

# Mathematical Models for Angiogenic and Metabolic Activity in Solid Tumors

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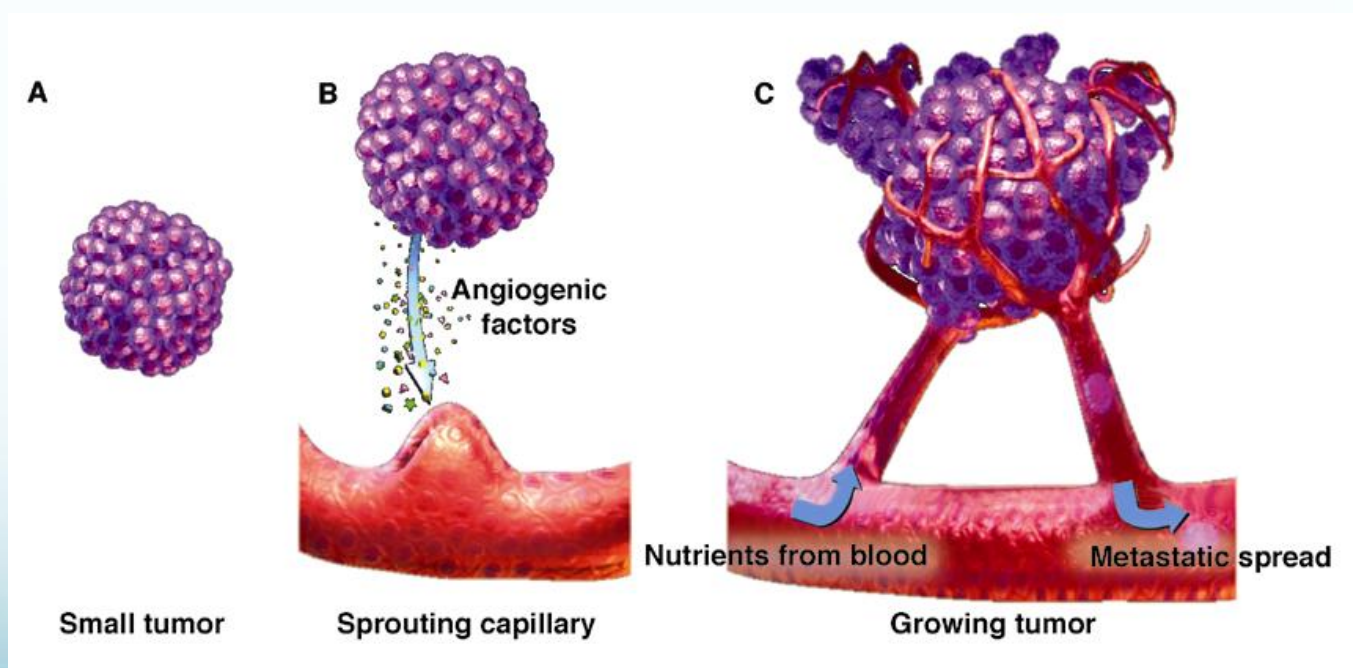
KITP

# Outline

- Background
- Mathematical model
  - Angiogenic growth factor transport
  - Interstitial fluid pressure
- Results
- Other projects
  - Microvessel model for metabolism
  - Predicting cell viability from protein expression

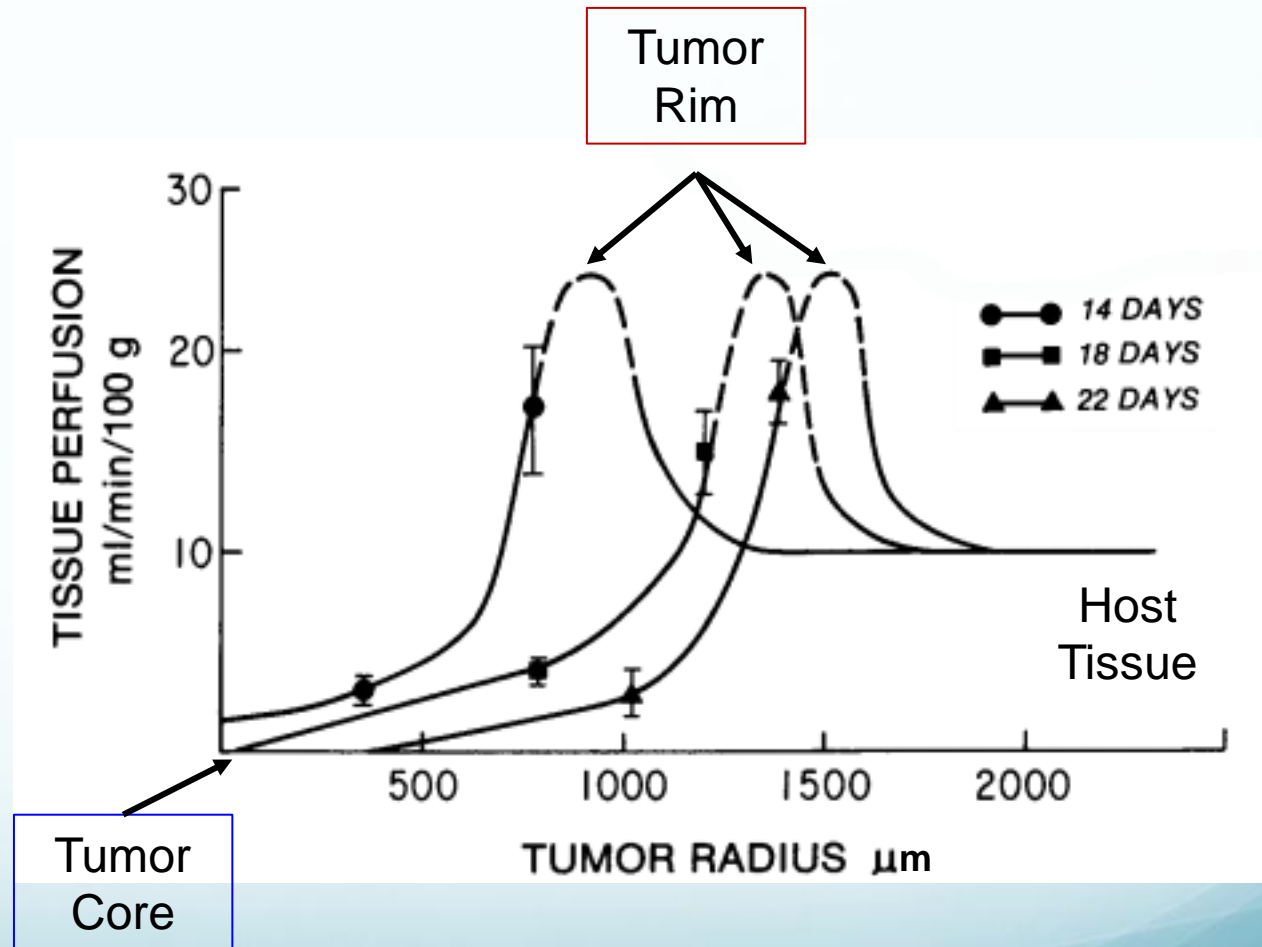
# Tumor angiogenesis

- Tumors initiate angiogenesis: formation of new blood vessels from pre-existing vessels
  - Hypoxia causes cells to produce angiogenic growth factors (AGFs)
    - e.g. proangiogenic: VEGF, antiangiogenic: angiostatin
  - Disrupts balance between proangiogenic and antiangiogenic factors
  - Blood vessels send sprouts toward tumor




# Tissue perfusion

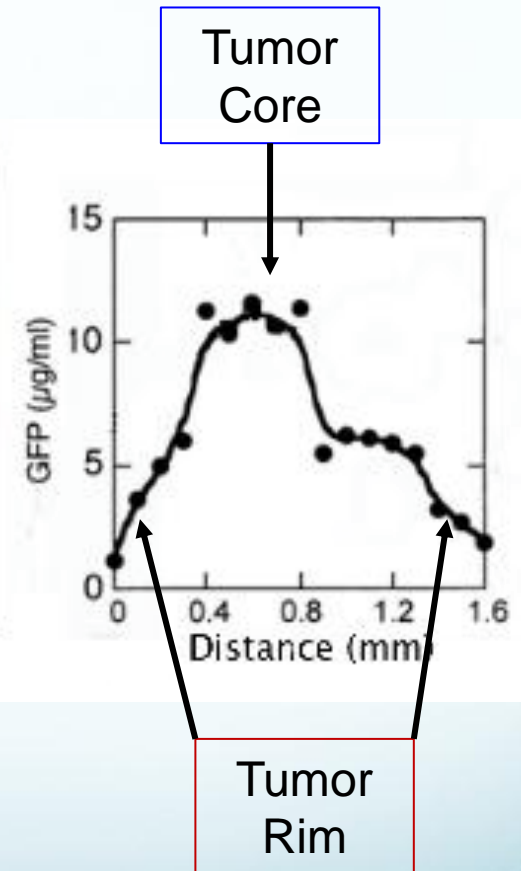
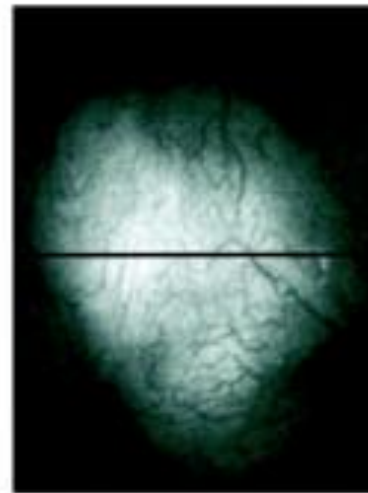
- Blood flow:
  - Highest at tumor rim
  - Lowest at tumor core
  - Reaches steady state in host tissue



Endrich et al. Tissue inhomogeneity during early tumor growth in rats. *J. Natl. Cancer Inst.*, 62 (1979), 387-395.

# Angiogenic growth factors

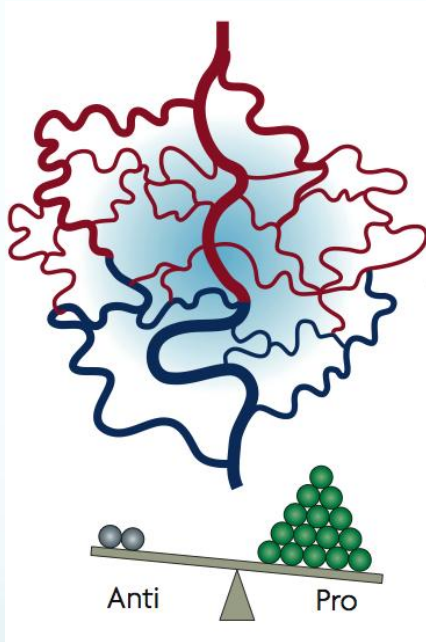
- VEGF concentration:
  - Highest at tumor core
  - Decreasing at rim
  - Reaches steady state outside tumor
- [GFP]  $\propto$  [VEGF promoter]
- Observation: [vasculature]  $\propto$  [VEGF] 
- Goal: Propose 'next' simplest relation



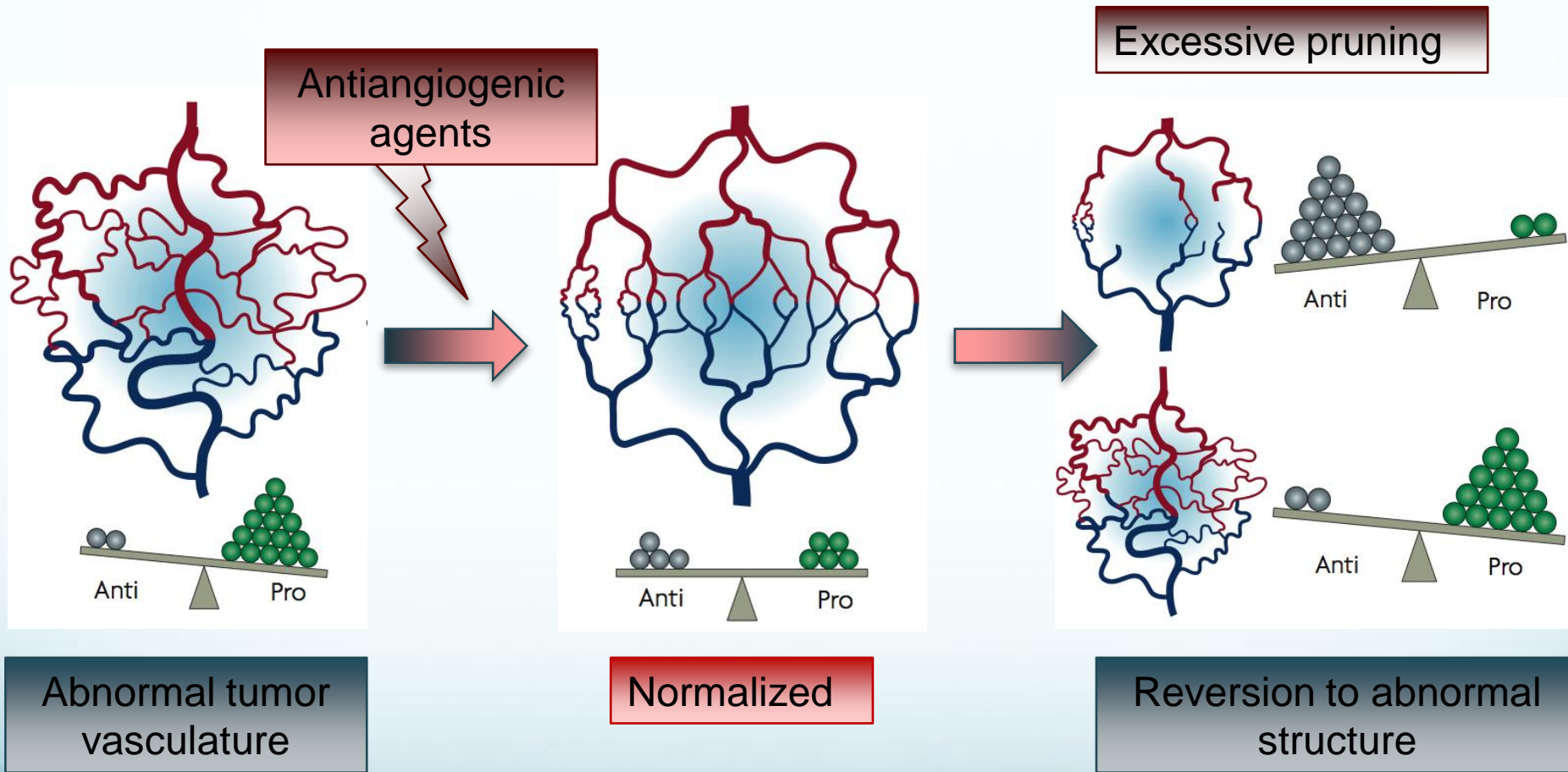
Fukumura et al. Hypoxia and acidosis independently upregulate VEGF... Cancer research (2001)

# Tumor vessels

- Tumor angiogenesis:
  - poorly regulated and hasty
- Tumor blood vessel networks:
  - Tortuous, unorganized, inefficient and experience inconsistent perfusion
  - Hinders drug delivery
- Tumor vessels are leaky
  - Leads to elevated interstitial fluid pressure and outward convective transport
  - Helps short-term delivery
- Clinical goals
  - Normalize tumor blood vessel network
  - Alleviate elevated interstitial fluid pressure

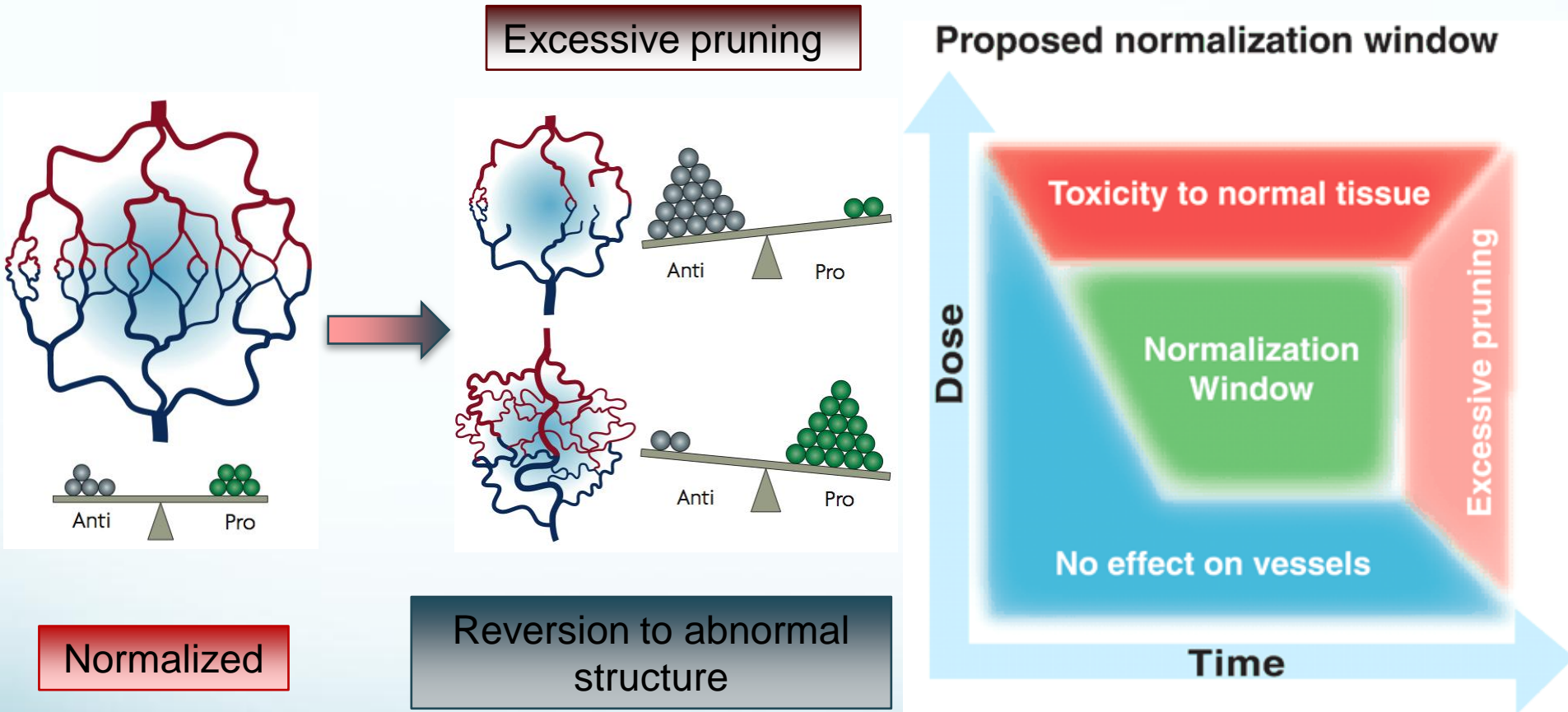


# Tumor normalization



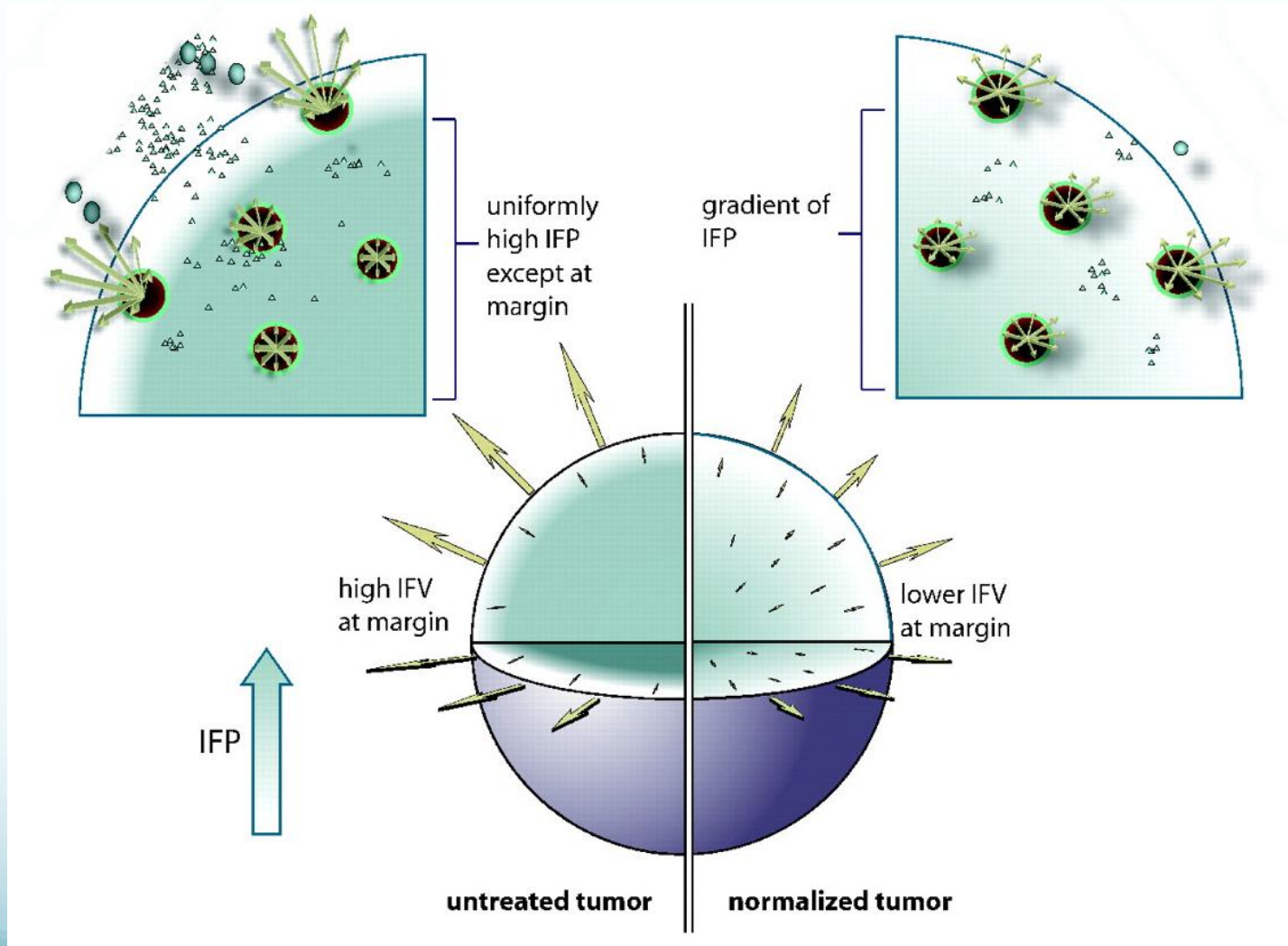


# Normalization window





# Pressure normalization



# Interstitial flow and angiogenesis

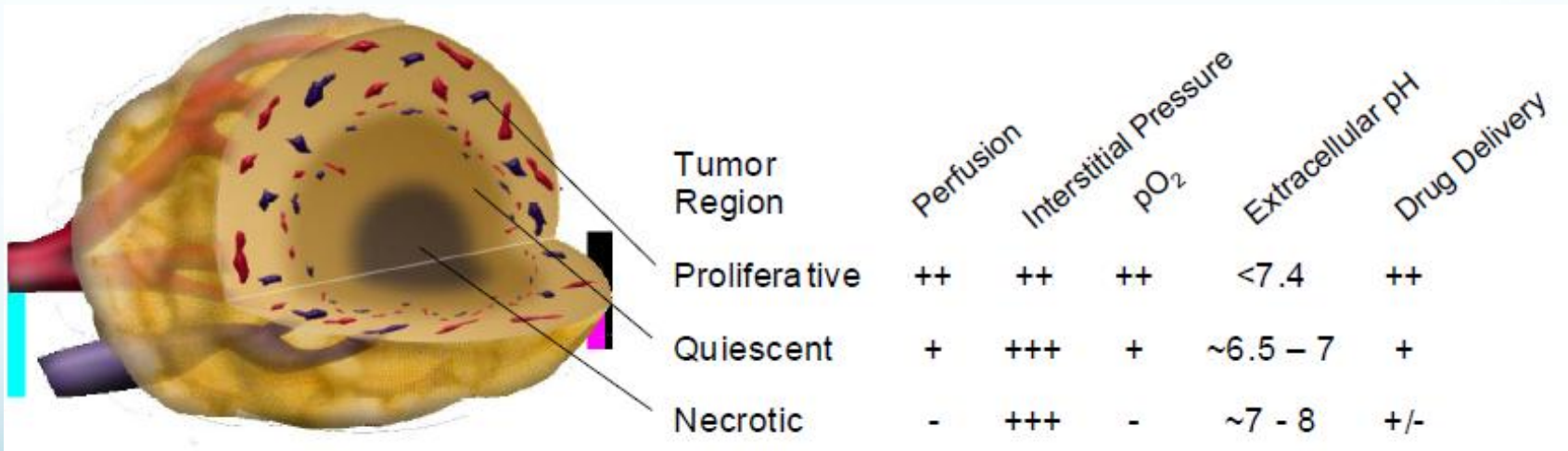
- Interstitial flow is crucial to capillary formation (Helm et al., PNAS, 2005):
  - VEGF or interstitial flow alone do not lead to capillary formation
  - BOTH must be present in order for capillaries to form
- Molecular weights of these factors suggest that convection could be important to their interstitial transport

**Table 1:** Molecular weights of common proangiogenic and antiangiogenic growth factors

Molecule	Angiogenic category	Size (kDa)
VEGF <sub>165</sub> dimer	Proangiogenic	45 (Ferrera <i>et al.</i> , 1989)
FGF family	Proangiogenic	17-34 (Ornitz <i>et al.</i> , 2001)
TSP-1	Antiangiogenic	140 (Rastinejad <i>et al.</i> , 1989)
angiostatin	Antiangiogenic	38 (O'Reilly <i>et al.</i> , 1994)
endostatin	Antiangiogenic	20 (O'Reilly <i>et al.</i> , 1997)

# Tumor microenvironment

- High interstitial pressure
- Perfusion/Diffusion-limited hypoxia
- Low pH (altered metabolism)



# Macromolecule transport

- Continuity equation for concentration of a macromolecule  $c$ , with flux  $J$  and production rate  $P$

$$\frac{\partial c}{\partial t} + \nabla \cdot J = P$$

- Flux is sum of diffusive and convective fluxes:

$$J = J_D + J_C = -D\nabla c + \vec{v}c$$

- Concentration in steady state:  $\nabla \cdot (-D\nabla c + \vec{v}c) = P$

- Constant diffusion coefficient and macromolecule velocity proportional to interstitial fluid velocity:  $\vec{v} = r\vec{u}$

$$-D\nabla^2 c + r\nabla \cdot (\vec{u}c) = P$$

- Constant production and degradation rates:

$$-D\nabla^2 c + r\nabla \cdot (\vec{u}c) = g - kc$$

# AGF equation

- Geometry: spherical tumor of fixed radius  $R$ , embedded in host tissue
- Two AGF categories: antiangiogenic, with concentration  $f_a$ , and proangiogenic, with concentration  $f_p$ , assume  $r_j \approx 1$

$$-D_j \nabla^2 f_j + \nabla \cdot (\vec{u} f_j) = g_j - k_j f_j, \quad j = p, a$$

where  $D_j$ ,  $g_j$  and  $k_j$  differ inside and outside the tumor,

e.g.

$$D_j = \begin{cases} D_j^t & 0 \leq r < R \\ D_j^h & r \geq R \end{cases}$$

- Spherical Laplacian and divergence:

$$-\frac{D_j}{r^2} \frac{d}{dr} \left( r^2 \frac{df_j}{dr} \right) + \frac{1}{r^2} \frac{d}{dr} \left( r^2 u f_j \right) = g_j - k_j f_j, \quad j = p, a.$$

# Boundary conditions

- Spherical symmetry at core:

$$-D_j^t \frac{df_j}{dr} + uf_j = 0, \quad r = 0.$$

- Continuity of concentration and flux at boundary:

$$f_j(R^-) = f_j(R^+) \quad \left( -D_j^t \frac{df_j}{dr} + uf_j \right) \Big|_{r=R^-} = - \left( D_j^h \frac{df_j}{dr} + uf_j \right) \Big|_{r=R^-}$$

- Far-field no flux:

$$-D_j^h \frac{df_j}{dr} + uf_j = 0, \quad r \rightarrow \infty.$$

- Concentrations approach steady state in host tissue:

$$f_j^s = \frac{g_j^h}{k_j^h}$$

# Interstitial fluid pressure and velocity

- Model for interstitial fluid pressure (IFP)
  - Continuity equation for steady state incompressible flow

$$\nabla \cdot u = f(r)$$

- Starling's Law

- Darcy's Law

$$u = -K \nabla p$$

- $u$  - interstitial fluid velocity (IFV)
- $p$  - interstitial fluid pressure
- $K$  - hydraulic conductivity of interstitium

$$f(r) = L_p \Phi (p_v - p - \sigma (\rho_v - \rho_i))$$

- $L_p$  - hydraulic conductivity of vessel wall
- $\Phi$  - surface area of vessel wall per unit volume of tumor
- $p_v$  - vascular pressure
- $\pi_{v(i)}$  - plasma osmotic (interstitial) pressure
- $\sigma$  - average osmotic reflection coefficient for plasma proteins

- Pressure equation

$$\nabla^2 p = -\frac{L_p \Phi}{K} (p_e - p)$$

- Effective pressure  $p_e = p_v - \sigma (\rho_v - \rho_i)$

- In tumors:  $p_e \gg p_v$

- LT Baxter, RK Jain, Microvascular Research 37 (1989)



# Interstitial fluid pressure and velocity

- Spherical Laplacian:

$$\frac{1}{r^2} \frac{d}{dr} \left( r^2 \frac{dp}{dr} \right) = -\frac{a^2}{R^2} (p_e - p), \quad a = R \sqrt{\frac{L_p F}{K}}$$

where  $a$  and  $p_e$  can differ between tumor and host tissue

- Spherical symmetry at core:  $\left. \frac{dp}{dr} \right|_{r=0} = 0$

- Continuity of concentration and velocity at boundary:

$$p(R^-) = p(R^+), \quad u(R^-) = u(R^+) \quad \text{i.e.} \quad -K^t \left. \frac{dp}{dr} \right|_{r=R^-} = -K^h \left. \frac{dp}{dr} \right|_{r=R^+}$$

- Far-field zero flux:

$$\left. \frac{dp}{dr} \right|_{r \rightarrow \infty} = 0,$$

- This leads to pressure approaching a steady state, assumed to be

$$p_e^h = 0$$

# Non-dimensionalized system

- Radial distance  $\tilde{r} = \frac{r}{R}$

- AGFs:

$$-\frac{1}{\tilde{r}^2} \frac{d}{d\tilde{r}} \left( \tilde{r}^2 \frac{d\tilde{f}_j}{d\tilde{r}} \right) + \frac{\tilde{K}_j}{\tilde{r}^2} \frac{d}{d\tilde{r}} \left( \tilde{r}^2 \tilde{u} \tilde{f}_j \right) = \tilde{g}_j - \tilde{k}_j \tilde{f}_j, \quad \tilde{f}_j = \frac{f_j}{f_j^s}$$

- Pressure  $\frac{1}{\tilde{r}^2} \frac{d}{d\tilde{r}} \left( \tilde{r}^2 \frac{d\tilde{p}}{d\tilde{r}} \right) = -a^2 (1 - \tilde{p}), \quad \tilde{p} = \frac{p}{p_e^t}$

- Velocity  $\tilde{u} = -\frac{d\tilde{p}}{d\tilde{r}}, \quad \tilde{u} = \frac{u}{K^t p_e^t / R}$

# Non-dimensional parameters

AGFs

$$\tilde{k}_j^t = R^2 \frac{k_j^t}{D_j^t}$$

$$\tilde{g}_j^t = R^2 \frac{g_j^t}{D_j^t f_j^s}$$

$$\tilde{K}_j^t = \frac{K^t p_e^t}{D_j^t}$$

$\tilde{r} = 0$

$\tilde{r} = 1$

Pressure

$$a^t = R \sqrt{\frac{L_p^t F^t}{K^t}}$$

$$\tilde{k}_j^h = R^2 \frac{k_j^h}{D_j^h}$$

$$\tilde{g}_j^h = R^2 \frac{k_j^h}{D_j^h} = \tilde{k}_j^h$$

$$\tilde{K}_j^h = \frac{K^h p_e^t}{D_j^h}$$

Production

Degradation

Molecular  
Transport

$$a^h = R \sqrt{\frac{L_p^h F^h}{K^h}}$$

Fluid  
Transport

# Angiogenic Activity

- Non-dimensional factor concentrations approach 1 in host tissue
- Measure of angiogenic activity from Stoll et al. (2003)

$$a(r) = \begin{cases} \frac{\tilde{f}_p}{\tilde{f}_a} - 1, & \tilde{f}_p \geq \tilde{f}_a \\ 1 - \frac{\tilde{f}_a}{\tilde{f}_p}, & \tilde{f}_p < \tilde{f}_a \end{cases}$$

- If  $a > 0$  angiogenesis is initiated
- If  $a = 0$  the vasculature is stable
- If  $a < 0$  vessels are regressing

- Angiogenic activity classes:
  - Global angiogenesis:  $a > 0$  in the tumor
  - Focal suppression:  $a < 0$  at tumor core,  $a > 0$  at tumor rim
  - Global suppression:  $a < 0$  in the tumor
- Purpose: Analyze sensitivity to physiological parameters

# Solution method

- Analytical solutions for pressure and velocity can be found:

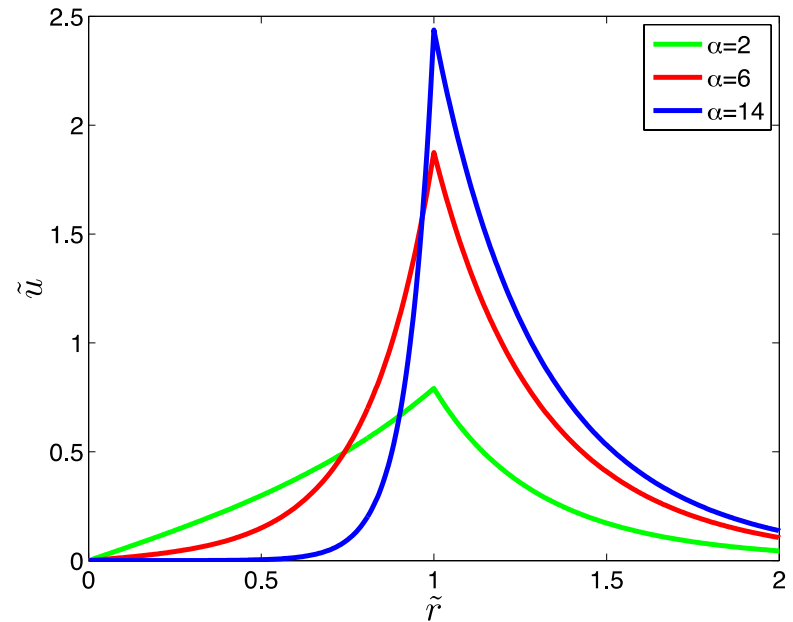
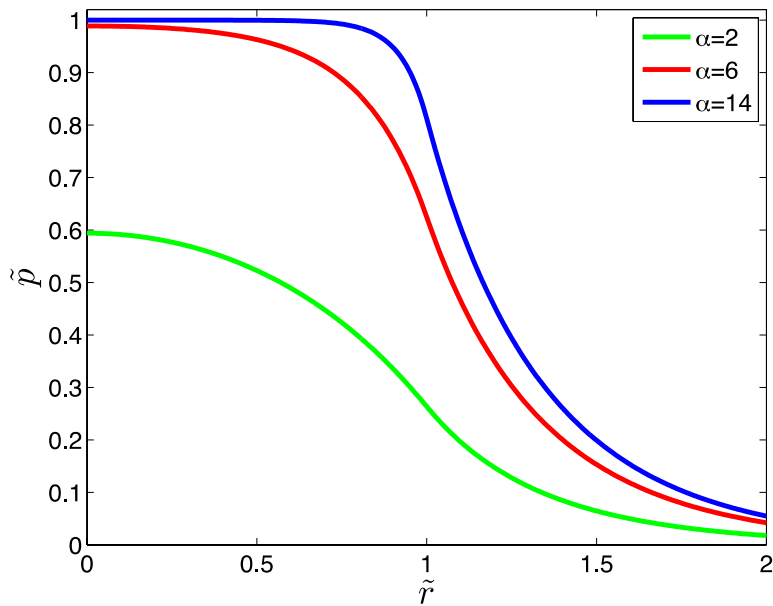
$$\tilde{u}(\tilde{r}) = \begin{cases} \frac{1 + a^h}{f + q} \frac{a^t \tilde{r} \cosh(a^t \tilde{r}) - \sinh(a^t \tilde{r})}{\tilde{r}^2} & 0 \leq \tilde{r} < 1 \\ \frac{q}{f + q} \frac{(a^h \tilde{r} + 1) \exp(a^h (\tilde{r} - 1))}{\tilde{r}^2} & \tilde{r} \geq 1 \end{cases}$$

- If  $q = \frac{K^t}{K^h} (a^t \cosh(a^t) - \sinh(a^t))$ ,  $f = (1 + a^h) \sinh(a^t)$  then we can find analytical solutions for AGFs [3], otherwise we rely on numerical integration schemes
- Determine measure of angiogenic activity

# Interstitial fluid pressure and velocity

$$\nabla^2 \tilde{p} = -a^2(1 - \tilde{p})$$

$$\tilde{u} = -\nabla \tilde{p}$$



$$a^t = R \sqrt{\frac{L_p^t F^t}{K^t}} - \text{D}$$

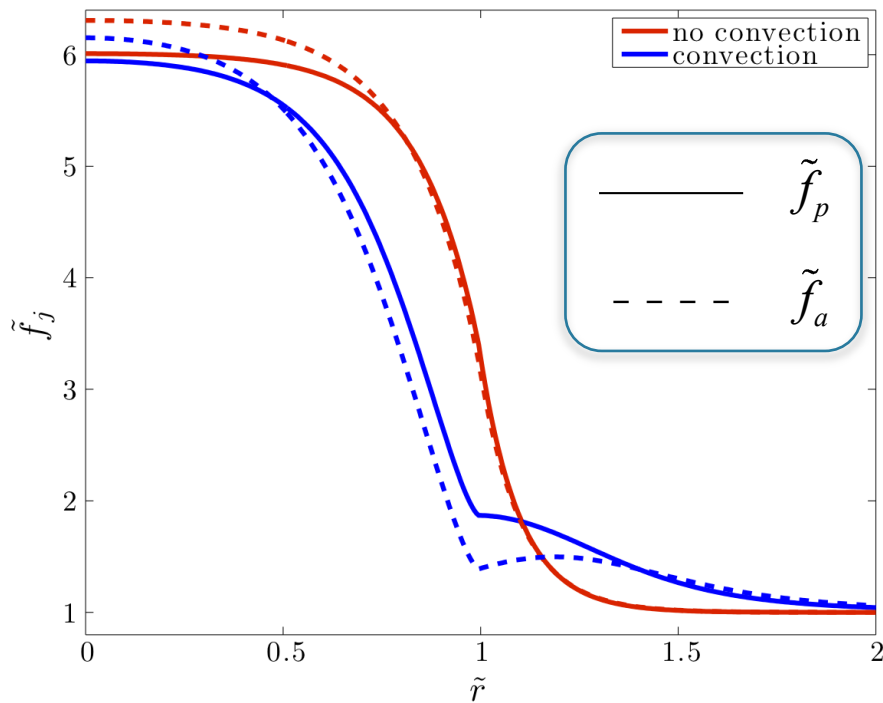
pressure rises in the tumor  
drastic pressure drop at rim

D

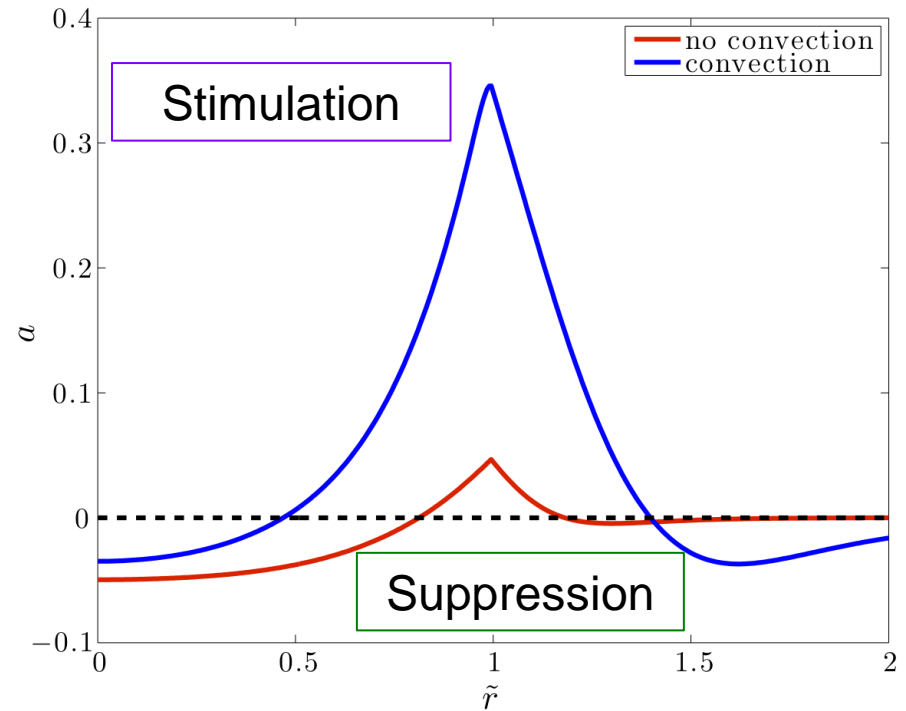
low velocity in tumor core  
high velocity at tumor rim

# Convection

## AGF concentrations



## Angiogenic activity



add convection  $\supset$

factors pushed out of tumor  
 $\supset$  large difference at rim

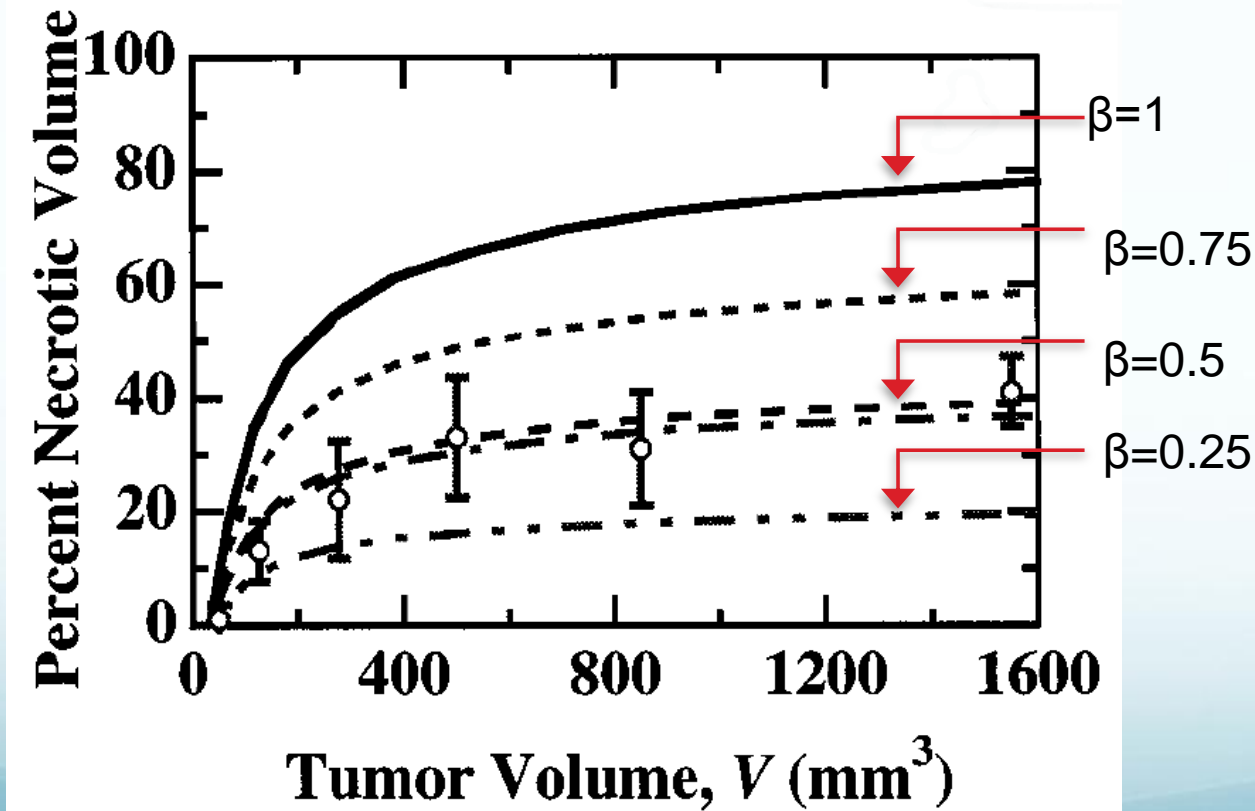
$\supset$

angiogenic suppression at core  
 $\supset$  high angiogenic activity at tumor rim



# Correlation with necrotic volume

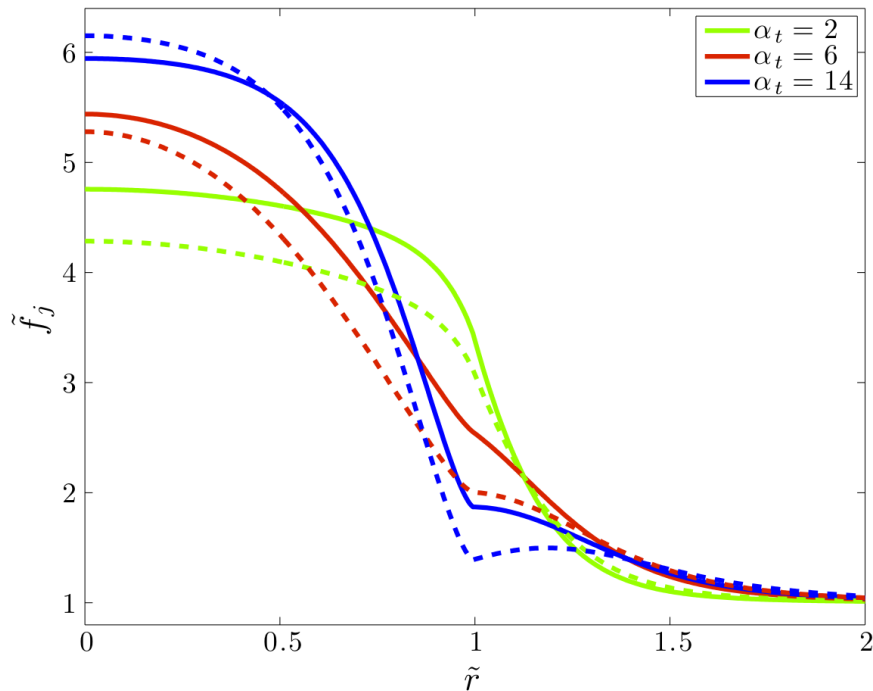
- Assume necrotic volume is proportional to suppressed volume:  $V_n = \beta V_s$
- Best fit for quantitative agreement:  $\beta = 0.47$



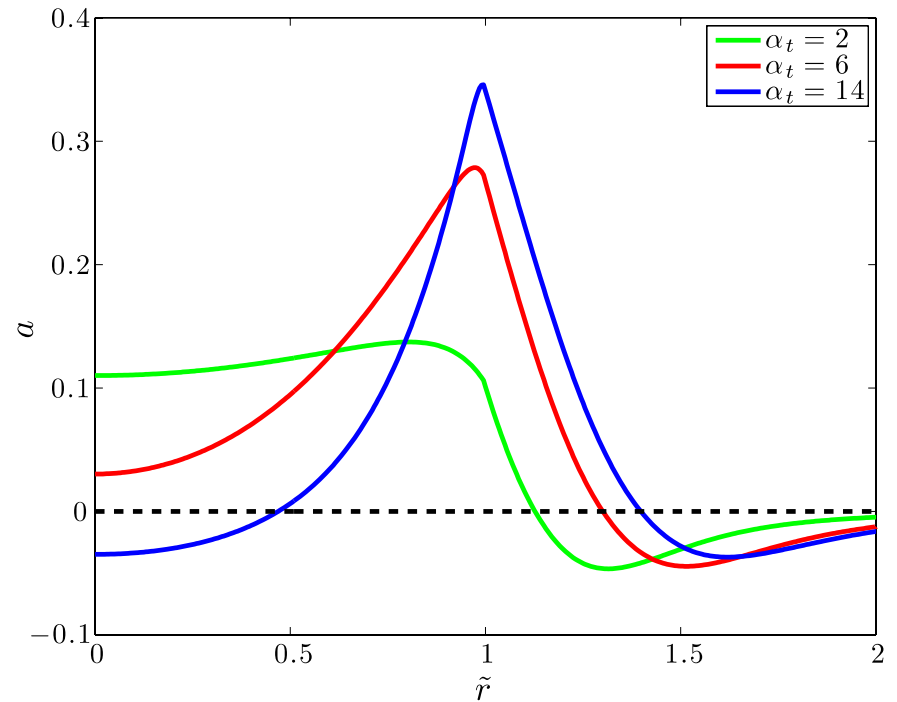
# Pressure parameter

$$a^t = R \sqrt{\frac{L_p^t F^t}{K^t}}$$

AGF concentrations



Angiogenic activity



$L_p$  or  $F - \rho a^t - \rho$

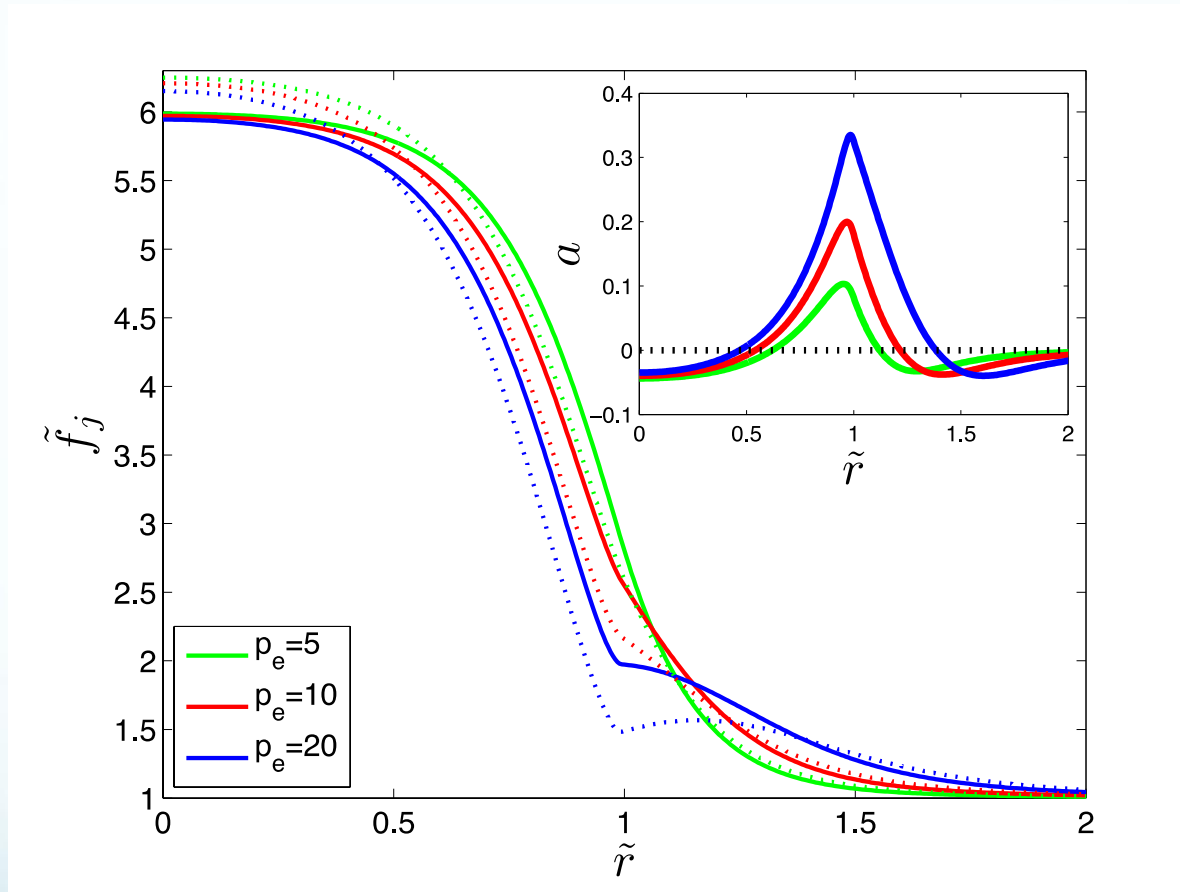
low velocity in tumor core  
high velocity at tumor rim

$\rho$

AGFs stay in tumor  
AGFs pushed into host tissue

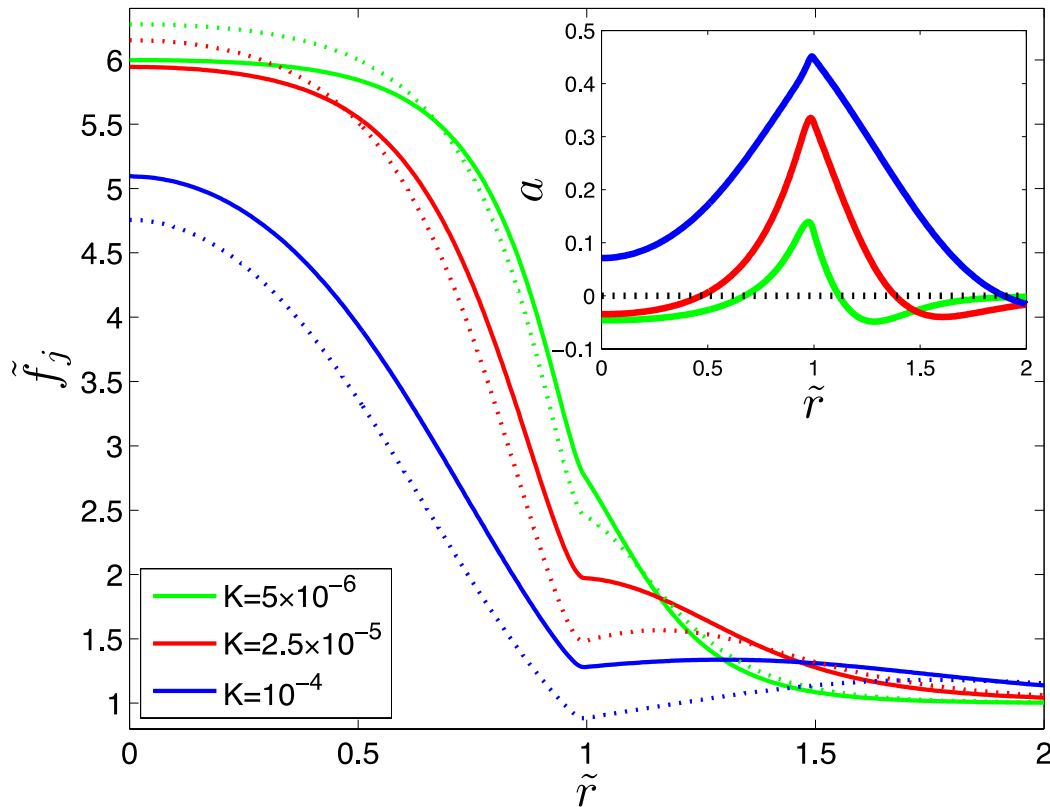
# Convection parameter

$$\tilde{K}_j = \frac{K p_e^t}{D_j}$$



$p_e^t \rightarrow \tilde{K} \rightarrow$  high velocity at tumor rim  $\rightarrow$  AGFs pushed into host tissue

# Hydraulic conductivity



$$a^t = R \sqrt{\frac{L_p^t F^t}{K^t}}$$

$$\tilde{K}_j = \frac{K p_e^t}{D_j}$$

$K - \supset$

$a^t - \supset$   
 $\tilde{K}_j -$

high velocity in tumor core  
high velocity at tumor rim

$\supset$  AGFs pushed into host tissue

# Therapeutic implications

- Antiangiogenic treatment

- Proangiogenic inhibition

- Increased proangiogenic deactivation:

$$\tilde{k}_p^t -$$

- Antiangiogenic factors

- Increased antiangiogenic production:

$$\tilde{g}_a^t -$$

- Vessel normalization

- Decreased vessel permeability:

$$L_p^t - \Downarrow a^t -$$

- Decreased resistance:

$$p_v - \Downarrow \tilde{K} -$$

- Decreased surface area:

$$F - \Downarrow a^t -$$

- Matrix-degrading enzymes

- Less dense tissue

- Increased hydraulic conductivity:

$$K - \Downarrow \tilde{K} -$$

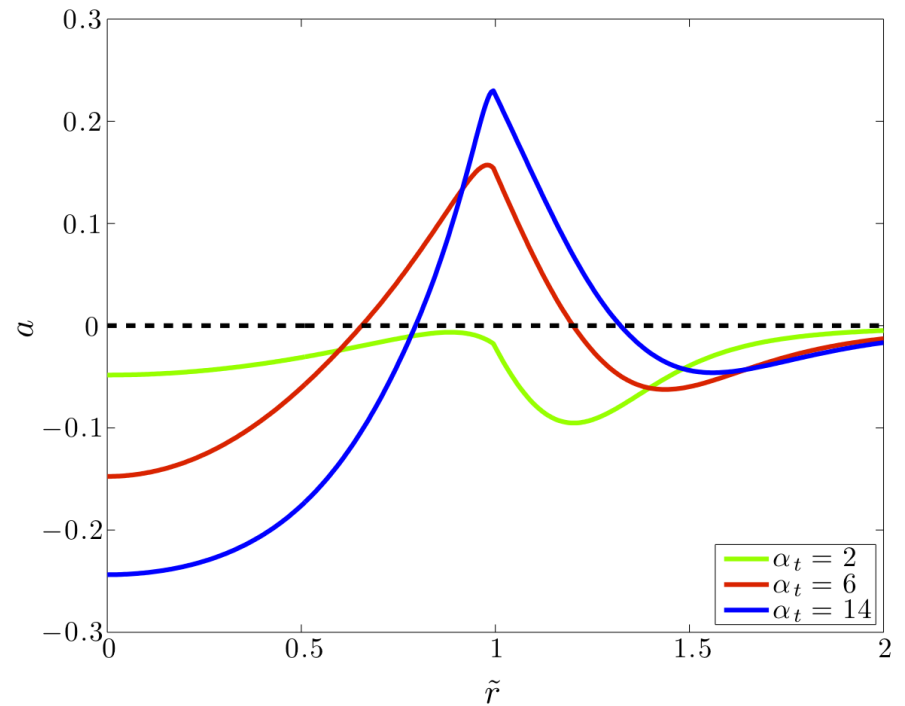
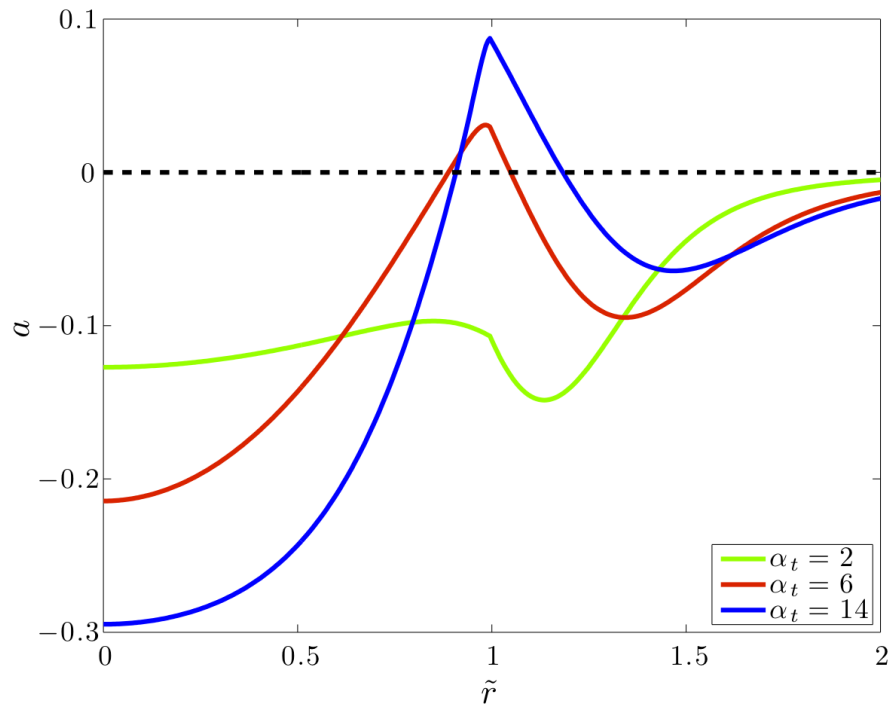
$$a -$$

- Cytotoxic agents

- Decreased proangiogenic production:

$$\tilde{g}_p^t -$$

# Other therapy effects



$$k_p^t - \mathcal{D} \tilde{k}_p^t -$$

$$g_a^t - \mathcal{D} \tilde{g}_a^t -$$

# Conclusions

- Angiogenic growth factors concentrations are influenced by convective transport in solid tumors
- Angiogenesis can be partially suppressed by modifying physiological parameters
- Treatments: decreasing vascular pressure or vessel permeability may discourage angiogenesis (along with improved chemotherapy delivery)
- Despite reducing IFP, decreasing hydraulic conductivity may be detrimental to treatment efficacy
- AGF imbalance is a simple way to incorporate angiogenesis into a tumor model, hopefully can be used as a predictive tool for treatment combinations



# Tumor metabolism

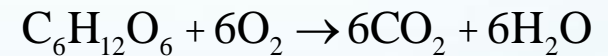
- Hypoxia: Leads to increased glycolysis
- Warburg effect: Aerobic glycolysis

## Glycolysis

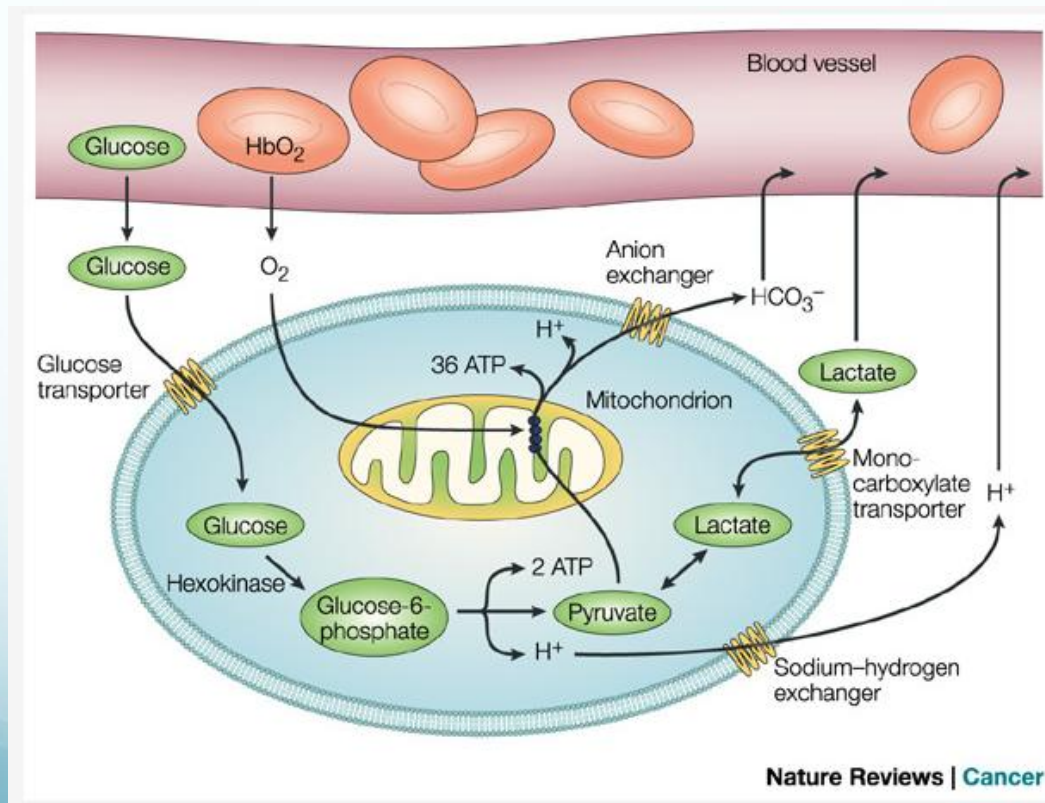


Yield: 4-2=2 ATP

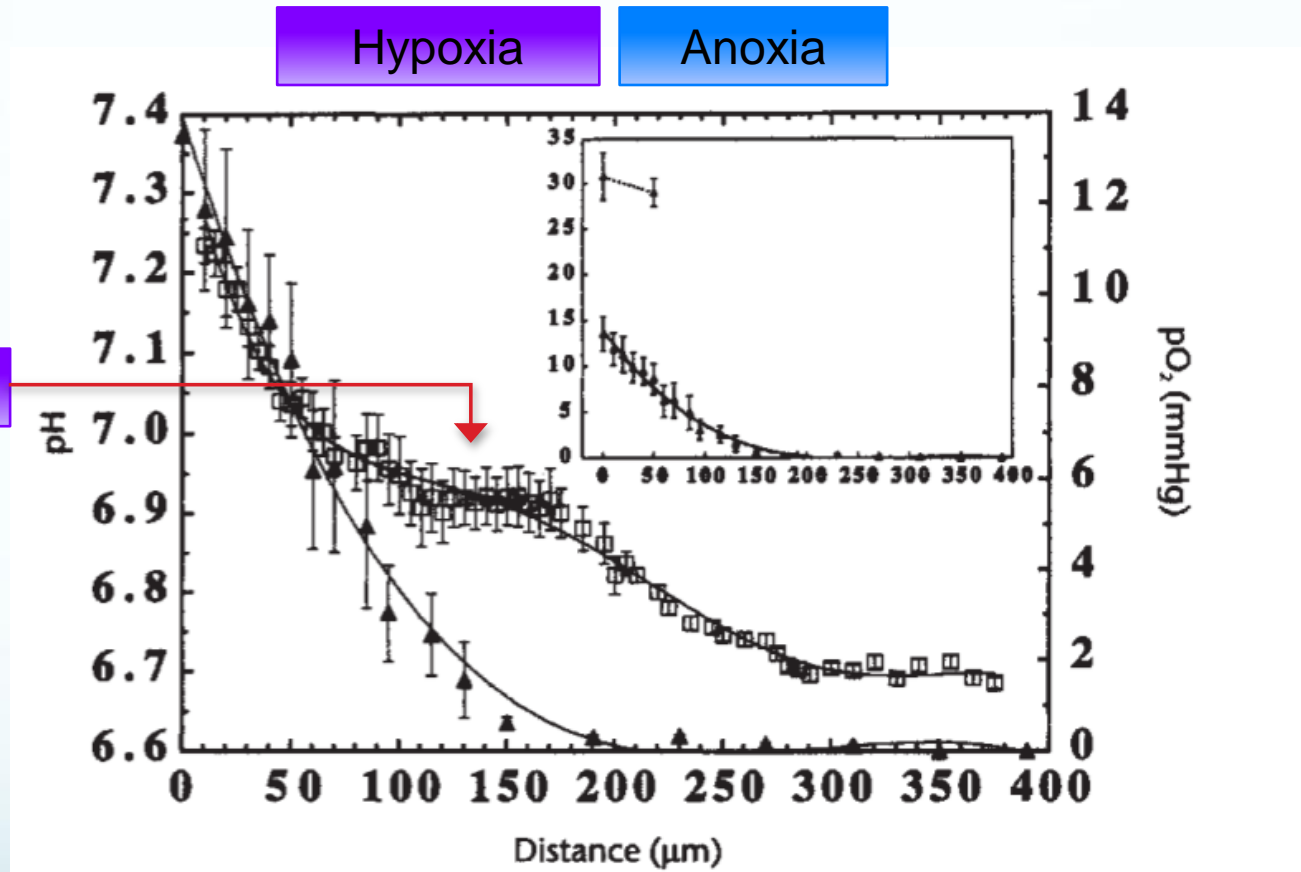
## Respiration



Yield:  $\leq 38$  ATP



# Microvessel experiment



Hypothesis:

Respiration-  
dominated

Downregulated  
Respiration/Glyc  
olysis

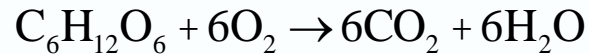
Glycolysis-  
dominated

Image from: Helmlinger et al.,  
Nature Med 3, 1997.

# Mathematical model

- Metabolism:

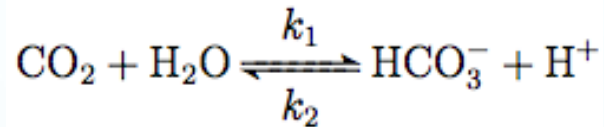
Respiration



Glycolysis



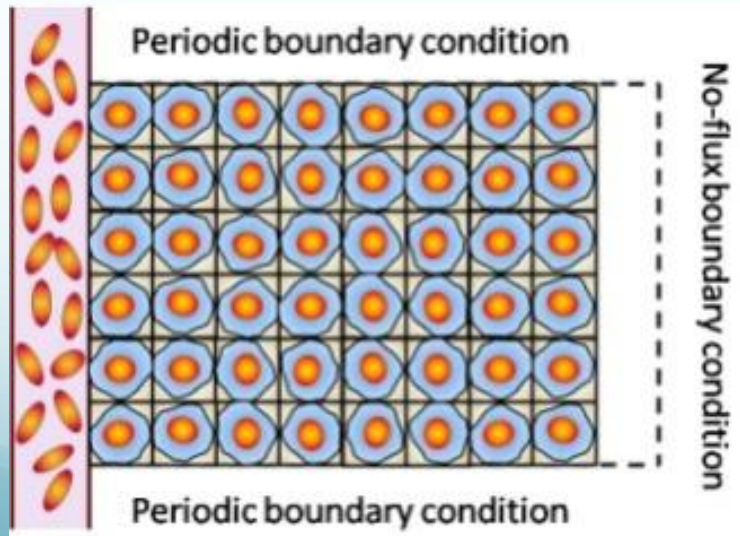
- Buffer:



- Na<sup>+</sup> and Cl<sup>-</sup> to ensure electroneutrality

- Interested in pH = -log(c<sub>H</sub>)

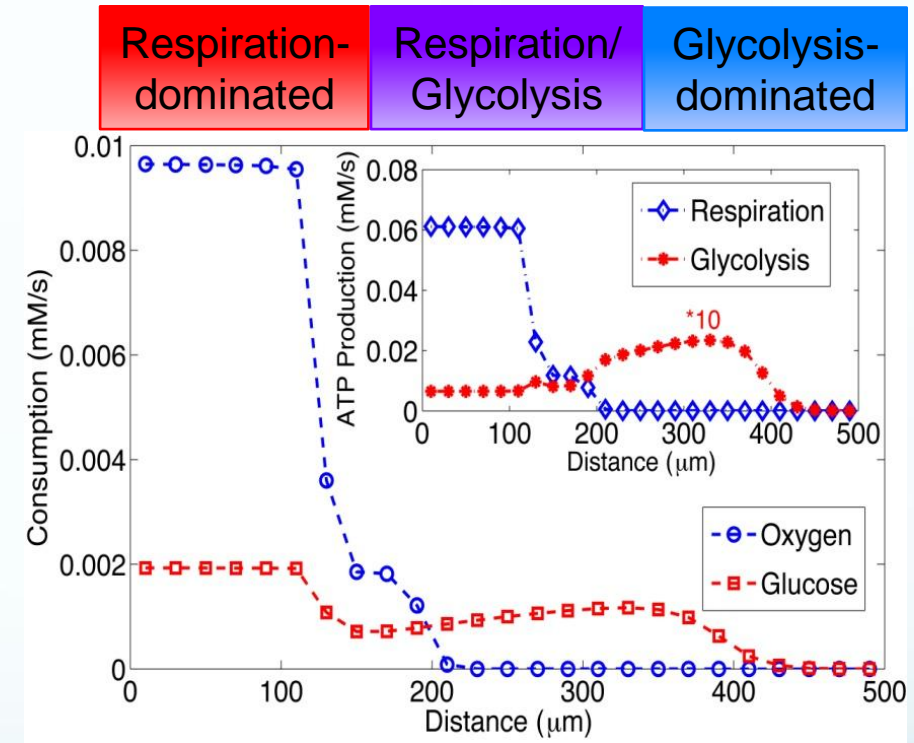
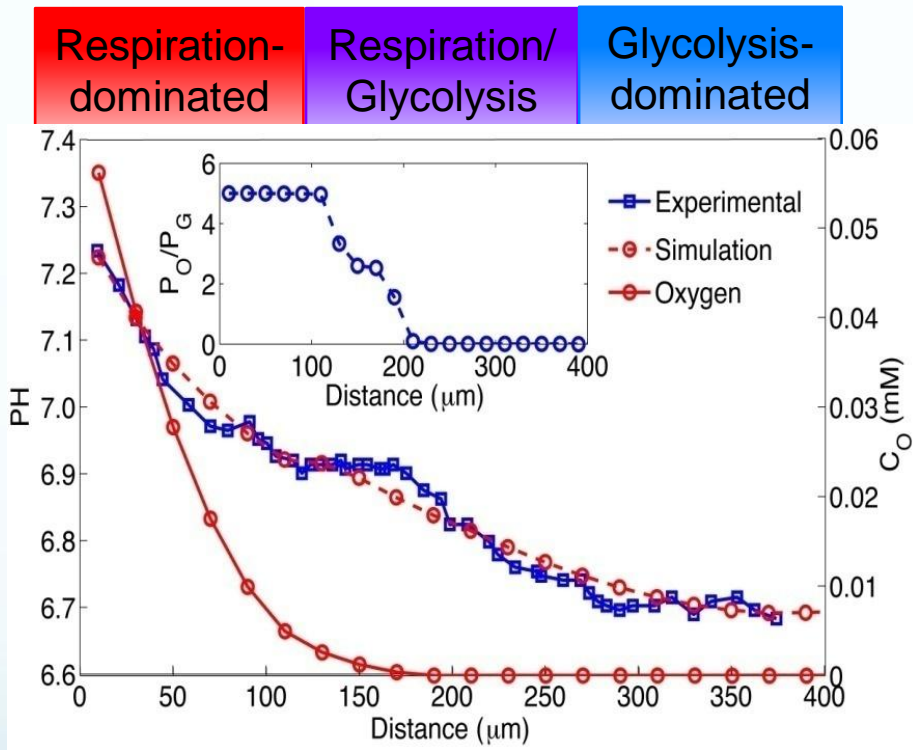
$$\frac{\partial C_i}{\partial t} = D_i \nabla^2 C_i + P_i$$



Molavian et al. Cancer Res 69 (2009).

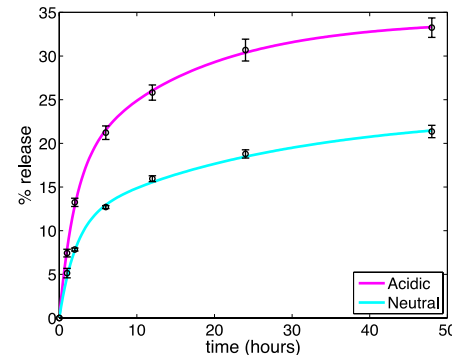
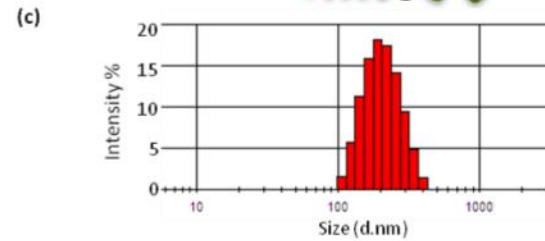
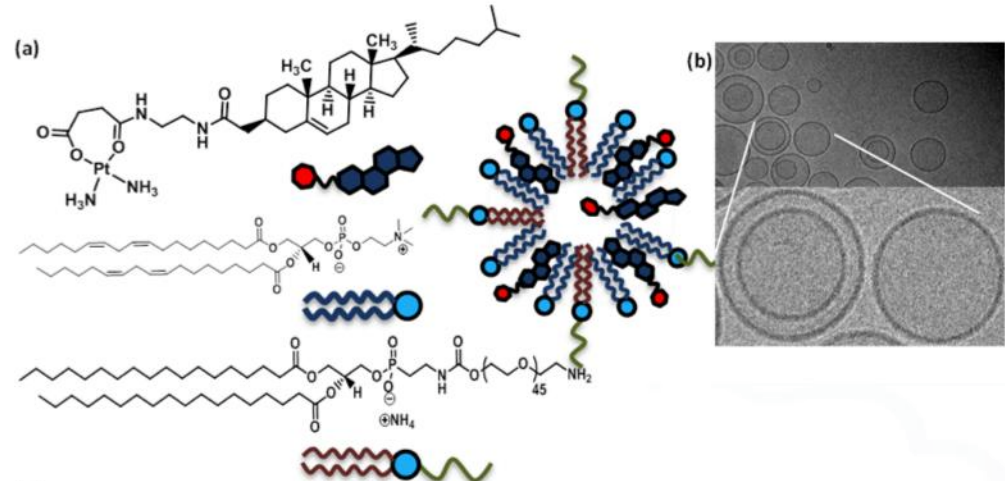
Compound	<i>i</i>	$P_i$ (10 <sup>-2</sup> mM/s)
Cl <sup>-</sup>	Cl	0
Na <sup>+</sup>	Na	-
Glucose	G	$-p_G \frac{C_G}{C_G + k_G} f_1(C_O)$
O <sub>2</sub>	O	$R P_G \frac{C_O}{C_O + k_O} f_2(C_O)$
Lactate <sup>-</sup>	L	$-2P_G + \frac{1}{3}P_O$
CO <sub>2</sub>	CO	$k_r C_H C_B - k_f C_{CO}$
Bicarbonate <sup>-</sup>	B	$k_f C_{CO} - k_r C_H C_B - P_O$
H <sup>+</sup>	H	$k_f C_{CO} - k_r C_H C_B - P_O + P_L$

# Results



# Cell viability prediction

- Experiments measure pAkt (pro-survival) and Caspase-3 (apoptosis trigger) proteins
- Cisplatin nanoparticles
  - Increase caspase-3
  - Increase pAkt
- Potential solution:
  - PI3K inhibitor: PI828
  - Prevents Akt phosphorylation
- Mathematical goal:
  - Optimize treatment combination



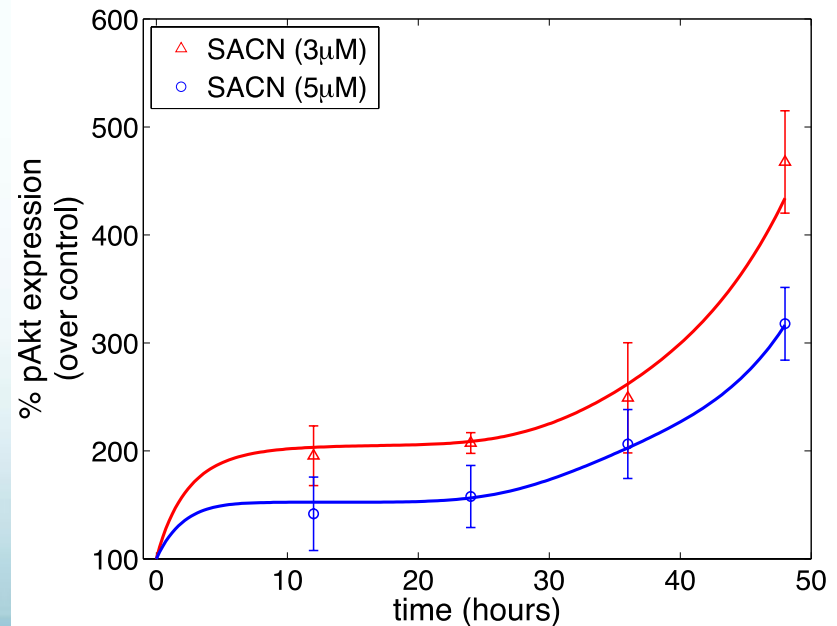
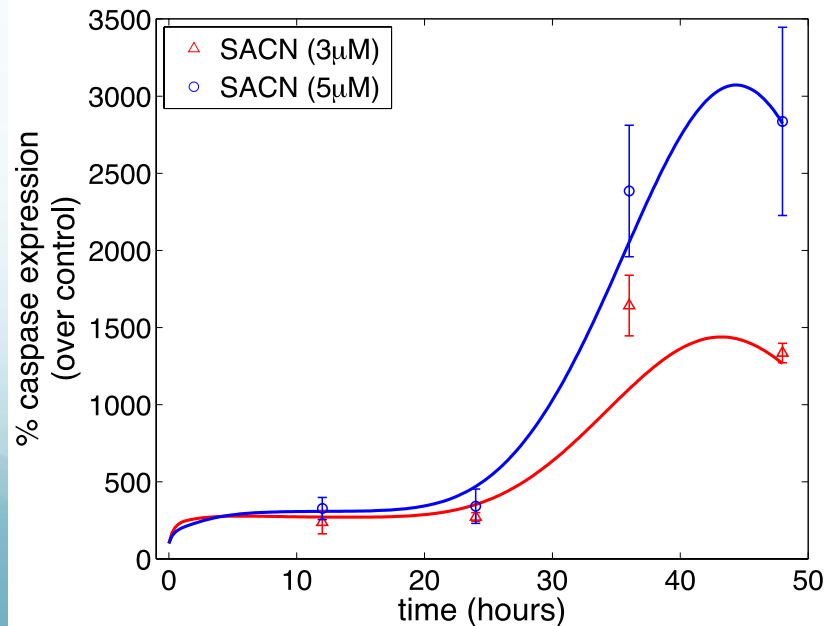
# Protein expression

- Protein equations:
  - Mutually inhibitory
  - Treatment effects

$$\frac{dP}{dt} = \frac{k_p + I_p f_p(t)}{1 + a_p C + g_p g_i(t)} - d_p P$$

$$\frac{dC}{dt} = \frac{k_c + I_c f_c(t)}{1 + a_c P} - d_c C$$

- Chemotherapy increases Caspase-3 and pAkt

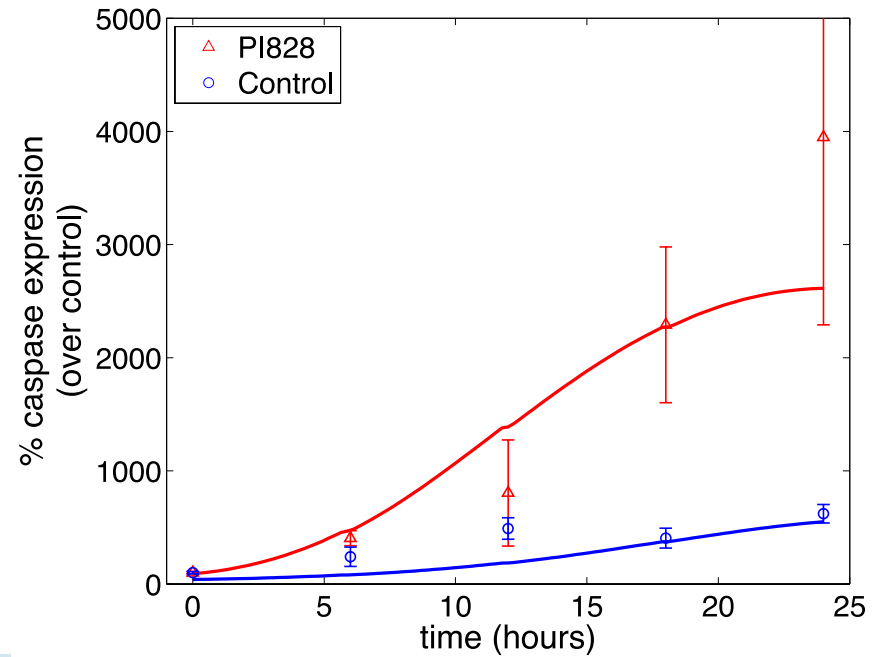
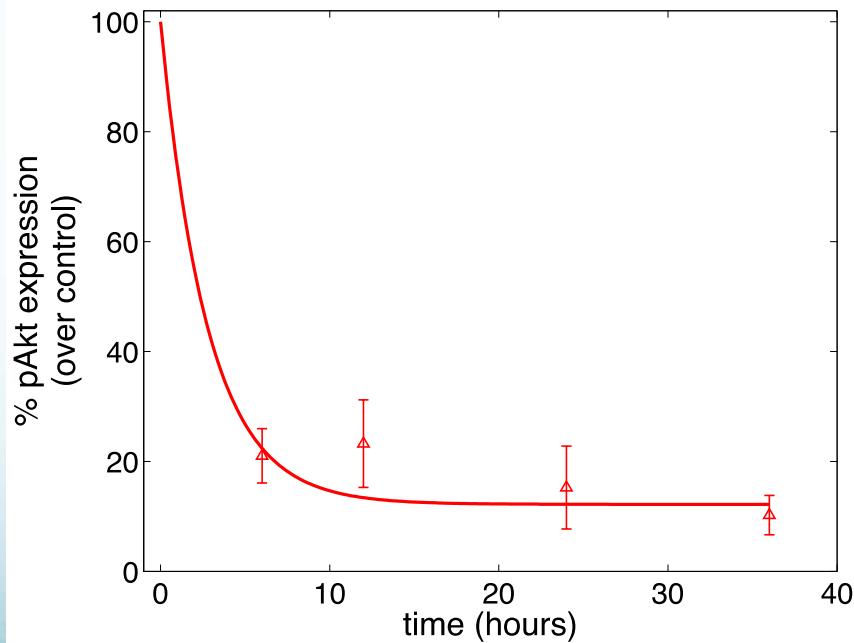




# Protein expression

- PI828 blocks Akt phosphorylation

- CisNP + PI828 has synergistic effect

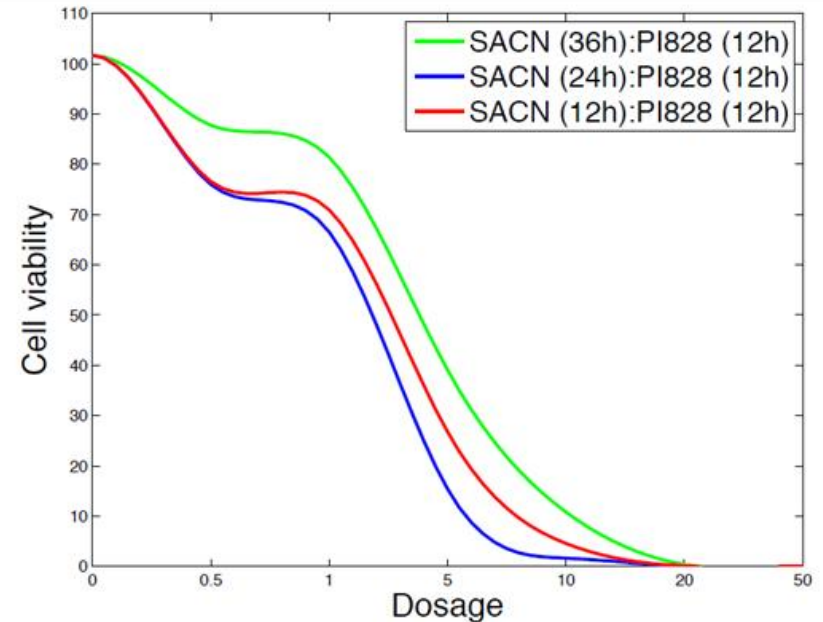
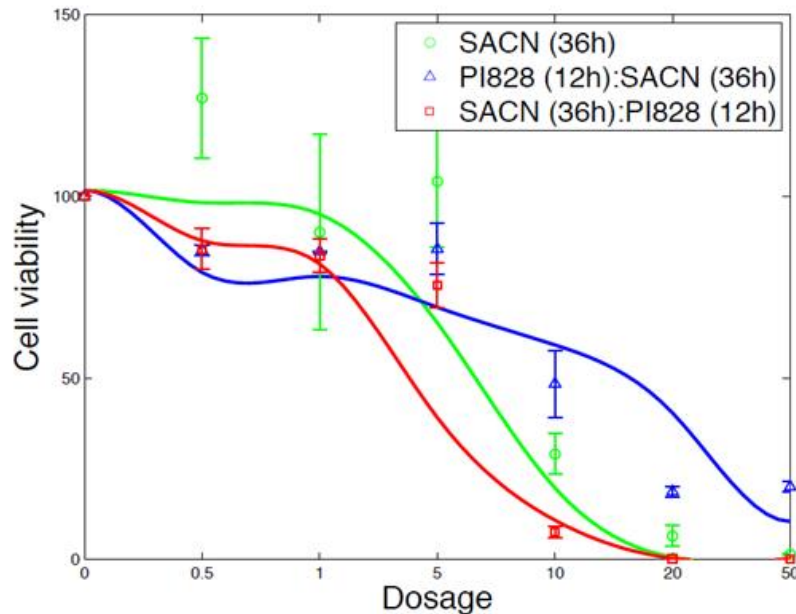




# Cell viability

- Equation:

$$\frac{dN}{dt} = [I_N P - d_N C] N$$



- Prediction: Synergistic effect if PI828 administered after 24 hours
  - 12 hours: limited cisplatin exposure
  - 36 hours: cisplatin effect is wearing off

- Collaborators
  - Mohammad Kohandel (supervisor, U. Waterloo)
  - Hamid Molavian (U. Waterloo)
  - Ambarish Pandey (U. Texas Southwestern)
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