

Data set integration for defining comprehensive models of gene regulation

Joel Smith

MBL, Woods Hole, MA



- GRN methodology for developmental systems
- [Optional] The results, examples from the sea urchin
- GRN methodology in cancer systems

20,000-30,000

2,000-3,000

200-300

20-30

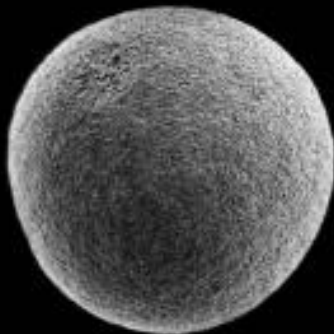
2-3

0.001

0.01

100

1,000-10,000





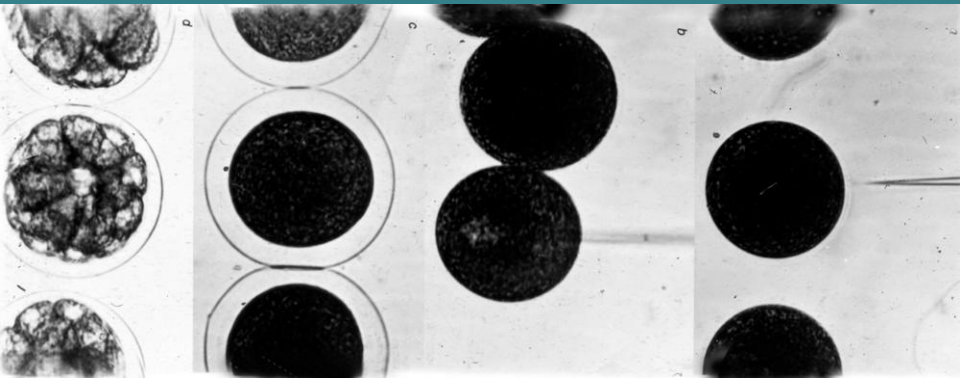
The starlet sea anemone

“Stella”

- Insights into basic animal development
- Tractable



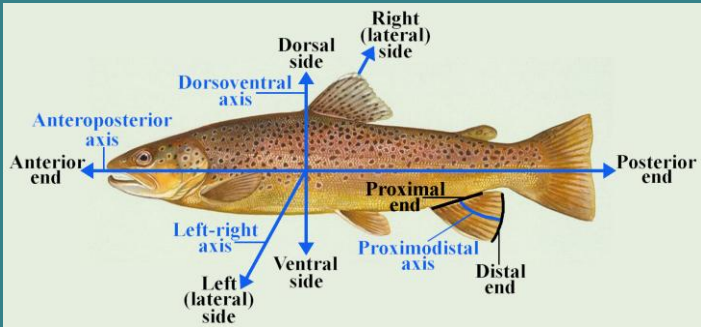
Nematostella vectensis

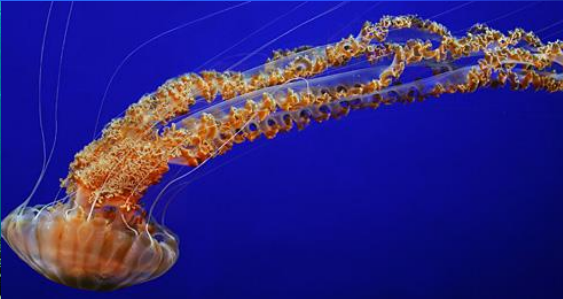


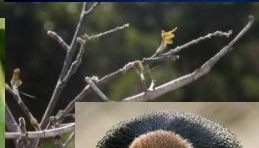
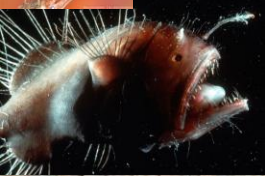




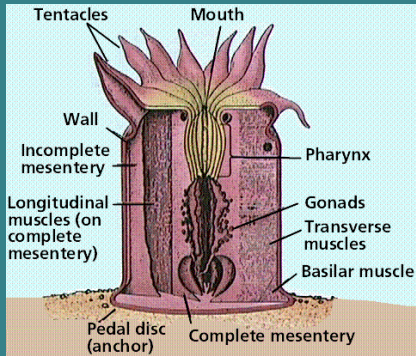
Orthogonal body axes and germ layers underly the bilaterian body plan







“Radial” and “Simple” Animals



The cnidarian body plan bears the “bilaterian” molecular signature



Wnt and TGF β pattern 1 $^\circ$ / 2 $^\circ$ axes

Cohort of genes driving mesoderm
specification and EMT



Wnt's and TGF β 's expressed along 1 $^\circ$ / 2 $^\circ$ axes

Canonical “mesoderm” genes expressed

Cnidarians and Bilaterians are sister groups

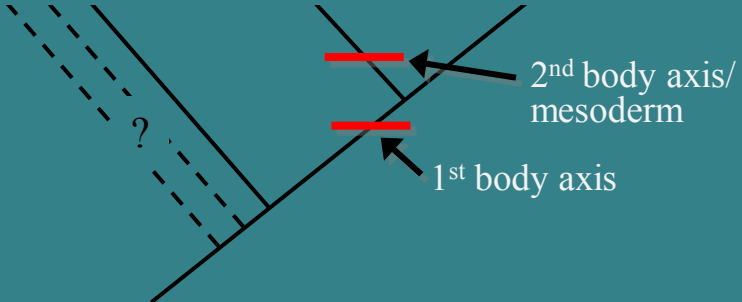
Porifera



Bilateria



Cnidaria



Cnidarians and Bilaterians are sister groups

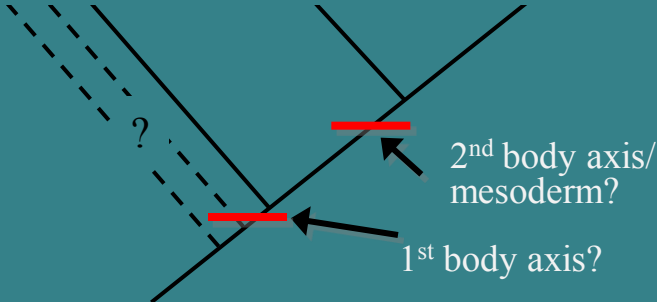
Porifera

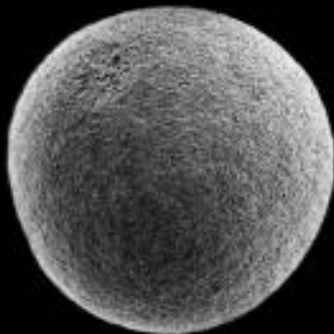


Bilateria

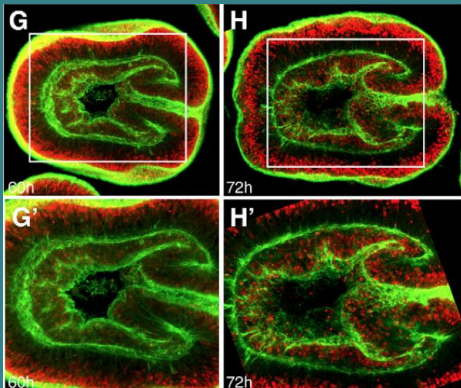
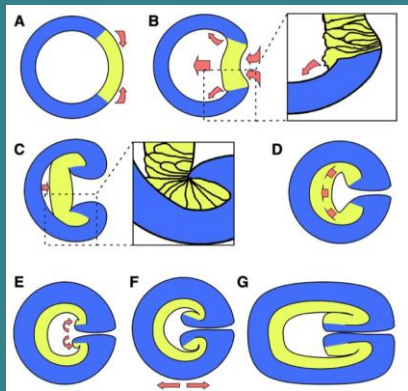


Cnidaria

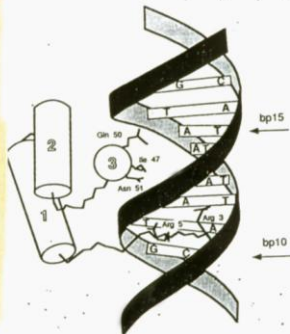
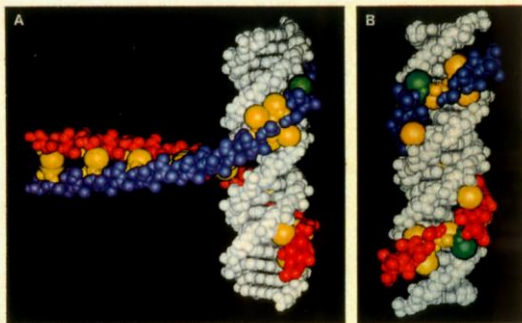






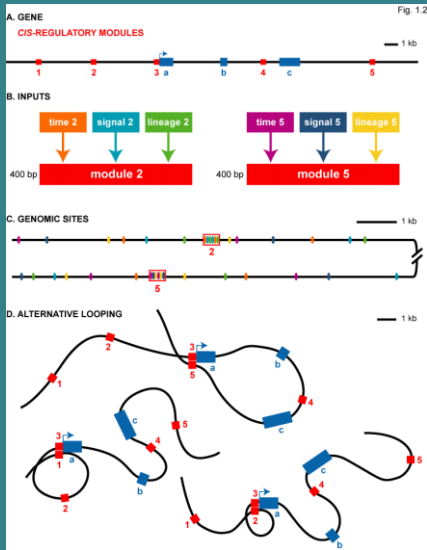


Magie et al., 2007



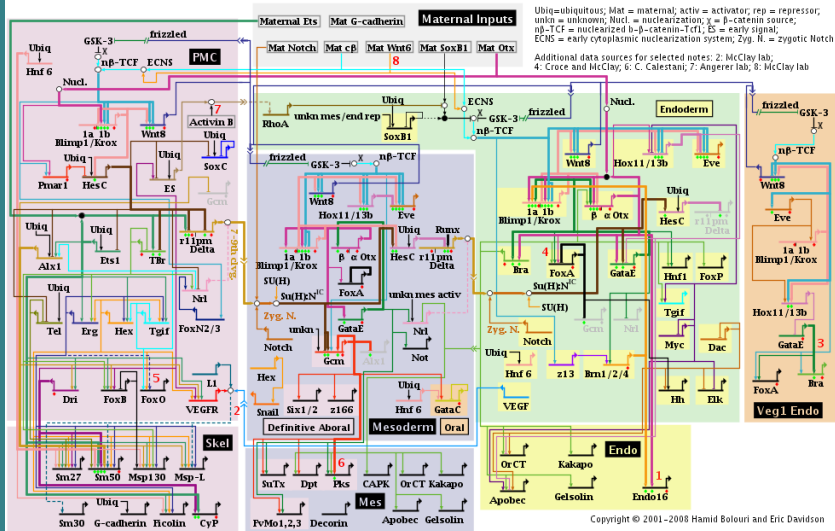
Combinations of different transcription factors

multiple “inputs” +
multiple “outputs”
= network

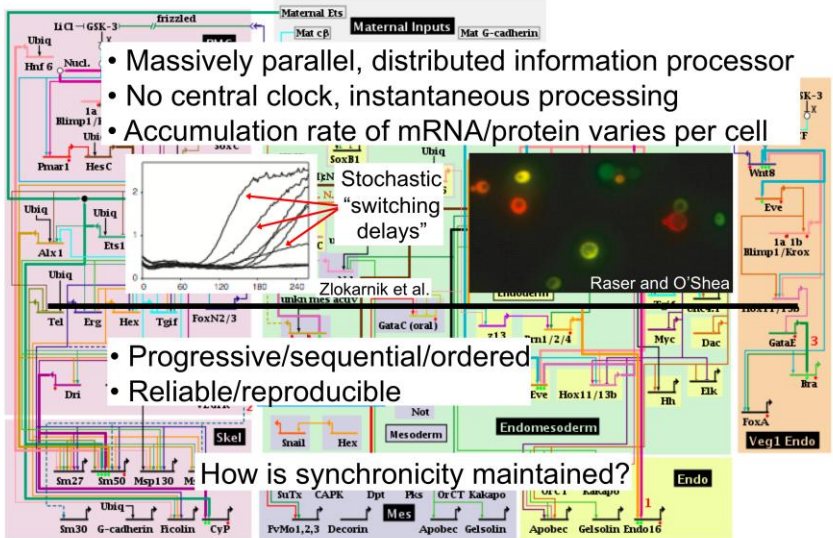


Endomesoderm Specification to 30 Hours

October 31, 2008



Copyright © 2001-2008 Hamid Bolouri and Eric Davidson



- Massively parallel, distributed information processor
- No central clock, instantaneous processing
- Accumulation rate of mRNA/protein varies per cell

- Progressive/sequential/ordered
- Reliable/reproducible

How is synchronicity maintained?

Ubiq=ubiquitous; Mat = maternal; activ = activator; rep = repressor; unkn = unknown; Nucl. = nuclearization; γ = β-catenin source; nb-TCF = nuclearized b-β-catenin-Tcf1; ES = early signal; ECNS = early cytoplasmic nuclearization system; Zyg. N. = zygotic Notch

Multi-scale problem

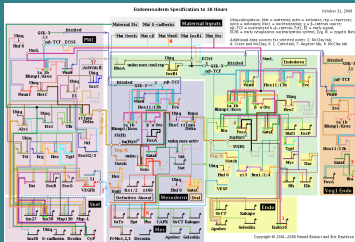
20,000-30,000 genes

2,000-3,000 “regulatory” genes

200-300 genes in control system

20-30 genes in spec. subcircuits

2-3 genes/interactions critical



Channels/gradients - milliseconds
 Signaling cascades - milliseconds
 to minutes
 Transcription - minutes to hours

Scale-integration

Which genes are relevant?

Transcriptome to Interactome

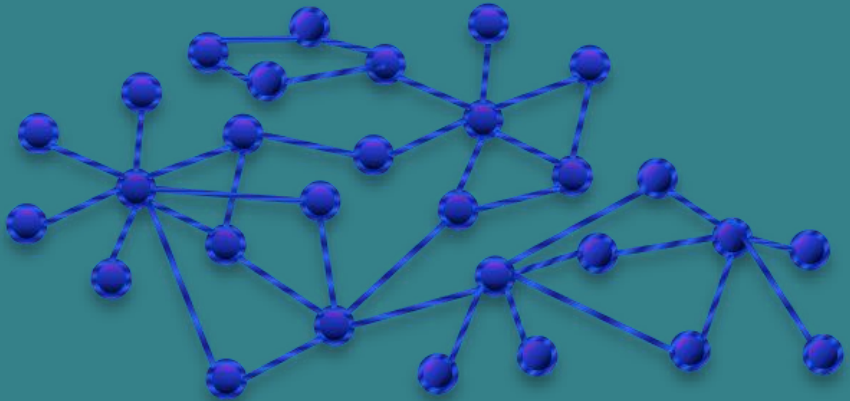
How are they functionally related?

Interactome to Network

Which interactions drive specific outcomes?

Network to Subcircuit

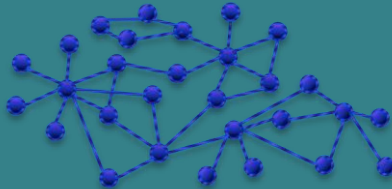
Who regulates the regulators?



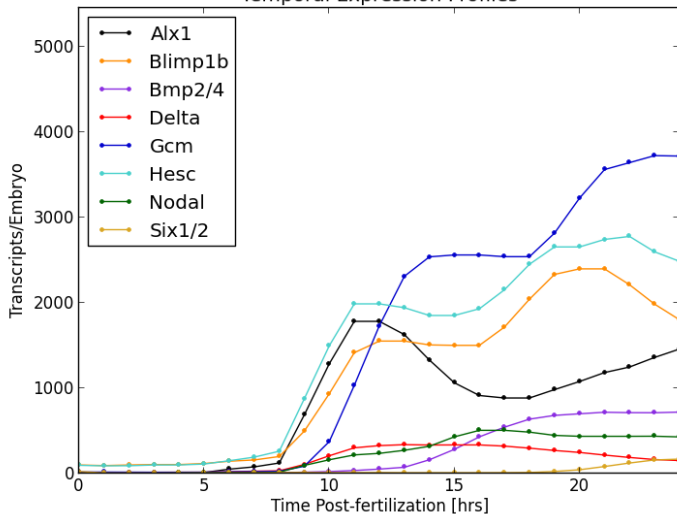
Who regulates the regulators?

Interactome “reverse-engineering”:

- the statistical dependency of each gene with every other gene
- requires hundreds of replicates
- successful with human disease biology



Temporal Expression Profiles



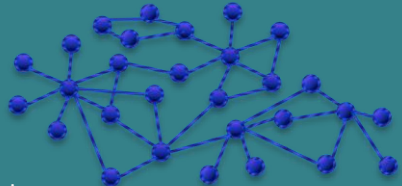
Who regulates the regulators?

Interactome “reverse-engineering”:

- the statistical dependency of each gene with every other gene
- requires hundreds of replicates
- successful with human disease biology



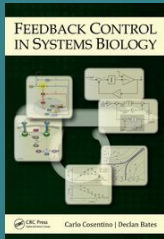
Carlo Cosentino
University of Magna Graecia,
Catanzaro, Italy



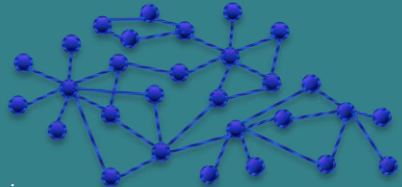
Who regulates the regulators?

Interactome “reverse-engineering”:

- the statistical dependency of each gene with every other gene
- requires hundreds of replicates
- successful with human disease biology



Carlo Cosentino
University of Magna Graecia,
Catanzaro, Italy



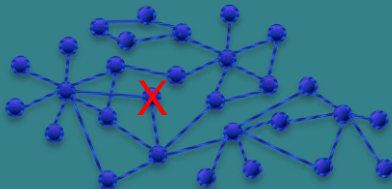
Scale-integration II: perturbations

Which genes are relevant?

Transcriptome to Interactome

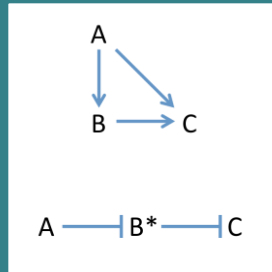
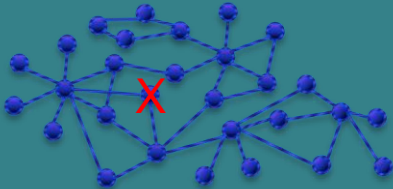
How are they functionally related?

Systematic perturbations



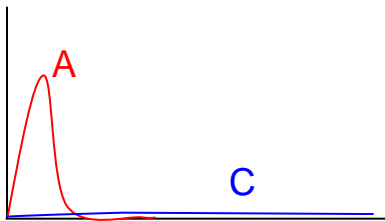
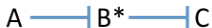
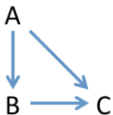
Scale-integration II: perturbations

Even exhaustive perturbations fail to resolve common, functionally important network structures



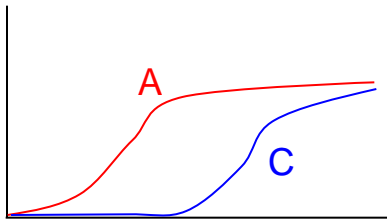
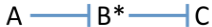
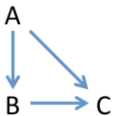
Scale-integration II: perturbations

Perturbation analysis failure



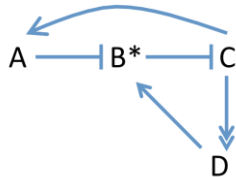
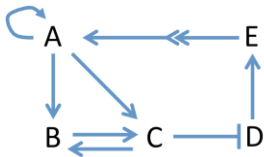
Scale-integration II: perturbations

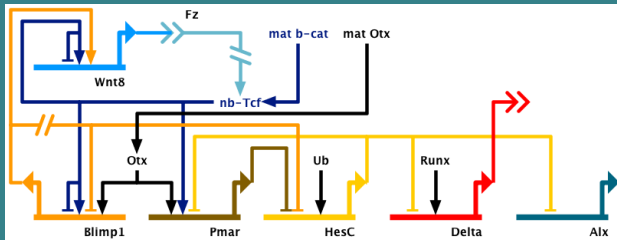
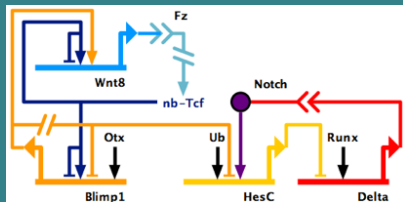
Perturbation analysis failure

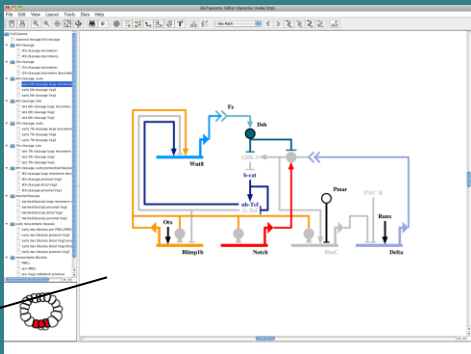
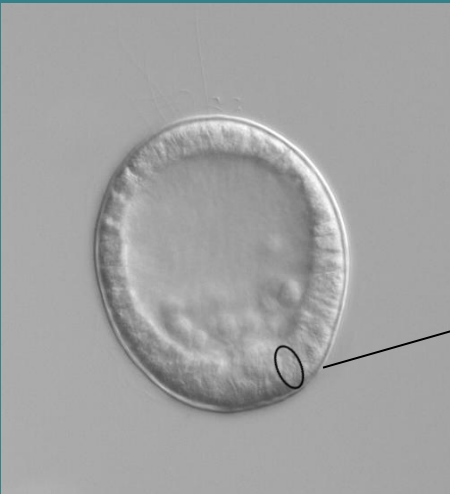


Scale-integration II: perturbations

Perturbation analysis failure

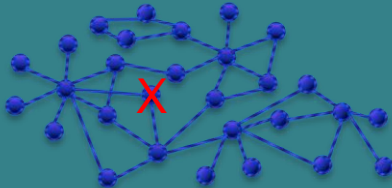






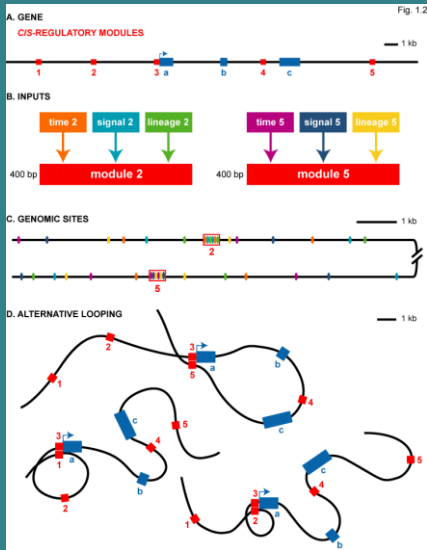
Scale-integration II: perturbations

Even exhaustive perturbations fail to resolve common, functionally important network structures



+

Complementary
data sets

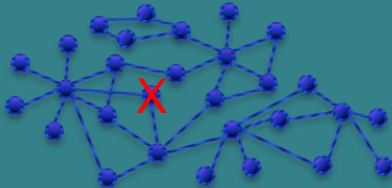


Genomic *cis*-regulatory elements

TF binding preferences

Scale-integration II: perturbations

Even exhaustive perturbations fail to resolve common, functionally important network structures



+

Complementary
data sets

Scale-integration III: testing network switches

Which genes are relevant?

Transcriptome to Interactome

*How are they functionally
related?*

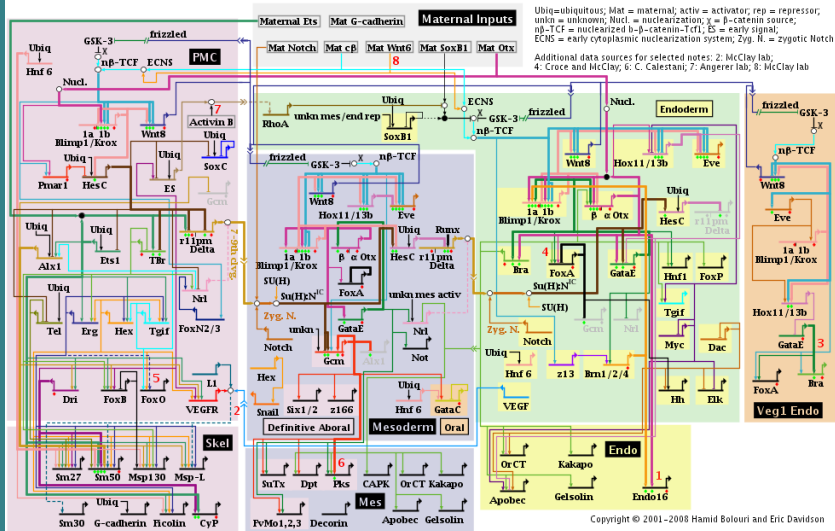
Interactome to Network

*Which interactions drive
specific outcomes?*

Network to Subcircuit

Endomesoderm Specification to 30 Hours

October 31, 2008



Copyright © 2001-2008 Hamid Bolouri and Eric Davidson

Scale-integration approach

Which genes are relevant?

First-pass “reverse engineering” the interactome

RNA-seq HD expression analysis, perturbation settling



Interactome

How are they functionally related?

Systematic perturbation analysis

Exhaustive pert-seq, complementary assays, merging diverse data sets



Network

Which interactions drive specific outcomes?

Hypothesis testing, kinetic modeling

CRNT modeling, *cis*-regulatory validation, network reliability testing



Dynamic Network

Scale-integration approach

Which genes are relevant?

Observation

Interactome

How are they functionally related?

Hypothesis generation

Network

Which interactions drive specific outcomes?

Hypothesis testing

Dynamic Network

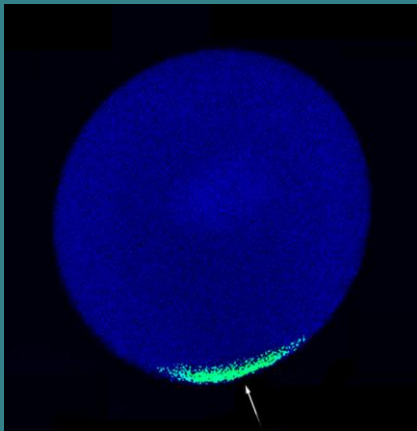
High Sensitivity
(Low Specificity)

Increasing
Specificity

Cis-regulatory analysis

- BAC reporters capture genomic context
- *Cis*-reengineering tests subcircuit function within endogenous regulatory context

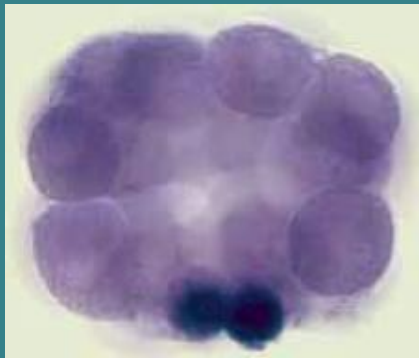
Vegetal Dsh



Ettensohn, et al.

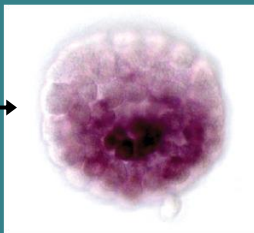


Vegetal *wnt8*

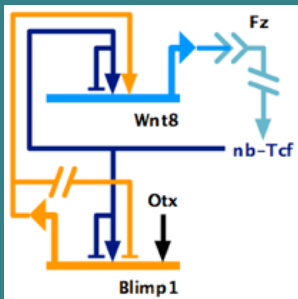


Wikramanayake, et al., Minokawa, et al.

The “Torus Subcircuit”



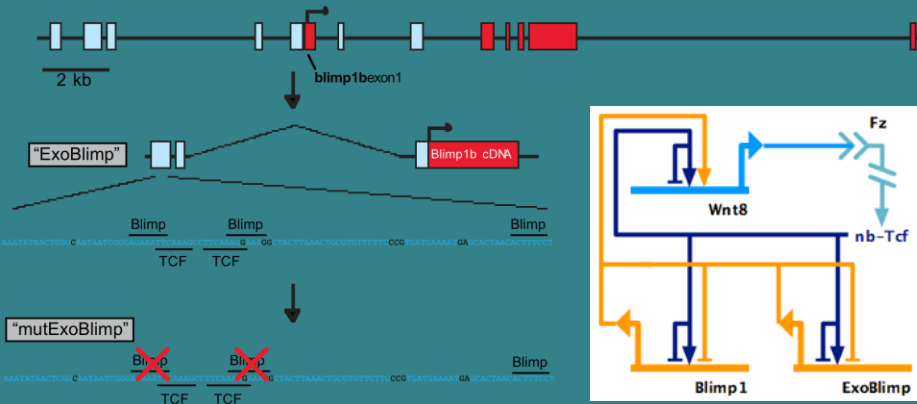
The “Torus Subcircuit”



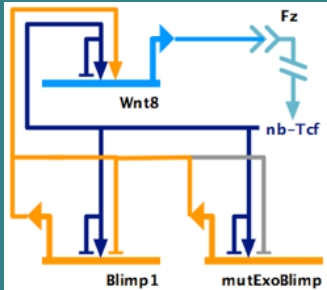
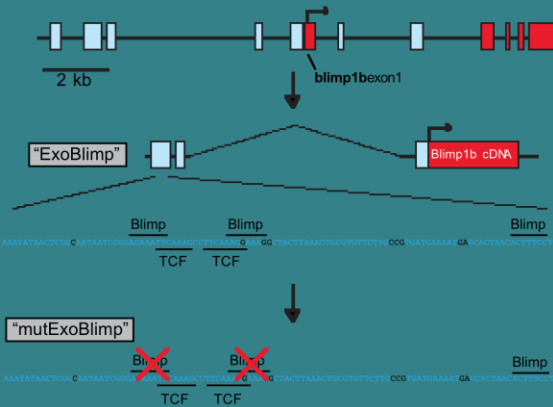
Expansion by ligand diffusion

S
h
u
t
d

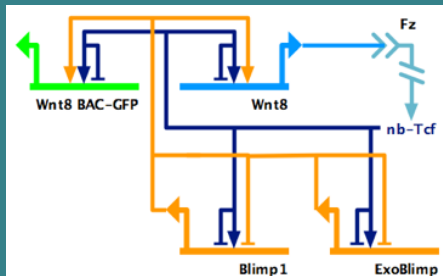
Testing the Torus Subcircuit: ExoBlimp



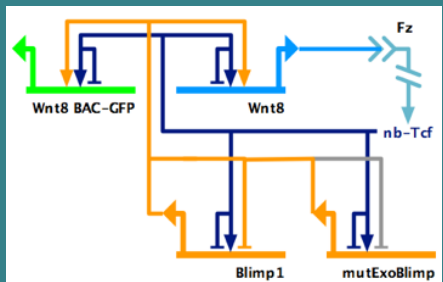
Testing the Torus Subcircuit: mutExoBlimp



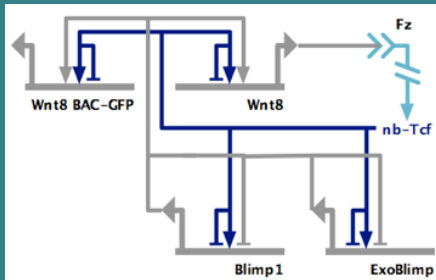
Control



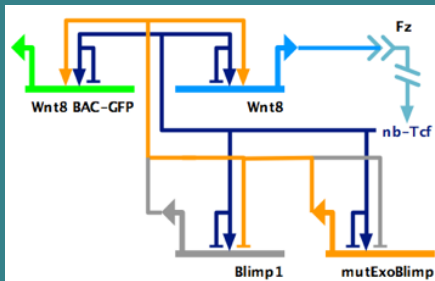
Test



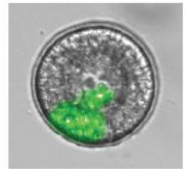
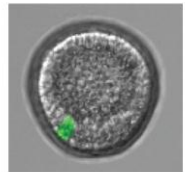
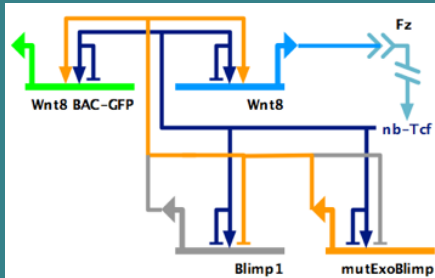
Control



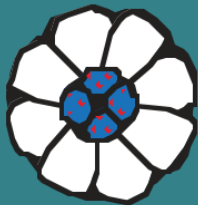
Test



mutExoBlimp assay (I): Wnt8 BAC-GFP



Wnt8 and Delta domains remain spatially aligned



**Cleavage/
early blastula**



Blastula



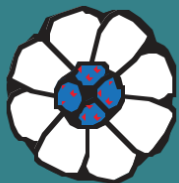
**Mesenchyme
blastula**

Blue = Wnt8/Blimp1

Red = Delta

Gene regulatory network subcircuit controlling a dynamic spatial pattern of signaling in the sea urchin embryo

Smith and Davidson, PNAS, **105**, 20089 (2008)



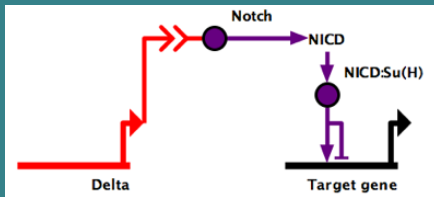
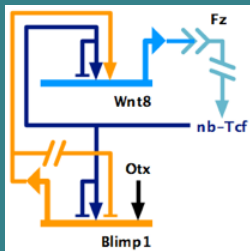
Cleavage/early blastula



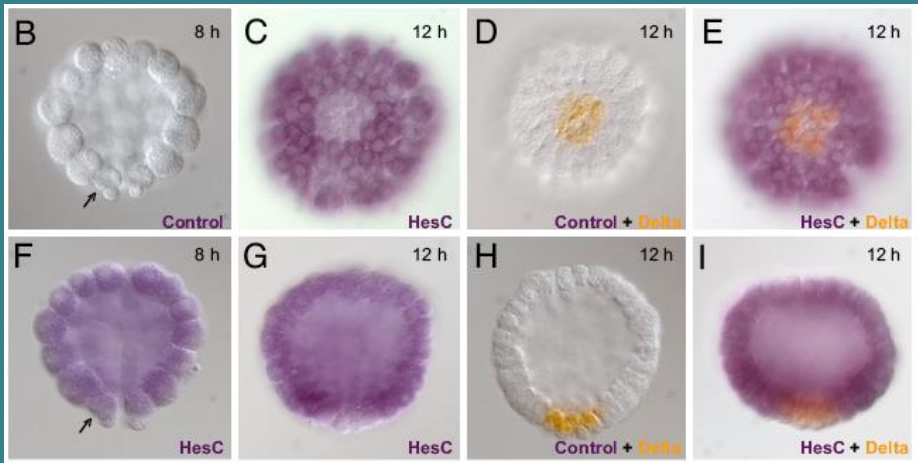
Blastula



Mesenchyme blastula



hesC and *delta* expression domains are exclusive



Revilla-i-Domingo *et al.*, 2007



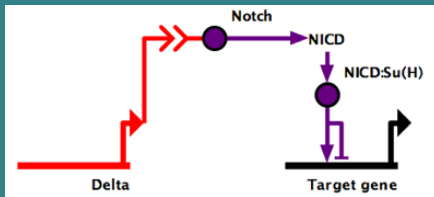
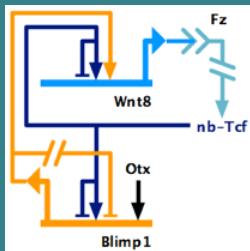
Cleavage/early blastula

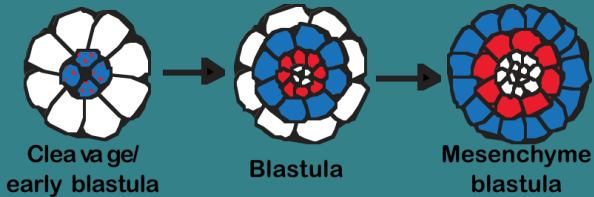
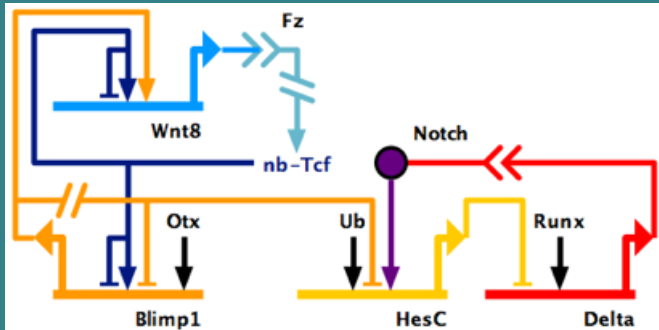


Blastula

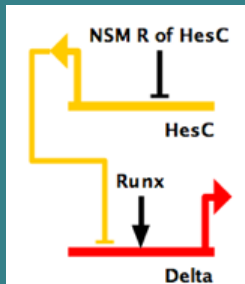


Mesenchyme blastula





cis-Regulatory analysis of *hesC*

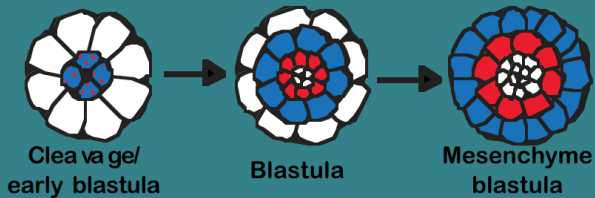
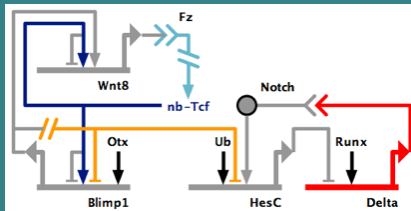
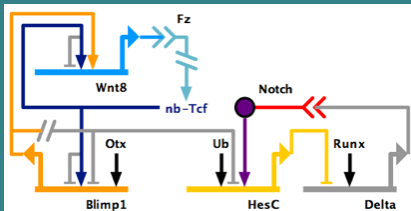


ACGCTCGAGCGCCACGGGCACGTGCTGTCTATATATAGCGGTCGTCTGCCGGGTAGTAAGTCATAACA
 ACCGCTCTCAGCGCTCGTCCACAAGATCCATACTCCTCTTCGTGGATACCAGTGACTATCTCTCTGCCTC
 TAAGGAATCTAACTCCTGTGATTATACTTCTTGCTTGTGCATTGAGACCATCTATATCAGCAATCATG
 CTTACTTCATCTGGATACCAACAAATGGACATGTGCTCTAACAGACCTAGGGTAAGTTCTTCTCAACC
 ATTTCTTCAATCACACATTTTCTCTTTTGAATTCACAAGTTAAGTCGAAACCGTGCATCAGTAA
 CCATGTTTACTTAGTAAGTTAACAGTAATCACGTTAACTTGTTCATCATGATCATTGATCTAATCATT
 GATCATTGATTAATAGTTGATTTCAGCAAAGGCCAAATACCTTCAATCTACTTTCAACTTAAC TGACA
 ATAACATCAAGGAACGAACATCAGACTCGTATGTTGATAAATGATTCATATTGATGATTCCAT
 GCACTTGATTTTCAATTGAGGACTGTTGACTTATTCTGTACCCTTCTTGTATTTCATCTACA

end exon 1 | begin intron 1

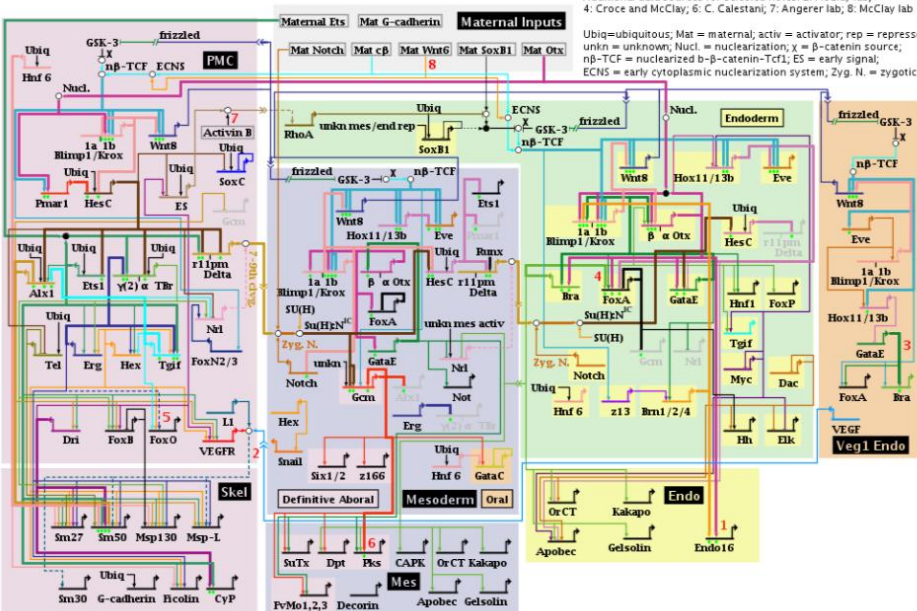
end intron 1

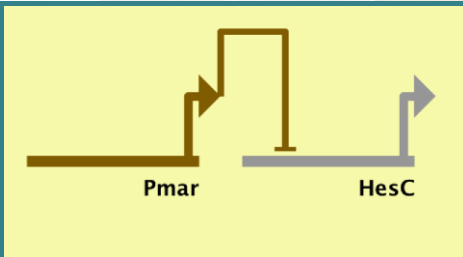
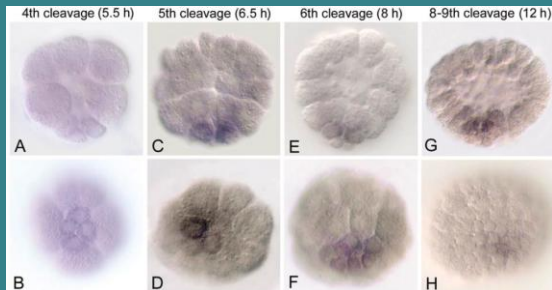
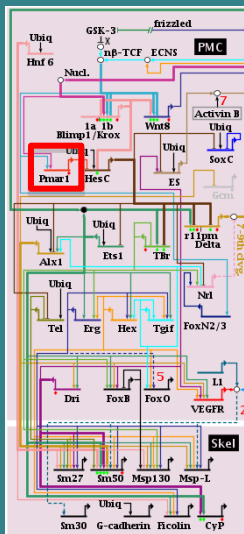
5' (A/C/T)(A/G)(G/T)NGAAAG(G/T)(A/G/T)-3'



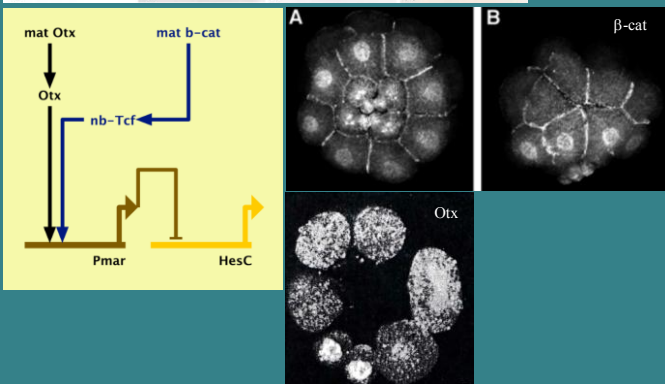
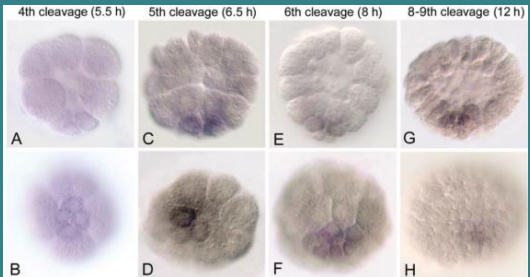
Additional data sources for selected notes: 2: McClay lab; 4: Croce and McClay; 6: C. Calestani; 7: Angerer lab; 8: McClay lab

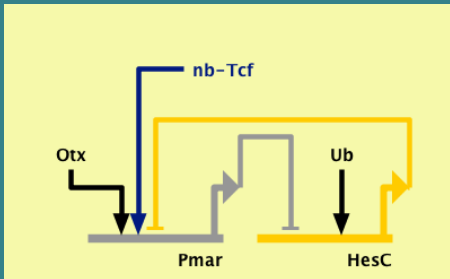
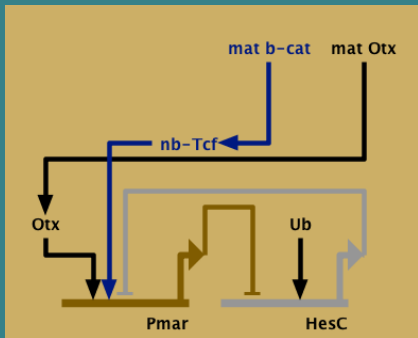
Ubiq=ubiquitous; Mat = maternal; activ = activator; rep = repressor; unkn = unknown; Nucl. = nuclearization; x = β -catenin source; n β -TCF = nuclearized b- β -catenin-Tcf1; ES = early signal; ECNS = early cytoplasmic nuclearization system; Zyg. N. = zygotic Notch





Revilla-i-Domingo *et al.*, 2007; Oliveri *et al.*, 2002, 2003

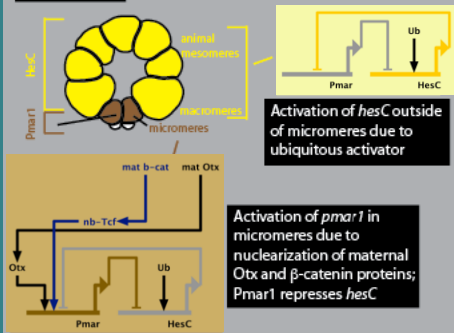




Pmar1-HesC subcircuit

A

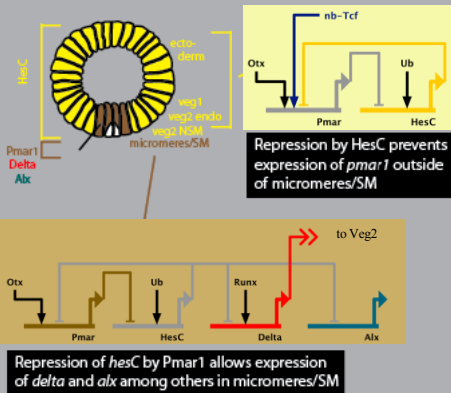
4-5th cleavage

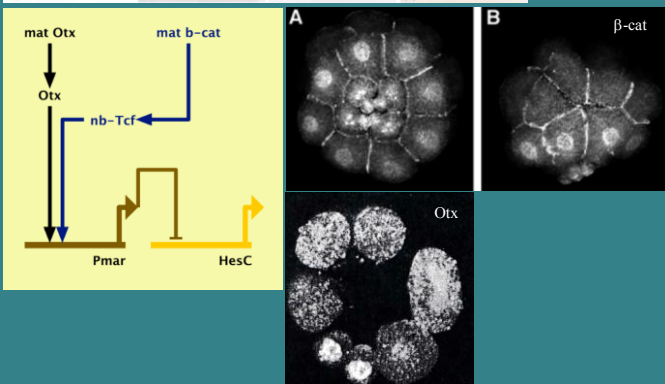
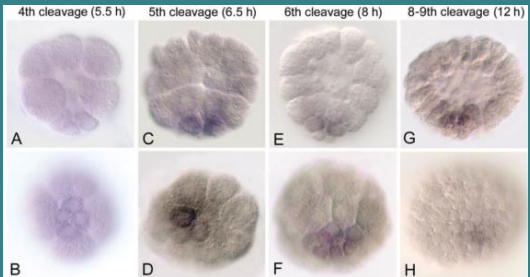


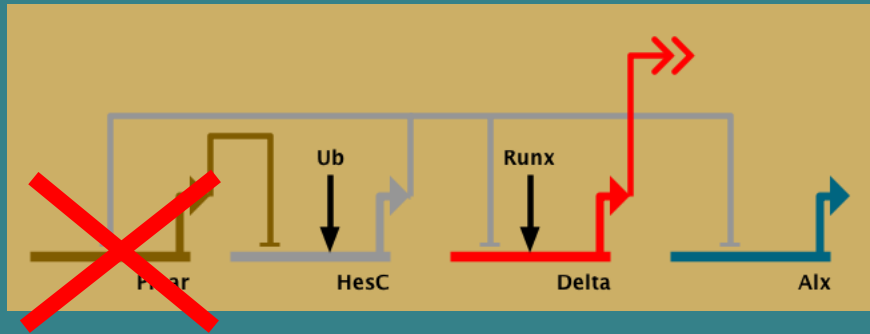
Reciprocal repression

B

Blastula







Control
MASO



24 h

53/55

Pmar
MASO



33/44



7/44

Pmar MASO:
shuts down
skeletogenic
mesoderm
specification
program

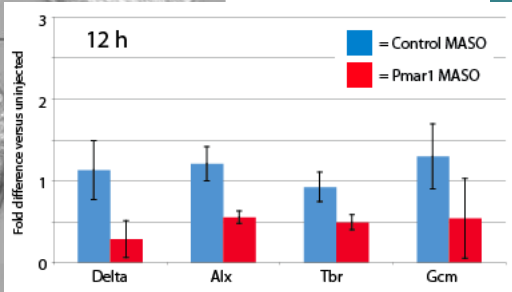
Control
MASO



24 h

53/55

Pmar
MASO



Pmar MASO:
shuts down
skeletogenic
mesoderm
specification
program



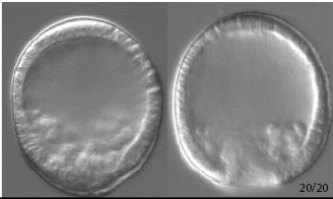
7/44



However: embryos recover from Pmar MASO by 30h!

30 h

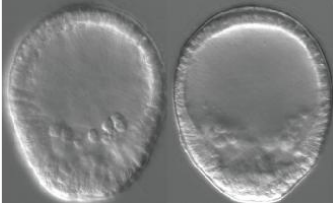
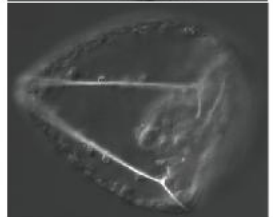
72 h



Control
MASO



Pmar
MASO

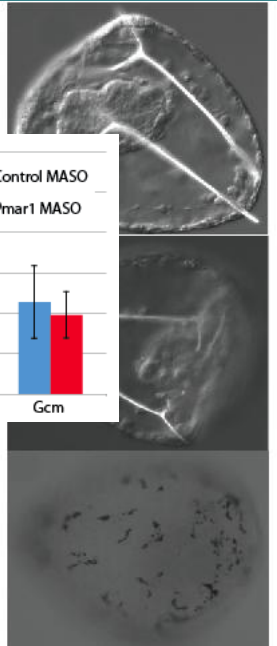
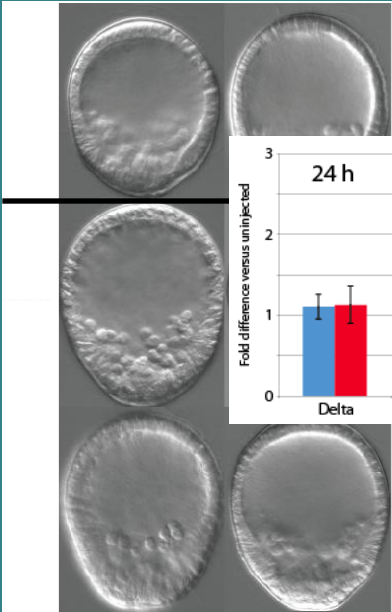
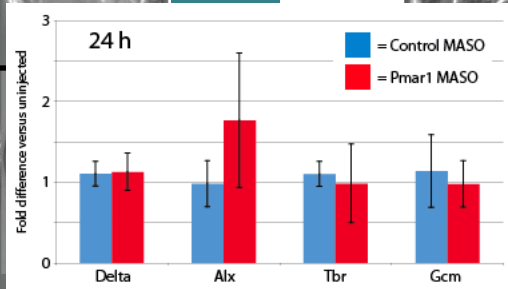


However: embryos recover from Pmar MASO by 30h!

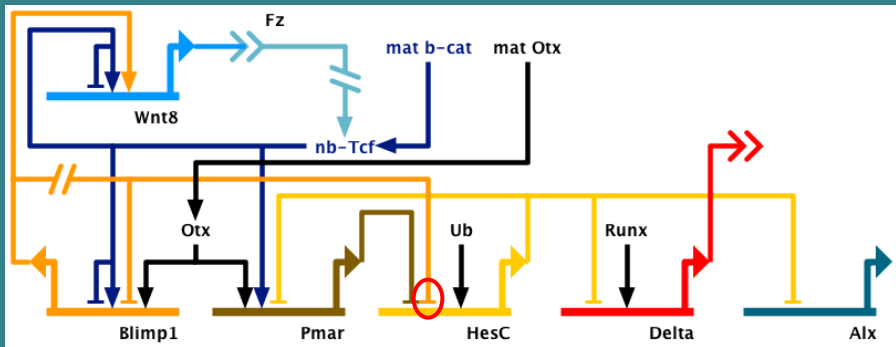
30 h

72 h

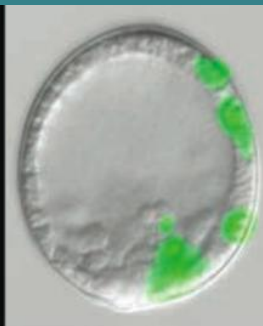
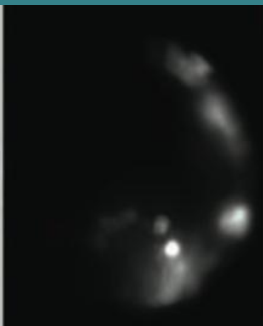
Control
MASO



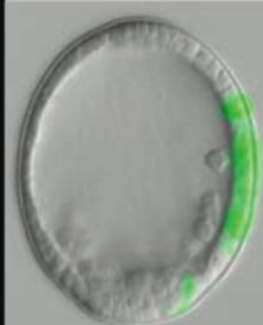
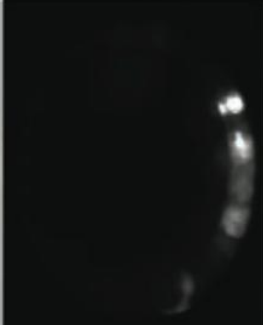
Does regulative recovery depend on repression of *hesC* by Blimp?



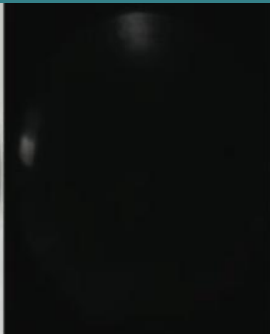
Control



Δ Blimp
HesC



HesC
 Δ Blimp



Δ Blimp
HesC



Two systems:
same maternal/early
inputs

The older one is
able to compensate
for disruption of the
newer one

