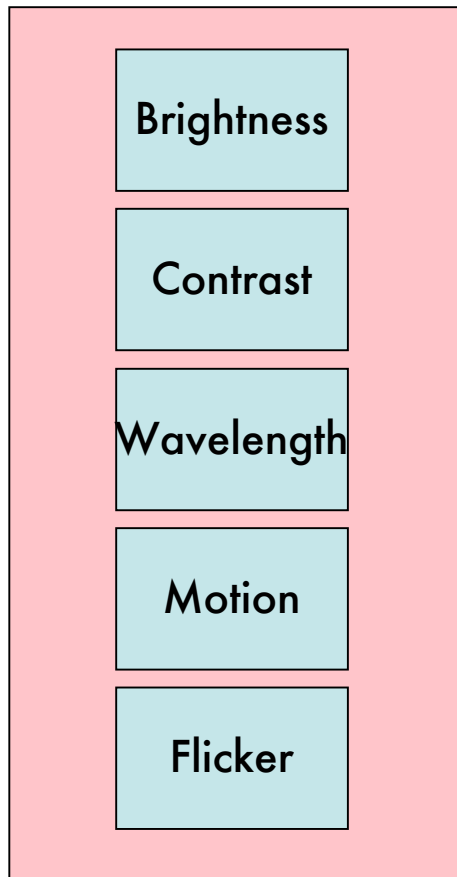


Unbiased genetic approaches to circuit assembly & function

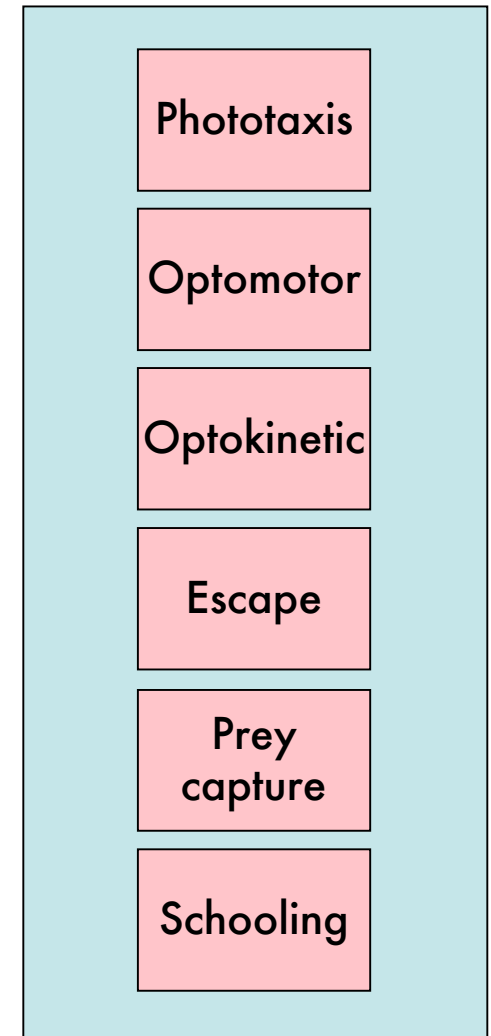
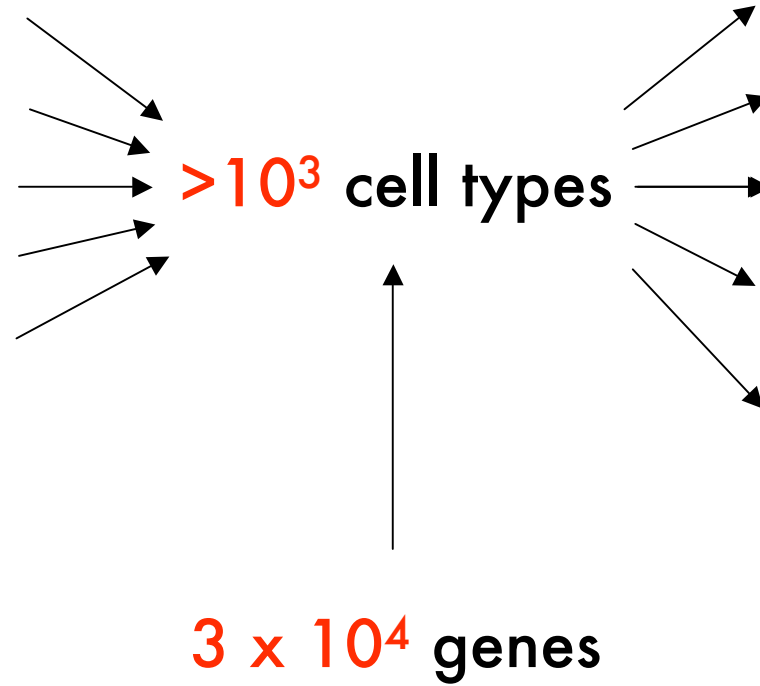


Phenotype-based systems neuroscience

Zebrafish visual system



Visual features



Behavioral responses

The zebrafish visual system develops on a tight schedule

- Day 0: Fertilization
- Day 1: Retinal ganglion cells born
- Day 2: Retinal axons leave eye
- Day 3: First visual responses
- Day 4: Retinotectal map complete
- Day 5: Optomotor behavior
- Day 6: Visually guided prey capture



How are specific synaptic connections formed in such a short time?

Optomotor response (OMR)

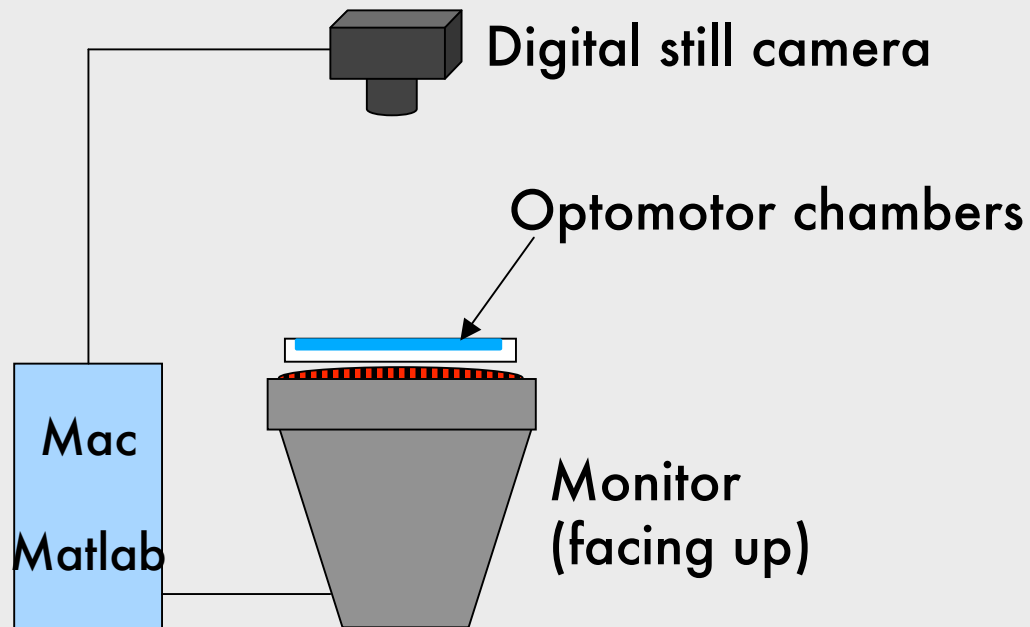
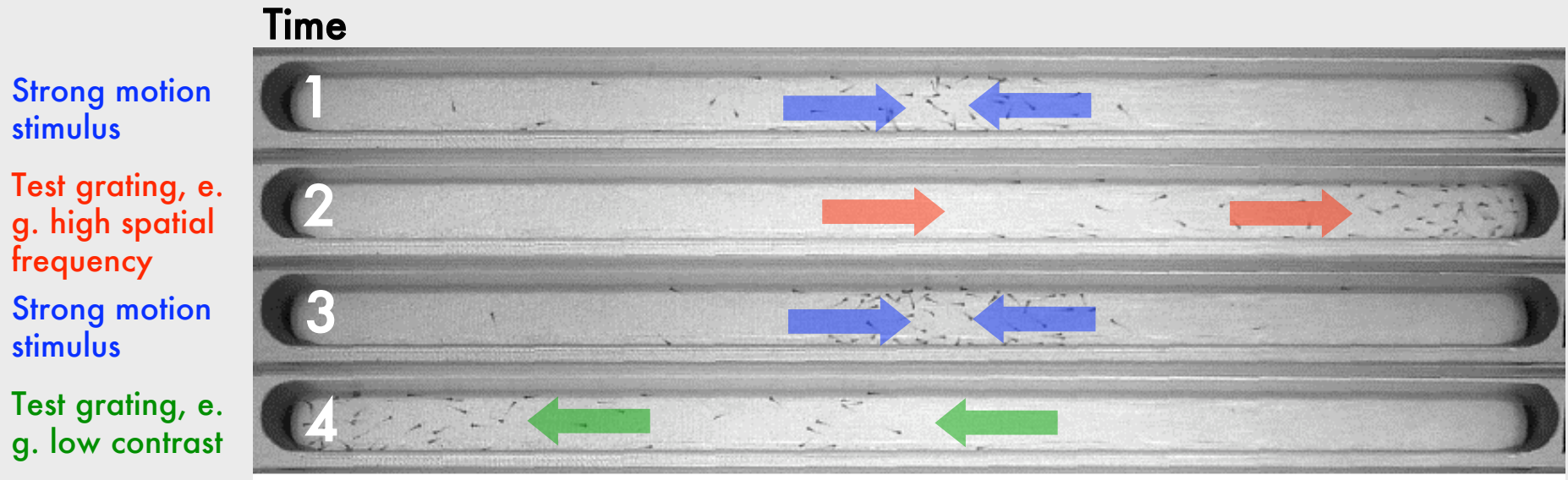
1 cm



2x speed

Mike Orger

High-throughput optomotor assay

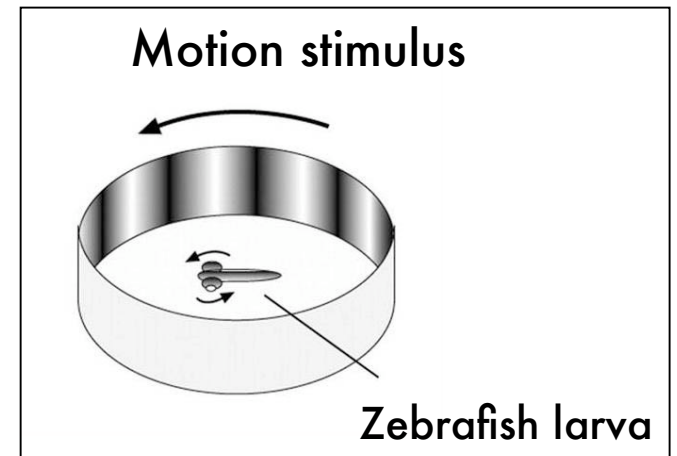
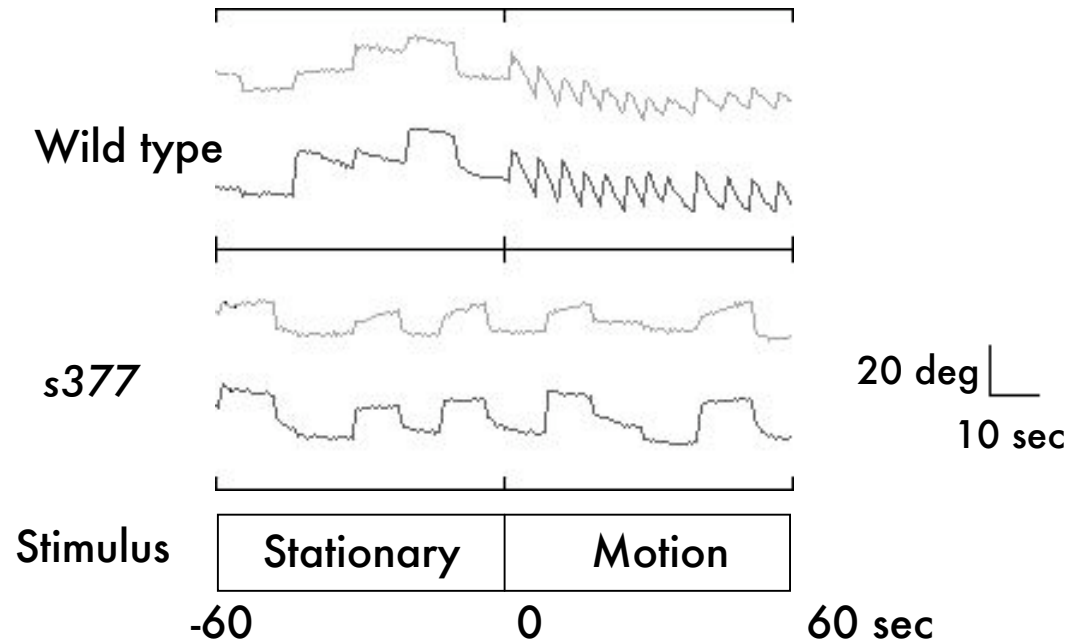


Mike Orger, Matt Smear

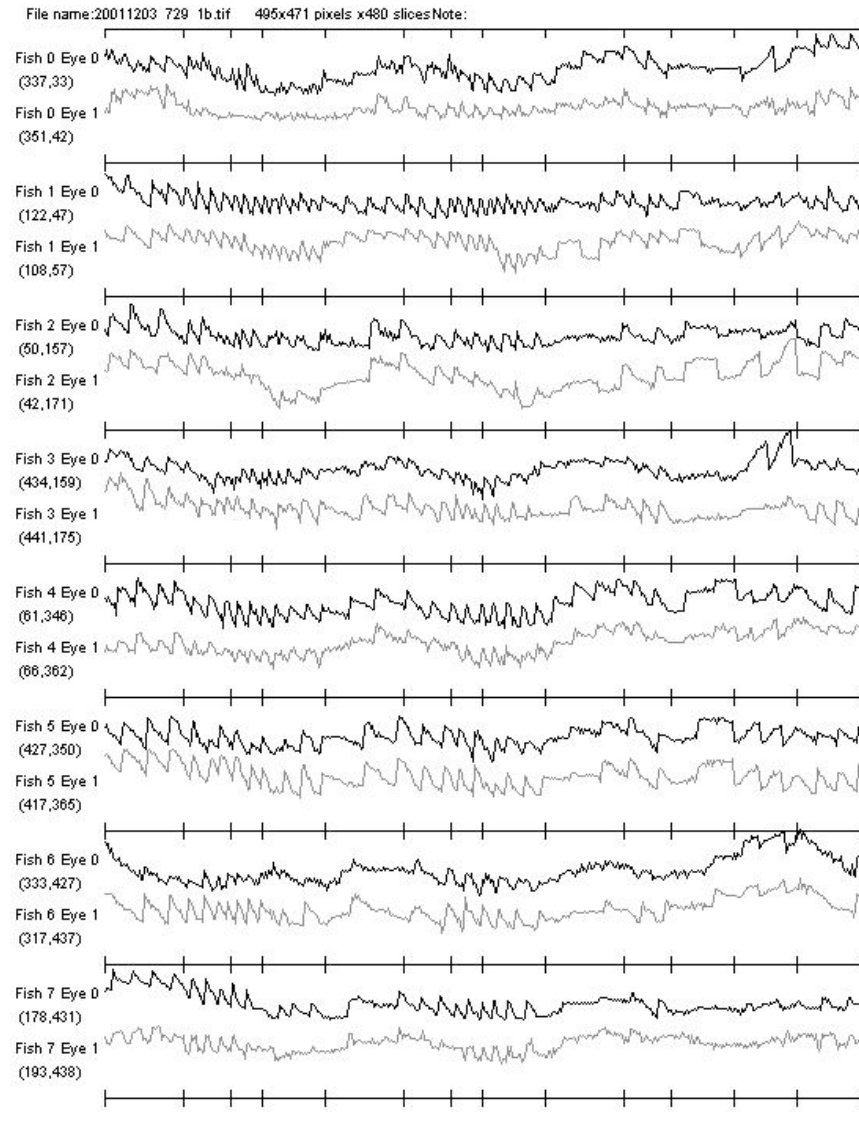
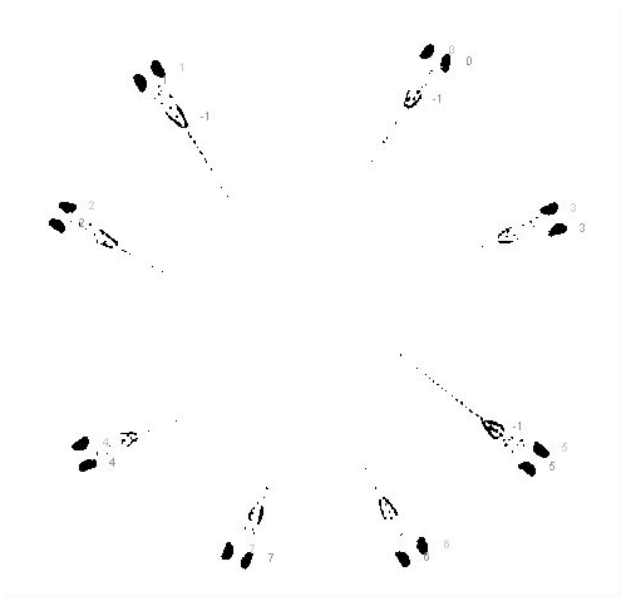
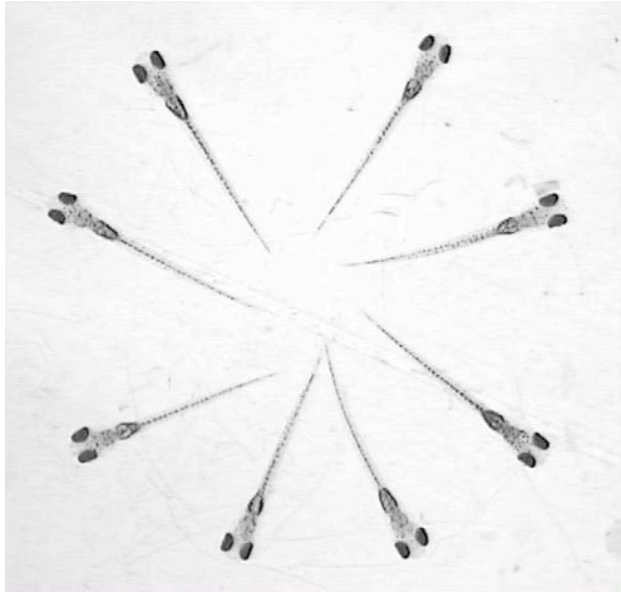
Optokinetic response (OKR)

Wild type

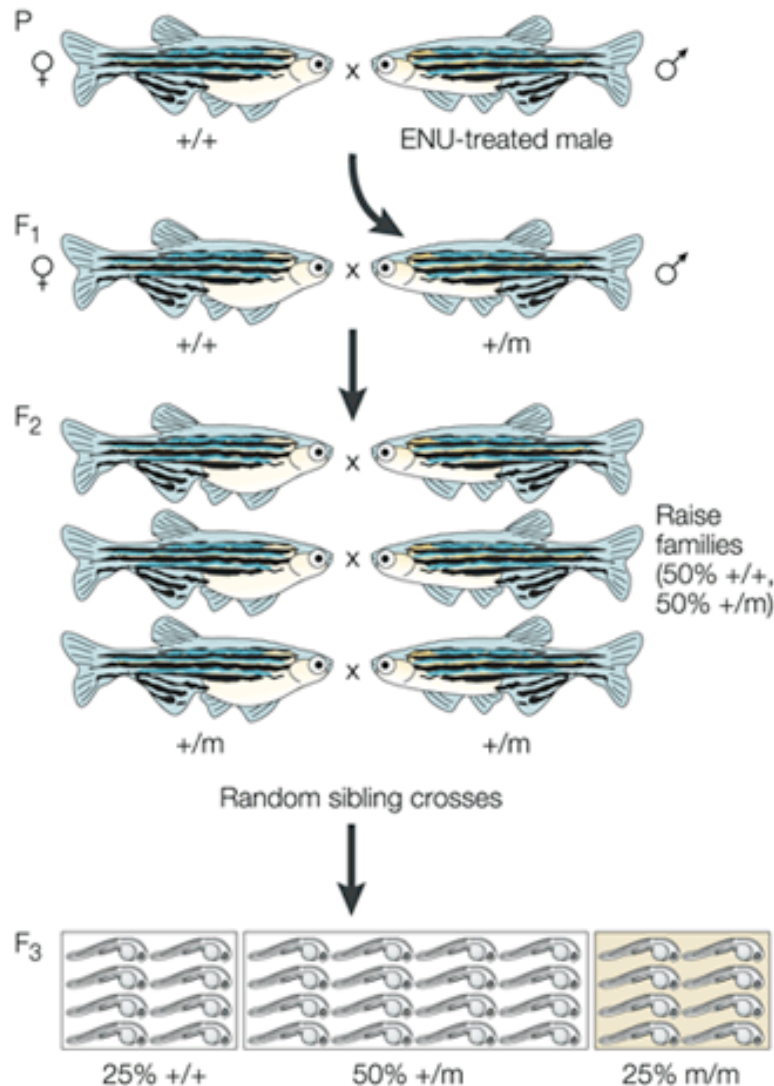
see no evil^{s377}



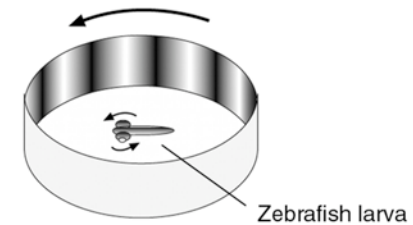
High-throughput OKR assay



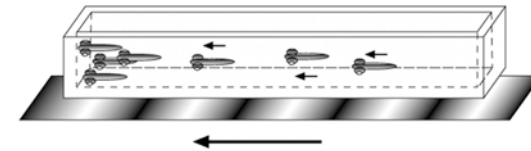
Large-scale ENU-mutagenesis screen for mutations disrupting visual responses



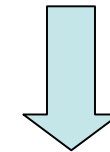
The optokinetic response (OKR)



The optomotor response (OMR)

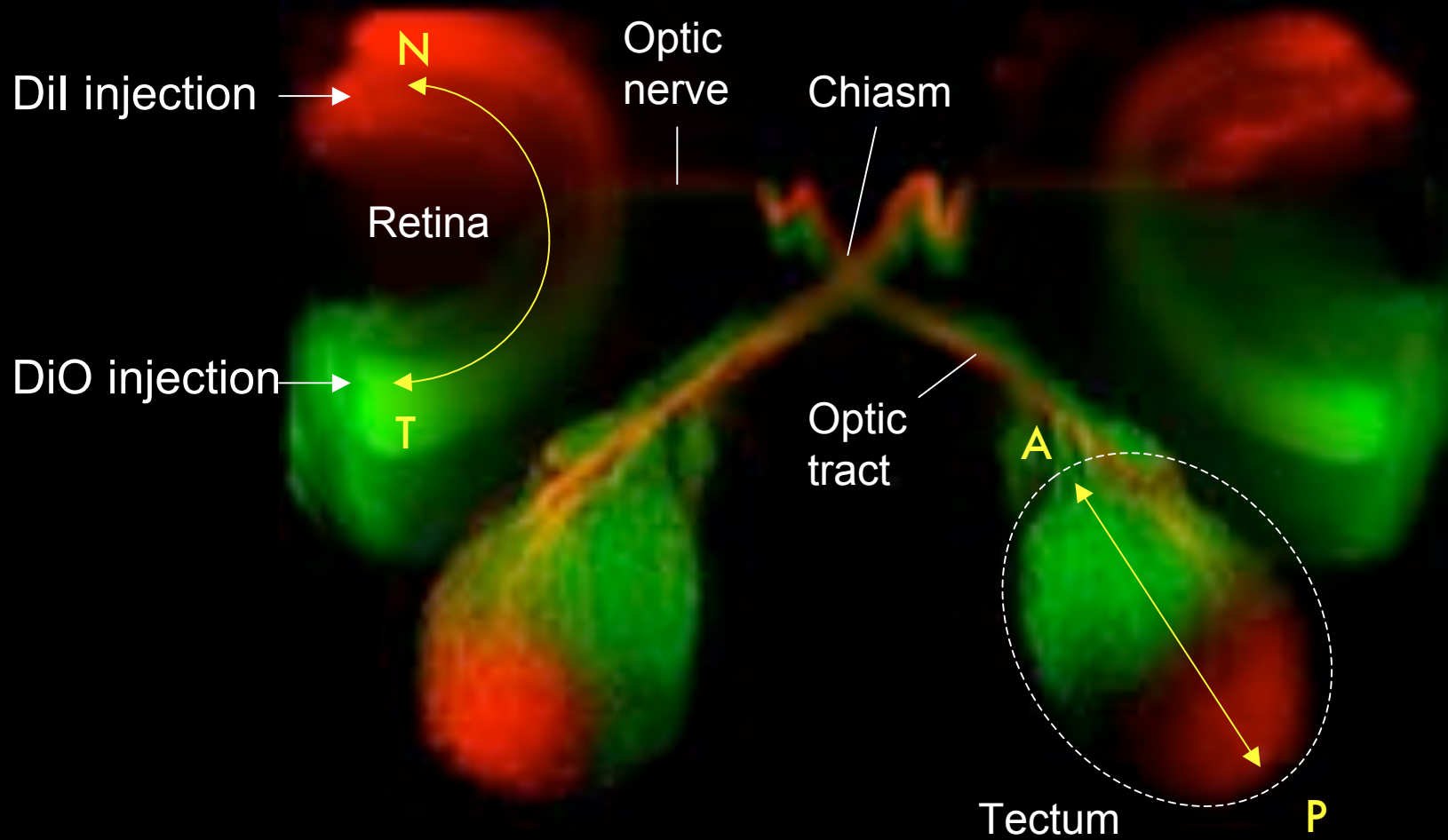


1,988 F₂ families (8,000 crosses;
>1,000,000 fish larvae)



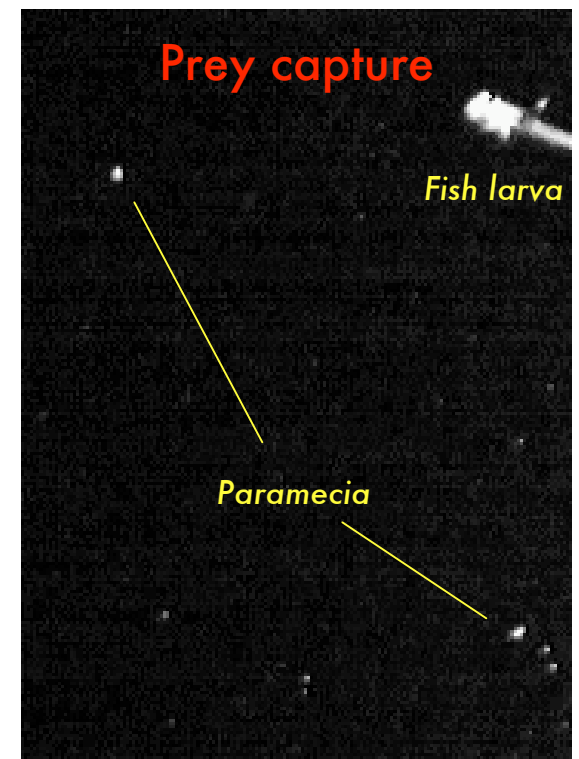
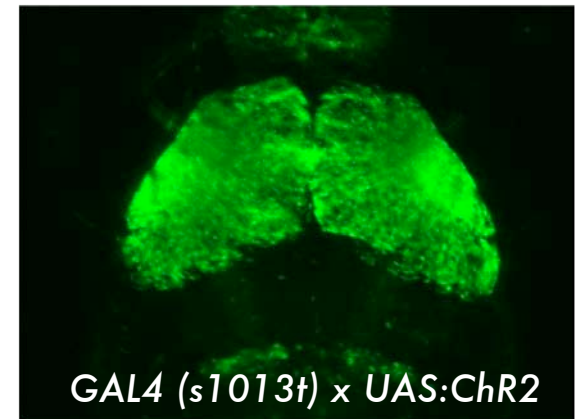
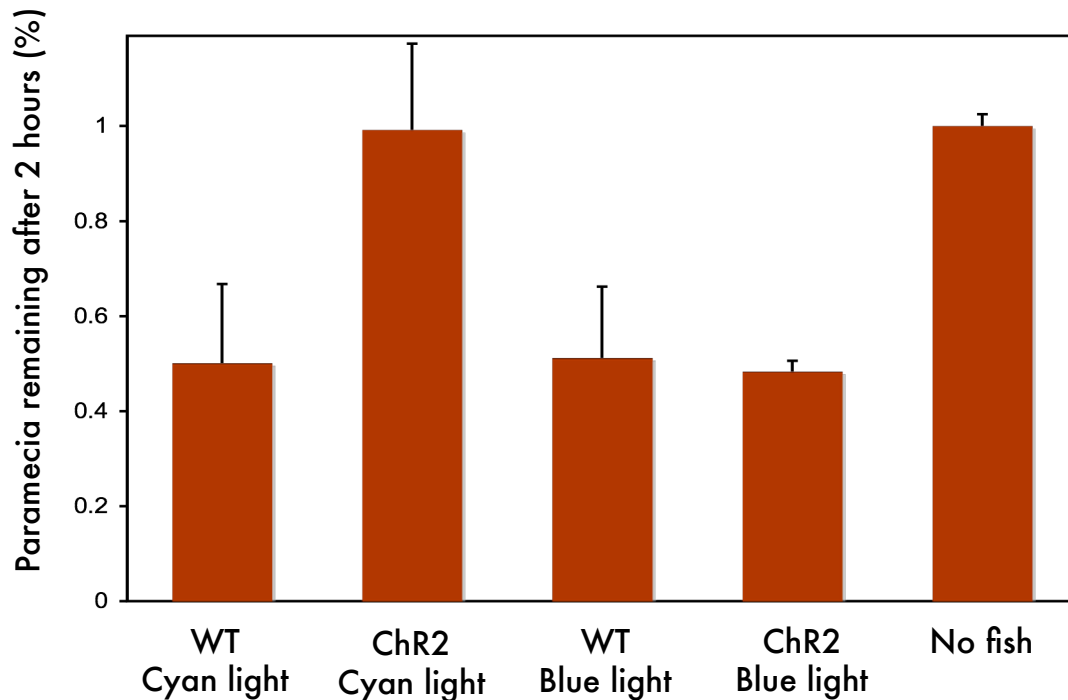
82 mutations in 55 genes
Degree of saturation: 25-40%

The zebrafish retinotectal map



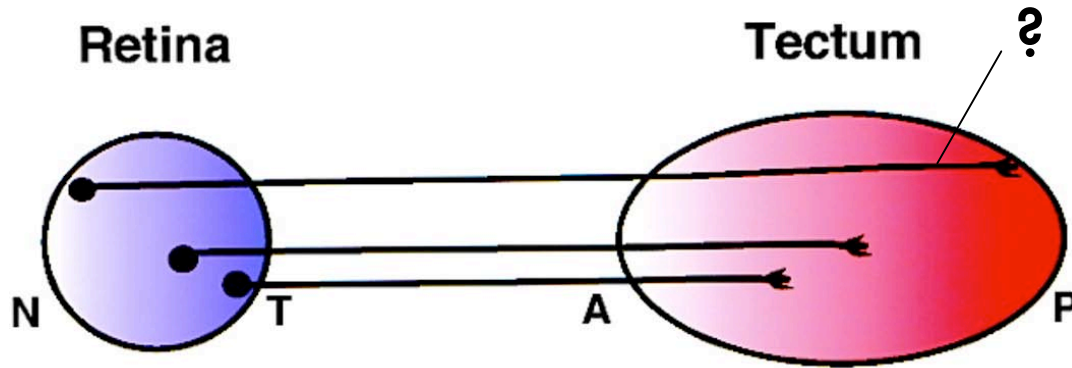
The retinotectal map is important for localizing small objects

- Tectum is required for prey capture (*not* for optomotor response) [Roeser et al., 2003; Gahtan et al., 2005]
- Precision of retinal axon targeting limits visual acuity [Smear et al., 2007]
- Perturbation of neuronal activity in the tectum with channelrhodopsin disrupts prey capture (*not* the optomotor response)

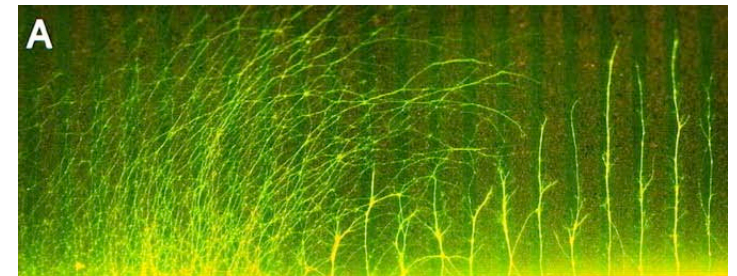


Filo Del Bene, Claire Wyart

What drives nasal axons into the posterior tectum?



Friedrich Bonhoeffer's stripe assay



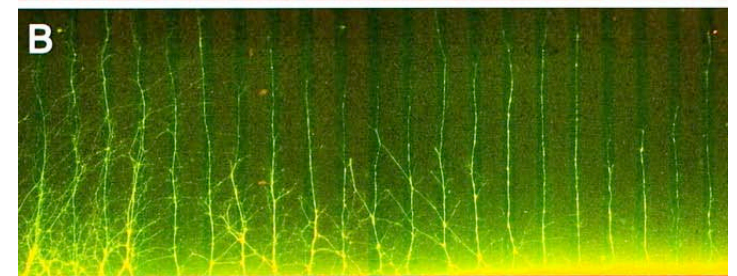
Ephrin-A2

Two possibilities:

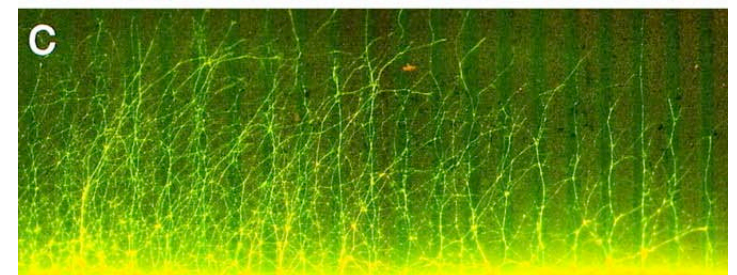
1) Axon-axon interactions
(e.g., competition)

2) Pure chemoaffinity
(e.g., a second gradient)

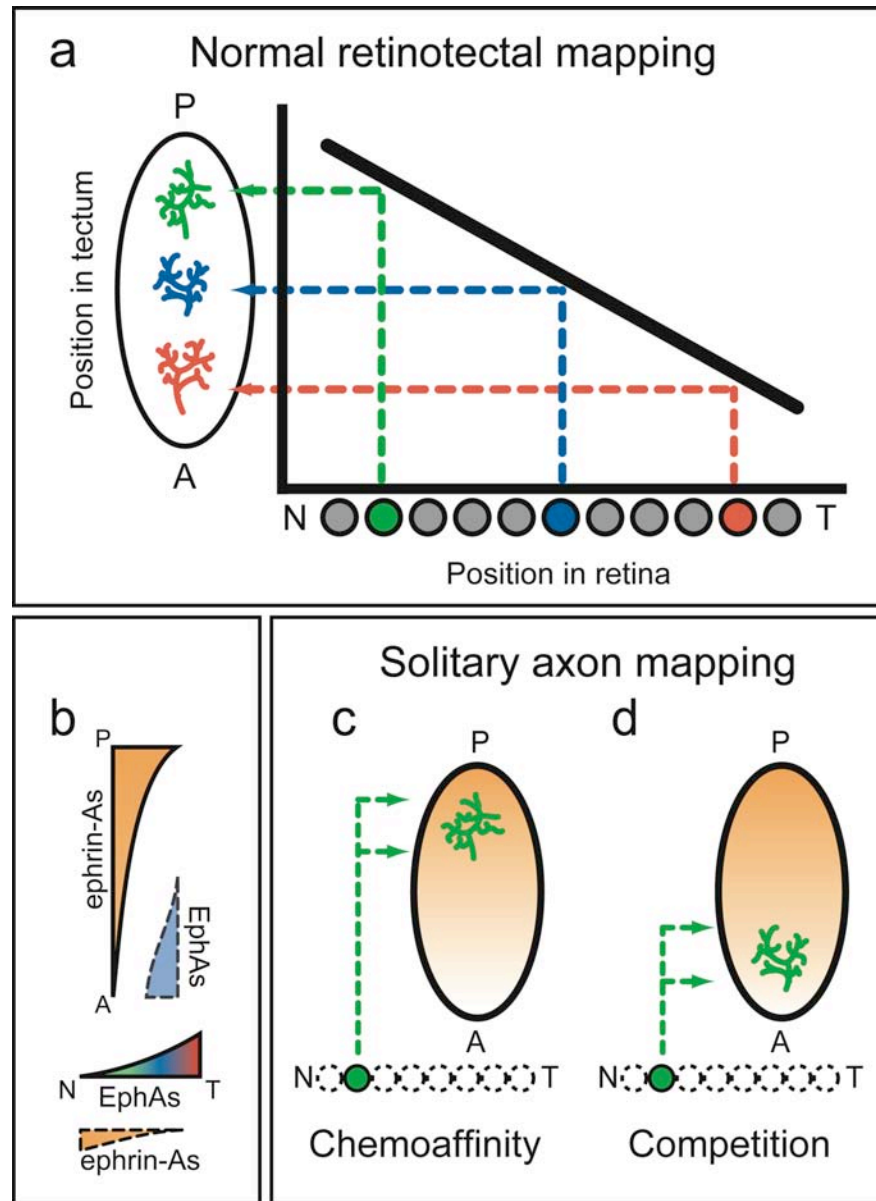
Ephrin-A5 (high)



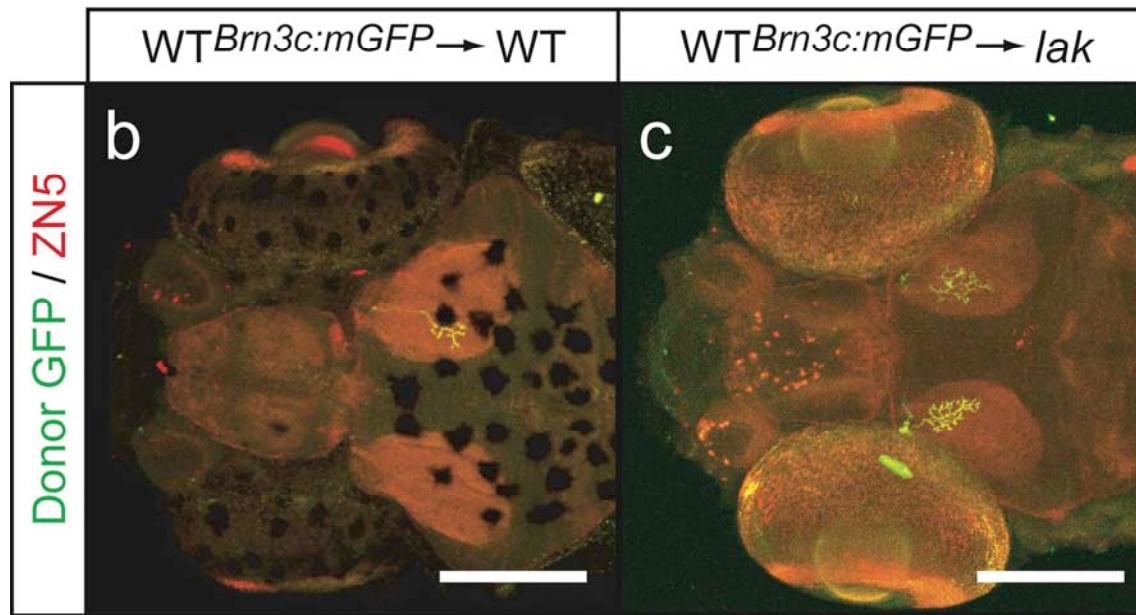
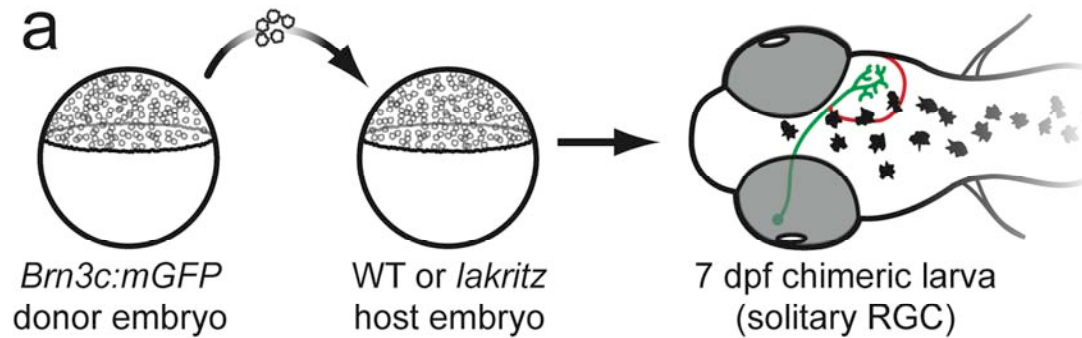
Ephrin-A5 (low)



Where will a solitary axon go in the tectum?

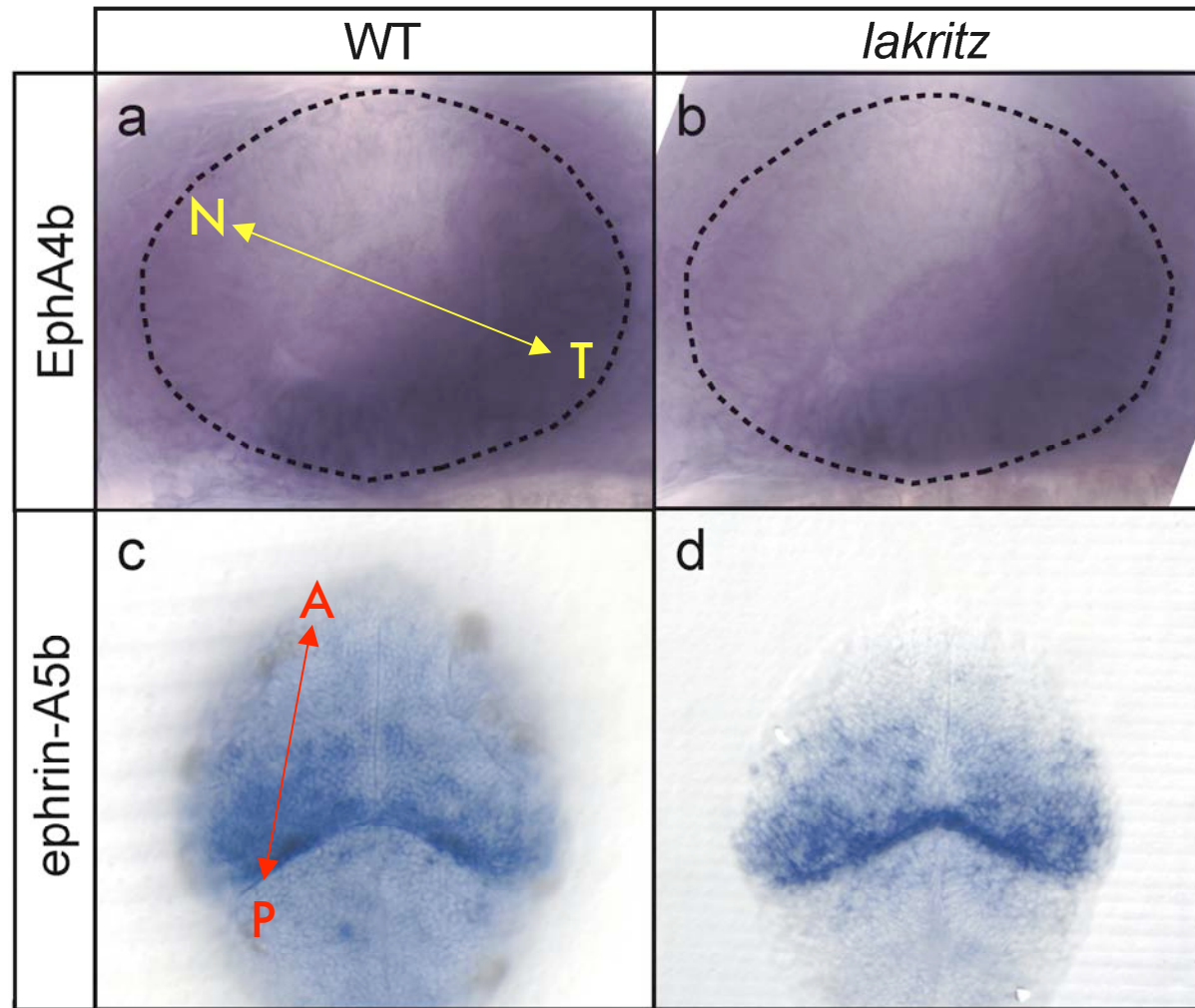


Experiment: Create retina with just one ganglion cell

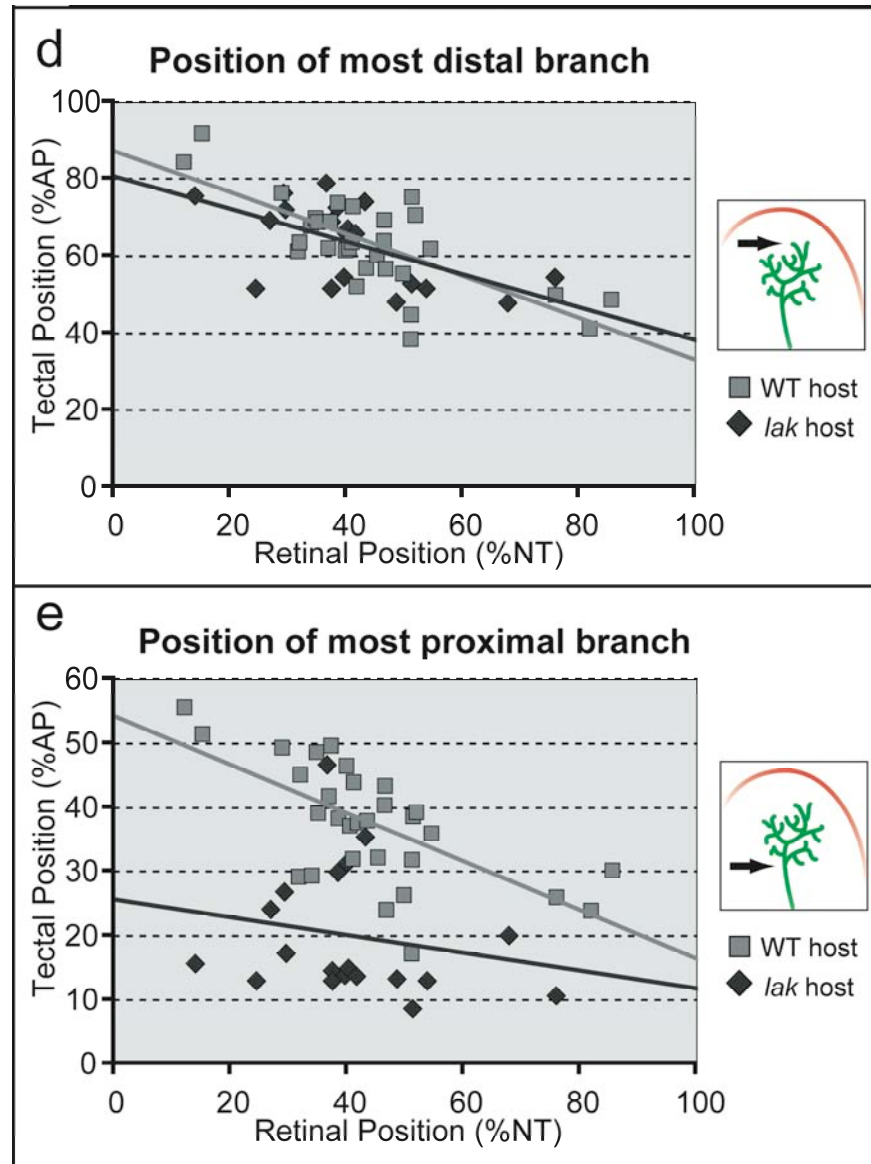


Zebrafish carrying the *lak* (*ath5*) mutation are unable to generate ganglion cells.

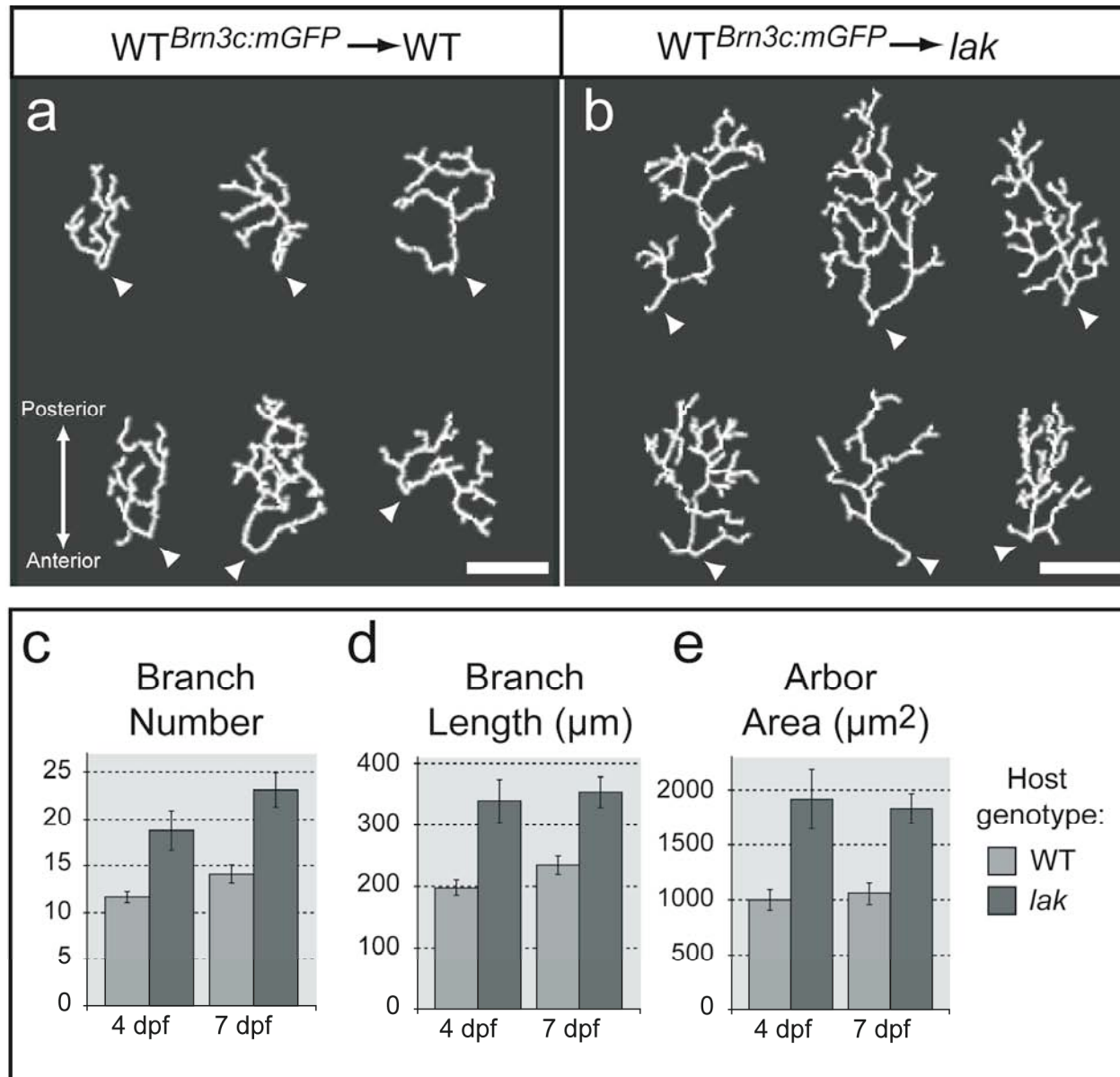
The *lak/ath5* retinal and tectal axes are properly patterned



Retinotopic order of solitary axons is maintained



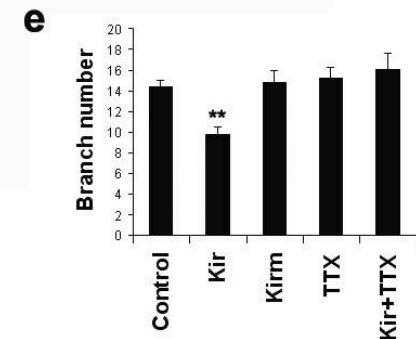
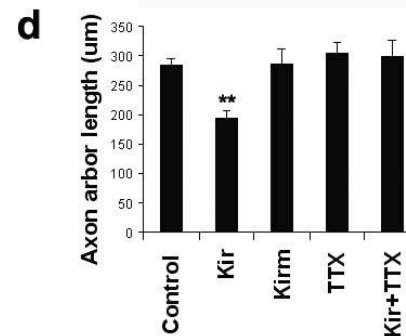
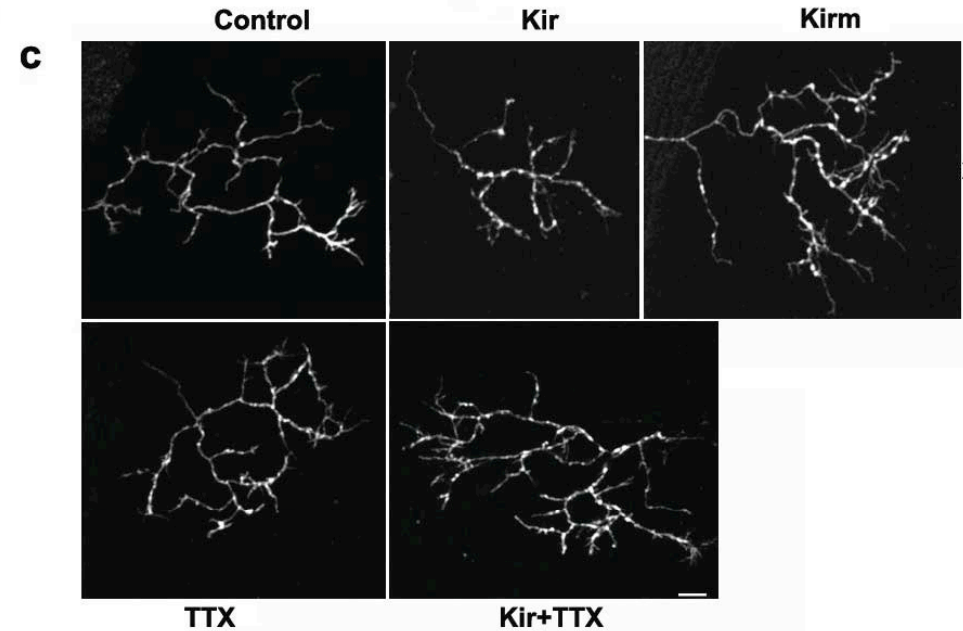
Axon arbors grow larger and more complex in the absence of other axons



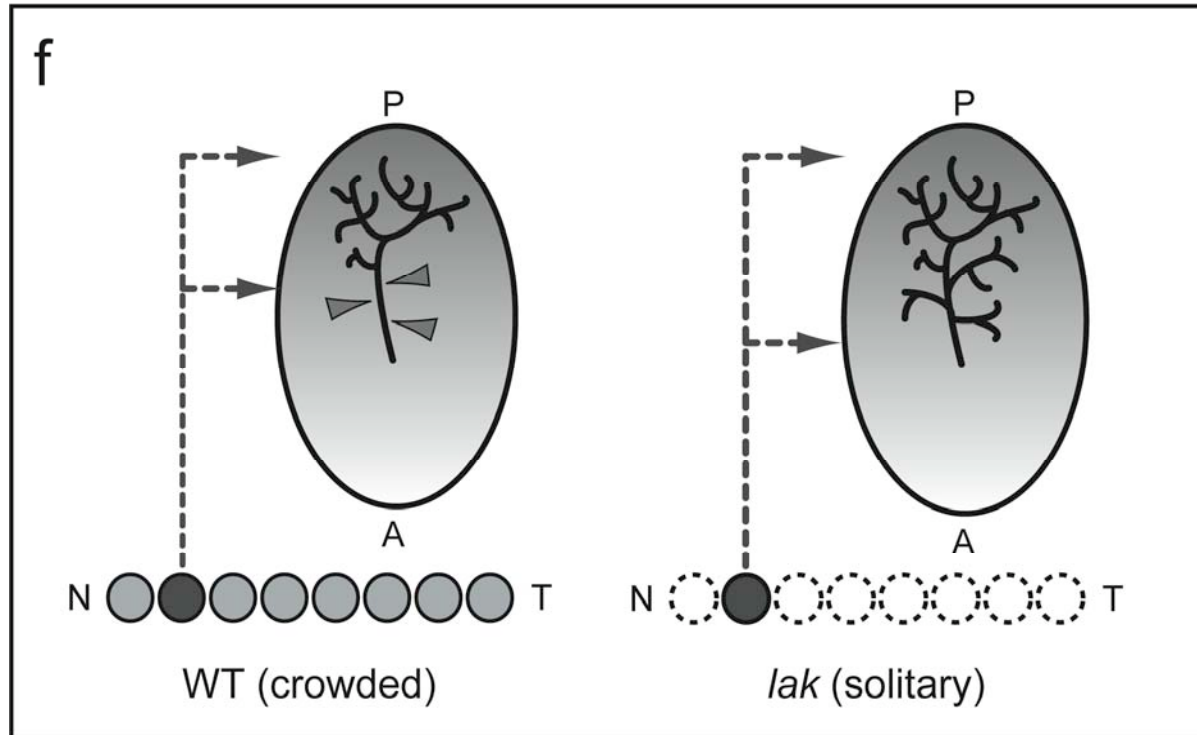
Triple role for activity in shaping axonal arbors

- Activity-based competition reduces arbor size of inactive axons among normally active axons (Hua et al., 2005).
- Homeostasis increases arbor size of less active axons in *blu/vglut2a* mutants (Smear et al., 2007).
- Correlated activity stabilizes overlapping arbors by Hebbian mechanism (Poo and Cline labs).

Either lack of competition and/or homeostasis could produce the solitary-axon phenotype.



Summary of this part

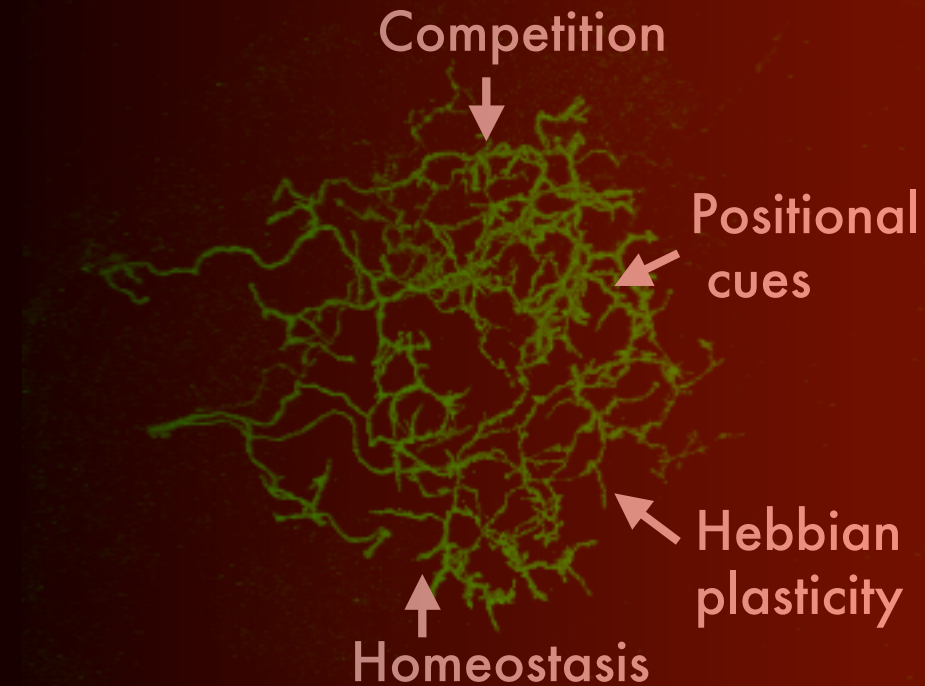


- Solitary axons project to their retinotopically appropriate targets.
- Competition or other axon-axon interactions are *not required* for map formation.
- Axon-axon interactions have strong effects on axon arbor size.

Discussion points/Caveats

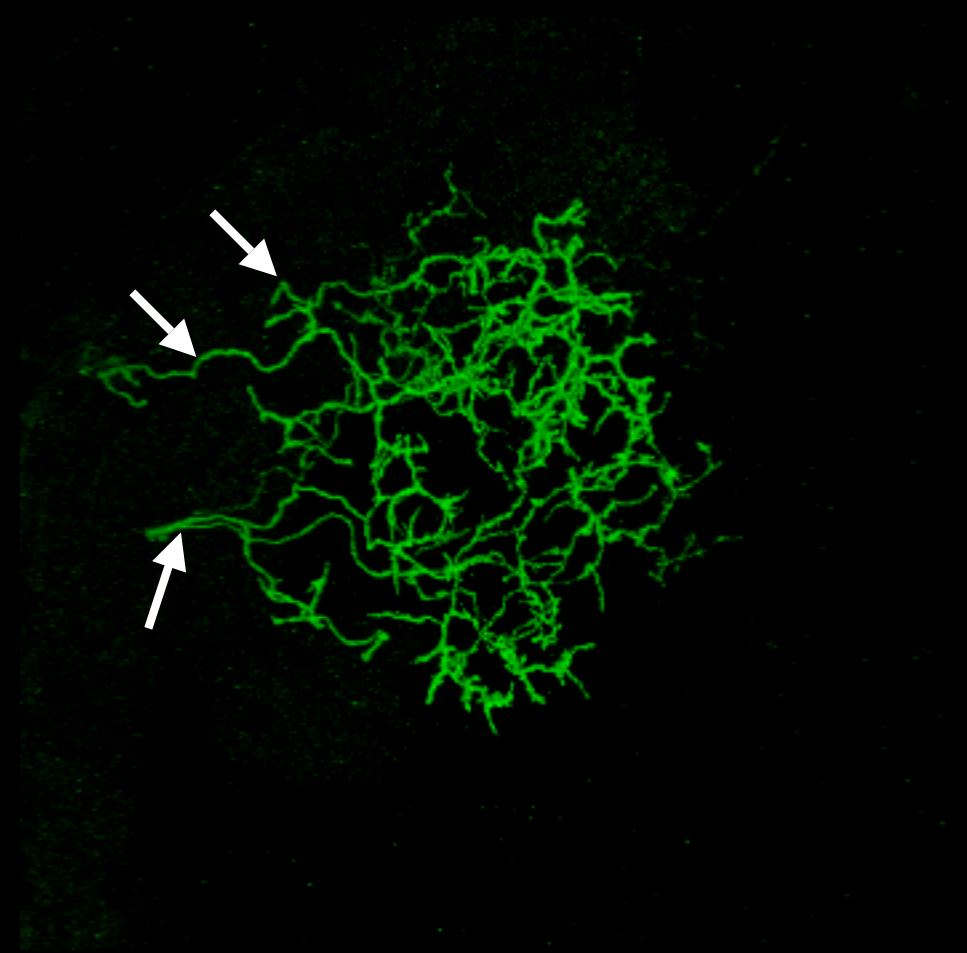
- Second gradient could be provided by EphA/ephrin-A signaling or other factors.
- Pathfinding to the tectum is not completely axon-autonomous (e. g., errors in the retina).
- Competition may still be important for mapping in crowded condition.
- Evolutionary view of species differences: Slow intrauterine development, delayed reliance on vision, and nocturnal lifestyle may have relaxed selection pressure on chemoaffinity mechanisms in mammals.

Retinotopic Mechanisms

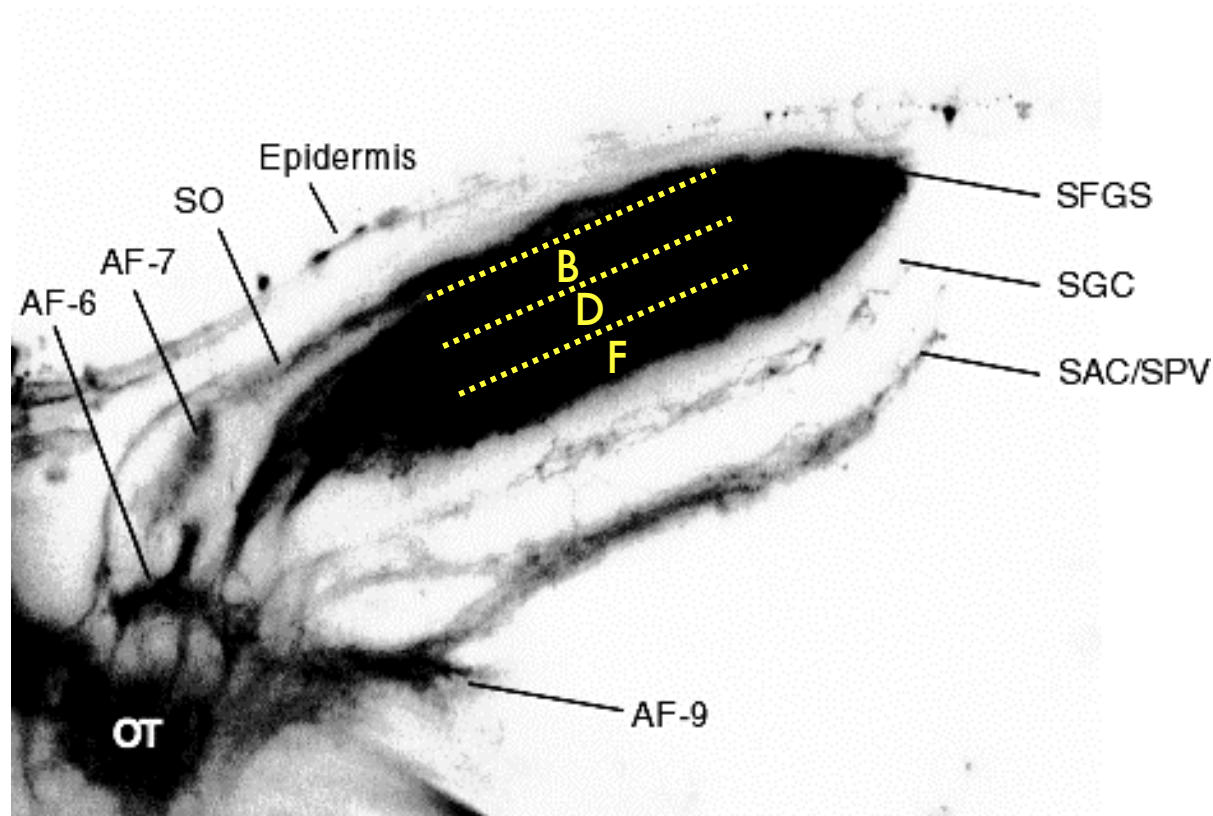


Ephrin-A

Planar organization of axonal arbors: Mechanisms?

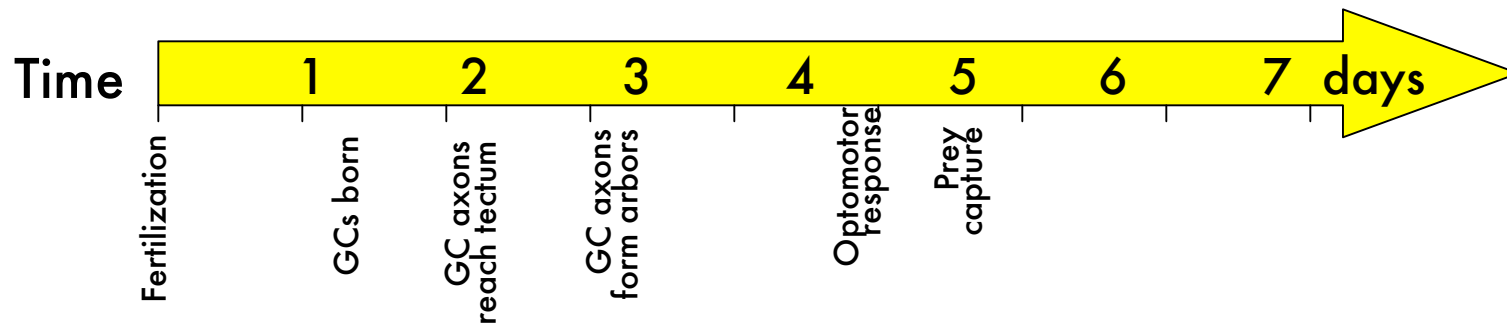
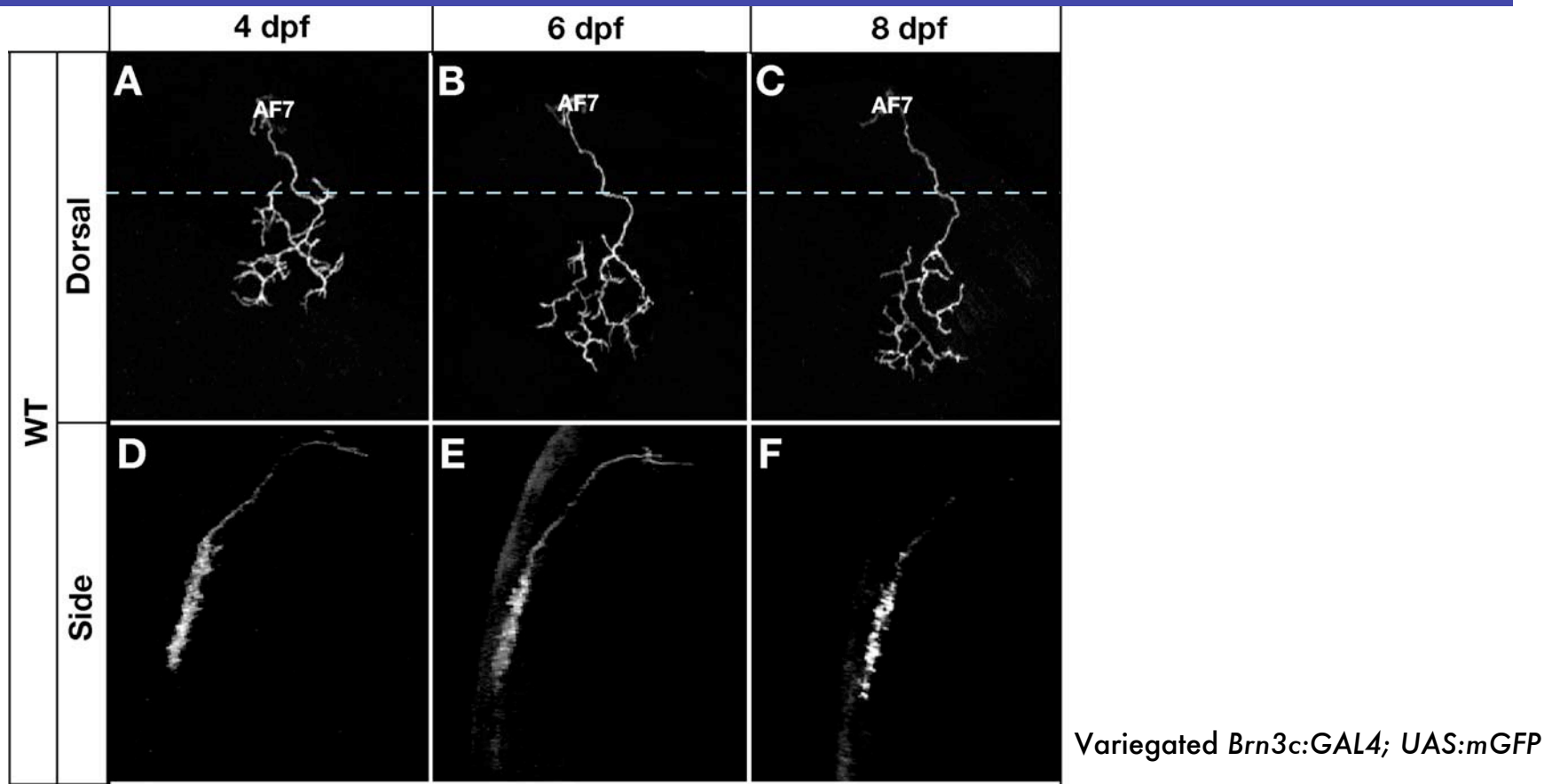


Retinal input to the tectum is organized in four layers

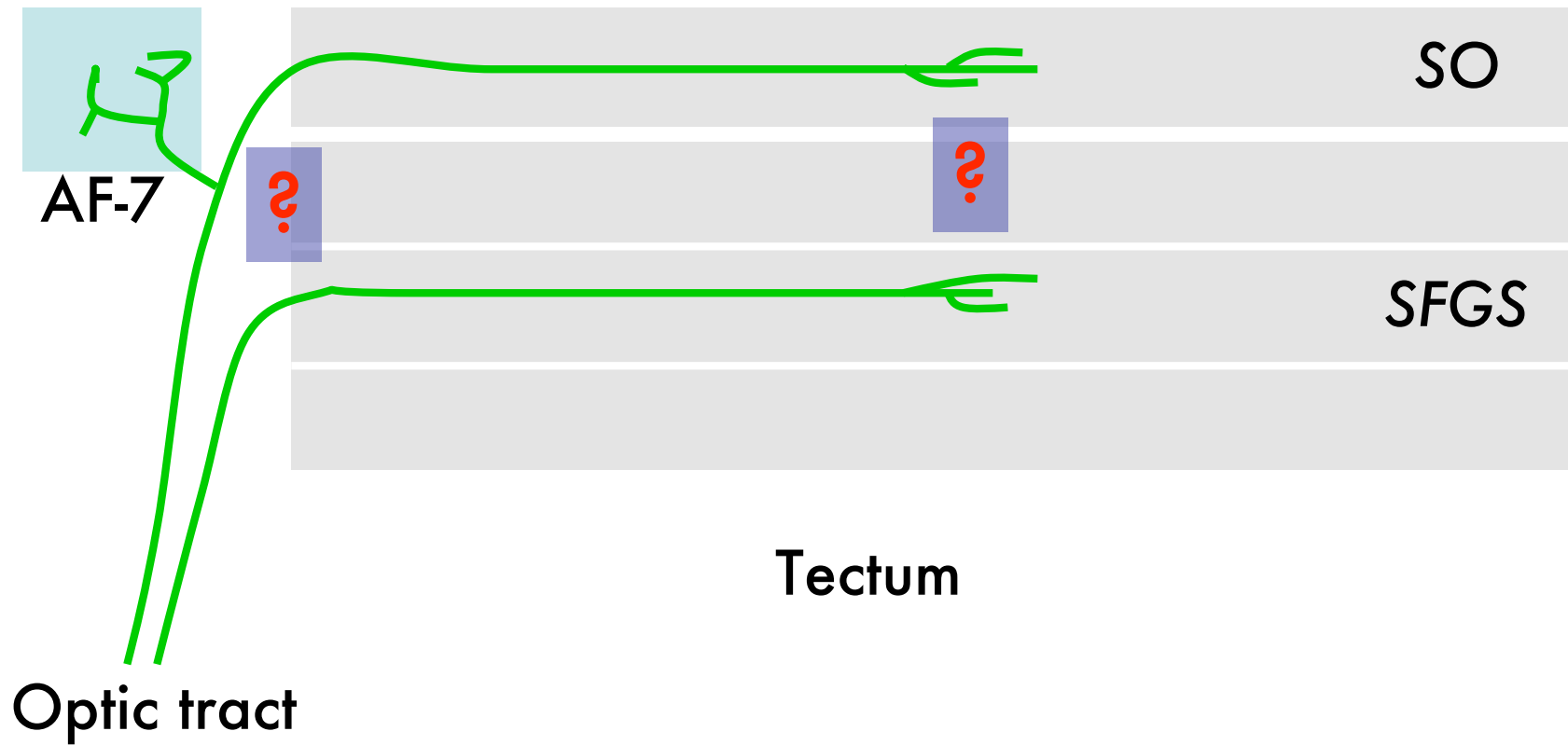


Whole-eye DiO fill of all ganglion cells

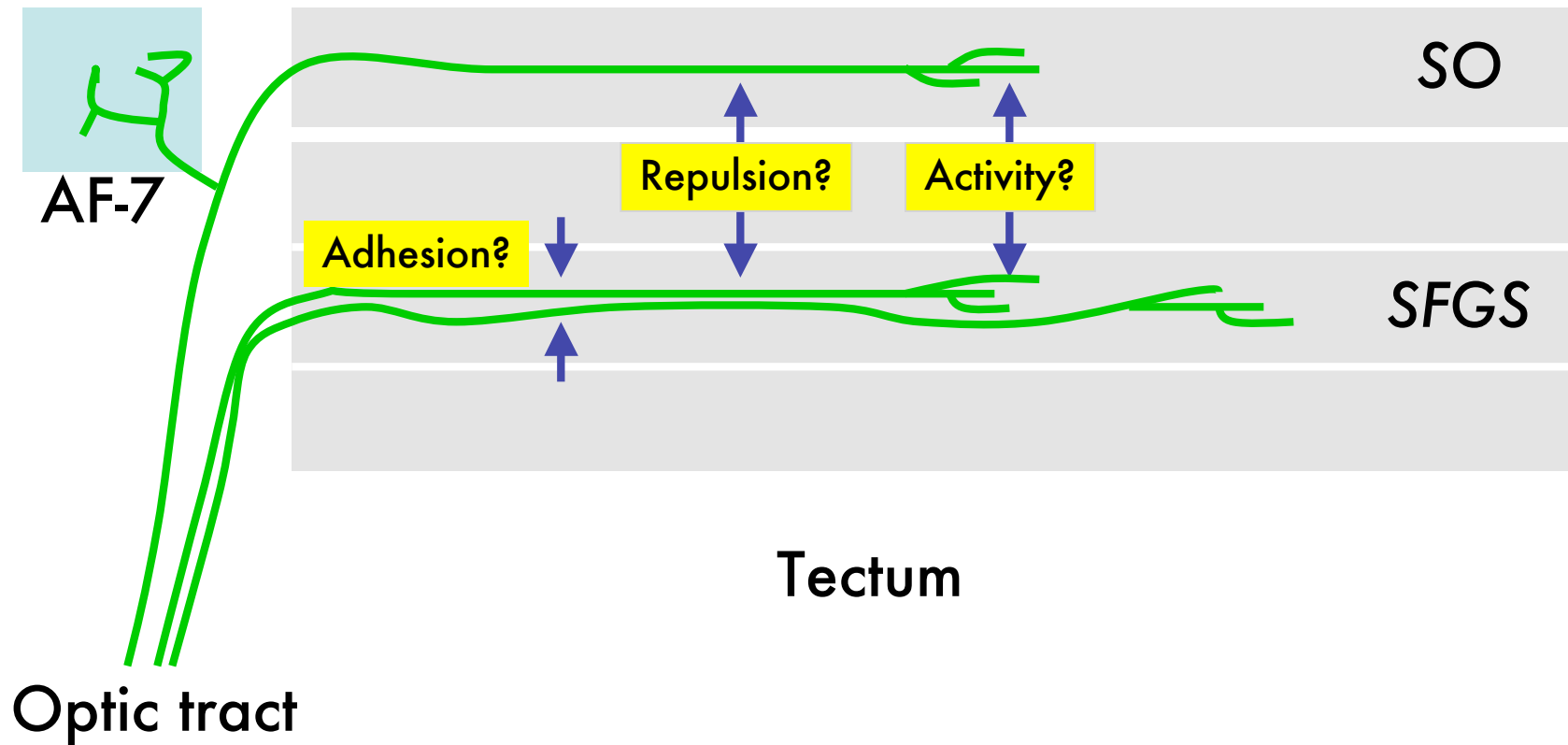
Retinal axon arbors are targeted precisely and remain confined to one lamina during remodeling and shifting



How is laminar specificity initiated and maintained?



Model 1: Lamina segregation by axon-axon interactions

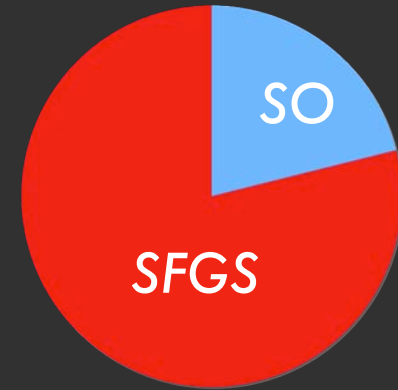
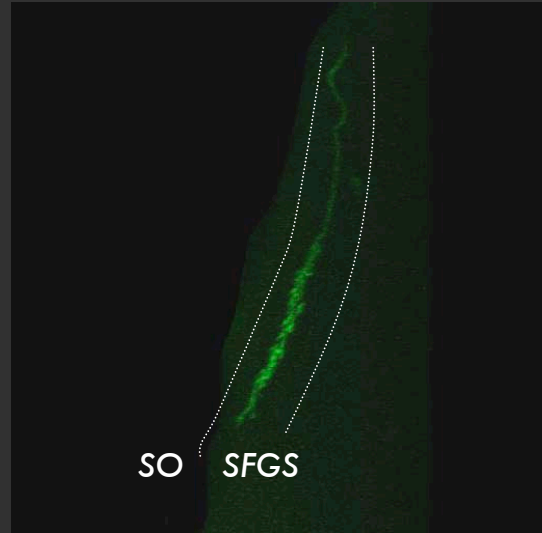
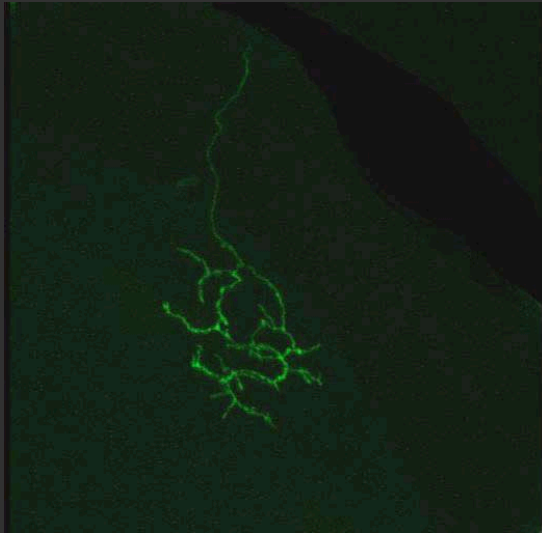


Precise lamina-specific targeting of solitary axons

Dorsal view

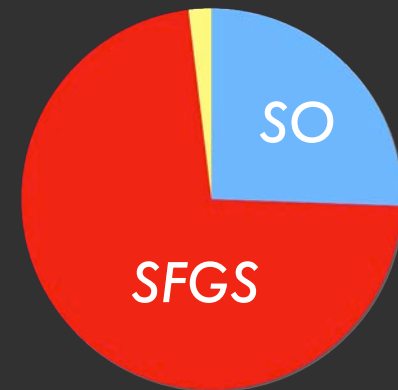
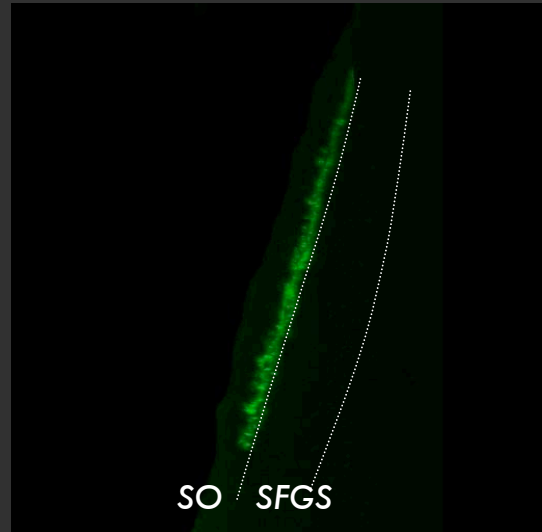
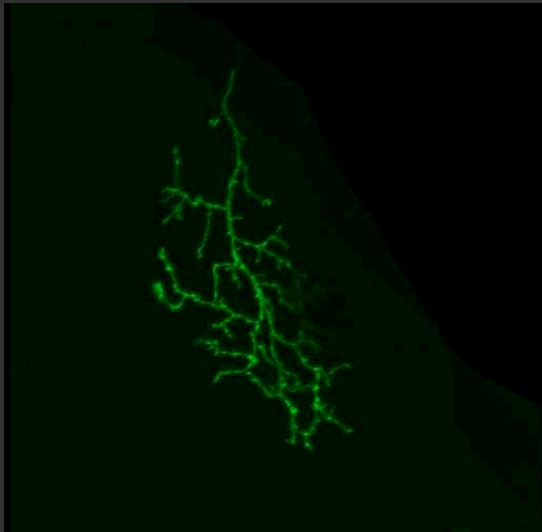
Side view

WT RGC in WT



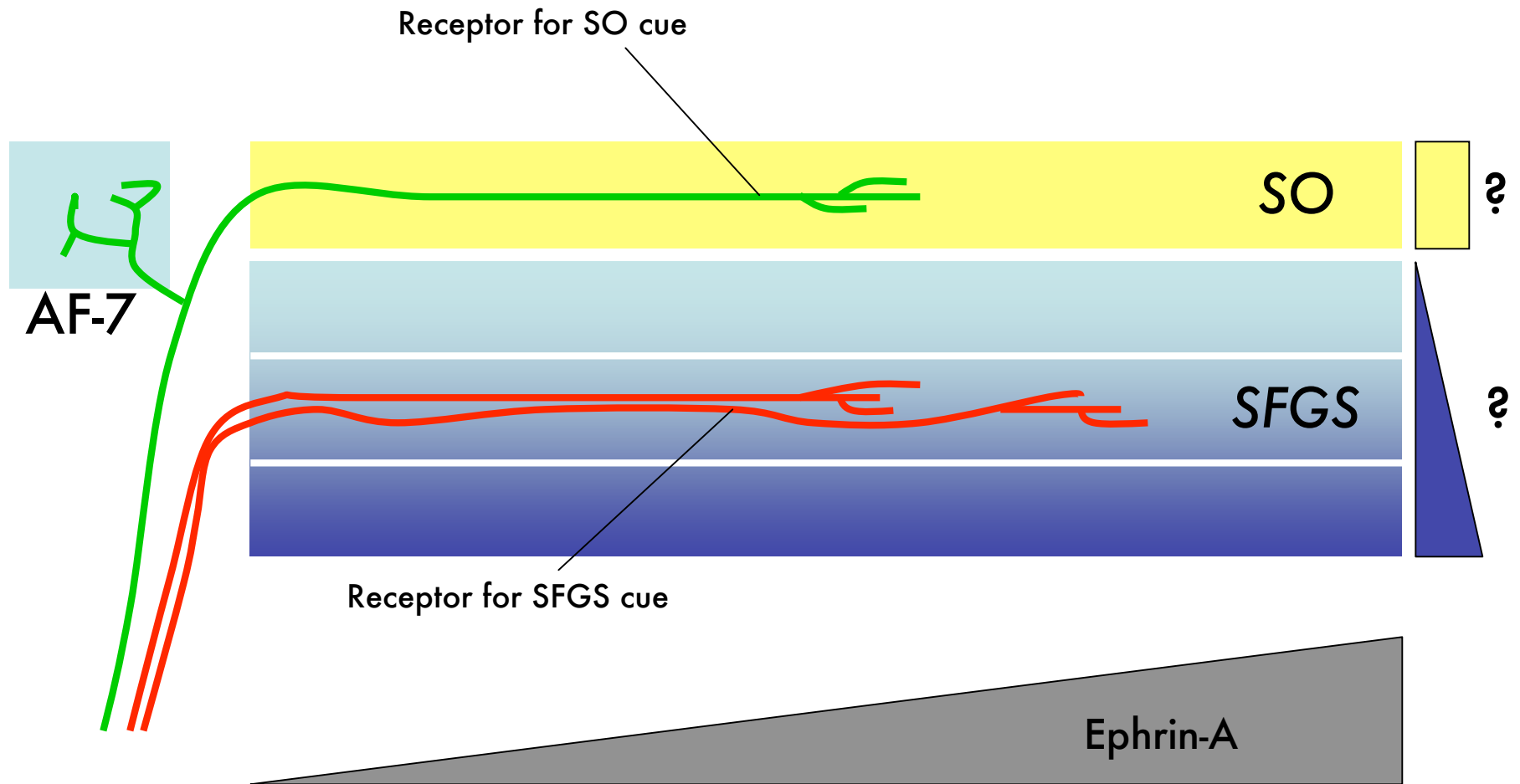
n = 104

WT RGC in *lakritz*



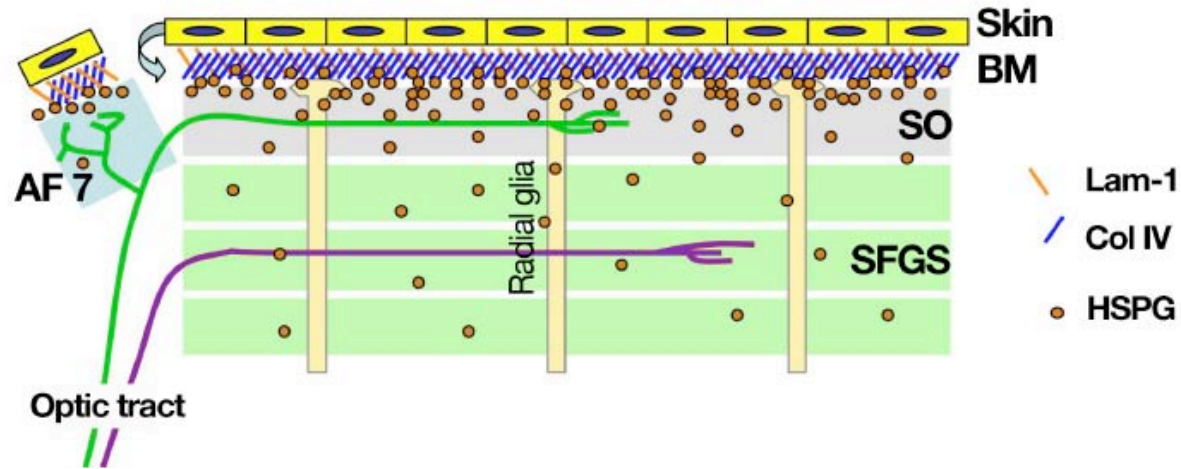
n = 51

Model 2: Segregation by laminar positional cues in the tectum

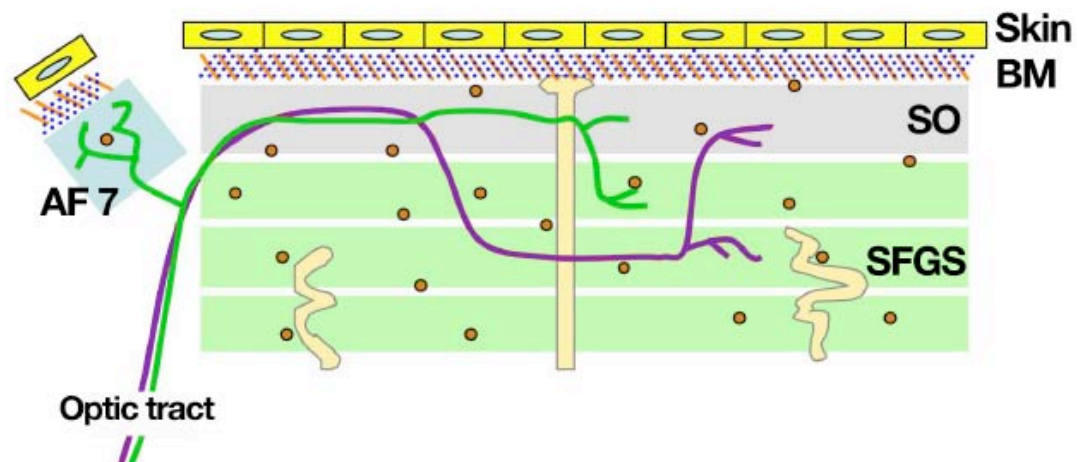


Laminar guidance by secreted guidance cues (HSPG, Slit) anchored to the surface of the tectum

a Wild type



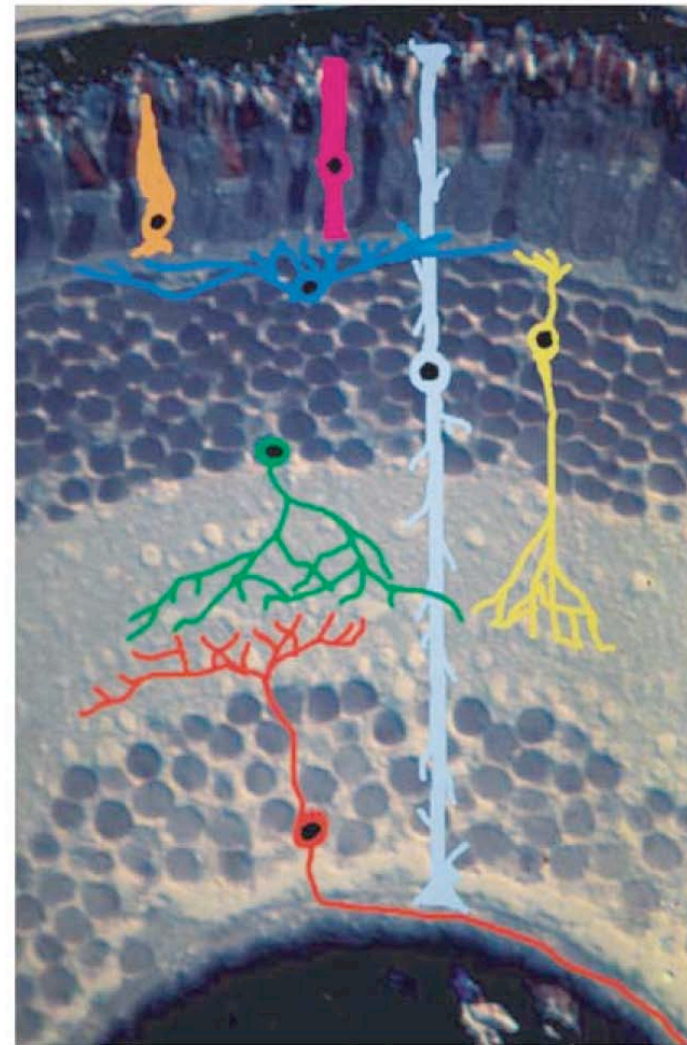
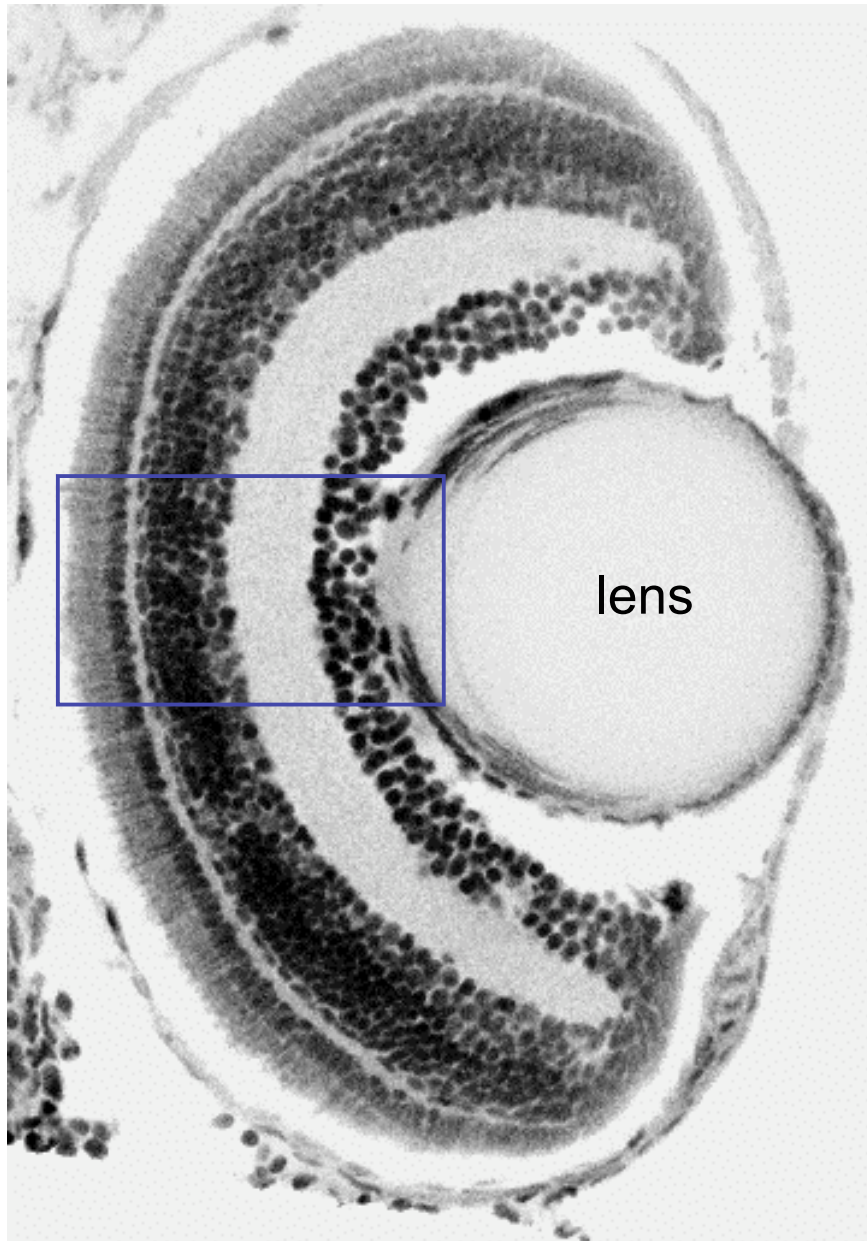
b *dragnet*



Conclusions

- Lamina-specific targeting is precise from the start; no evidence for exuberant branching and subsequent pruning.
- Retinal ganglion-cell axons show laminar specificity in the tectum even in the absence of other axons.
- Mutations reveal the function of secreted molecules and axonal receptors in lamina-specific targeting.
- Axonal laminar specificity is essential for visual processing.

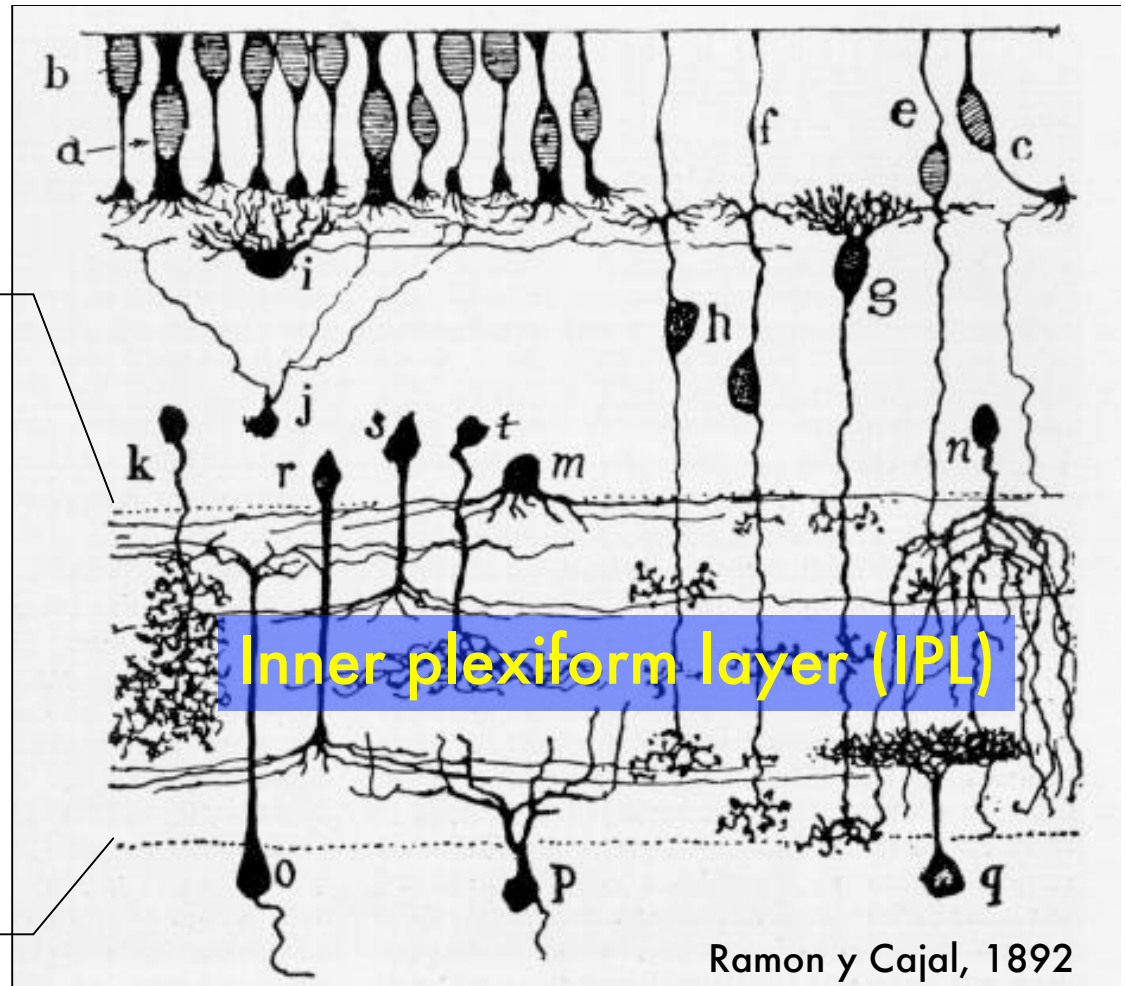
Assembly of synaptic layers in the retina



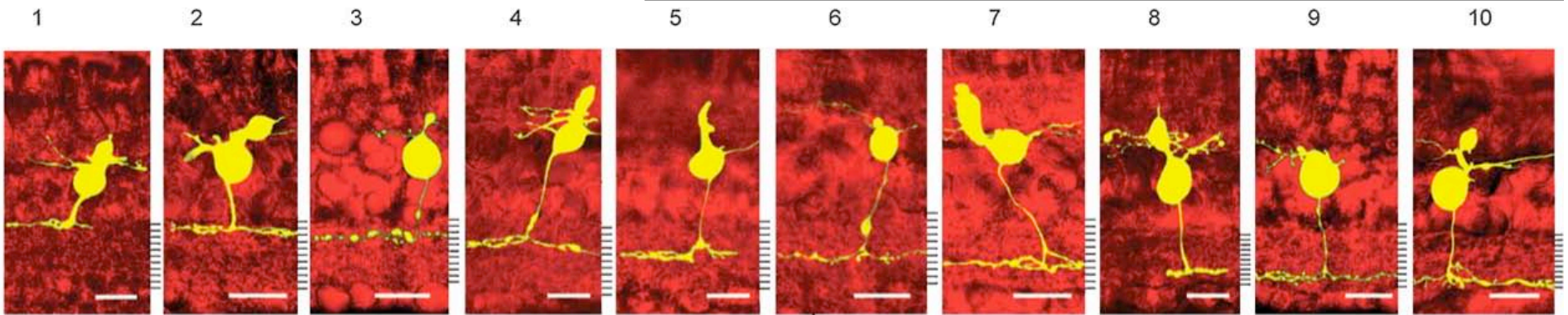
Bill Harris

The IPL is a stack of ca. 10 parallel circuits

- 1
- 2
- 3
- 4
- 5
- 6
- 7
- 8
- 9
- 10

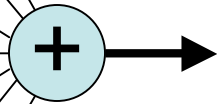


Ramon y Cajal, 1892



Pang, Gao & Wu (J. Physiol., 2004)

In the retina, the visual image is parsed into features by cells with distinct response properties

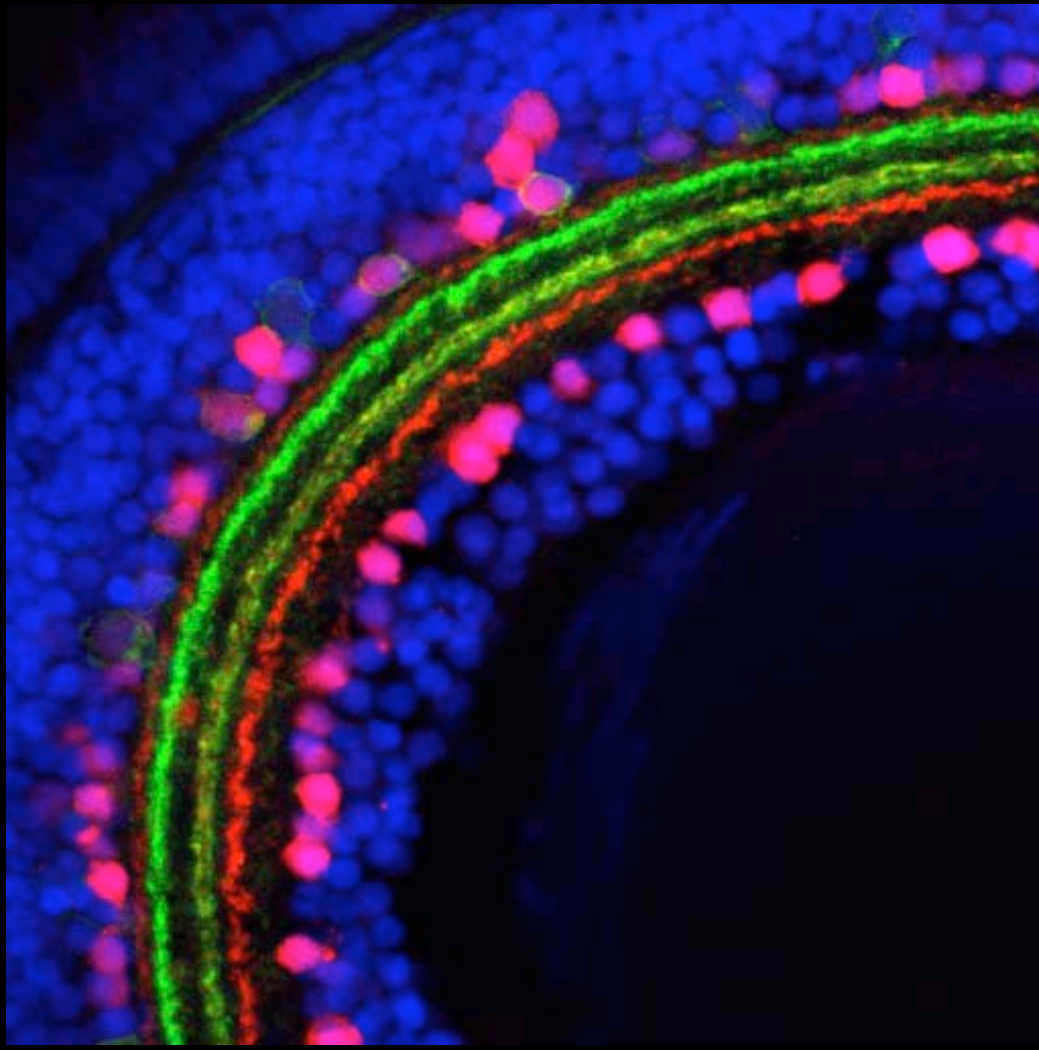


[The face of Frank Werblin; animated in original]

Channels of visual processing:

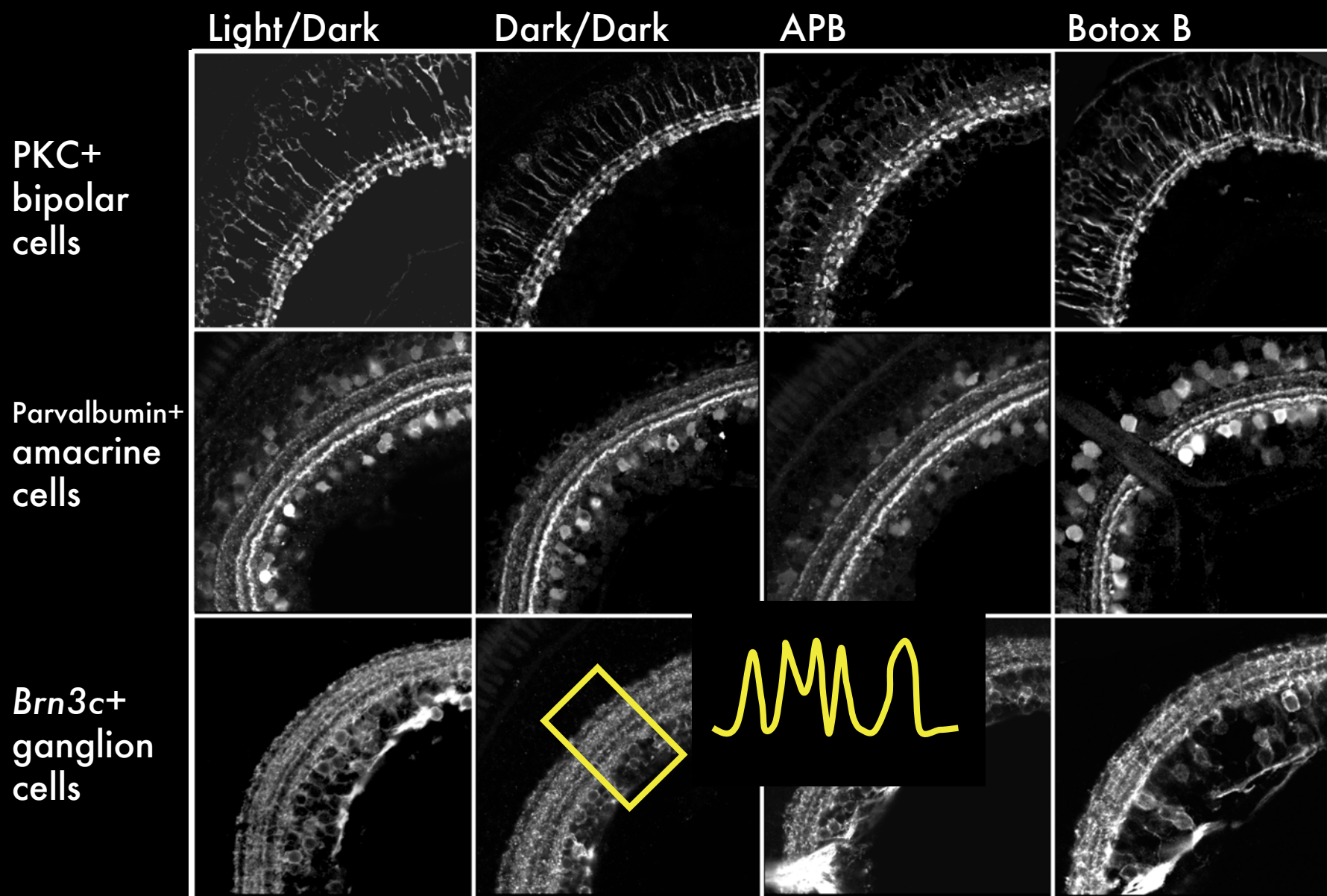
- brightness
- spatial contrast
- temporal contrast
- motion
- direction
- color

IPL sublaminae in zebrafish

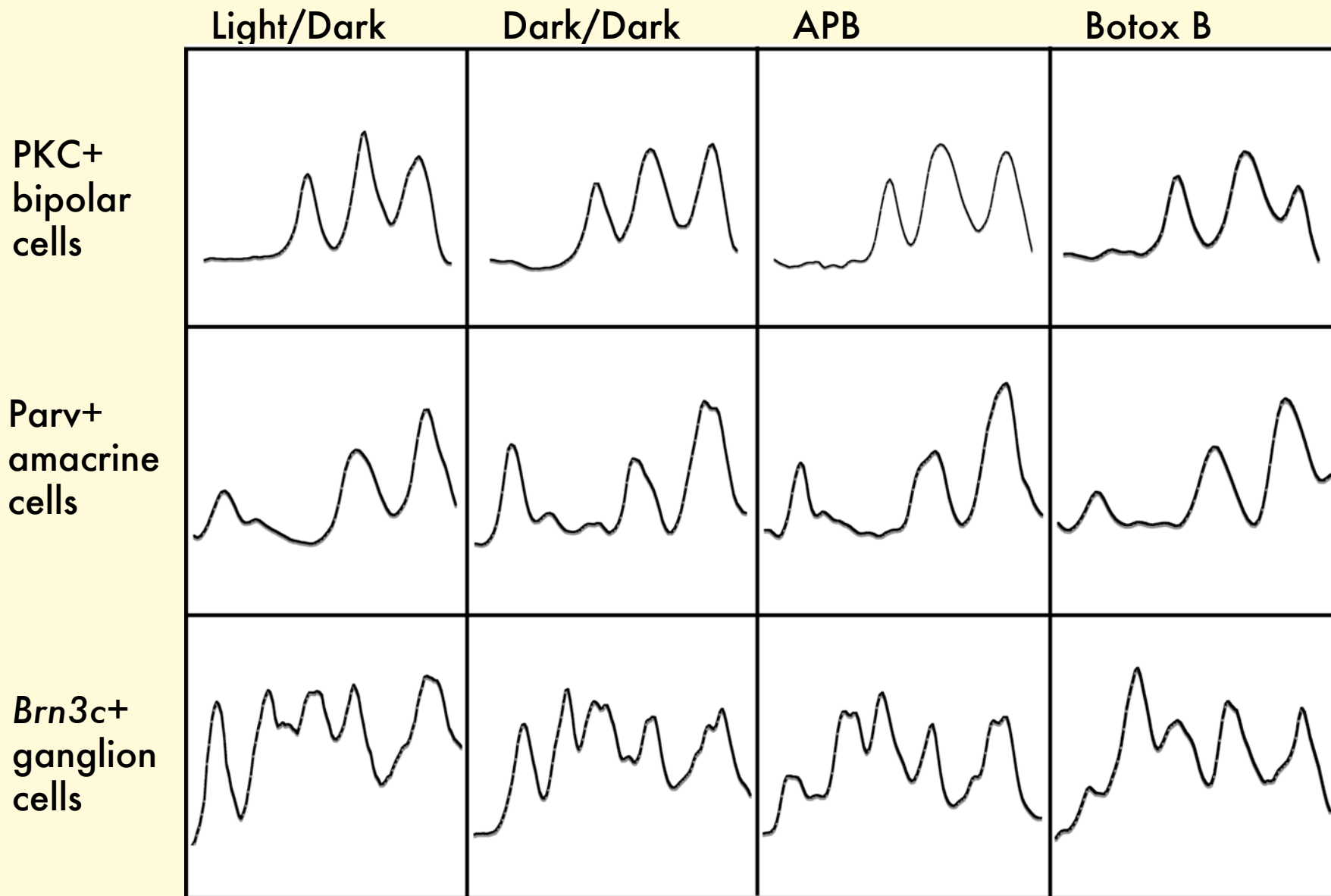


DAPI/*Pax6:mGFP*/Parvalbumin

Resilience of sublaminal circuitry to perturbations of activity



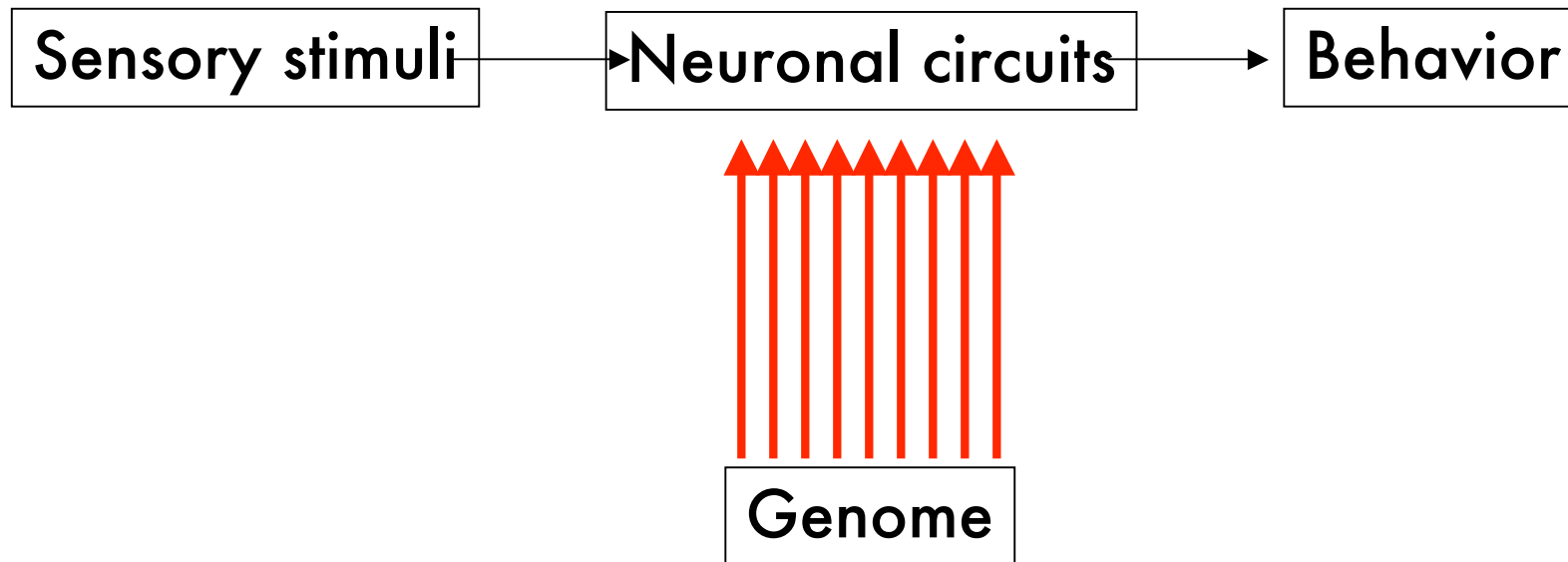
Resilience of sublaminal circuitry to perturbations of activity



Conclusions

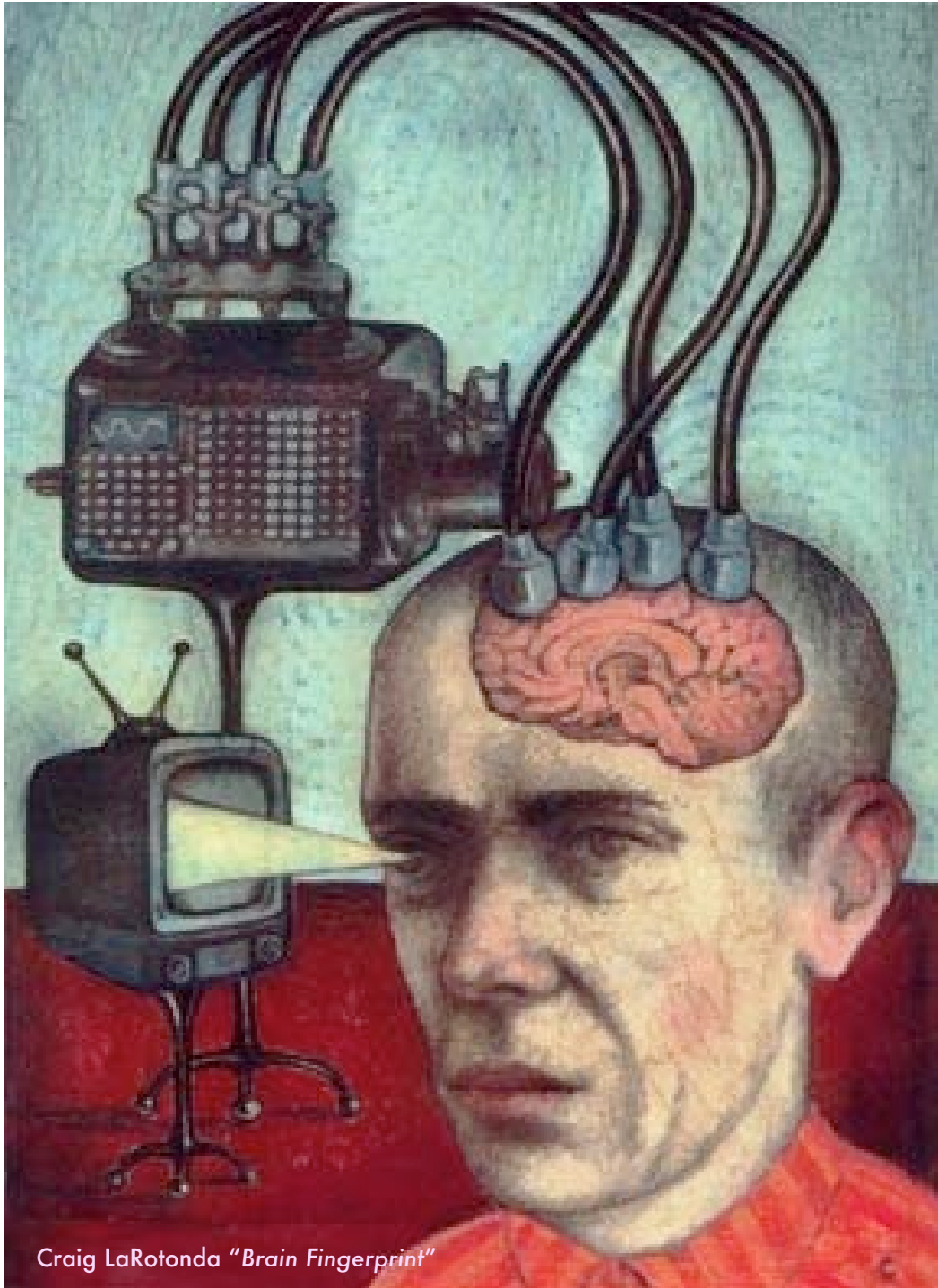
- Sublaminar targeting in the IPL is precise from the start; no evidence for exuberant branching and subsequent pruning.
- Lack of visual experience, lack of synaptic transmission, or imbalanced synaptic input do not prevent, delay, or otherwise alter synaptic sublamination.

Massive transfer of information from genes to synaptic circuitry



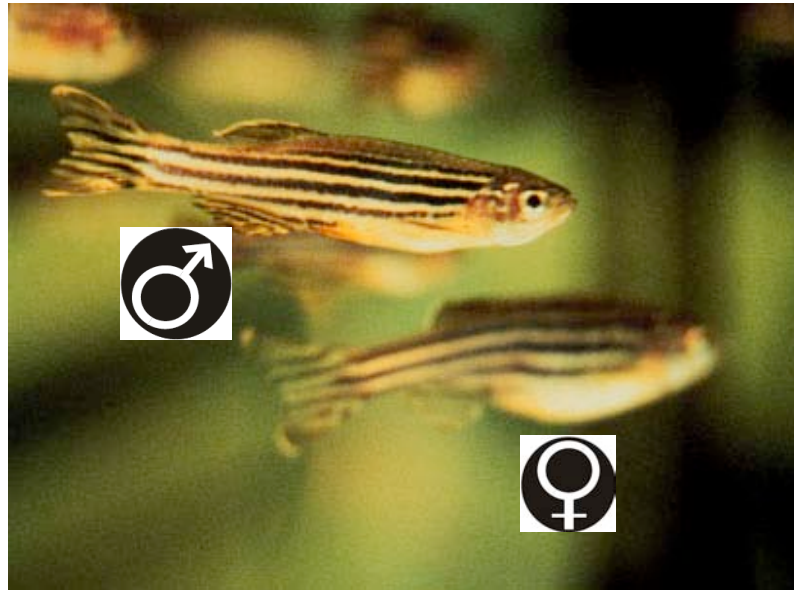
Remote optical control of neuronal activity

- Installation of a genetic switch in subsets of neurons by GAL4/UAS
- Optical manipulation of neural activity: channelrhodopsin, halorhodopsin, LiGluR, et al.
- Large-scale production of GAL4 enhancer-trap lines by Tol2 transposition.



Craig LaRonda "Brain Fingerprint"

GAL4/UAS



Tissue-specific promoter

gal4

X

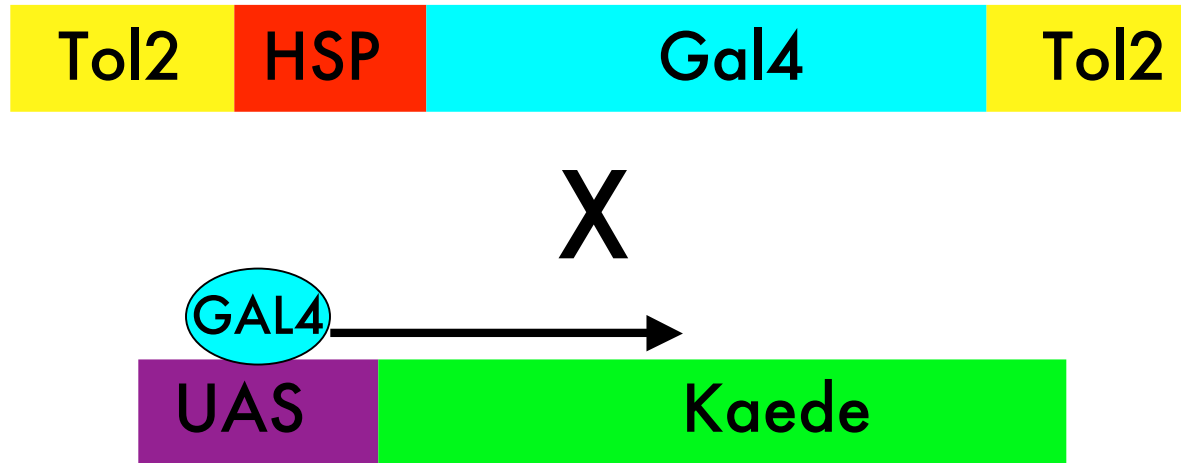
GAL4

UAS

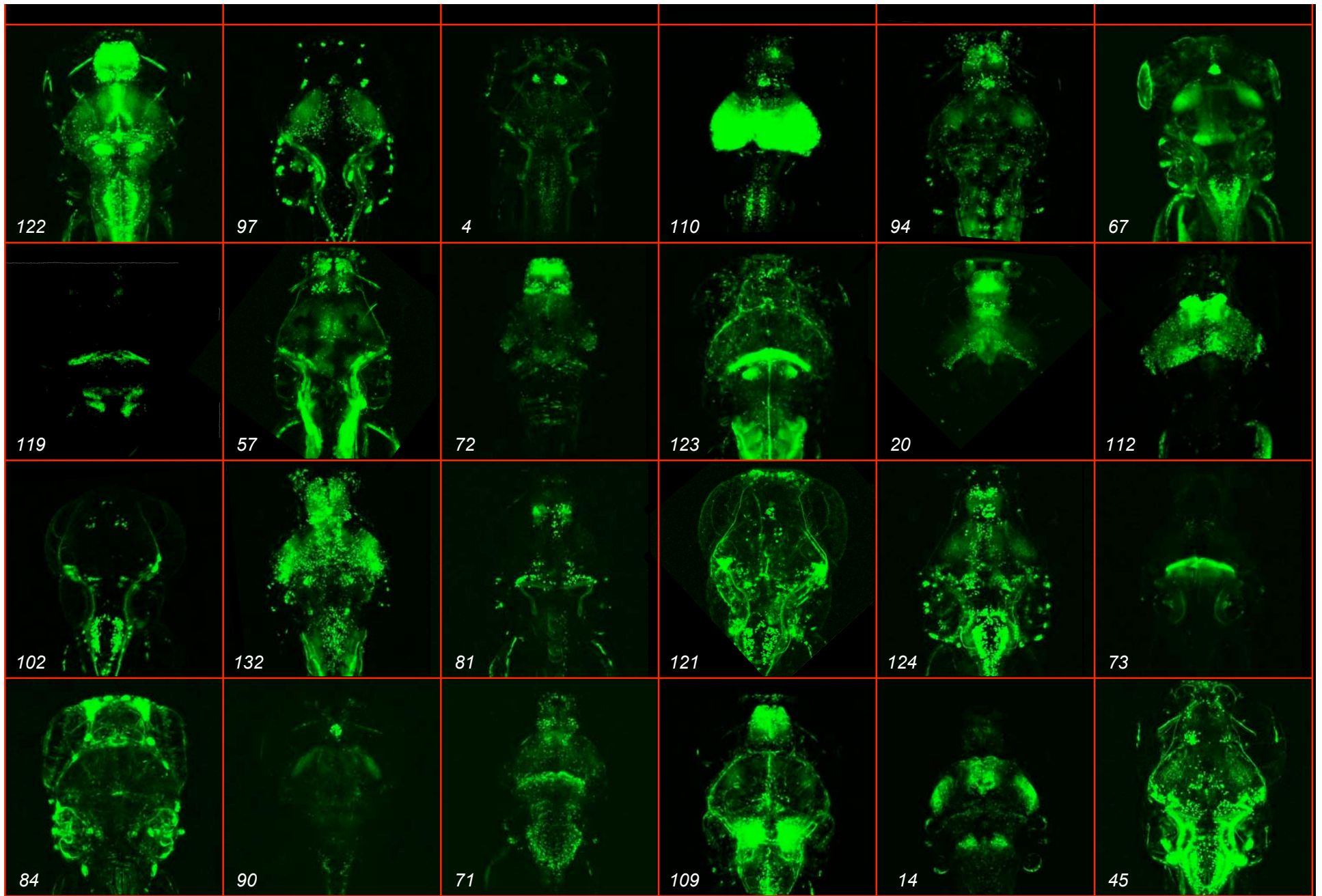
YFG

GFP, ChR2...

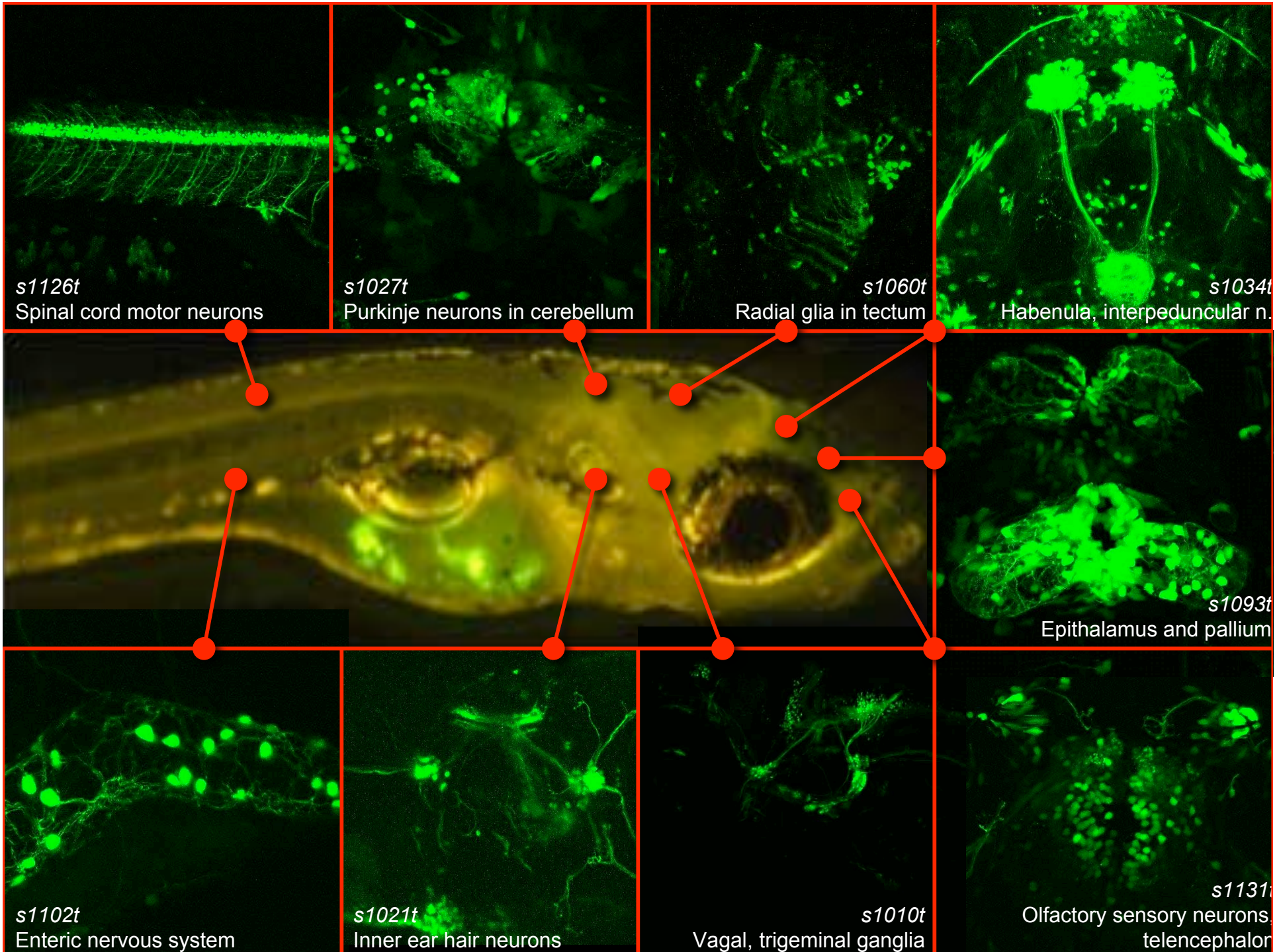
GAL4 Enhancer Trapping



- Inject GAL4 construct with transposase mRNA into 1-cell stage embryos
- Raise potential founders (F_0)
- Cross to UAS:Kaede to identify successful traps
- Raise offspring (F_1) to establish stable lines



A collection of 200 GAL4 enhancer trap patterns in the CNS



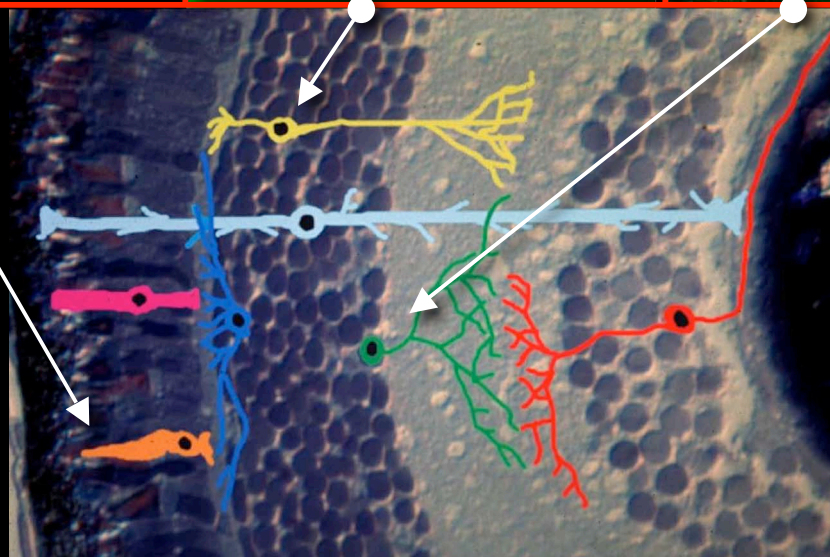
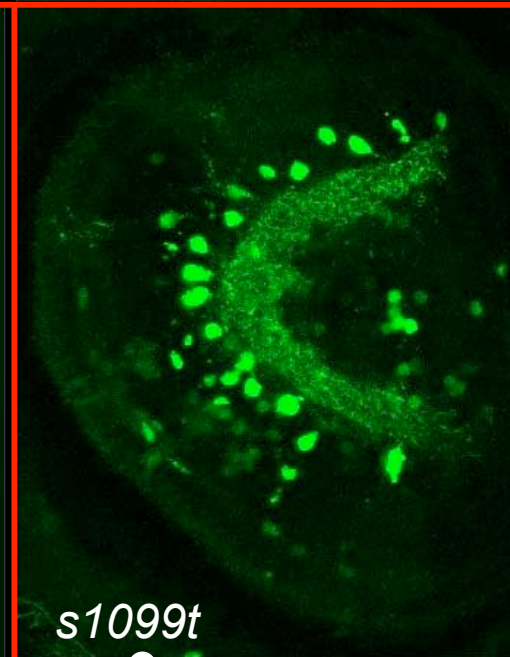
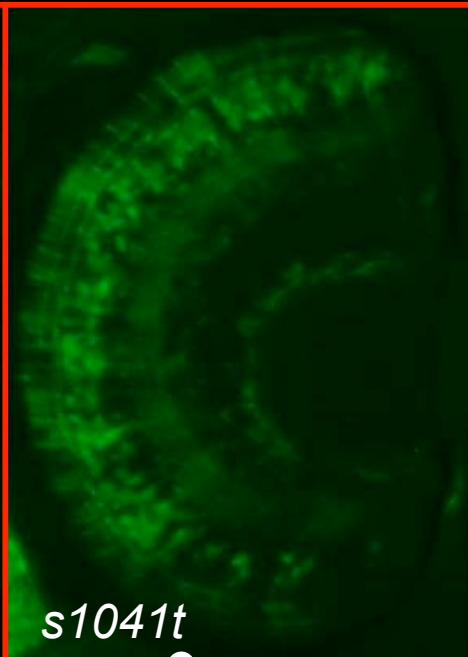
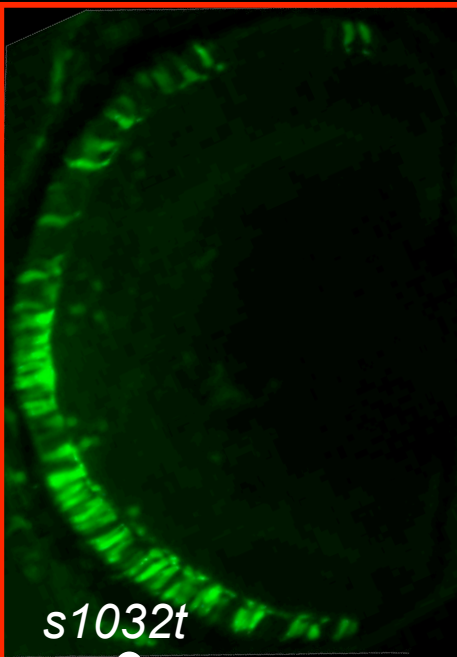
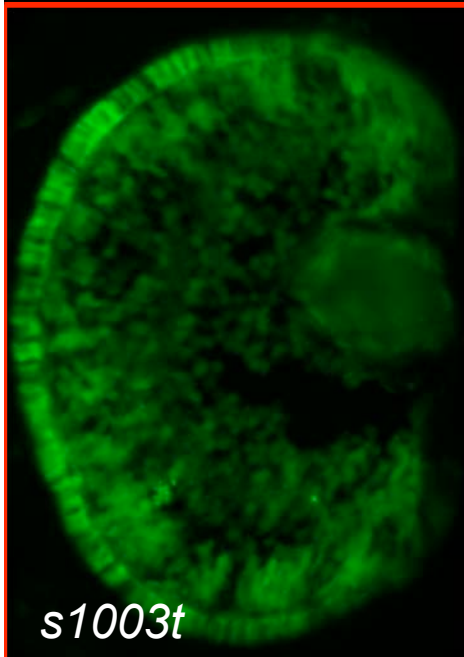
Retina-specific enhancer traps

All retinal cells

Cone photoreceptors

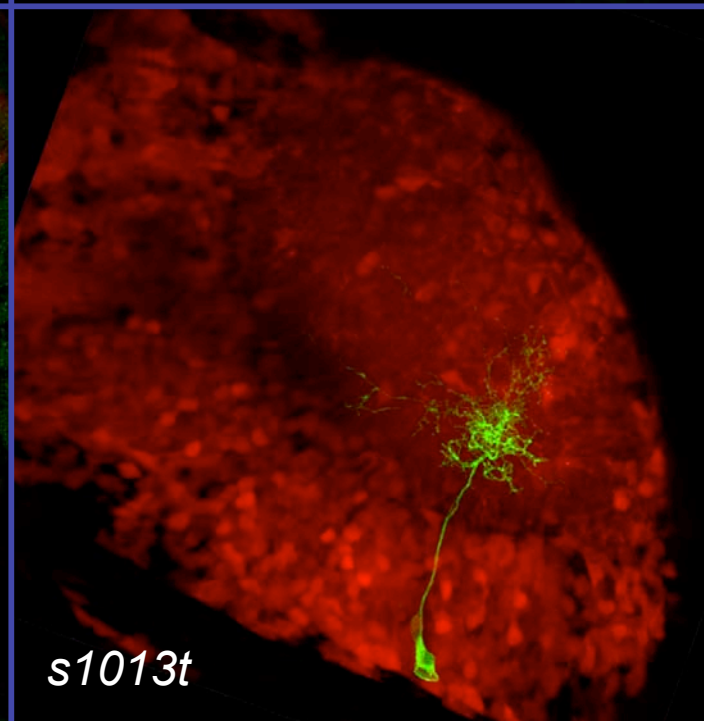
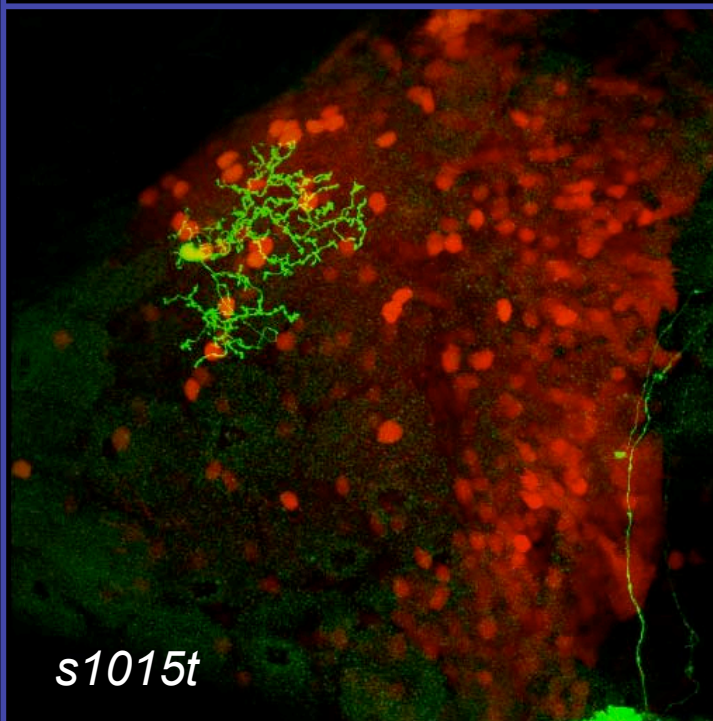
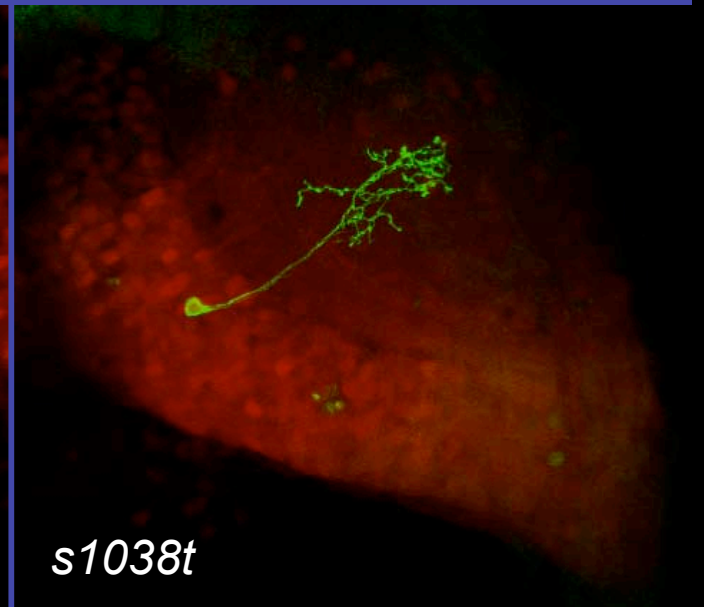
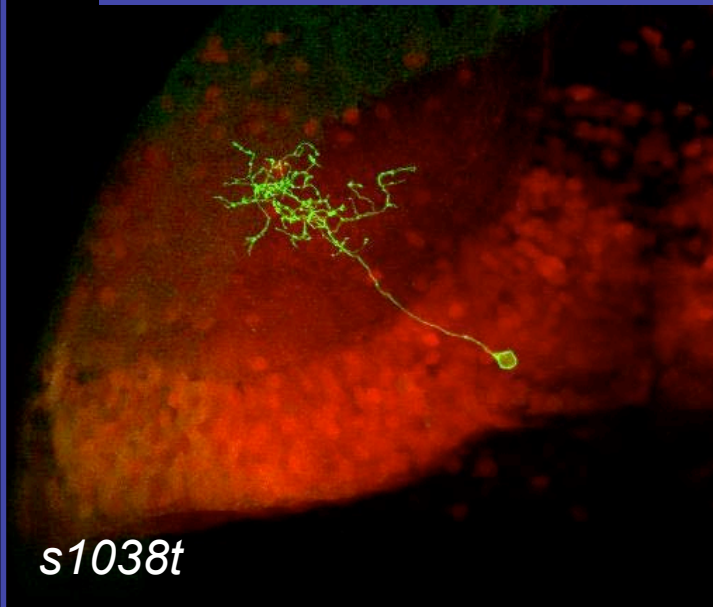
Bipolar cells

Subset of amacrine cells



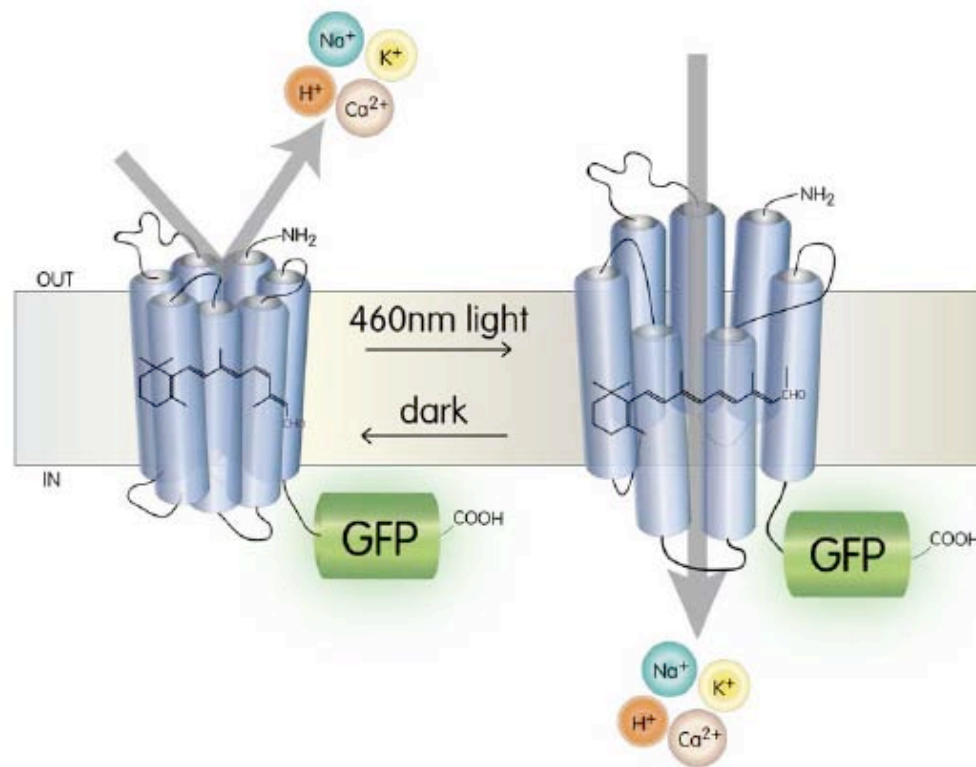
Overlap and specificity of cell type labeling

Et:GAL4; UAS:Kaede; UAS:mGFPvar
Kaede photoconverted / GFP



Channelrhodopsin-2

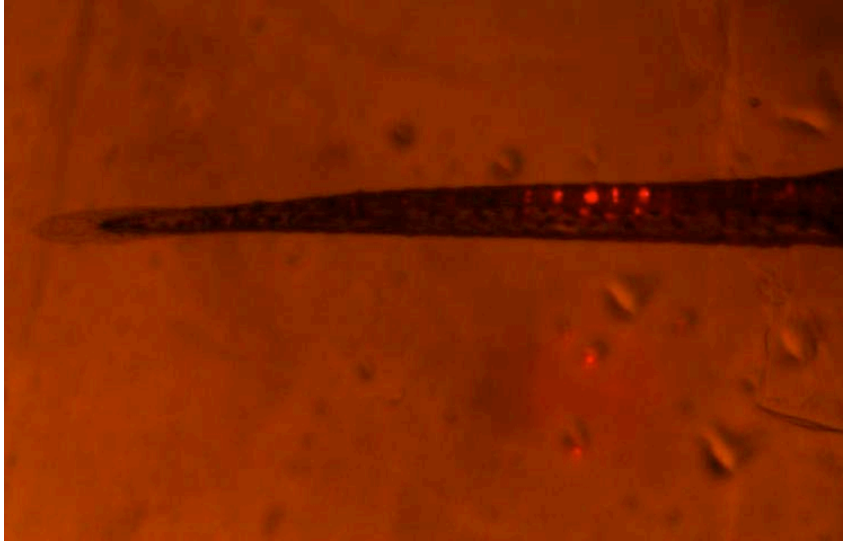
- ChR2 is a light-gated cation-selective membrane channel from *Chlamydomonas* (here tagged with mCherry).
- Misexpression in *C. elegans* and *Drosophila* neurons has allowed optical control of simple behavior.



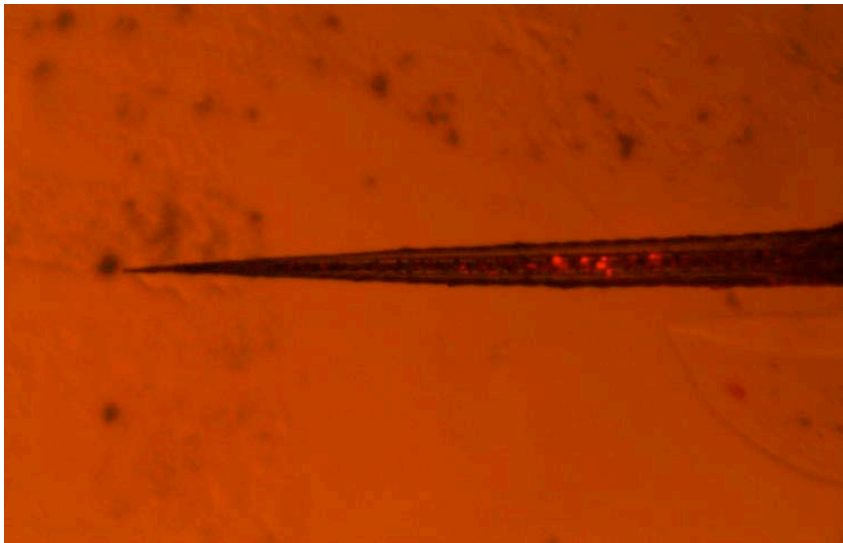
Flannery et al. 2006

Channelrhodopsin in motoneurons: Tail movement elicited by local (200 μm), 1 sec, 460nm pulses

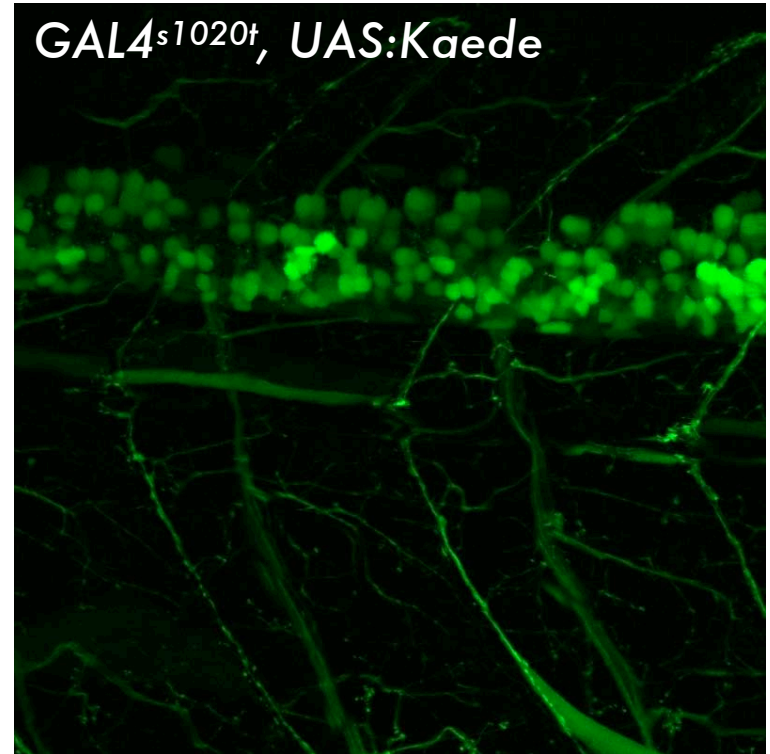
GAL4^{s1020t}, UAS:ChR2



Control: *GAL4^{s1020t}*



GAL4^{s1020t}, UAS:Kaede



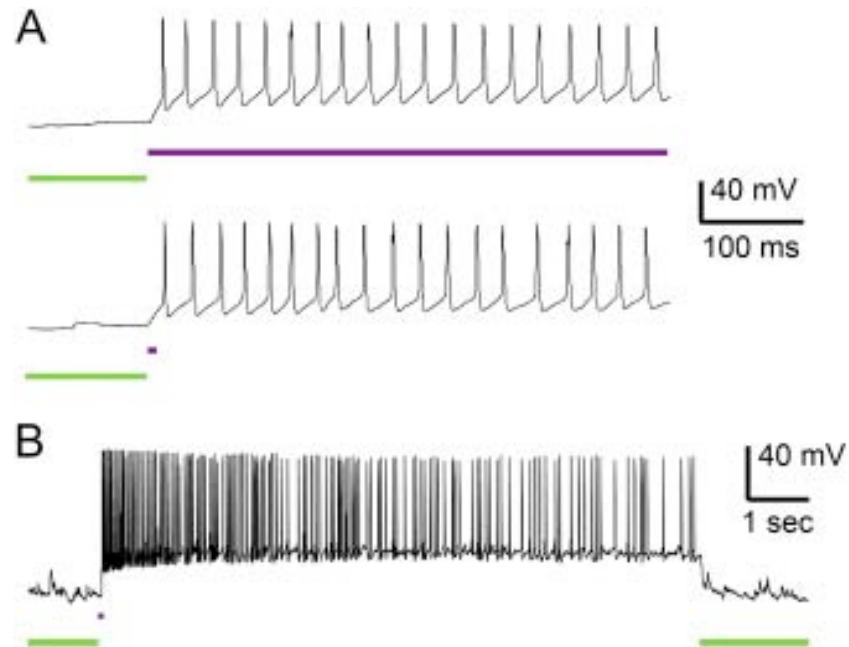
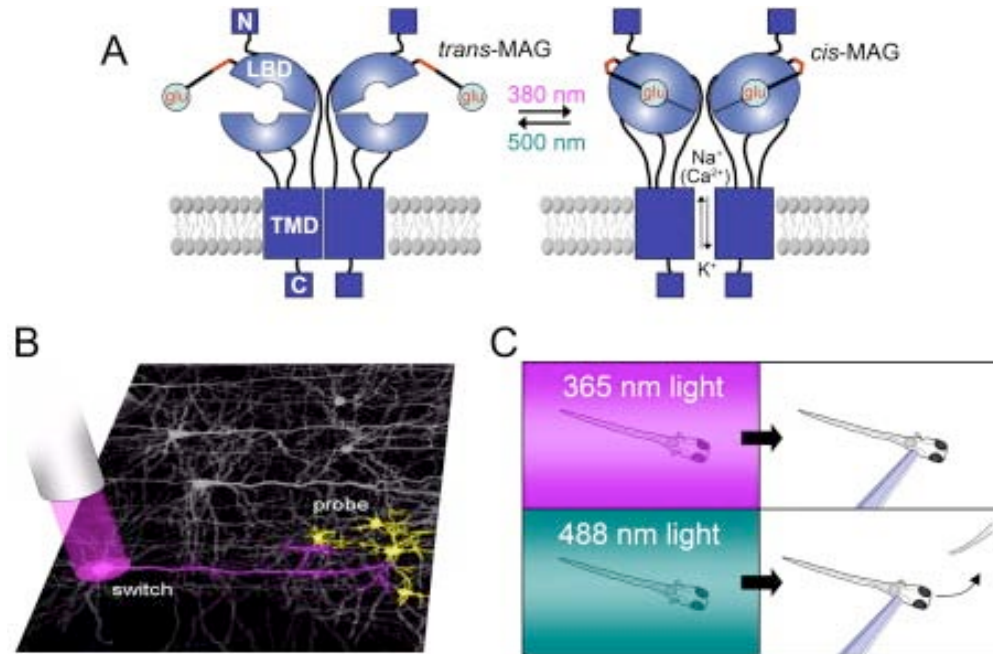
- Expression in primary and secondary motoneurons.
- 20 ms pulses sufficient
- Response in <100ms
- Accompanied by robust calcium responses

Light-gated glutamate receptor (LiGluR)

ON switch: 365 nm
OFF switch: 488 nm

Advantages for behavioral experiments:

- MAG provides additional conditionality
- **Precise timing**
- **Persistent activity in the dark**
(low rate of spontaneous isomerizations)



LiGluR in "sensory-specific" GAL4 line abolishes touch responses

after 365 nm
illumination



after 488 nm
illumination



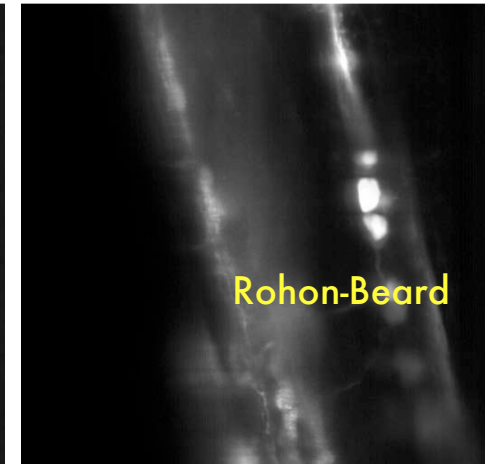
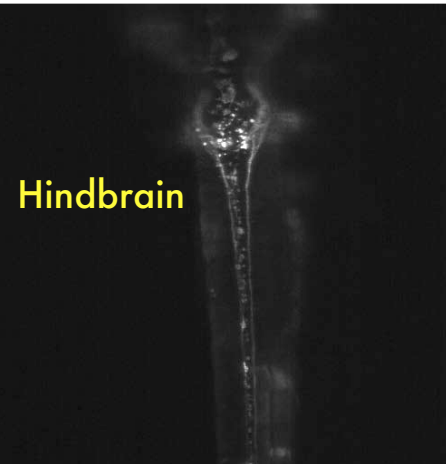
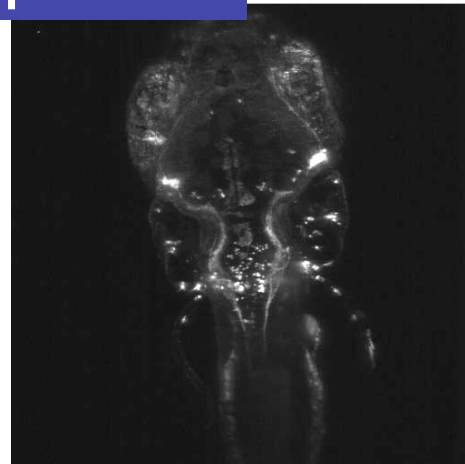
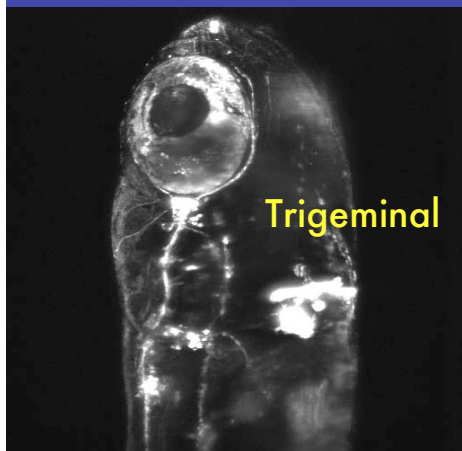
UAS:iGluR(L439C); MAG (1 mM) bath applied.

"If I were a fish, and somebody poked me in the side," (in this case, with a fine glass tip), Dr. Isacoff said, "I would escape." But when the translucent fish were strobed with violet light, the overstimulated creatures no longer detected being prodded. Blue-green light reversed the effect.

The New York Times

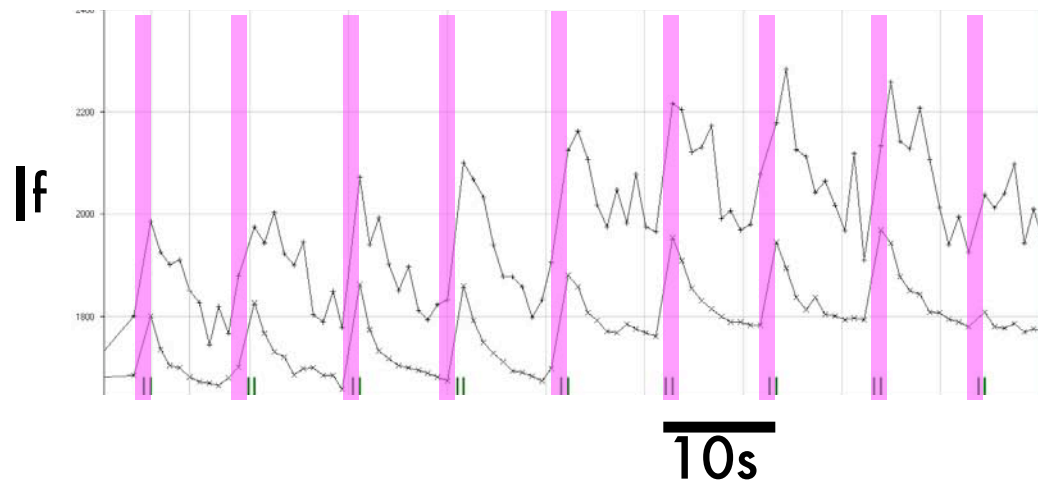
August 14, 2007

The *GAL4^{s102.1t}* pattern:

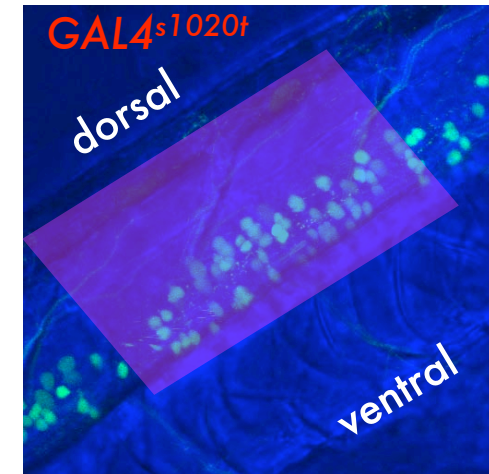


Szobota et al., *Neuron* (2007)

Calcium imaging confirms effects of LiGluR activation



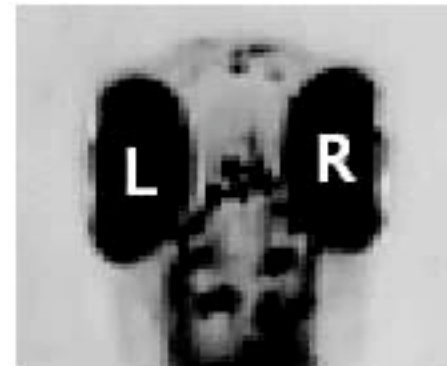
Spinal motoneurons



Halorhodopsin activation with optical fiber blocks behavioral responses



Sibling



NpHR expressor

Non-expressor
(control)



Pan-neuronal
NpHR expressor



Acknowledgements



Wendy Staub
Ethan Scott
Filippo Del Bene
Limor Ziv
Lindsay Mason
Mike Orger
Akira Muto
Jeremy Kay
Linda Nevin

Tong Xiao

Nathan Gosse

Linda Nevin

Peter Schoonheim
Ann Wehman
Karin Finger-Baier
Junko Kitamoto
Ari Arrenberg
Matt Smear
Ethan Gahtan

Funded by NIH, March of Dimes,
and the 2006 Byers Award