

Slow delayed rectifier current protects ventricular myocytes from arrhythmic dynamics across multiple species

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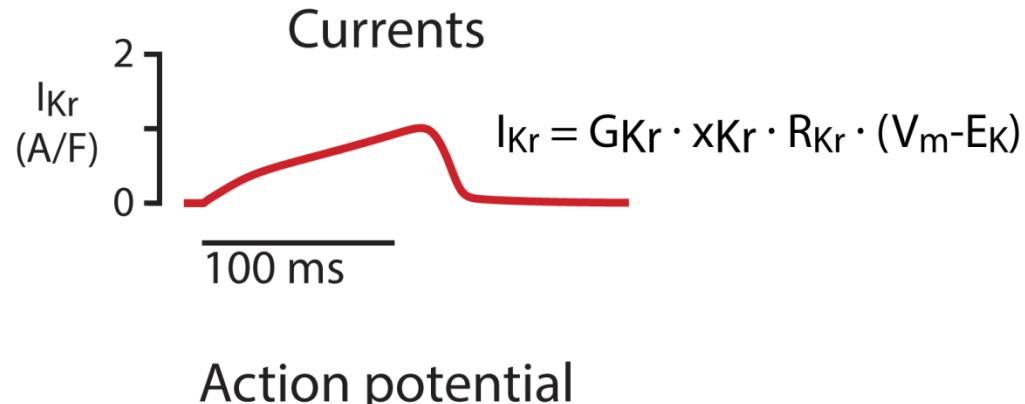
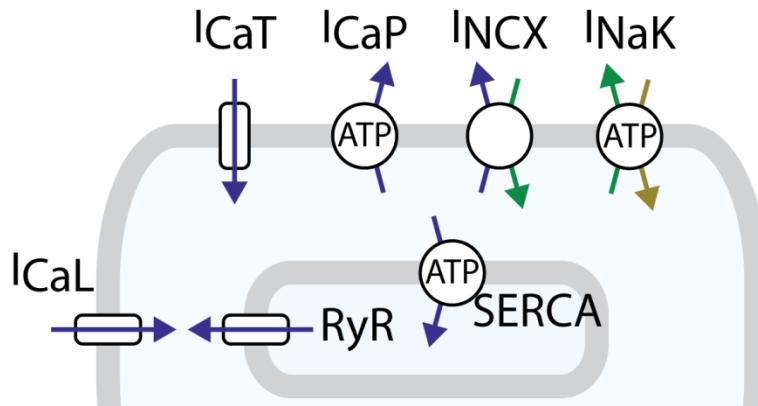
Icahn School
of Medicine at
**Mount
Sinai**



Integrative Cardiac Dynamics
July 3, 2018

Studying the cardiac myocyte with mathematical models

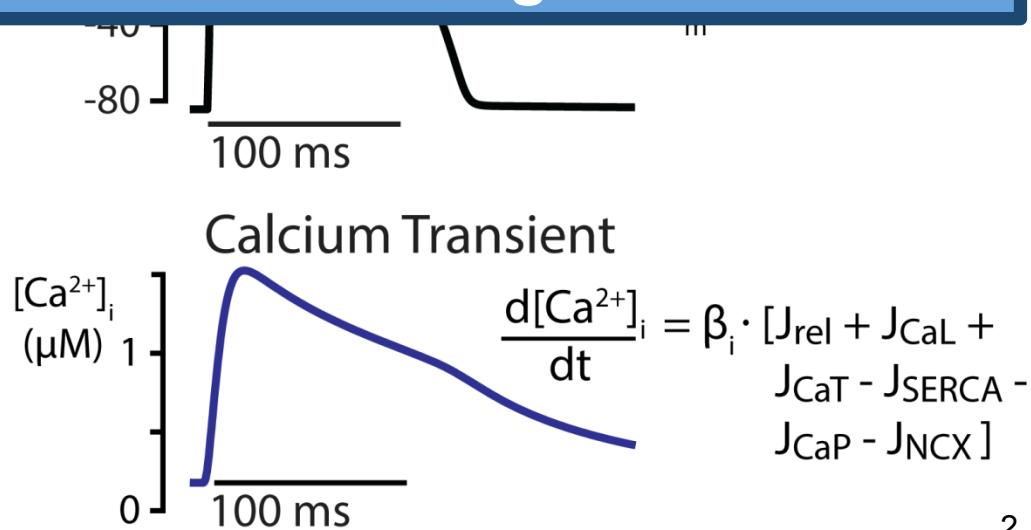
First cardiac cell model published in 1962 – hundreds of subsequent models



Overall Strategy: Exploit existing models to develop a systems-level understanding

I_{K1} I_{Kr} I_{Ks} I_{Kp} I_{Na}

- 10-20 ion channels, pumps, and transporters
- 20-60 ordinary differential equations



Outline

Control of ventricular action potential duration

Coupling of modeling and experiments (guinea pigs)

Developing more predictive mathematical models

Implications for cellular arrhythmia dynamics

Hypothesis: slow delayed rectifier I_{Ks} is stabilizing

Extending the I_{Ks} stabilization hypothesis

Testing multiple mathematical models

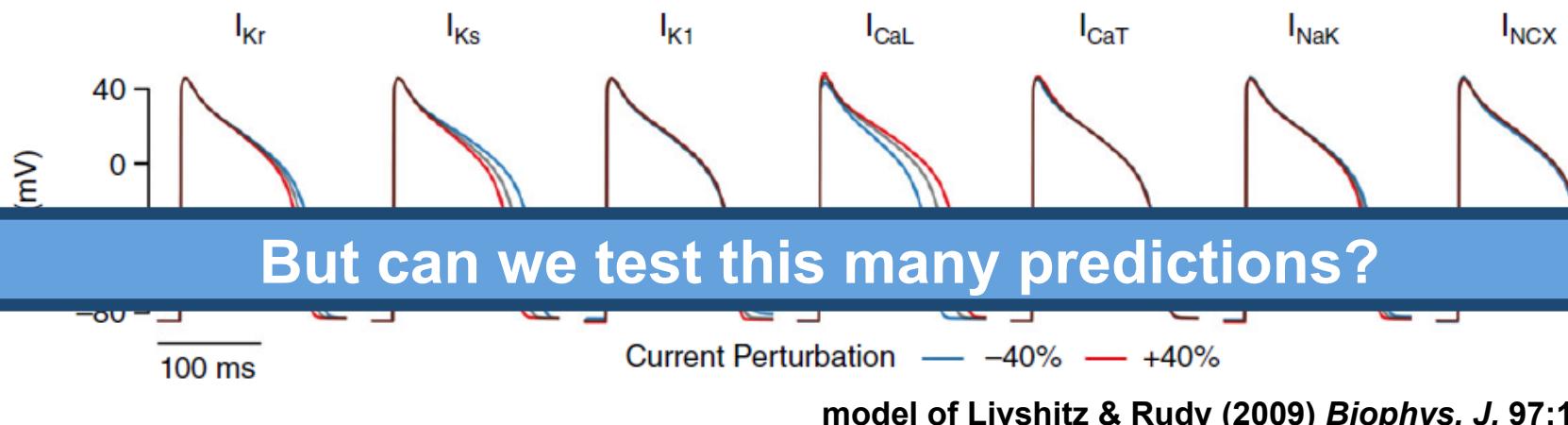
Mechanistic explanations

Physiological importance: β -adrenergic stimulation

How do we choose parameters and improve the existing models?

Test how the system responds to perturbations

Model predictions: augmenting/inhibiting guinea pig ionic currents



Ryan Devenyi, M.D., Ph.D.

J Physiol 000.00 (2016) pp 1–17

Differential roles of two delayed rectifier potassium currents in regulation of ventricular action potential duration and arrhythmia susceptibility

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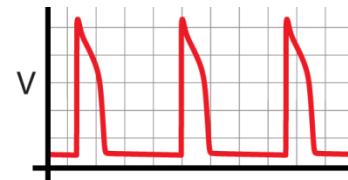
²Physiology, Biophysics, and Systems Biology Graduate Program, Weill Cornell Graduate School, New York, NY, USA

³Greenberg Division of Cardiology, Weill Cornell Medical College, New York, NY, USA

⁴IMEC, Holst Centre, Eindhoven, The Netherlands

Dynamic Clamp: a real-time, closed loop system for altering ionic current levels

1. Record V_m via patch electrode



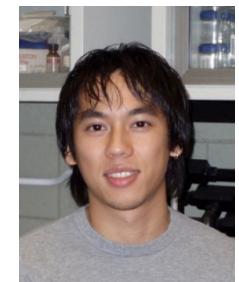
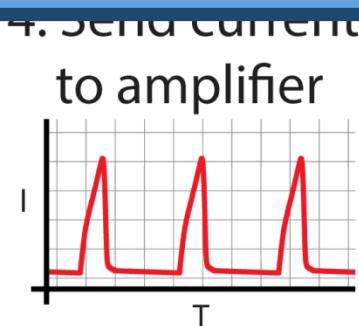
2. Send V_m to computer

3. Calculate current in real-time

$$I_{Kr} = G_{Kr} \sqrt{\frac{K_o}{5.4}} x_{Kr} R_{Kr} (V_m - E_K)$$

Digital implementation allows multiple perturbations to be tested in each cell

current

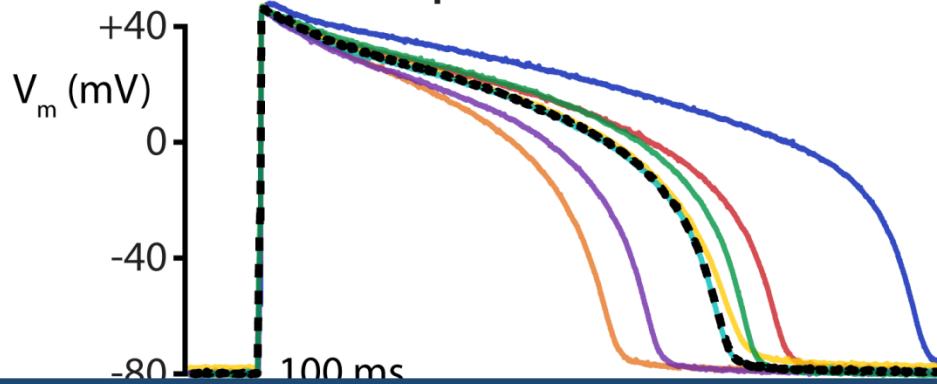


Prof. David Christini, Francis Ortega
Weill-Cornell Medical College

Larger changes in experiments than in model

Dynamic clamp in guinea pig myocytes

Experiment



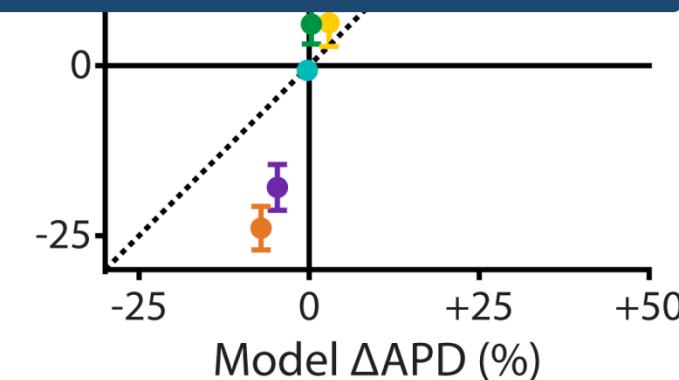
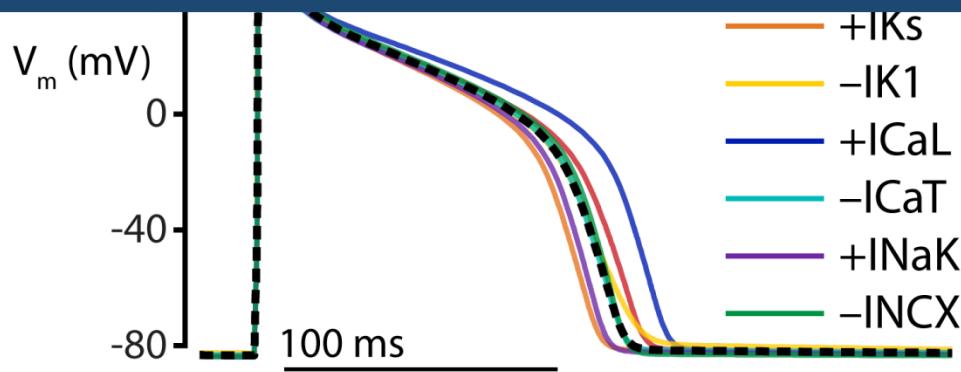
Model vs. Experiment
Changes in APD

Experiment
ΔAPD (%)

+50

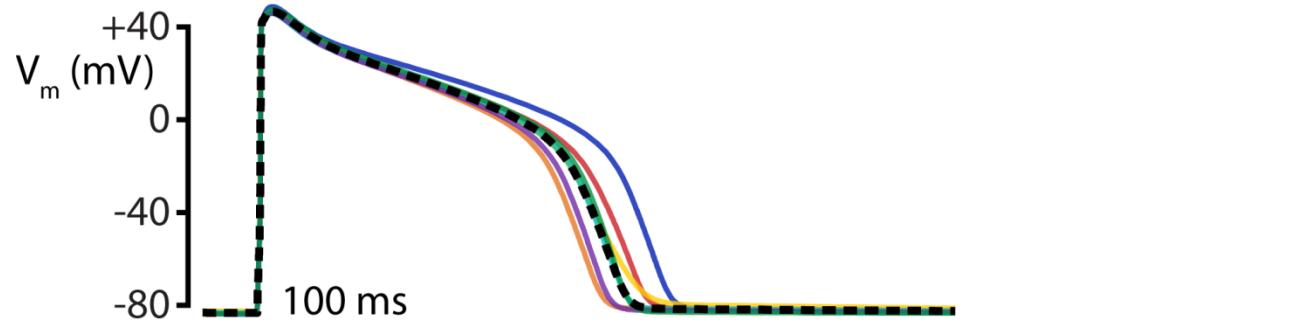
-40

Can we adjust the model to improve agreement with experiment?

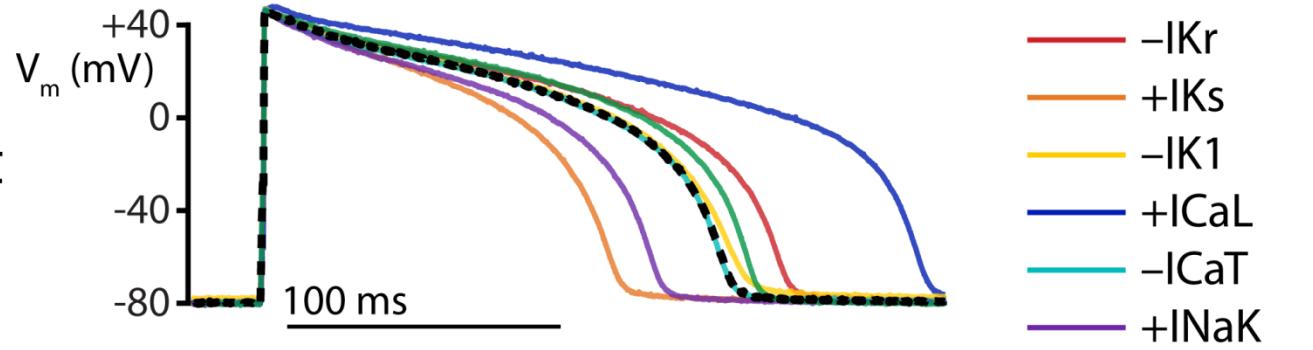


A Genetic Algorithm improves agreement between simulation and experiment

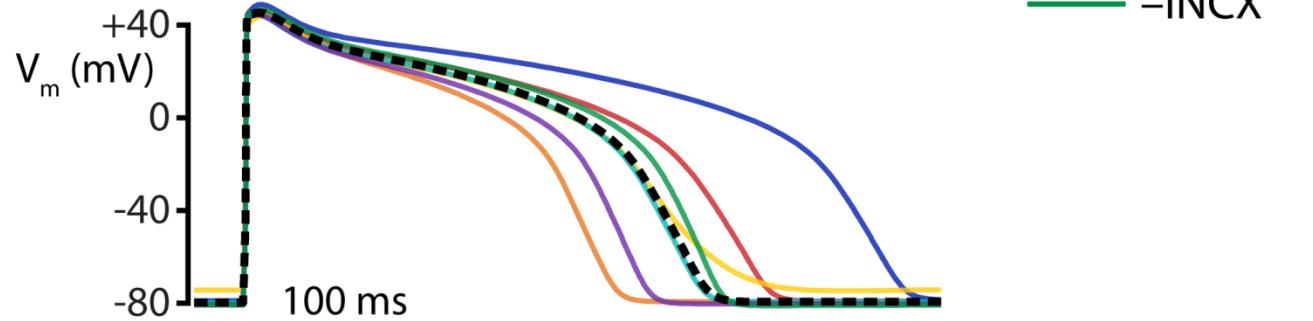
Original Model



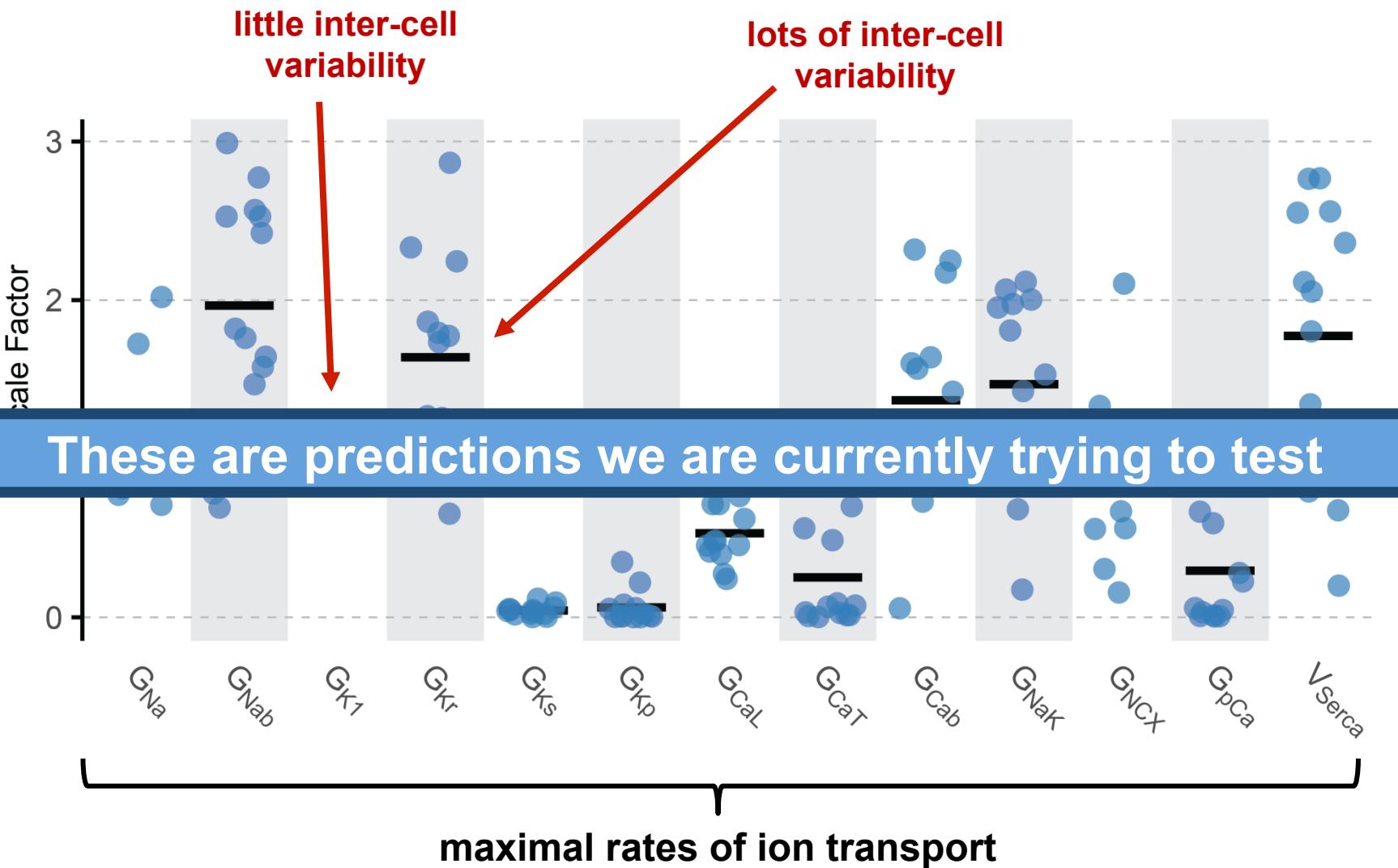
Experiment



Adjusted Model

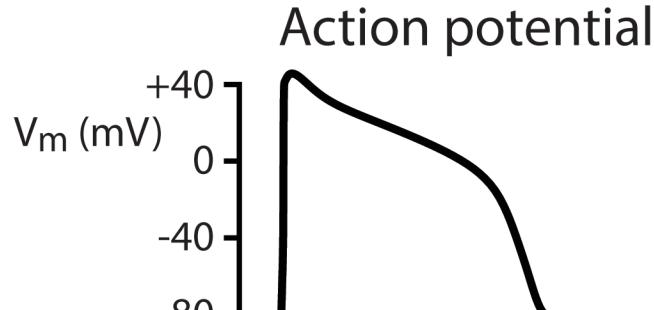


The GA fit generates a parameter set for each experimental sample

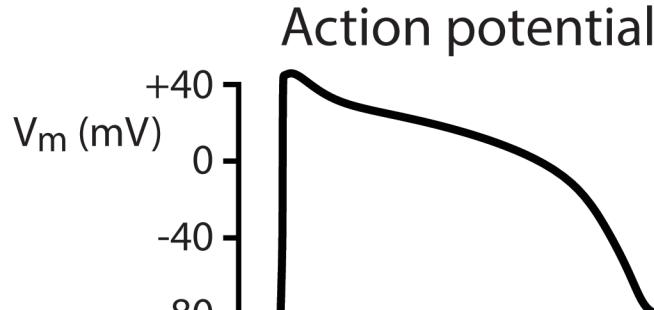


The new model behaves differently

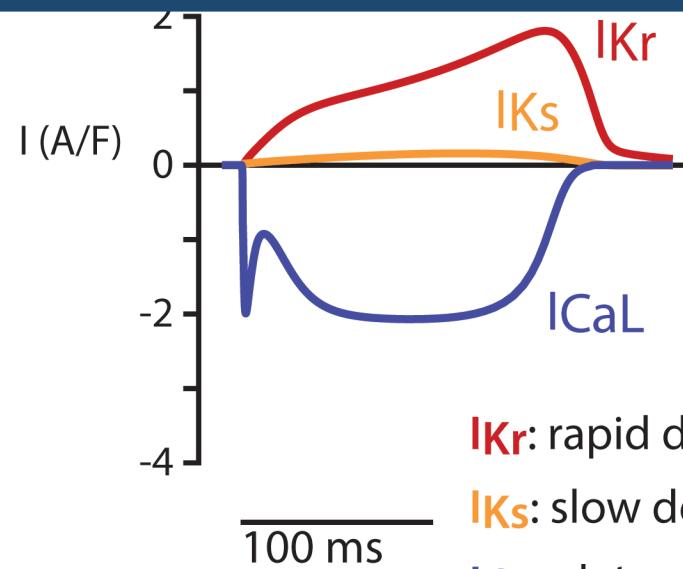
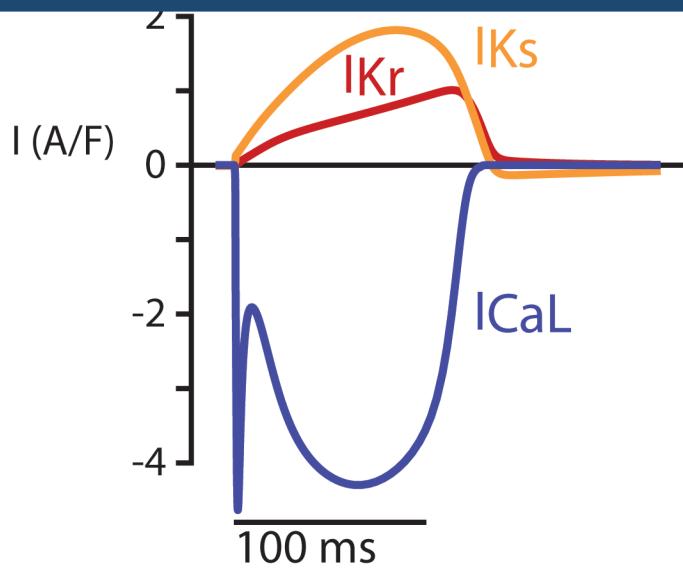
Original Model



Adjusted Model



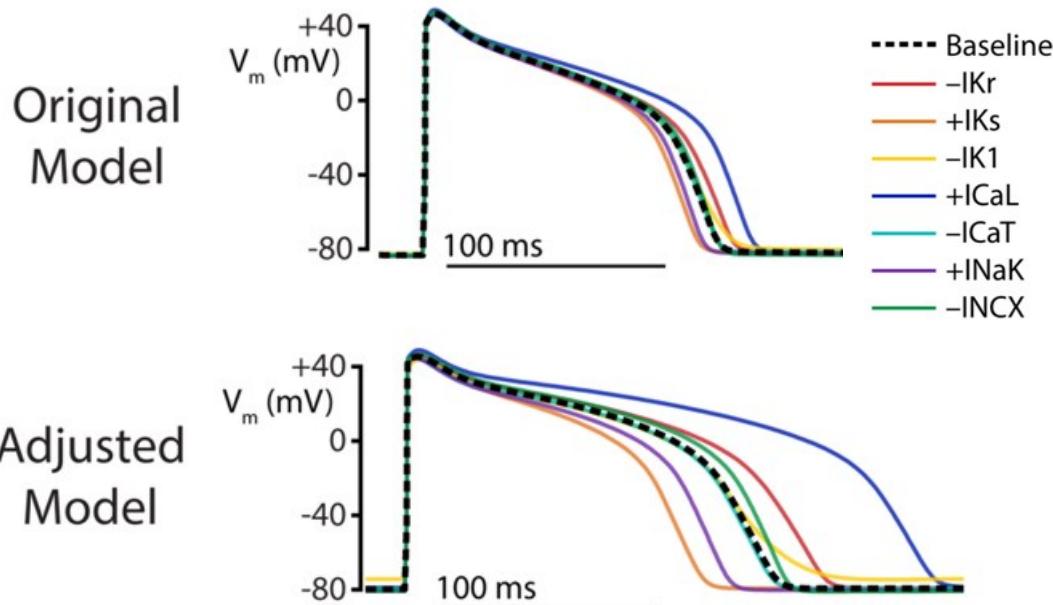
Adjusted model has different balance of K^+ currents



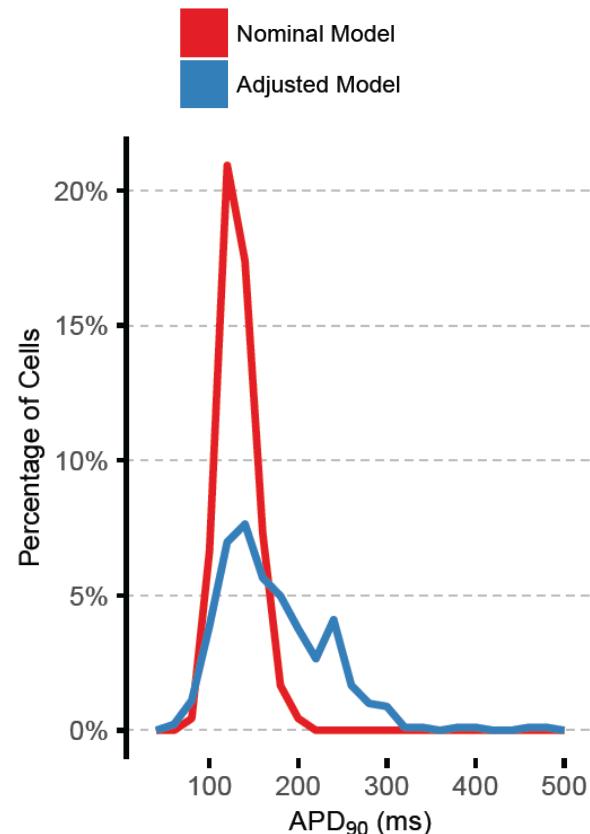
IKr : rapid delayed rectifier
 IKs : slow delayed rectifier
 $ICaL$: L-type Ca^{2+} current

Decreased stability in the adjusted model

Response to perturbations

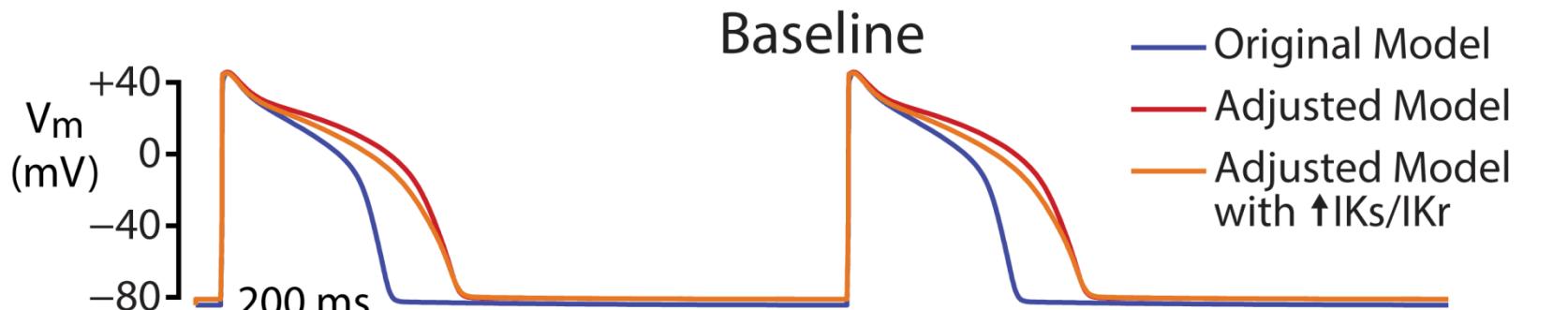


Population simulations

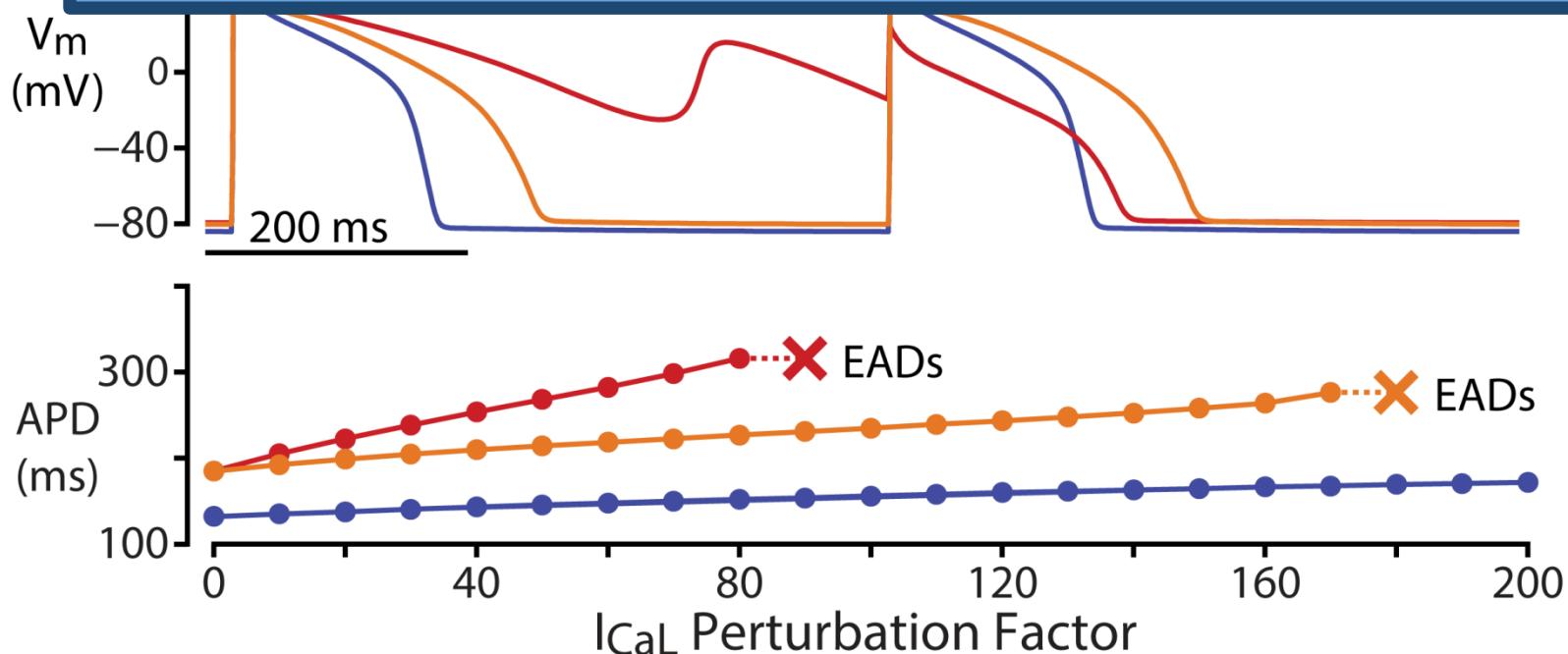


Hypothesis: decreased stability may predispose cells to arrhythmic dynamics

Different K⁺ currents have different effects on pro-arrhythmic EADs



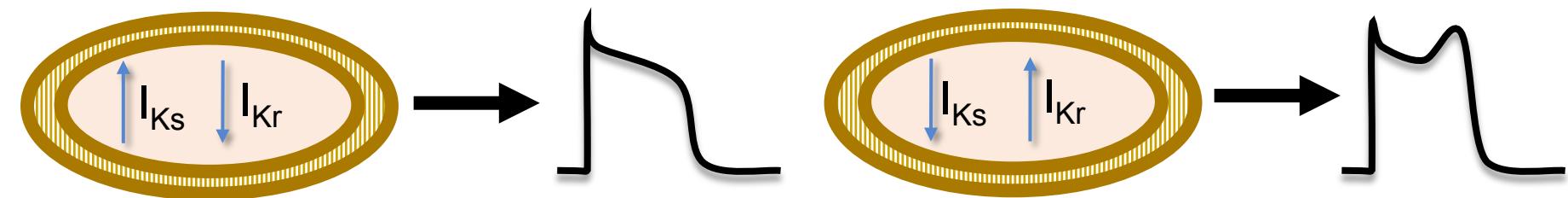
I_{Ks} prevents EADs better than I_{Kr} does



Summary

Combining perturbation modeling with thorough experimental tests gives insight into AP dynamics

1. Roles of complementary K⁺ currents, I_{Kr} and I_{Ks}
2. Predictions about pro-arrhythmic events (EADs)
3. A more predictive mathematical model



Goals

- (1) Is this concept true in other animal and human models?
- (2) What is the underlying mechanism that allows I_{Ks} to be protective?
- (3) Is this concept true only at baseline or during other physiological conditions such as during β -adrenergic activation?



Meera Varshneya
Ph.D. candidate

(1) Is this concept true in other species models?



(1) Fox et al.
DOI: [10.1152/ajpheart.00612.2001](https://doi.org/10.1152/ajpheart.00612.2001)

(2) Hund et al.
DOI: [10.1161/01.CIR.0000147231.69595.D3](https://doi.org/10.1161/01.CIR.0000147231.69595.D3)



(3) Shannon et al.
DOI: [10.1529/biophysj.104.047449](https://doi.org/10.1529/biophysj.104.047449)



(4) Livshitz et al.
DOI: [10.1016/j.bpj.2009.05.062](https://doi.org/10.1016/j.bpj.2009.05.062)

(5) Devenyi et al.
DOI: [10.1113/JP273191](https://doi.org/10.1113/JP273191)

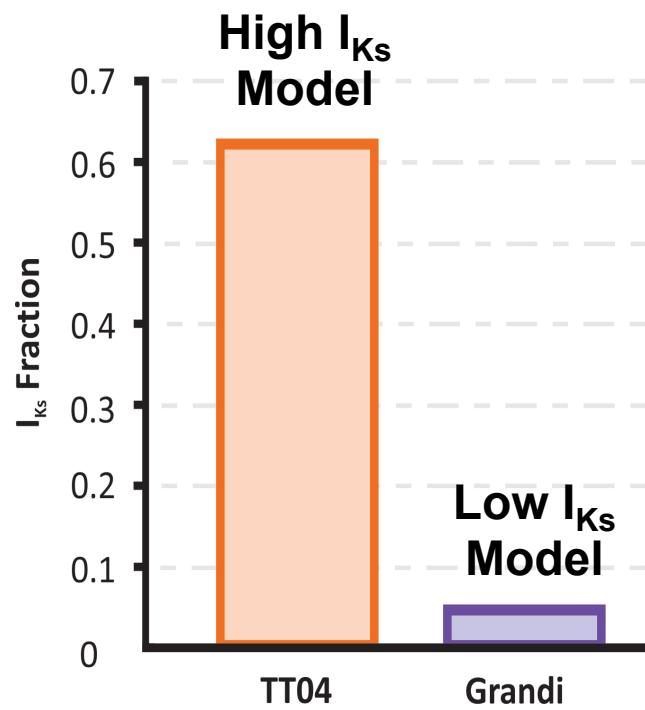
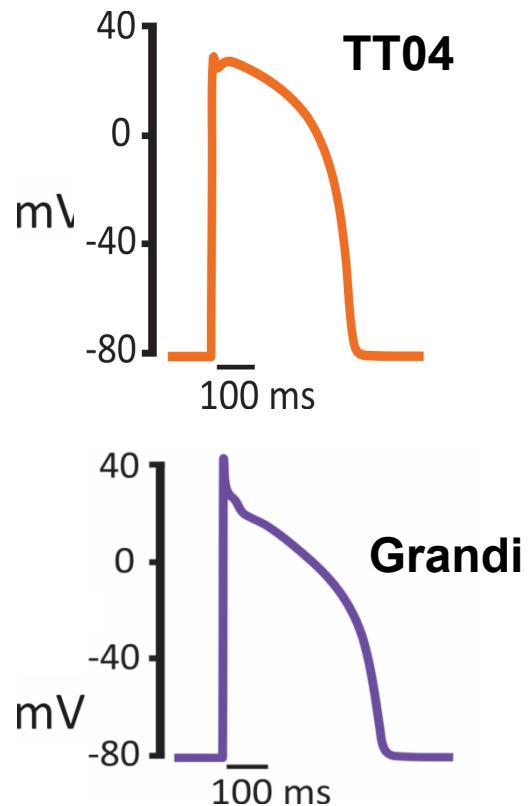


- (6) TT04 et al.
DOI: [10.1152/ajpheart.00794.2003](https://doi.org/10.1152/ajpheart.00794.2003)
- (7) TT06 et al.
DOI: [10.1152/ajpheart.00109.2006](https://doi.org/10.1152/ajpheart.00109.2006)
- (8) Grandi et al.
DOI: [10.1016/j.yjmcc.2009.09.019](https://doi.org/10.1016/j.yjmcc.2009.09.019)
- (9) O'Hara et al.
DOI: [10.1371/journal.pcbi.1002061](https://doi.org/10.1371/journal.pcbi.1002061)

Nine ventricular myocyte mathematical models

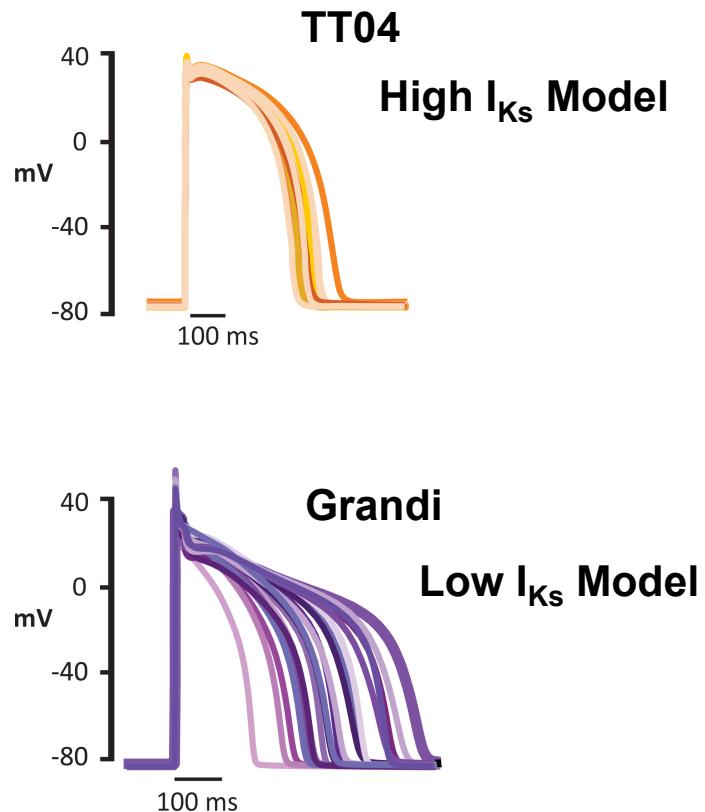
- Endocardial variant of models with multiple layers
- Constant pacing at 1 Hz

Two human ventricular with radically different I_{Ks} levels

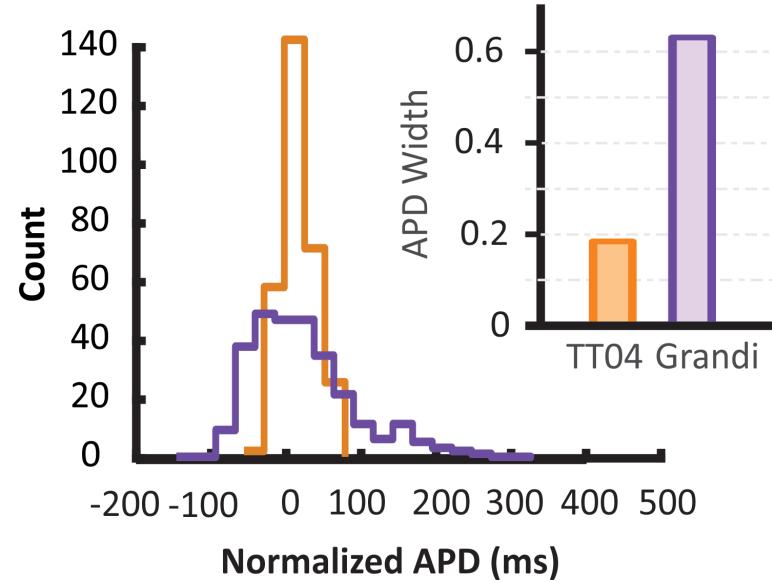


Examine variability in heterogeneous populations

Randomize parameters control ion transfer rates



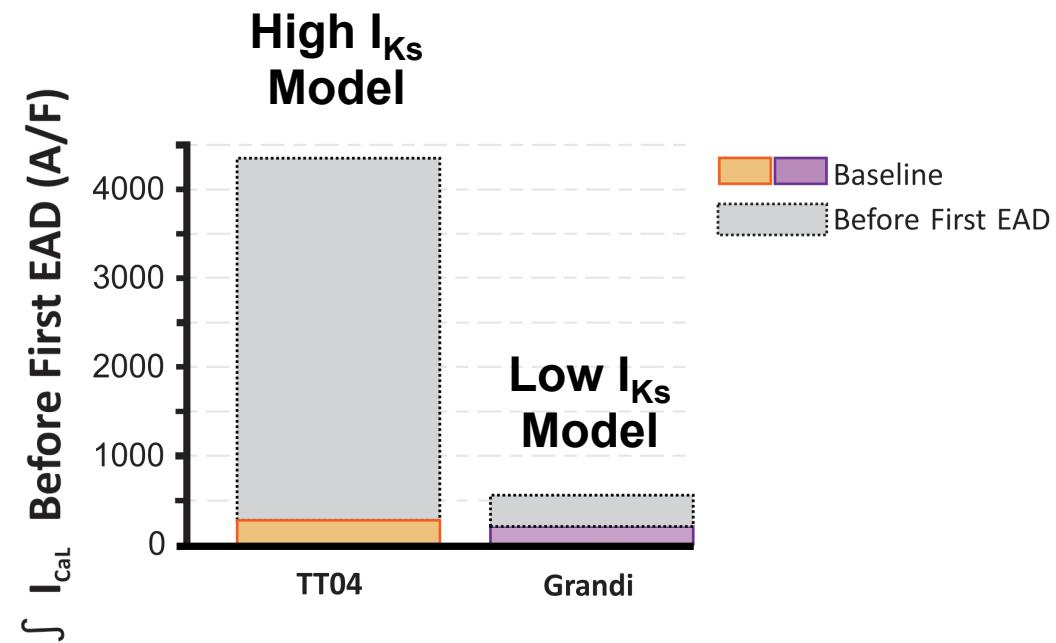
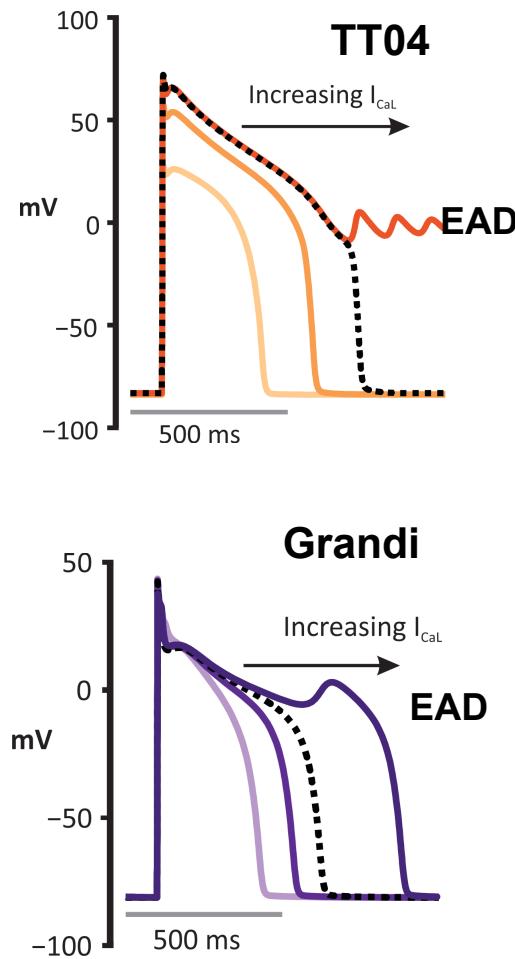
$$\text{APD Spread} = \frac{90^{\text{th}}\text{%ile} - 10^{\text{th}}\text{%ile}}{\text{median}}$$



High I_{Ks} model has less population variability

Test susceptibility to EADs

Increase inward current through L-type Calcium Channel (I_{CaL})



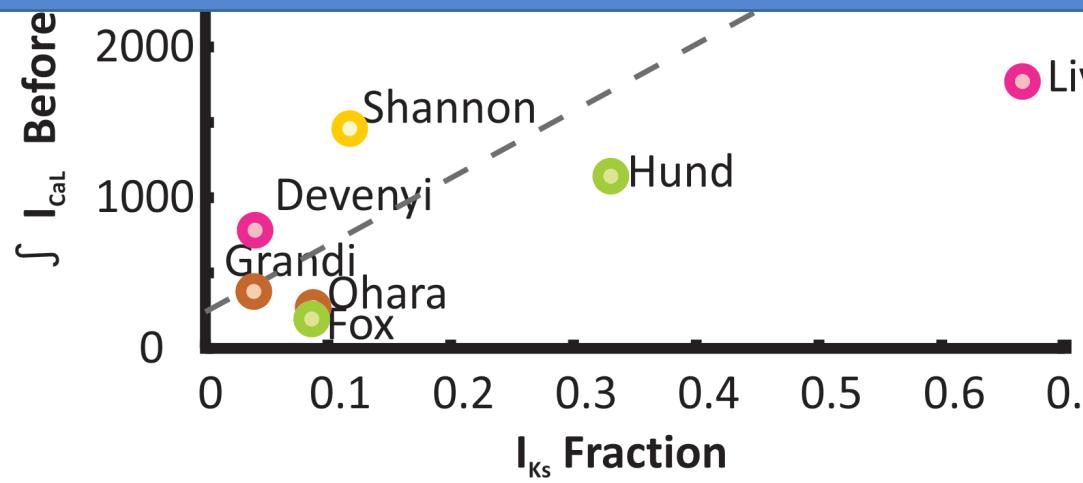
High I_{Ks} model is less susceptible to EADs

Study concept across multiple models

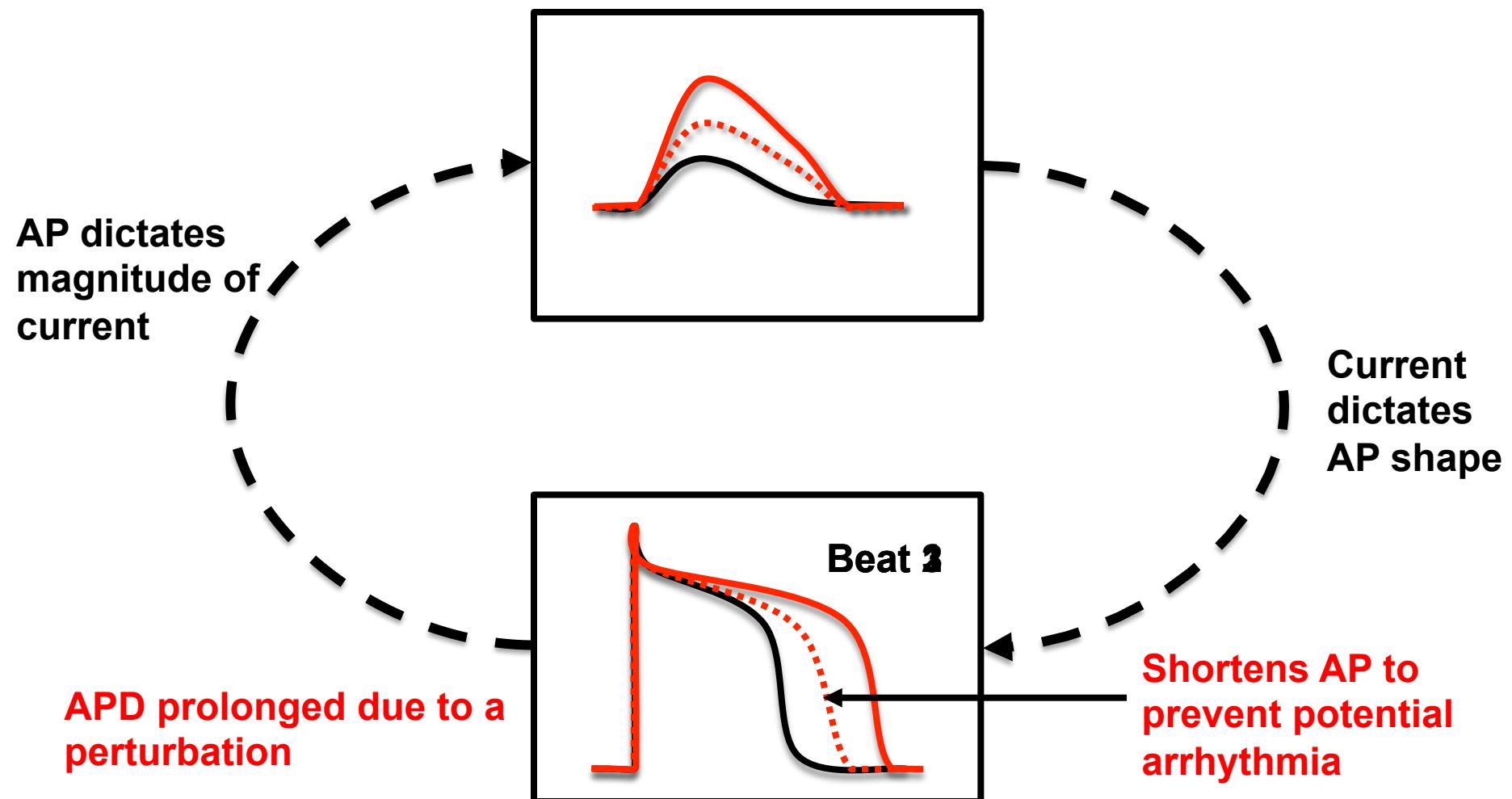
Examining variability with respect to a population



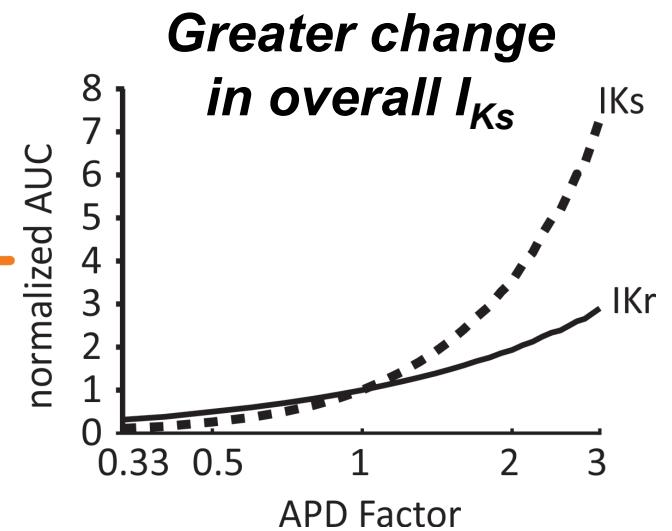
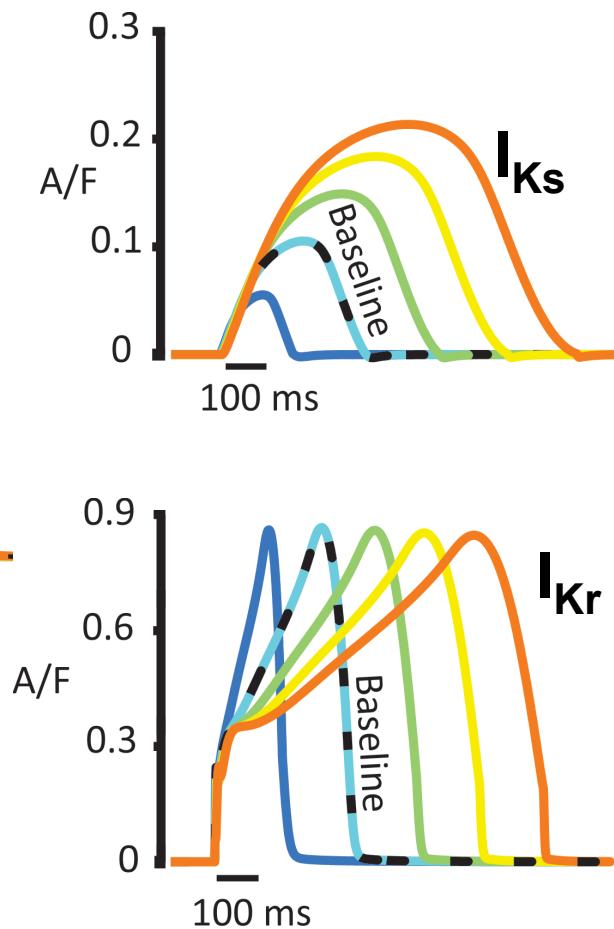
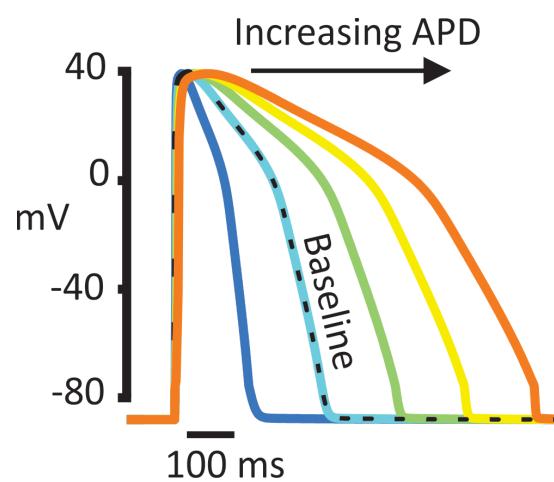
Same pattern across multiple species: Cells with higher I_{Ks} stabilize the action potential making them less susceptible to arrhythmic behavior



Cardiac currents & Negative Feedback

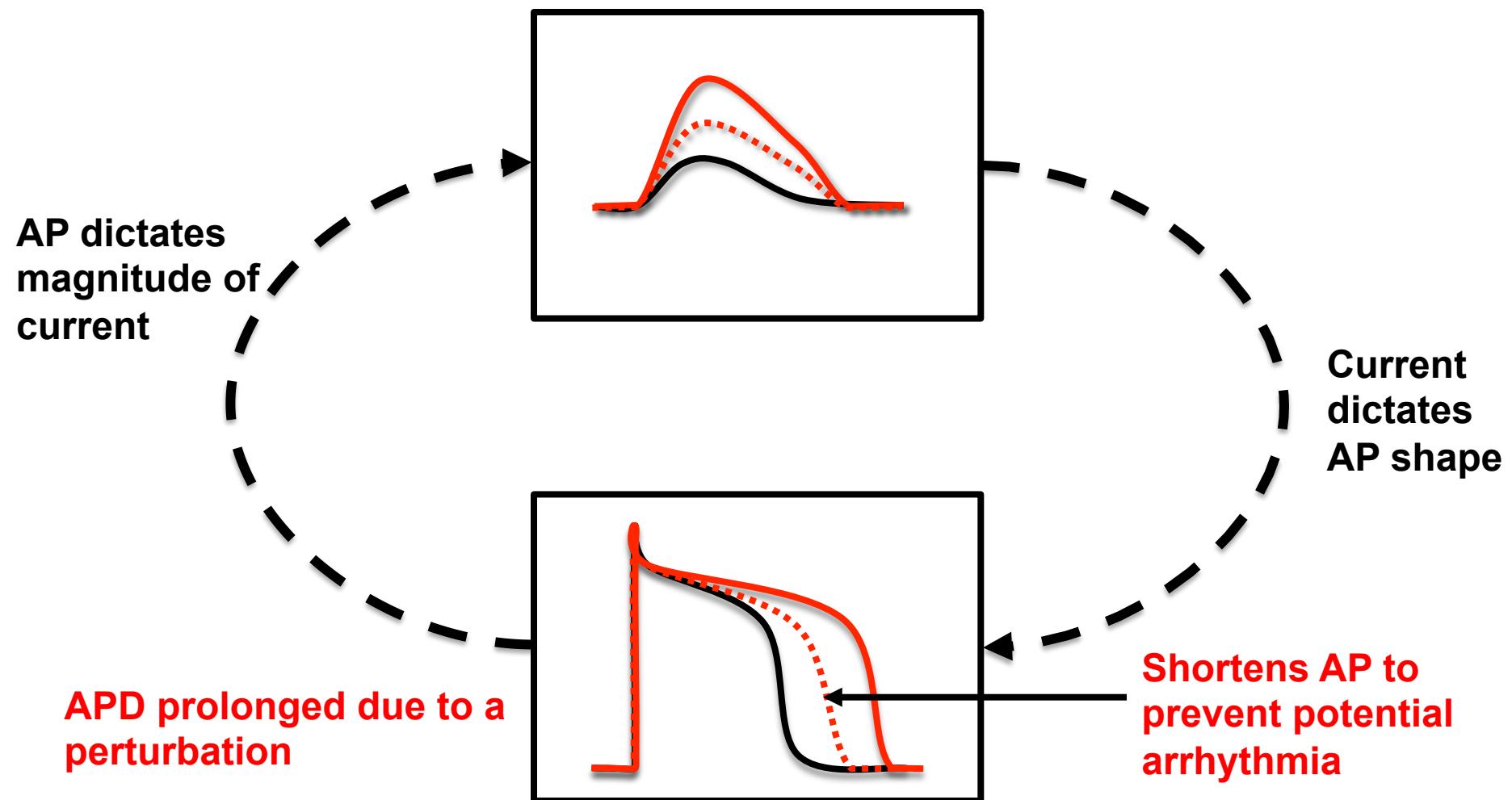


AP Clamp Simulation Results



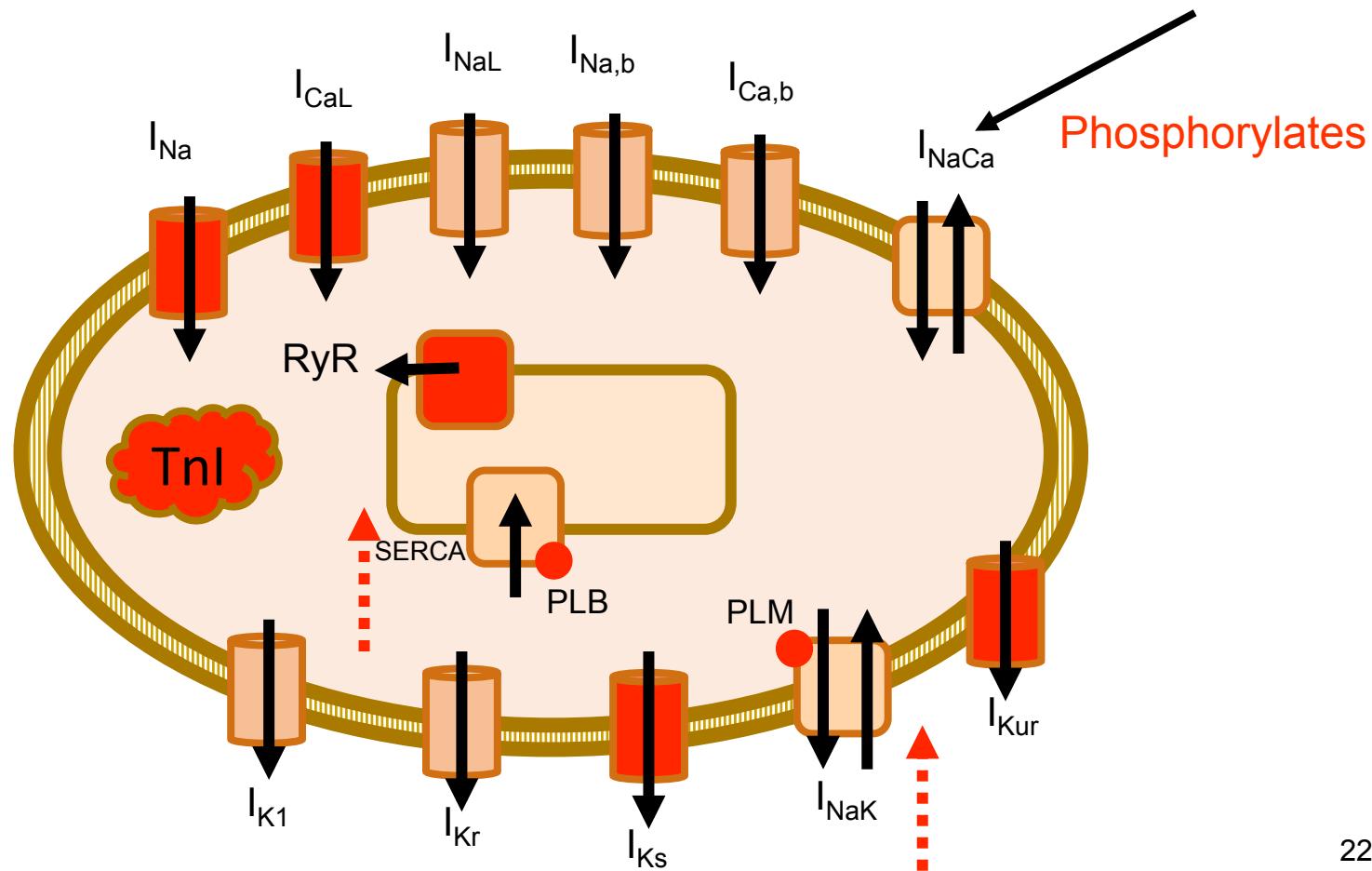
Mechanistically, there exists a more powerful negative feedback for I_{Ks} .

Cardiac currents & Negative Feedback



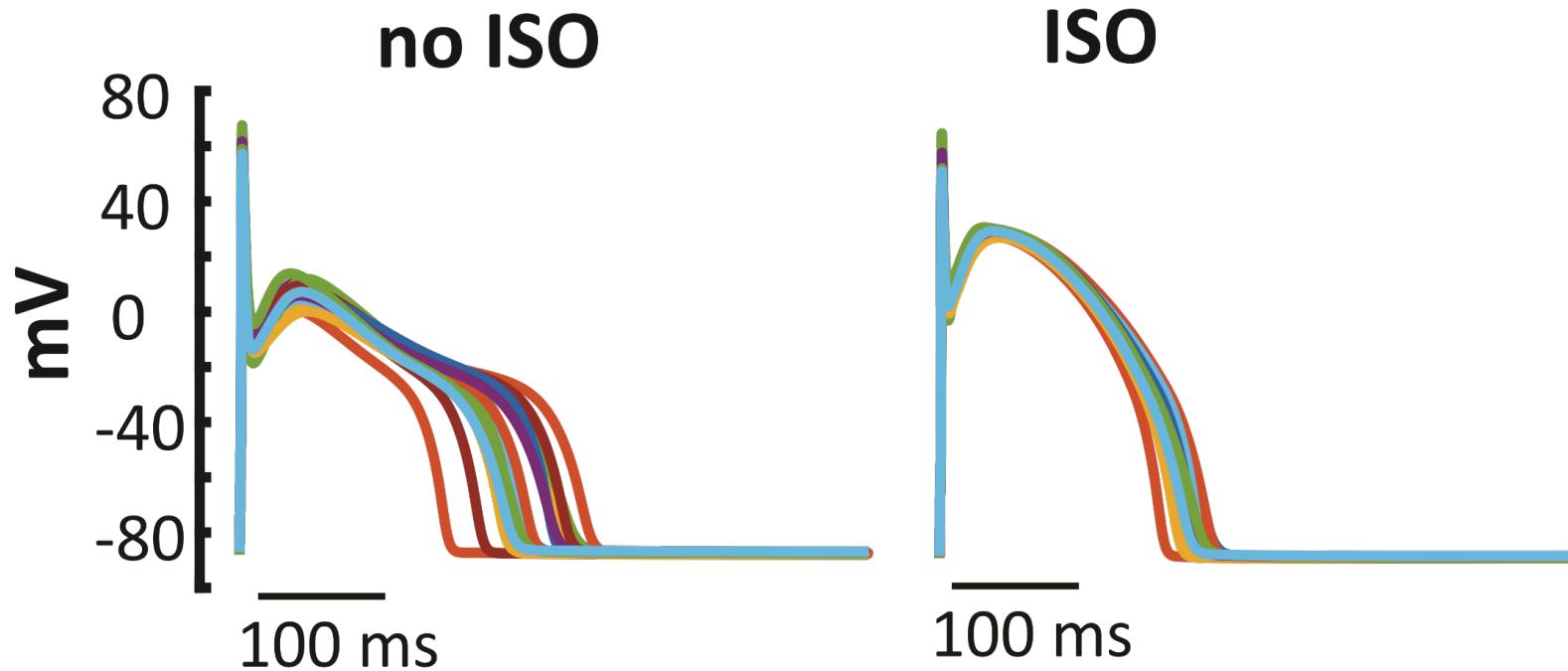
Is I_{Ks} upregulation during β -adrenergic activation important for arrhythmia suppression?

Sympathetic Nervous System Activation → β -adrenergic signaling pathway activated → Protein Kinase A (PKA) activated



Heterogeneous population variability

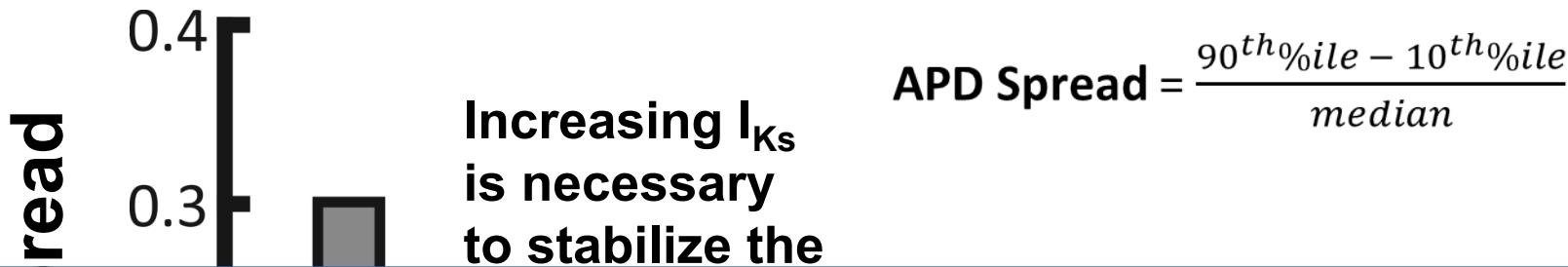
Randomize parameters that control ion transfer rates



Simulated in the Heijman Canine Electrophysiology + Signaling Model
Heijman et al (2011) *J Mol Cell Cardiol.* 50:863-71.

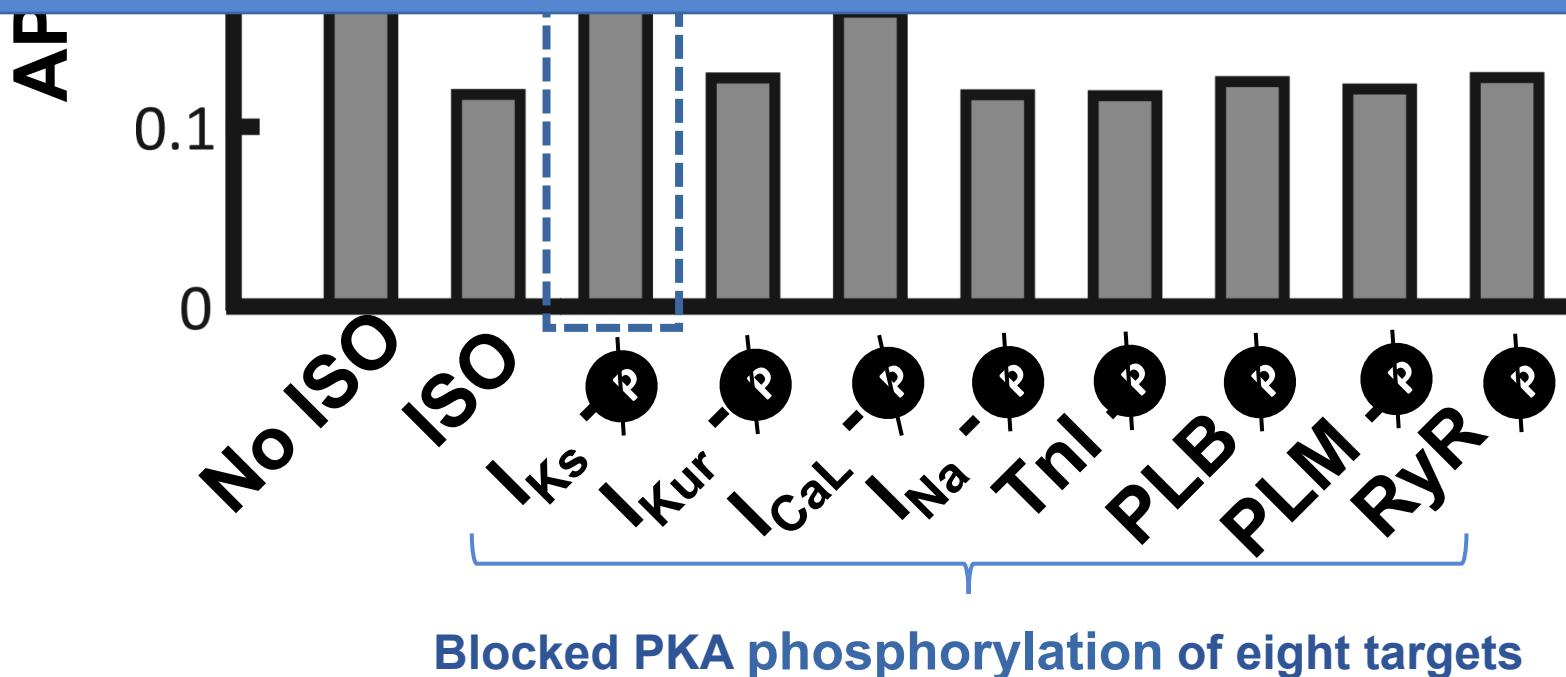
Adding β -agonist Isoproterenol (ISO) shortens the AP

I_{Ks} phosphorylation is necessary to stabilize the AP during β -adrenergic activation.



$$\text{APD Spread} = \frac{90^{\text{th}}\text{percentile} - 10^{\text{th}}\text{percentile}}{\text{median}}$$

Physiologically, I_{Ks} is important to maintain cell stability during repolarization.



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Current lab members:

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Jingqi Gong
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Tobias Holden
DeAnalisa Jones
Jaehee Shim
Meera Varshneya

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Ryan Devenyi
Megan Lancaster
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