

Does experimental evolution  
tell us anything useful?

KITP 2017

# Ask questions!



<http://catalog.fborfw.com/strips/98/FB112398.gif>

# And since time expired...

Yeast can evolutionarily learn to predict...

High salt induces arrest

Arrest prevents lethal DNA replication

Two ways of letting more  $\text{Na}^+$  in

Kill inducers of  $\text{K}^+$  entry ( $\text{K}^+/\text{Na}^+$  balance critical)

Alter sugar transporters to admit  $\text{Na}^+$



Nichole Wespe

# OUTLINE

Time travel (fossils and DNA sequence) extremely limited

Experimental evolution should be asking specific questions

Question 1: How does evolutionary novelty appear?

Question 2: How do cells adapt to the loss of beloved proteins?

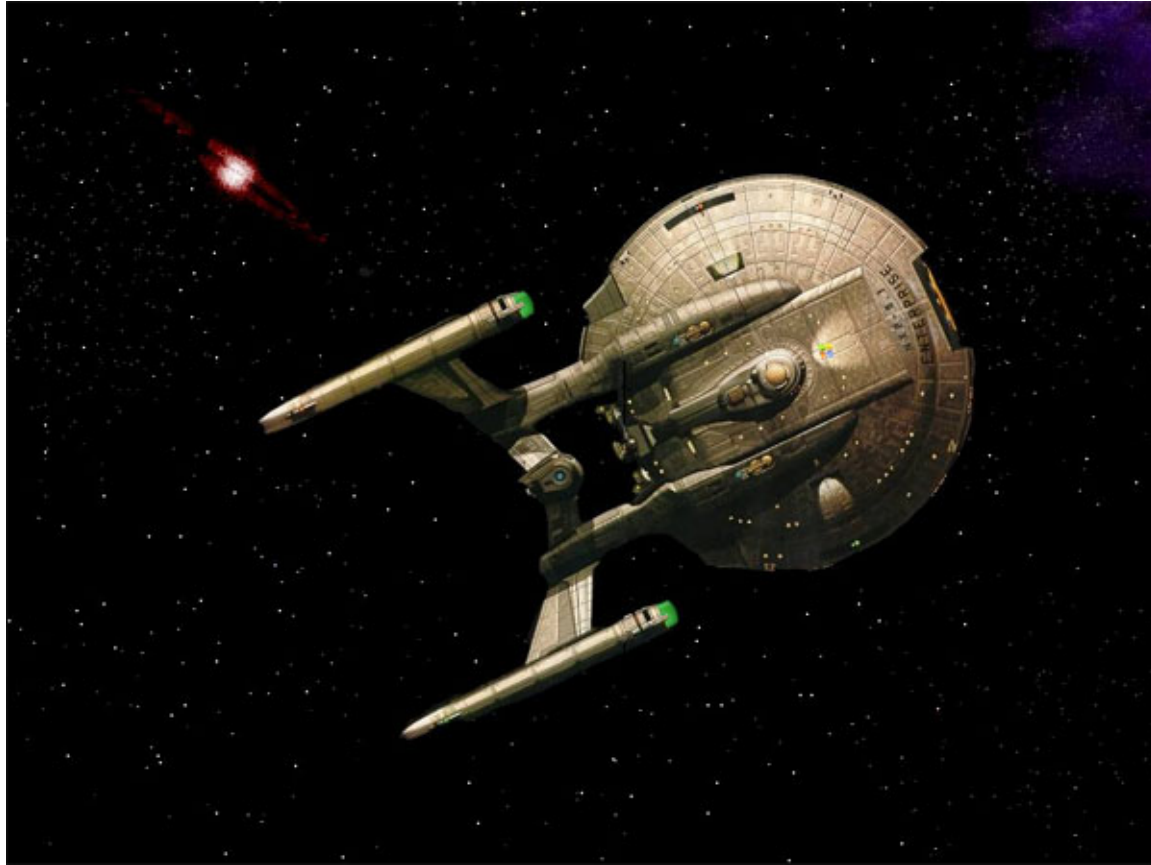
Question 3: Can evolution mimic Pavlov?



# No non-theological purpose



But organisms are...



...vehicles to disperse genes through time and space

# Yes, evolution happens, Dorothy

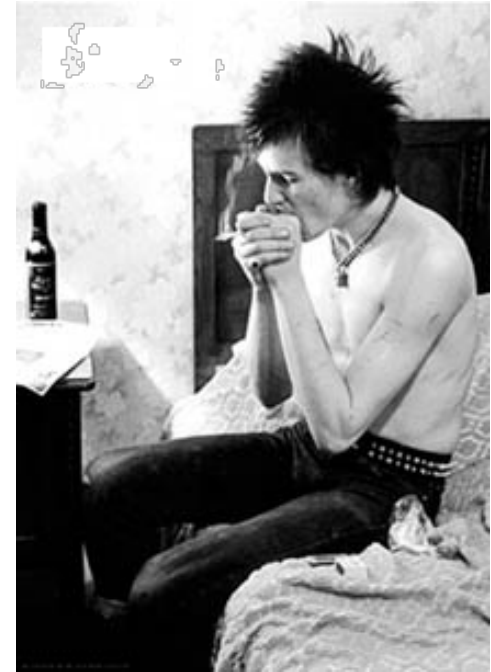


**Normal Cell**

Careful with chromosomes



At least 5  
mutations



**Cancer Cell**

Careless with chromosomes

# Evolution

Depends on

Inheritance: offspring like parents

Mutation: genetic variation constantly generated

Selection: some genotypes leave more progeny

We have trouble understanding it because

Evolution is dominated by successions of *very rare* events

Relative probability determines evolutionary path

Historical inference from fossils & DNA imperfect

# What we're missing



# Experimental versus “real” evolution

## Advantages

- Starting point known

- Selective pressure designed (not known!)

- Can keep “living fossil” record

- Ancestral and evolved can interbreed

- Multiple, parallel experiments possible

## Disadvantages

- Time and population size limited

- Natural environments are much, much more complex

- Unknown relevance to long term, natural evolution



# What is novelty and how do you get it?

Acquiring a qualitatively new, fitness-increasing property



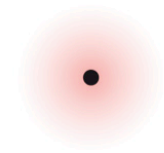
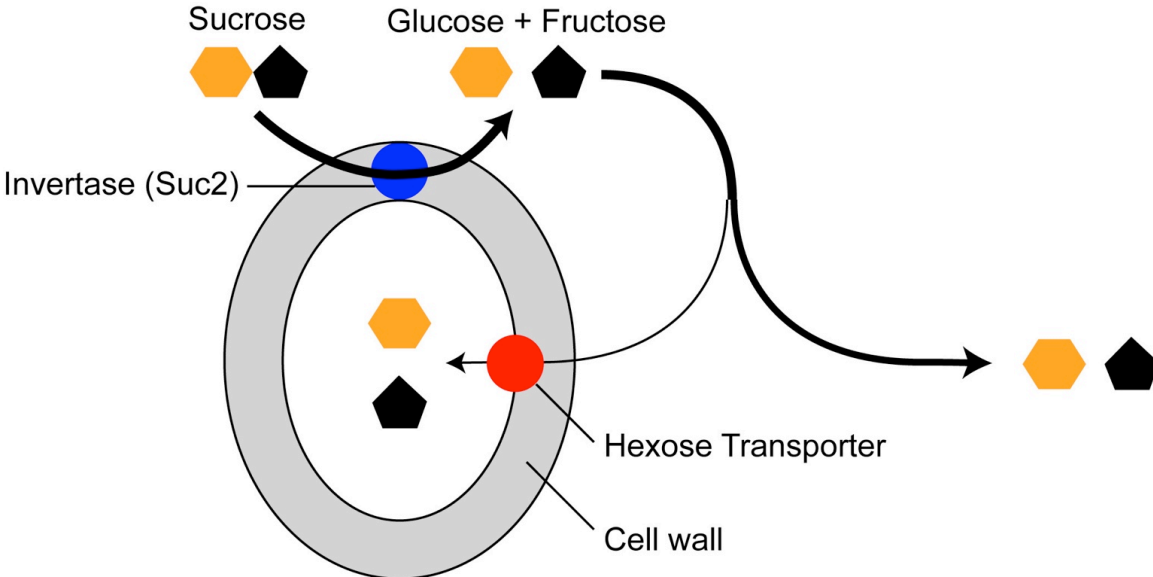
# Does nutrient capture select for multicellularity?



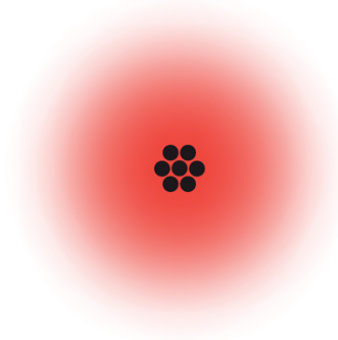
John Koschwanez



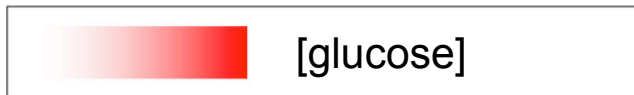
# Why be multicellular? To utilize public goods?



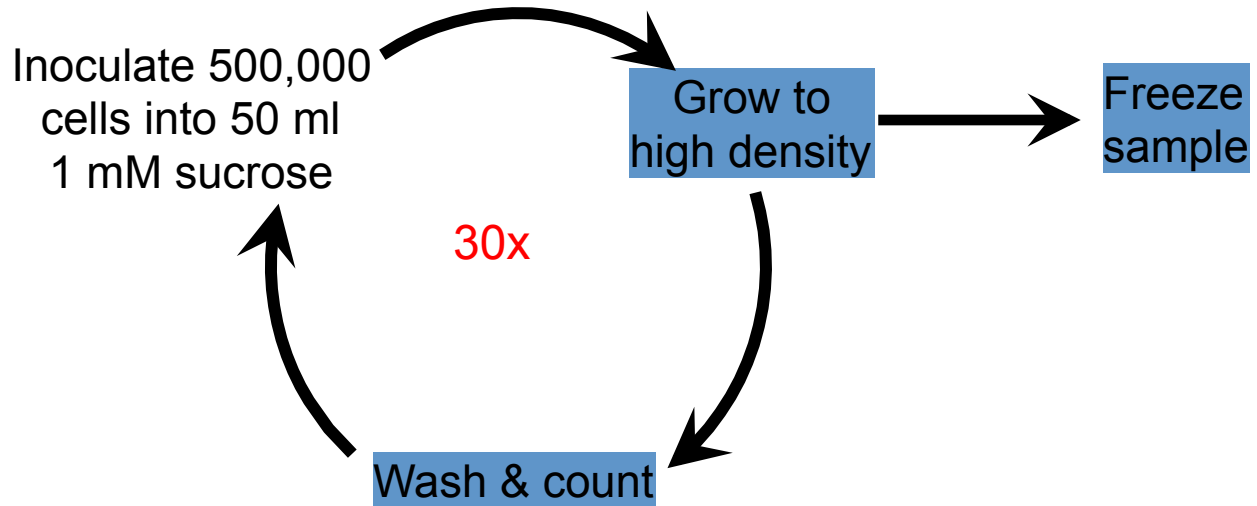
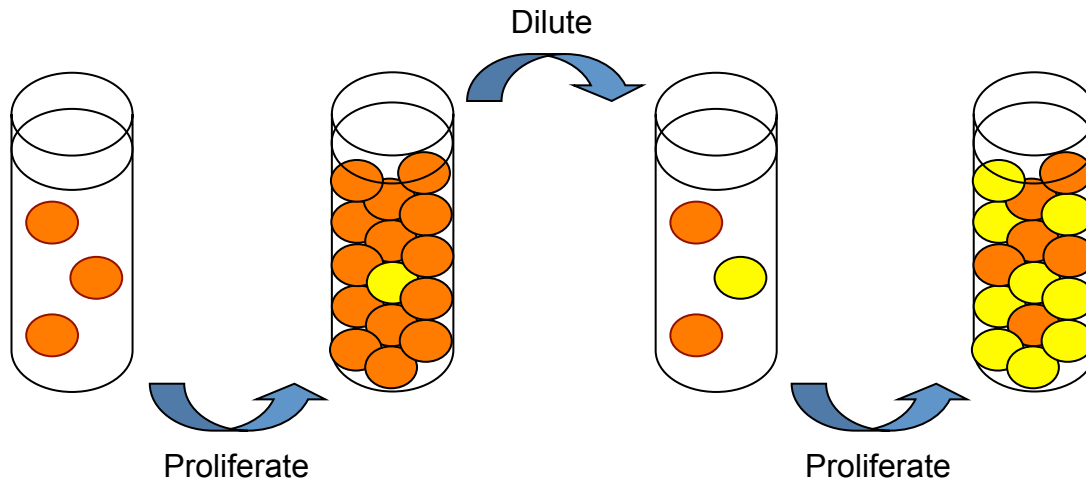
Single cell



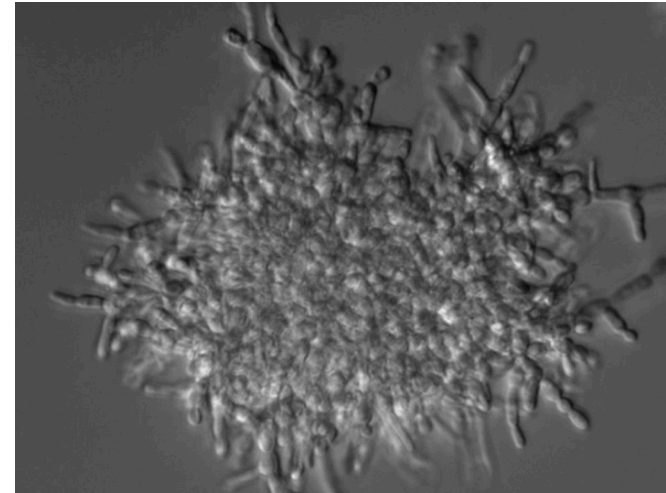
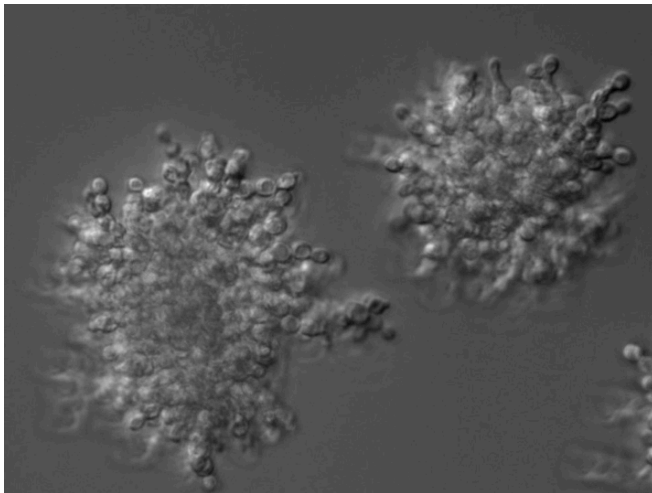
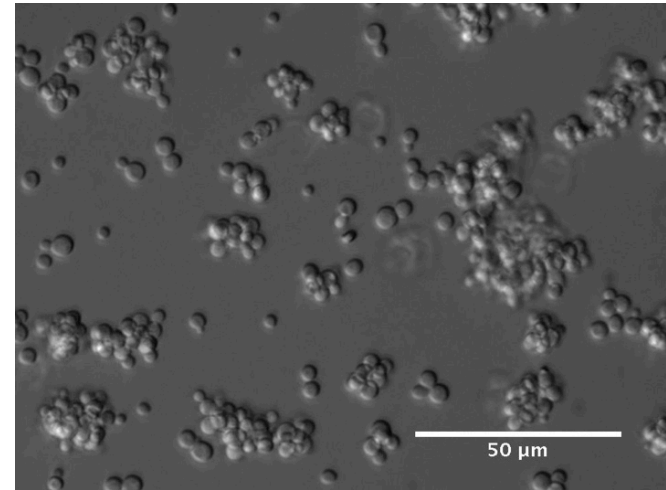
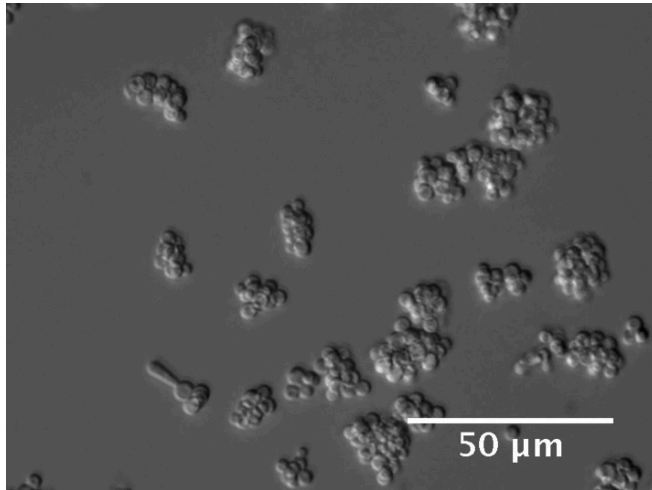
Cell clump



# WWED? :Evolving multicellularity



# Experimentally evolved multicellularity



# Who's mutated?

Times mutated	# Genes	Names
7	1	<i>ACE2</i>
6	1	<i>UBR1</i>
3	2	<i>RGT1, SNF3,</i>
2	11	<i>IRA1, IRA2, etc</i>
1	39	<i>many genes!</i>

80 mutations: 7 promoter, 19 stop, 12 indel, 42 missense

**Most (perhaps all) missense mutations are loss of function**

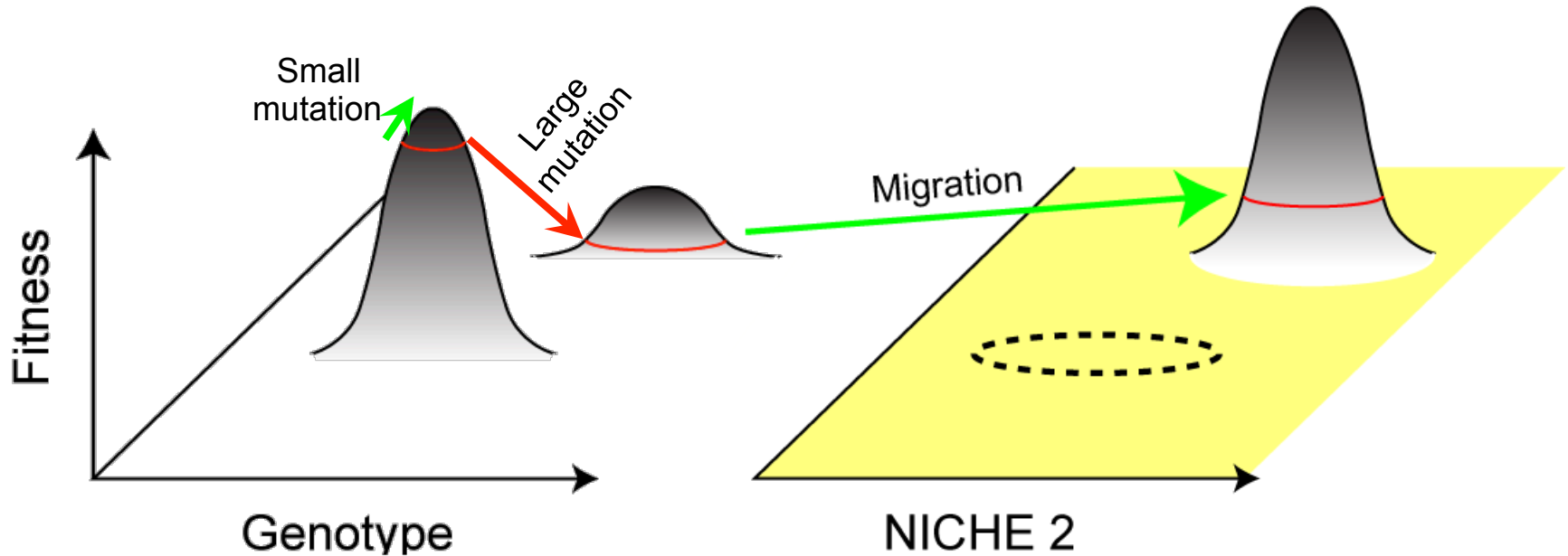
3 frequently mutated pathways

cAMP signaling (altered go/no go gambling setpoint)

catabolite repression (reduced glucose addiction)

mediator complex (complicated, confusing transcription control)

# A model for novelty



In a crowded ecosystem

Niche 2 occupied

Mutant outcompeted

Mutant = hopeless monster

In a virgin ecosystem

Niche 2 empty

Mutant survives if  $w_{abs} > 1$

Mutant = hopeful monster

# Evolving to live without important genes



Liedewij Laan (TU Delft)

# A seriously compromising mutation



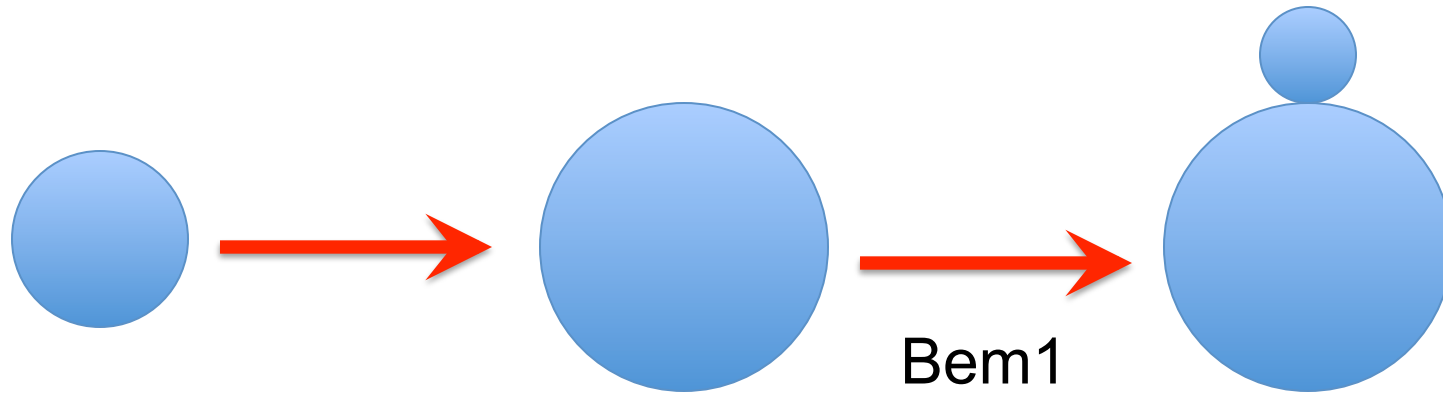


If cars evolved

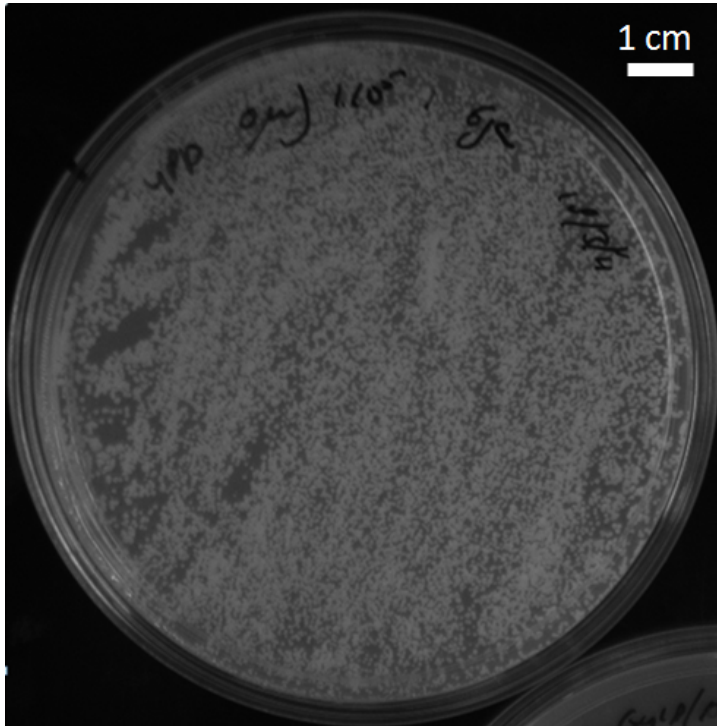




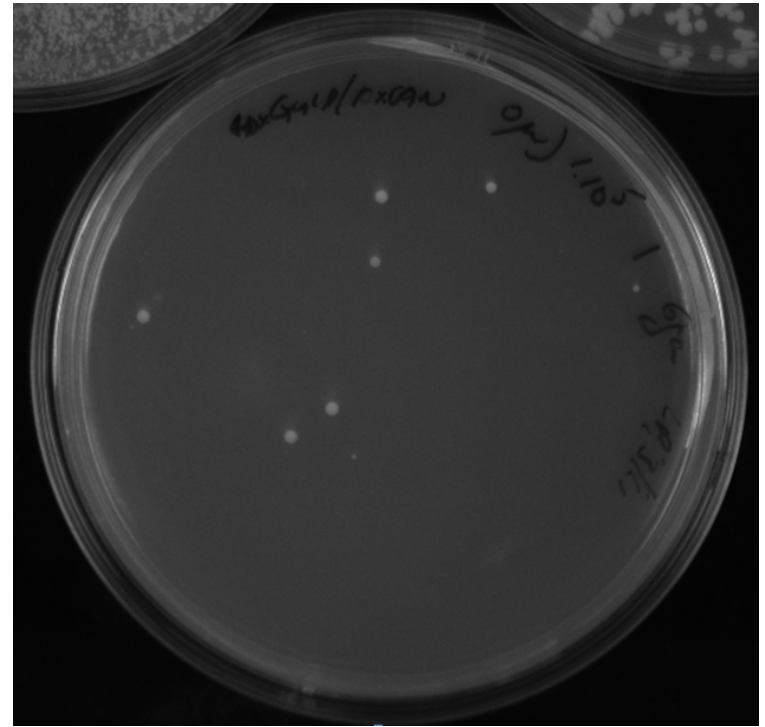
# Yeast breaks symmetry to proliferate



# Removing Bem1 is very bad

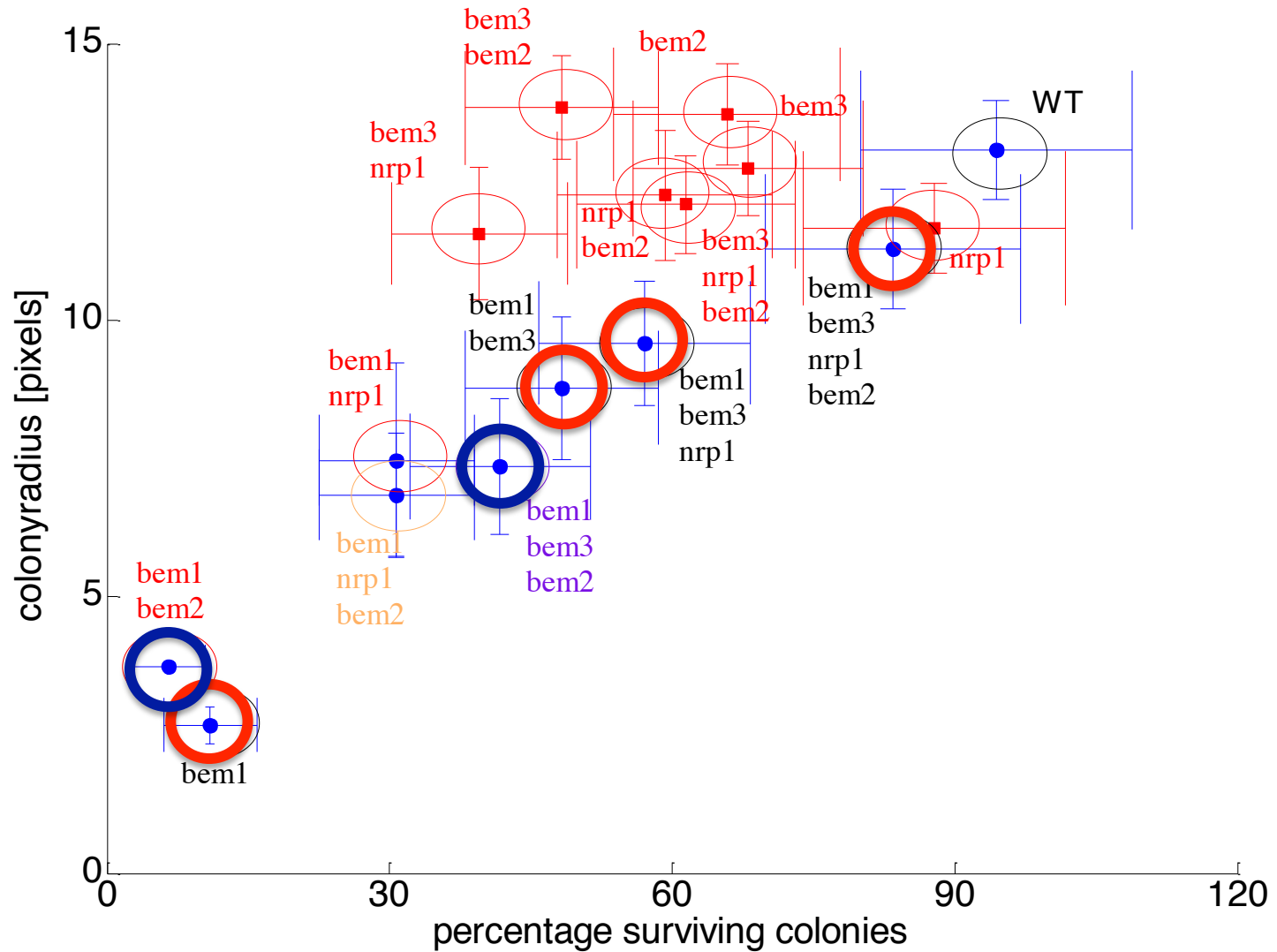


*BEM1*



*bem1*Δ

# Looking at all combinations explains order of mutations



# Previous selections

Multiple, poorly related trajectories

Novelty: Multicellularity (eLife '13)

Novelty: Circadian oscillator (eLife '14)

Single consistent trajectory

Loss important protein: (Bem1) (eLife '15)

**Loss of function mutations dominate**

Are these straws in the wind?

Repair trajectories more reproducible?

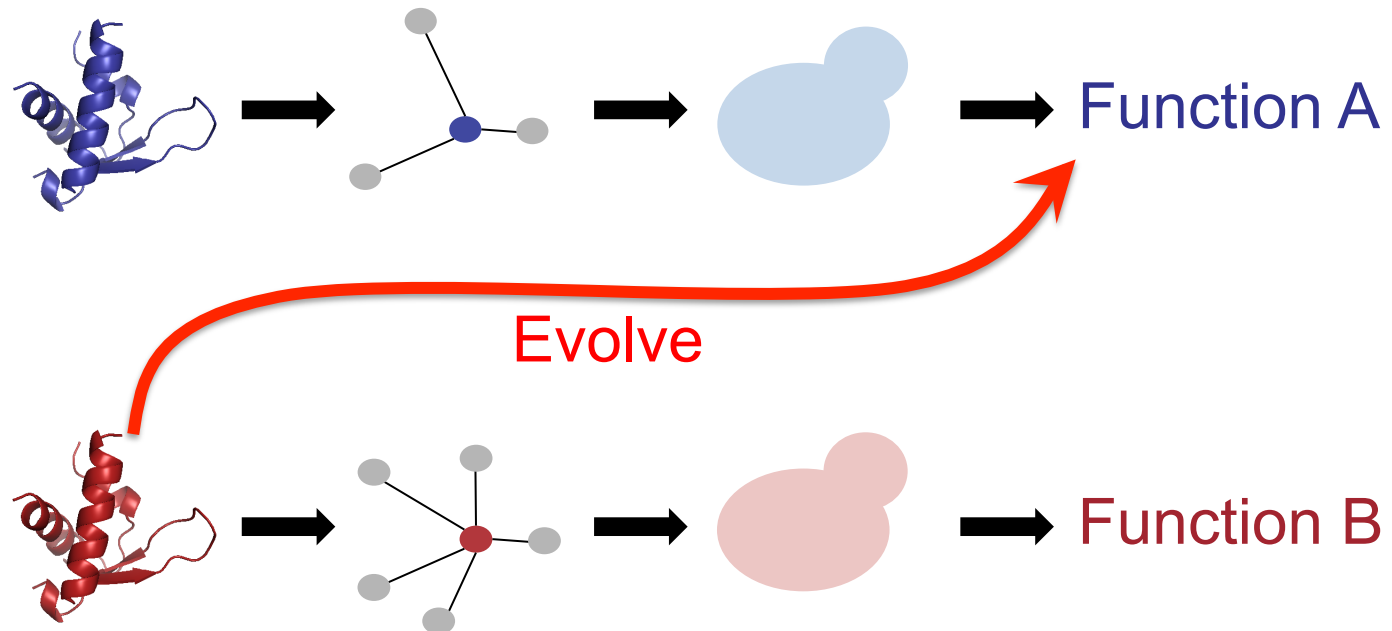
Loss of function dominates causal mutations?

# Learning to live with the wrong part



Phoebe Hsieh

# Paralogs: functionally specialized proteins



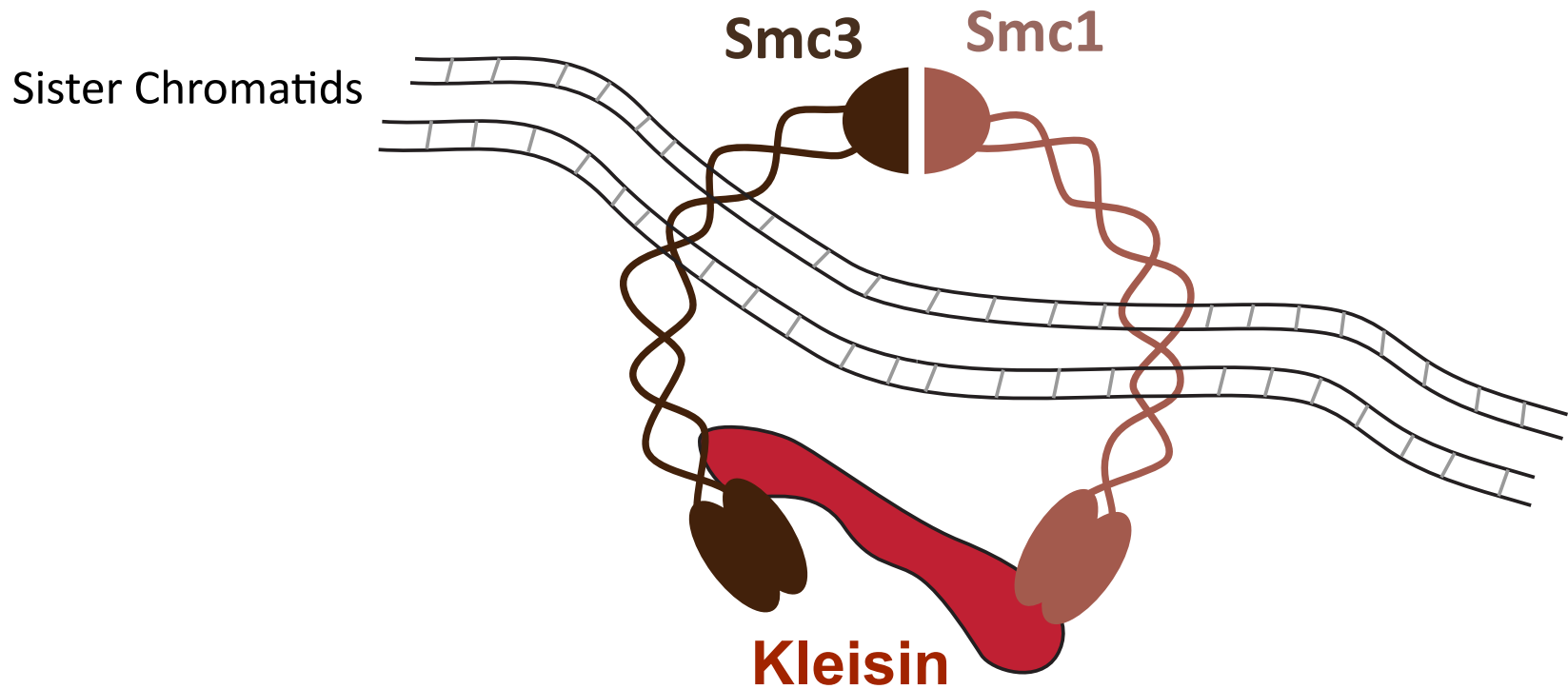
How does evolution substitute red for blue?

Change in protein itself?

Change in usual suspects (its interacting partners)?

Change in novel suspects ?

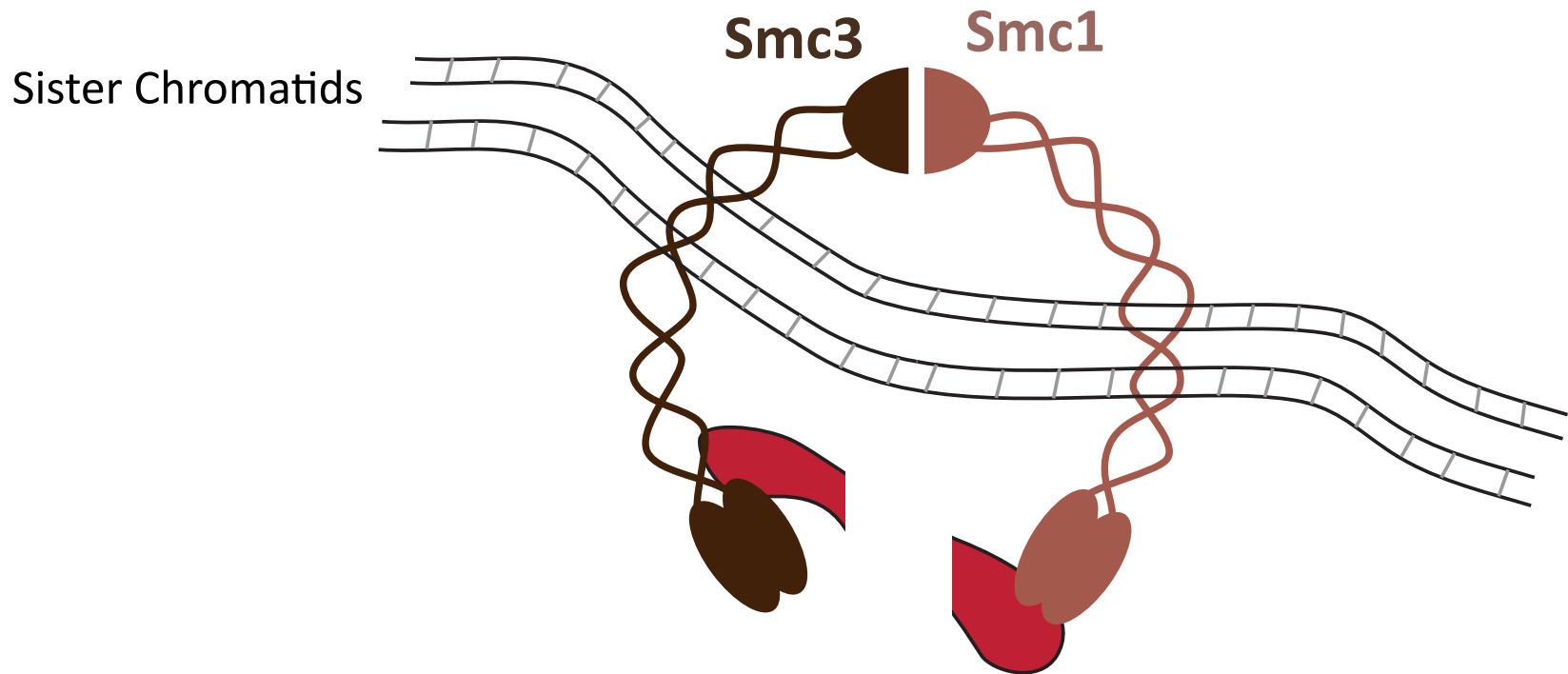
# Kleisin is a subunit of cohesin complex



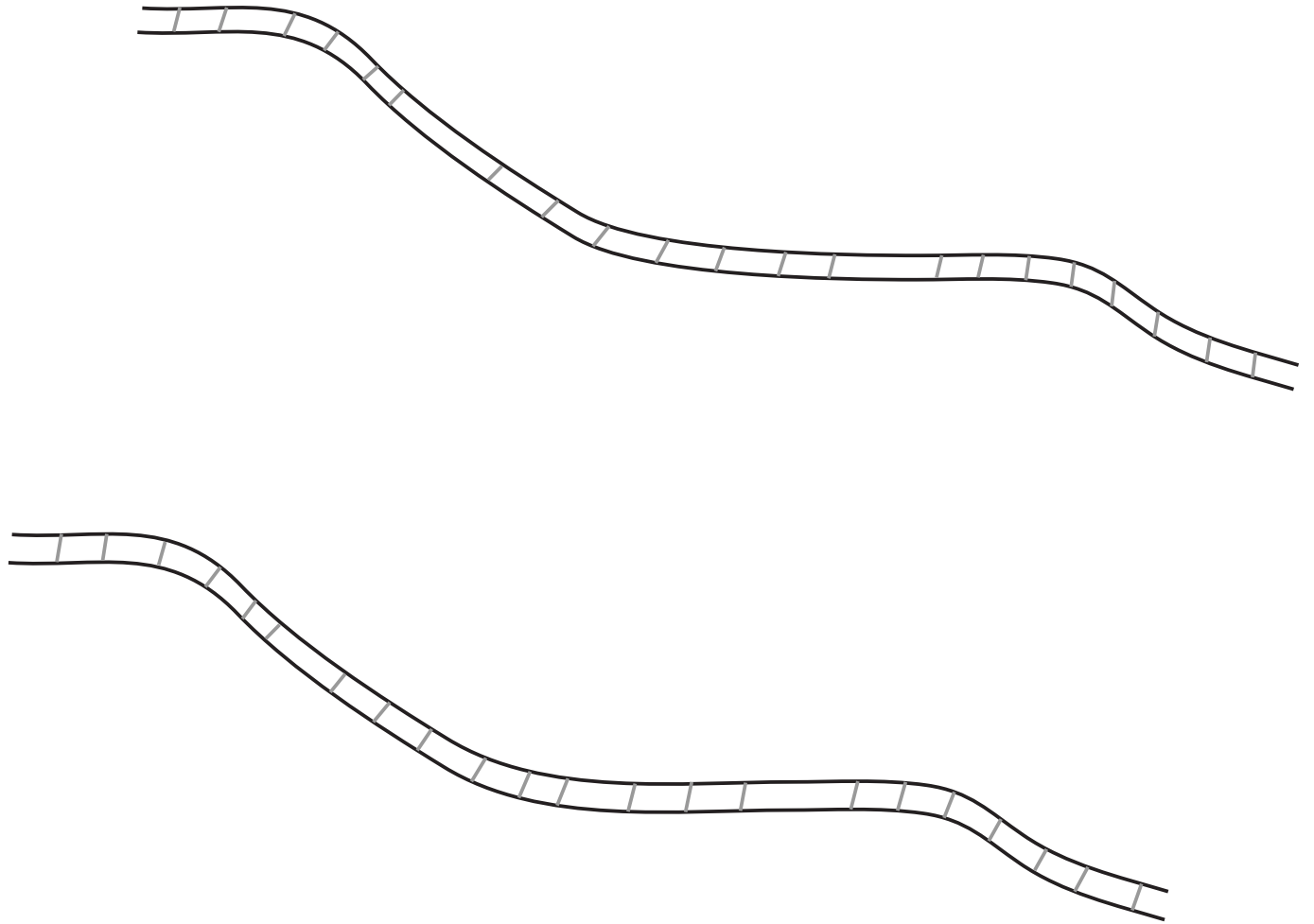
Conserved & essential for eukaryotic chromosome segregation



# Cleaving kleisin separates sister chromosomes

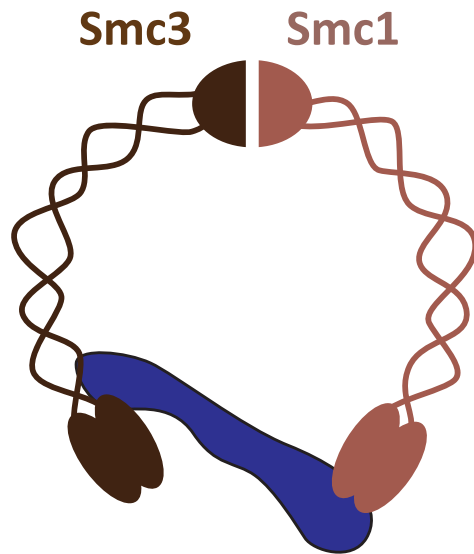


# Cleaving kleisin separates sister chromosomes



# Mitotic and meiotic kleisin paralogs

## Mitotic Cohesin Complex



Mitotic Kleisin: Scc1

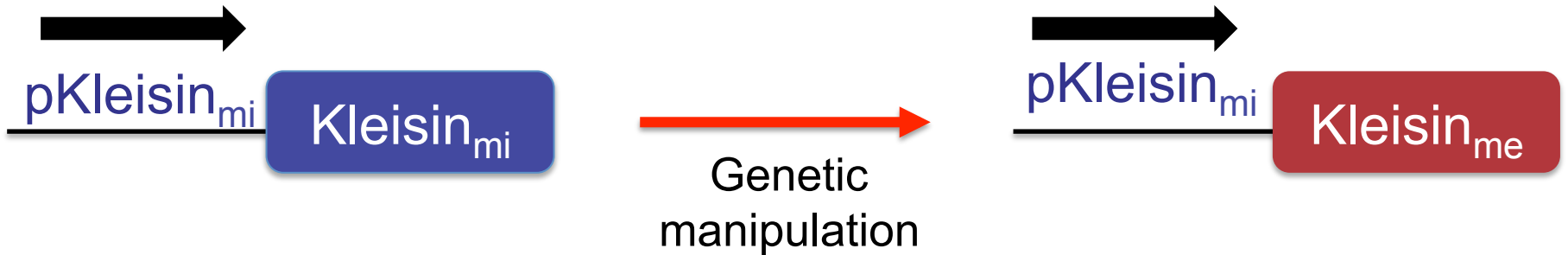
## Meiotic Cohesin Complex



Meiotic Kleisin: Rec8

Proteolysis different to allow different modes of chromosome segregation

# Meiotic kleisin is bad for mitotic cells

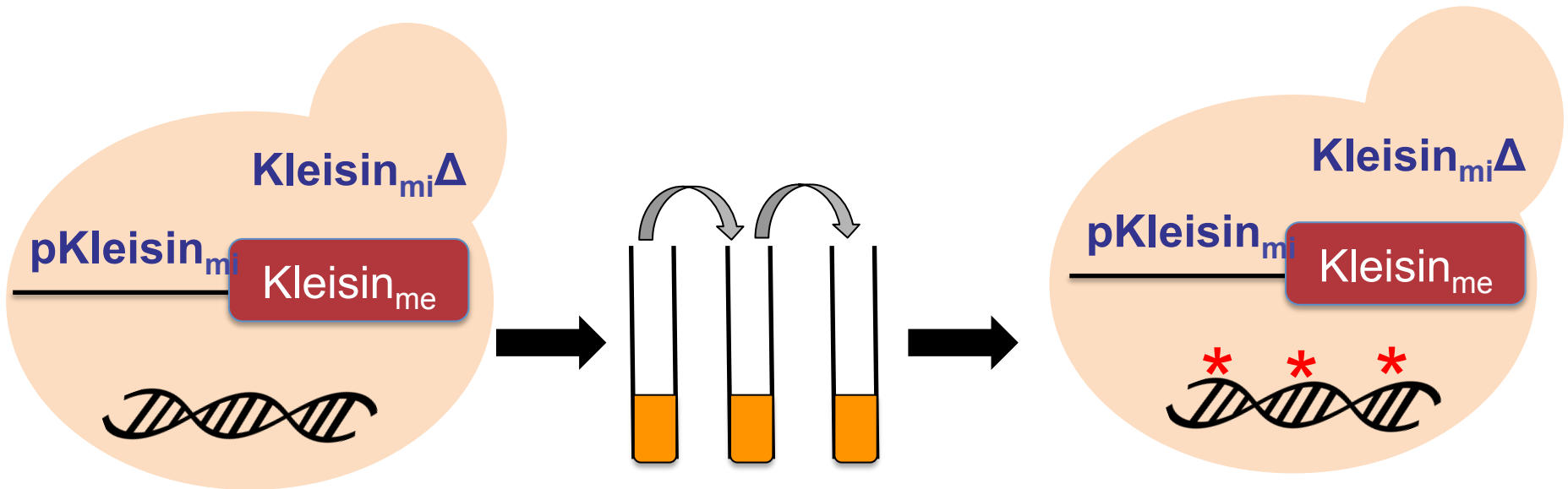


Slow growth

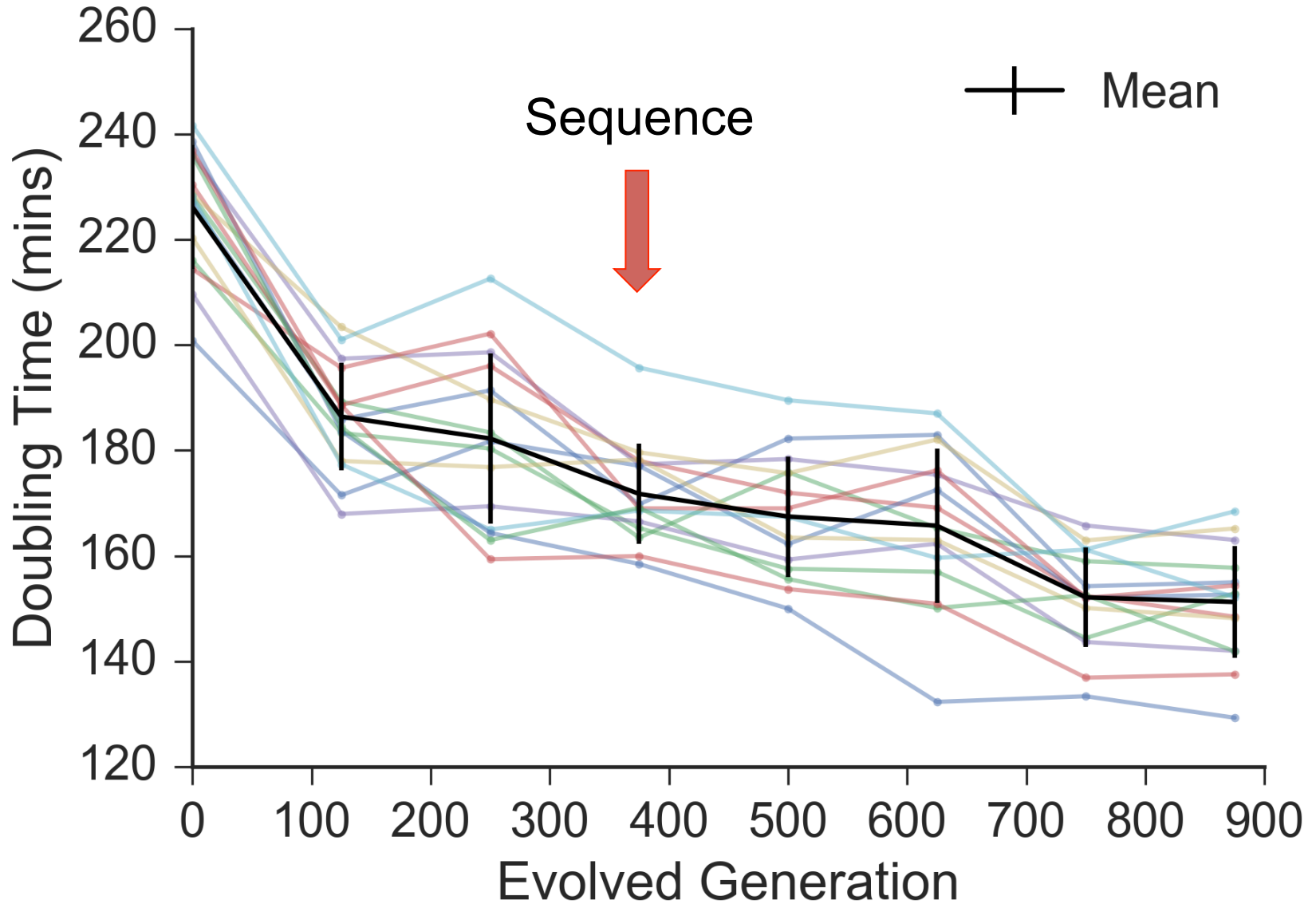
Reduced cohesin binding

Defective sister chromosome cohesion

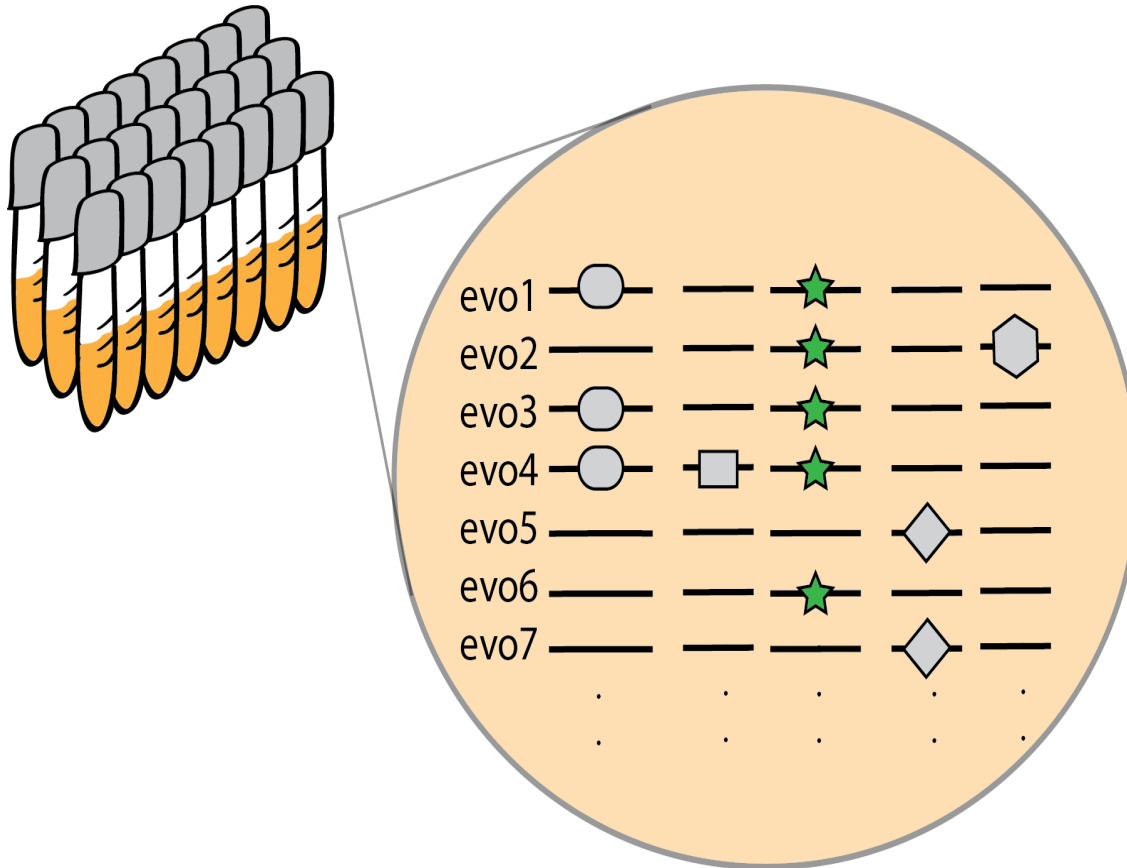
# Evolving cells to live with meiotic kleisin



# All populations evolve faster proliferation



# Sequencing finds putative causative mutations



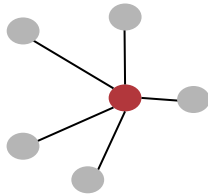
Experiments test their causality!!!

# Who are the putative causal mutations?

## Usual suspects



No mutation in the **Kleisin<sub>me</sub>**



Rare mutations in three kleisin-interacting partners  
Cohesin subunit: *SMC1*, *SMC3*  
Kleisin protease: *ESP1*

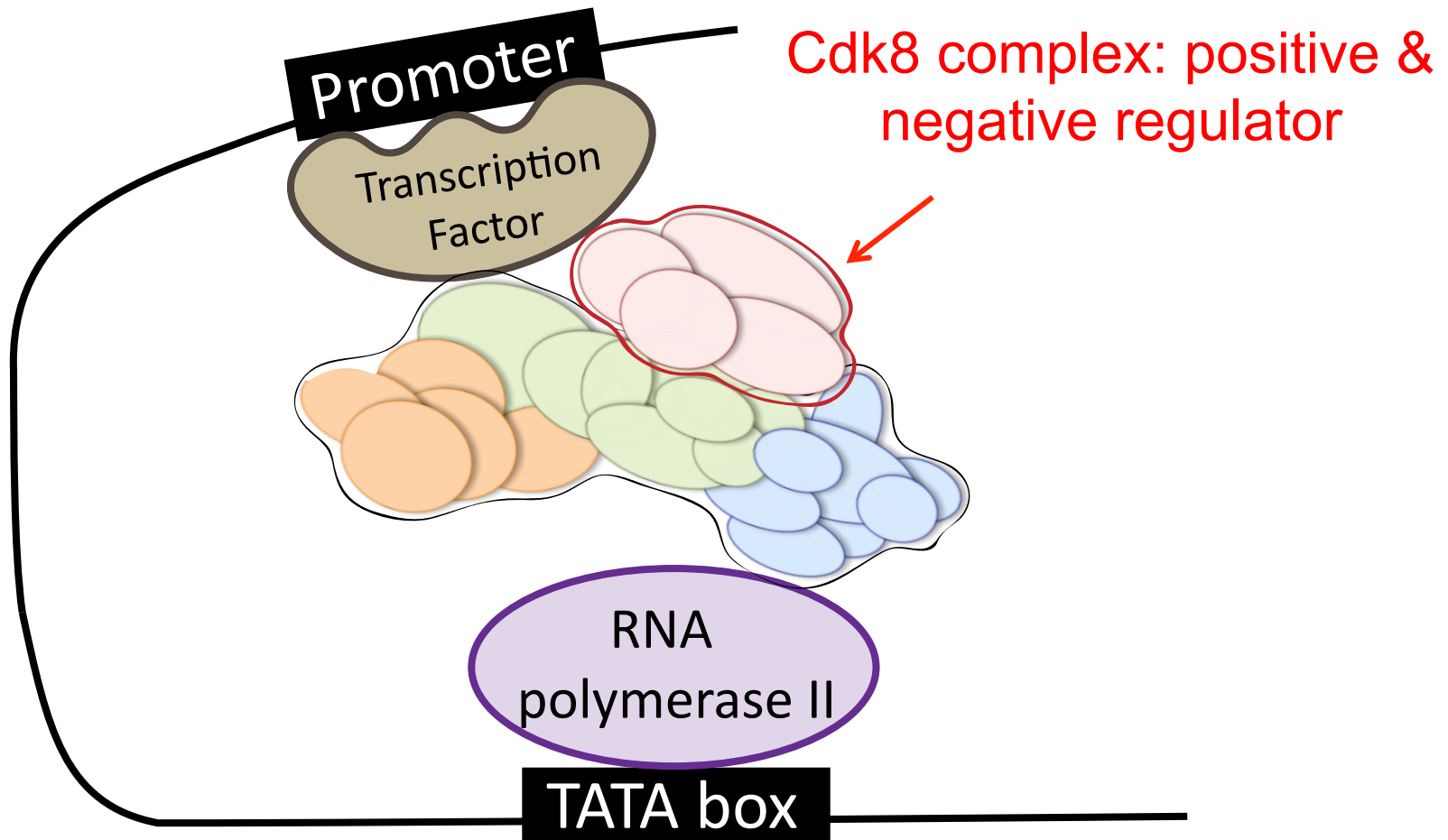
## Novel suspects



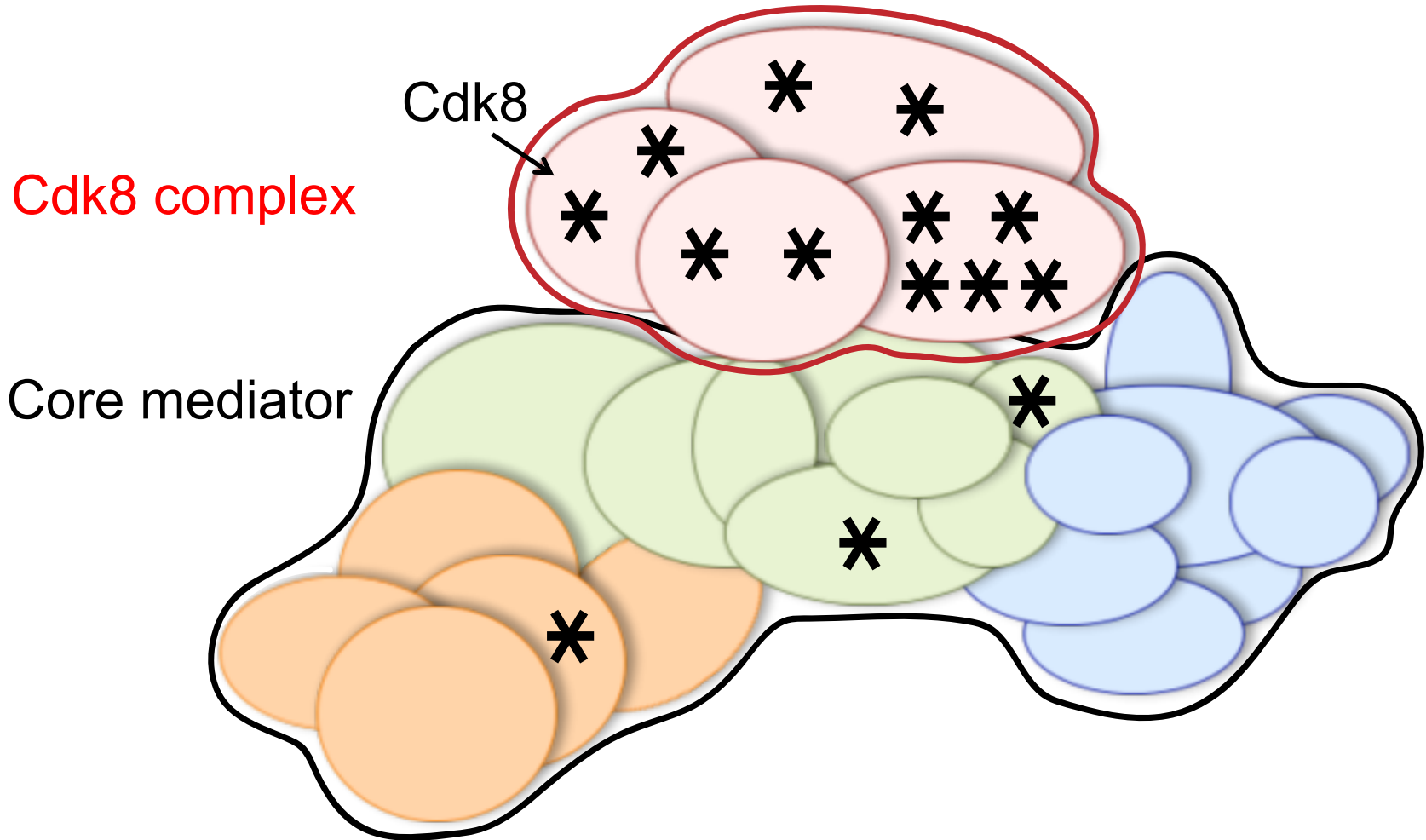
Transcriptional mediator complex



# Mediator links RNA polymerase II and transcription factors

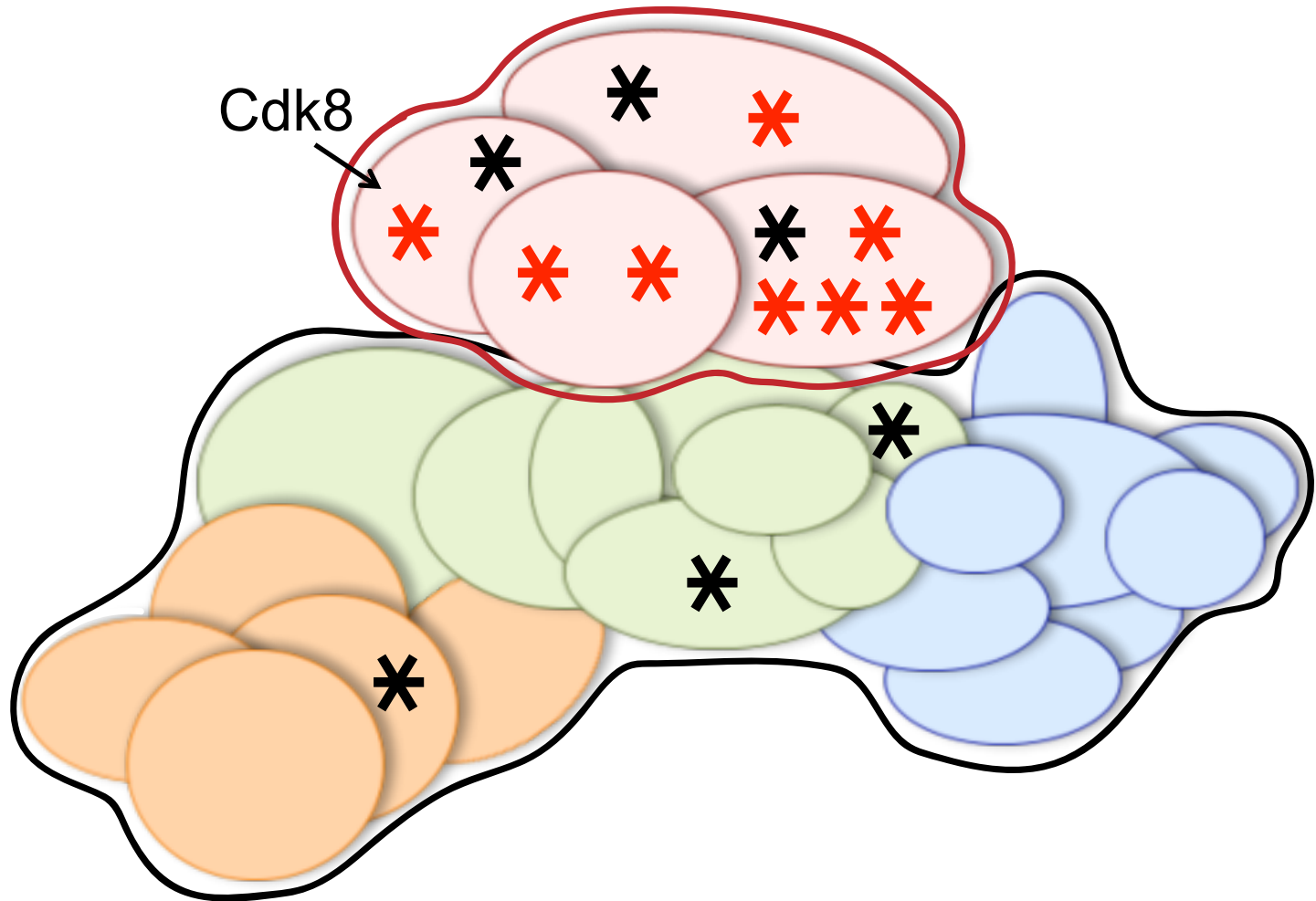


# The Cdk8 complex is highly mutated



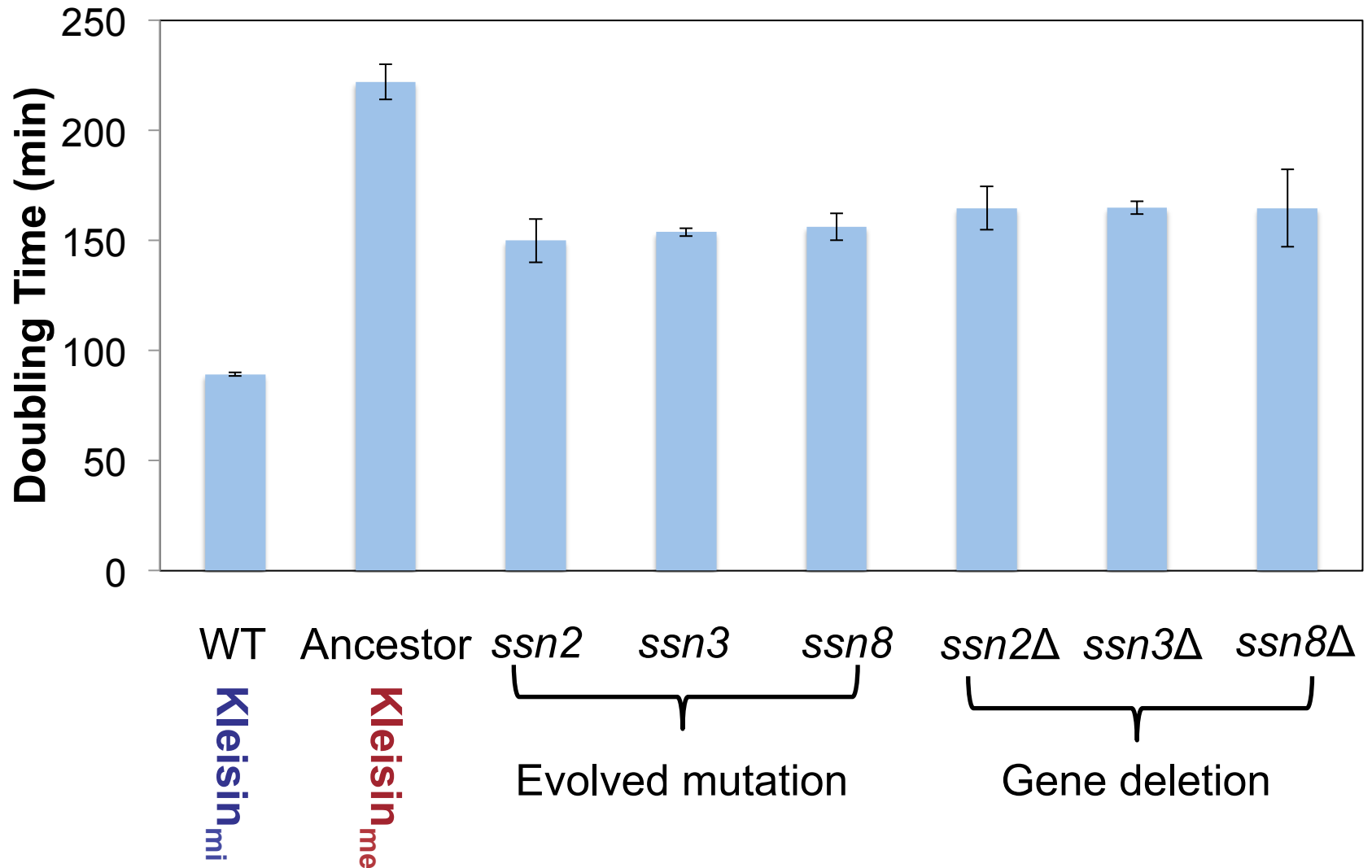
Mediator mutations in 14/15 independent lineages

# The Cdk8 complex is highly mutated



Majority of mutations: **early stop codon**

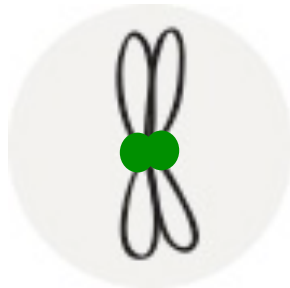
# Cdk8 complex mutations kill proteins



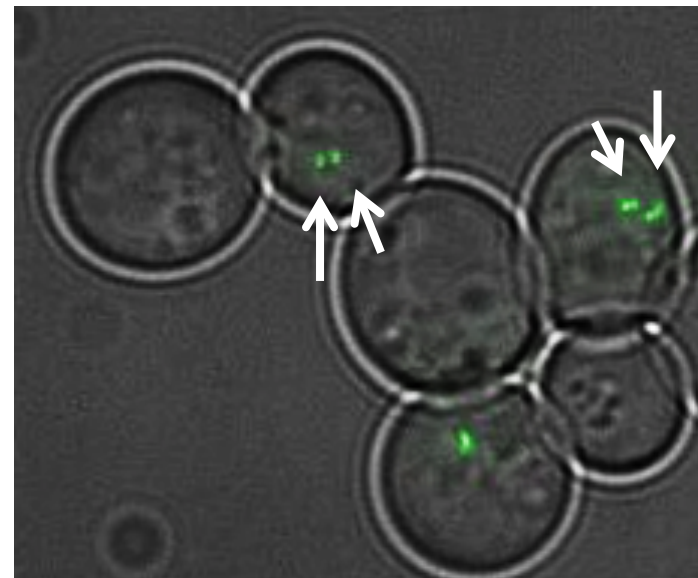
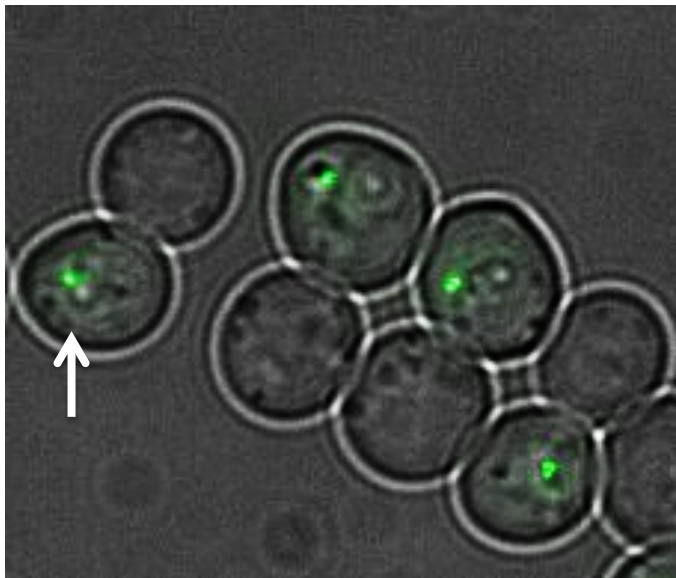
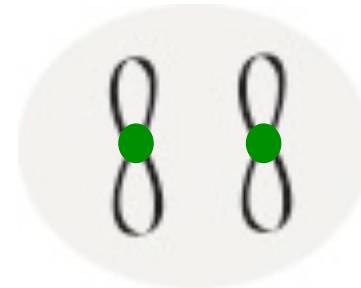
# Does loss of Cdk8 function fix sister cohesion?

The assay: mark one chromosome with little green dots

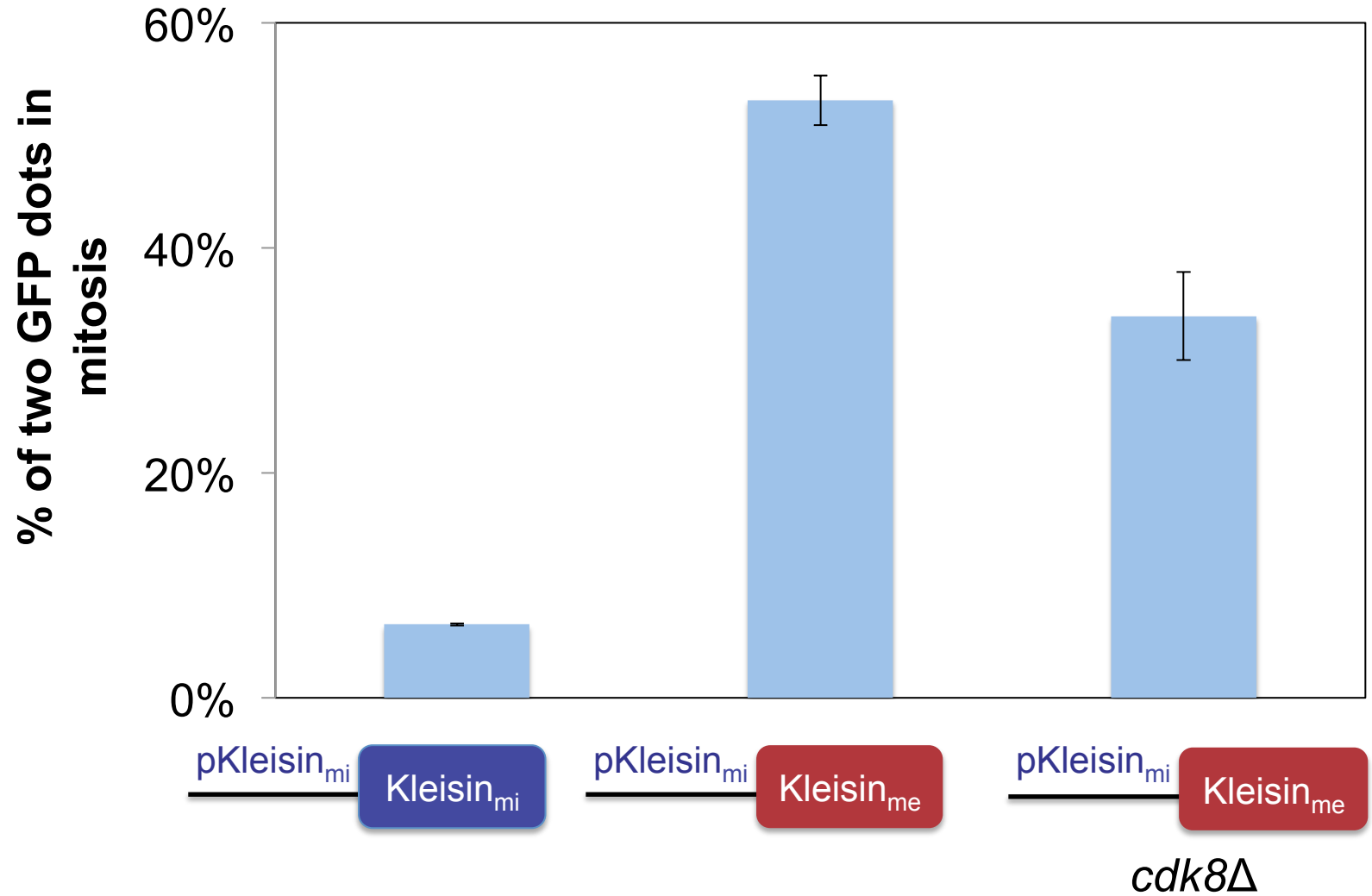
Sister cohesion



No sister cohesion



# Loss of Cdk8 partially fixes cohesion defect

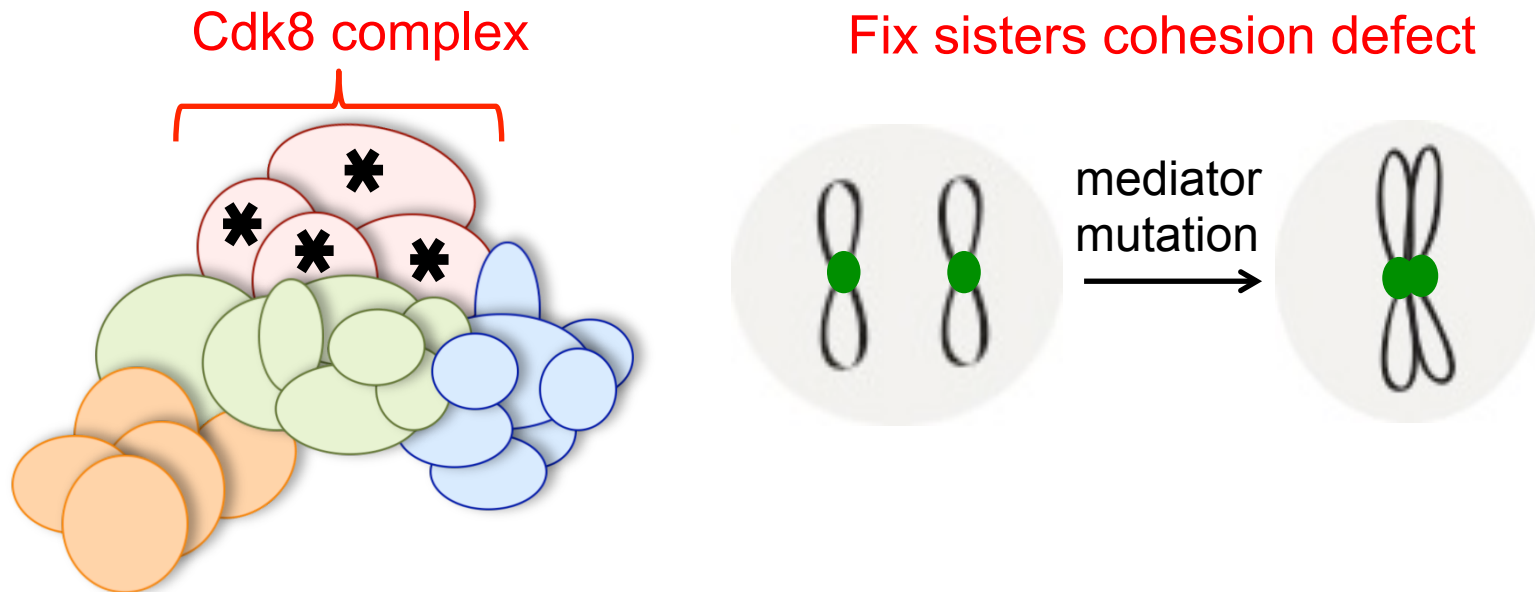


# Summary

Yeast cells adapt to use meiotic kleisin in mitosis

Primary target: transcriptional mediator complex

Killing Cdk8 reduces cohesion defect



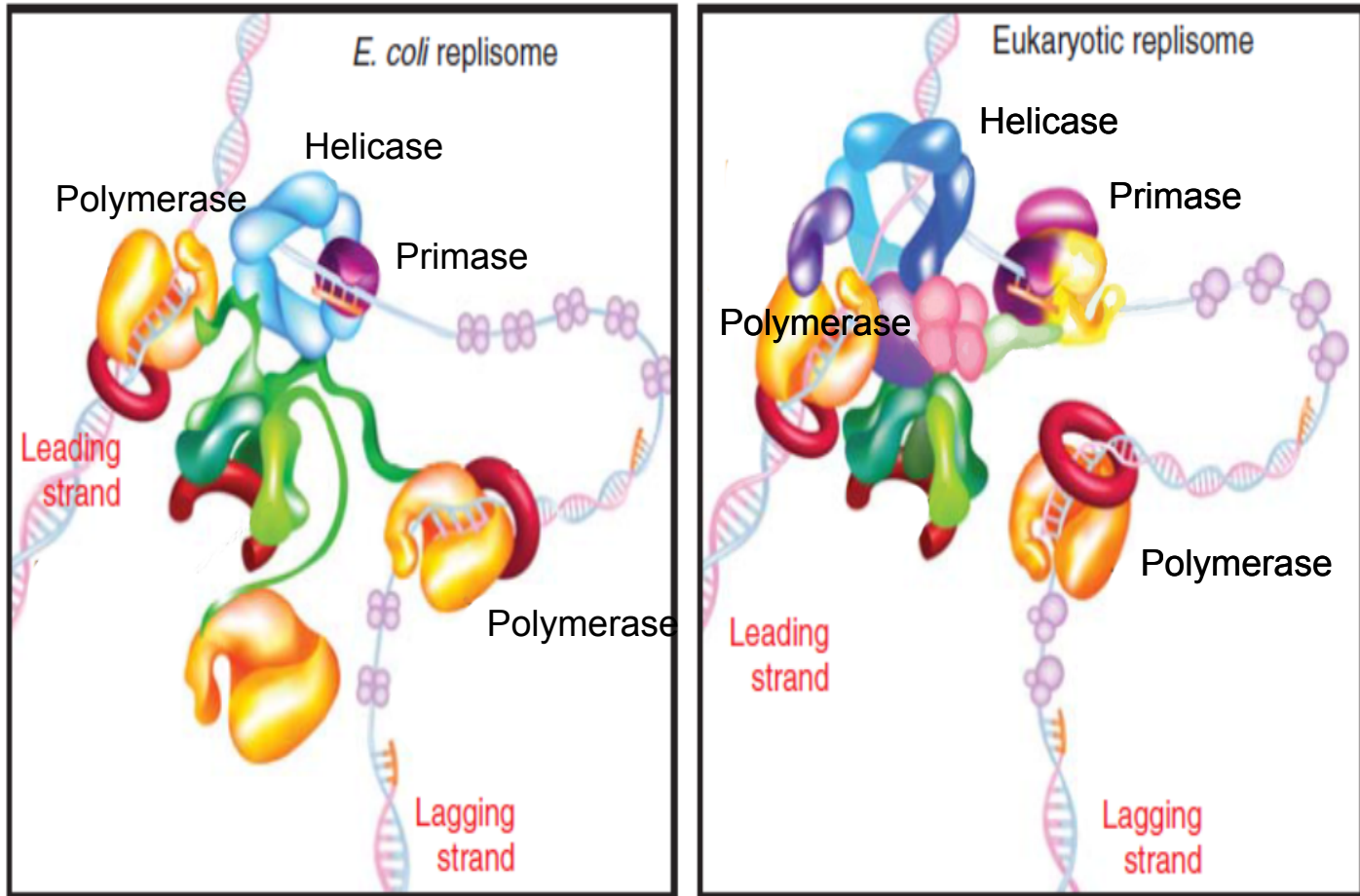
# Evolving to live with unstable replication forks



Marco Fumasoni

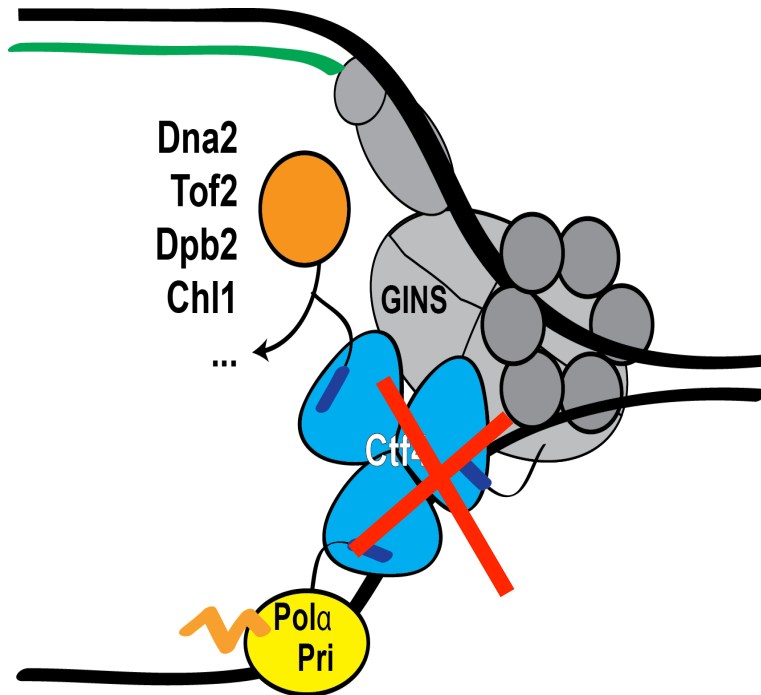


# Organization of bacterial and eukaryotic replisomes



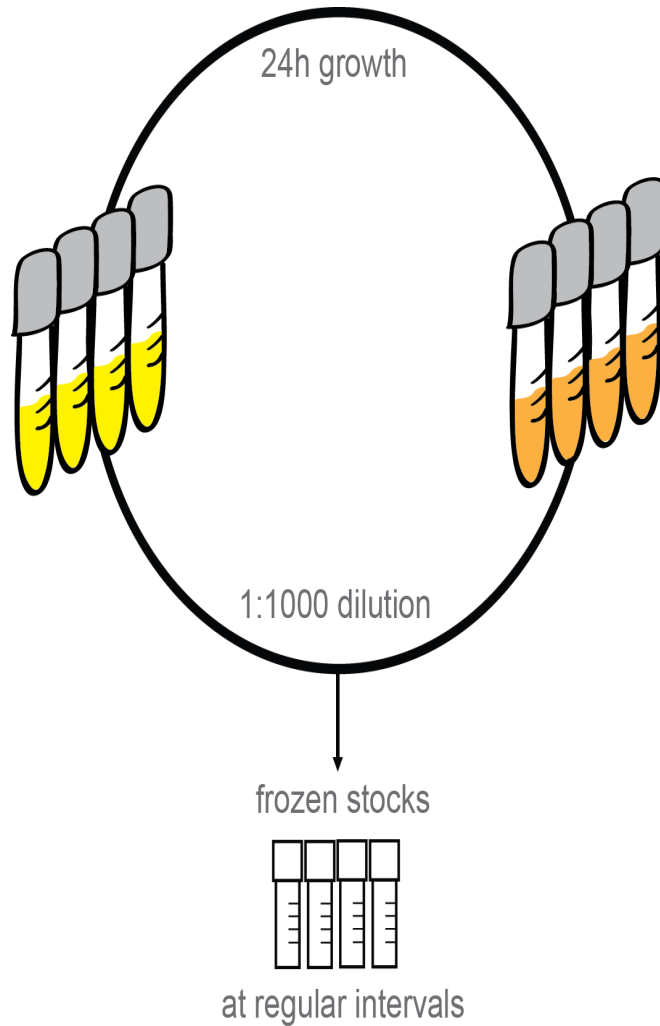
Adapted from O'Donnell et al., 2013

# Genetically perturbing the replisome: *ctf4*Δ

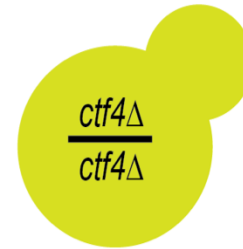


- Poor growth  
(Miles and Formosa, 1992)
- Premature sister chromatid separation  
(Hanna et al., 2001)
- Defect in Homologous Recombination  
(Ogiwara et al., 2007)
- Perturbed DNA damage tolerance  
(Fumasoni et al., 2015)

# Experimental evolution

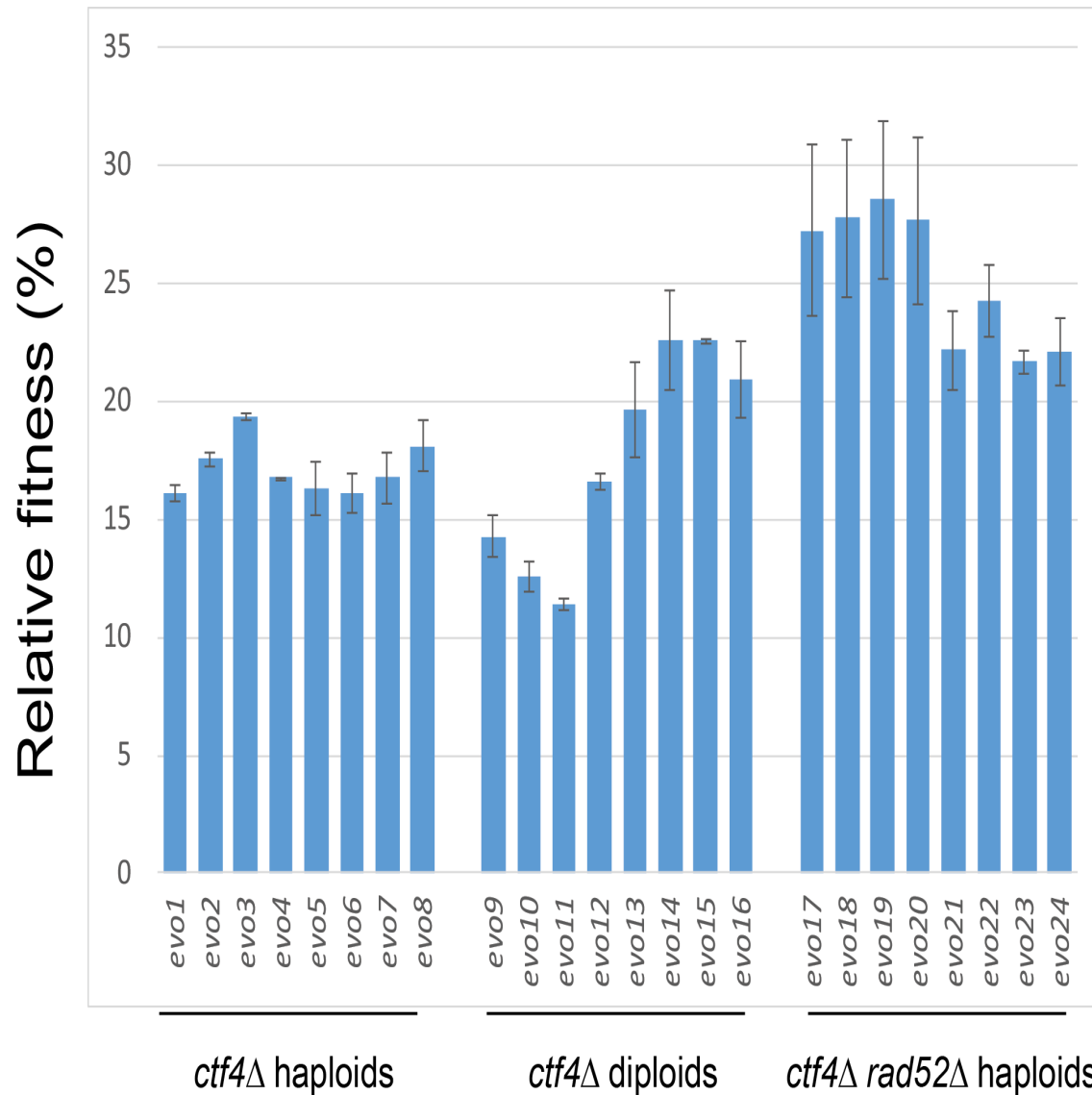


GENOTYPES EVOLVED



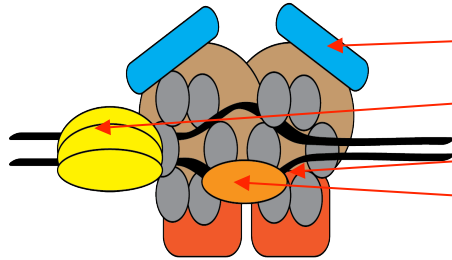
10 ~ Generations per day = 1000 ~ Generations in 4 months

# Relative fitness increase over 1000 generations



# Mutations affect the replication machinery

## Replication origin



## Recurrent gene hits

**DPB11** (3)

**ORC1** (1)

**MCM10** (1)

**FKH1** (1)

**Essential Genes**

## Replication fork

**SLD5** (6)

**RFA1** (4)

**CDC45** (3)

**MCM3** (3)

**TOF1** (3)

**CSM3** (3)

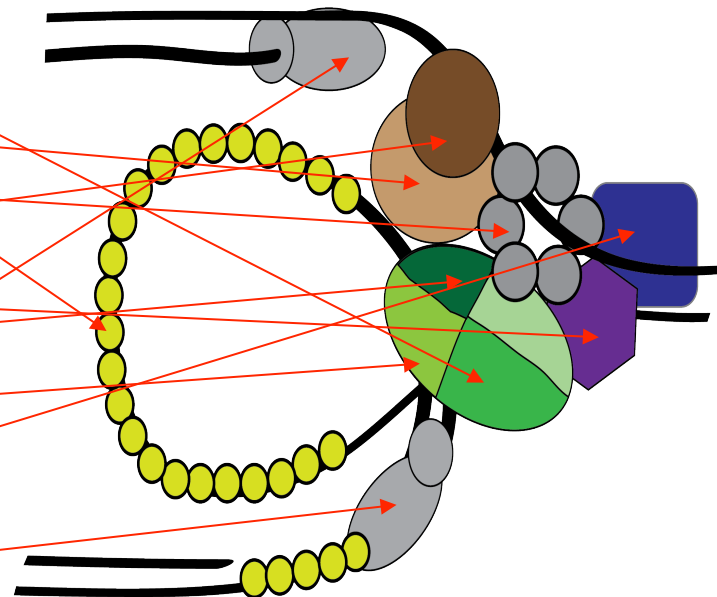
**PSF3** (3)

**PSF1** (2)

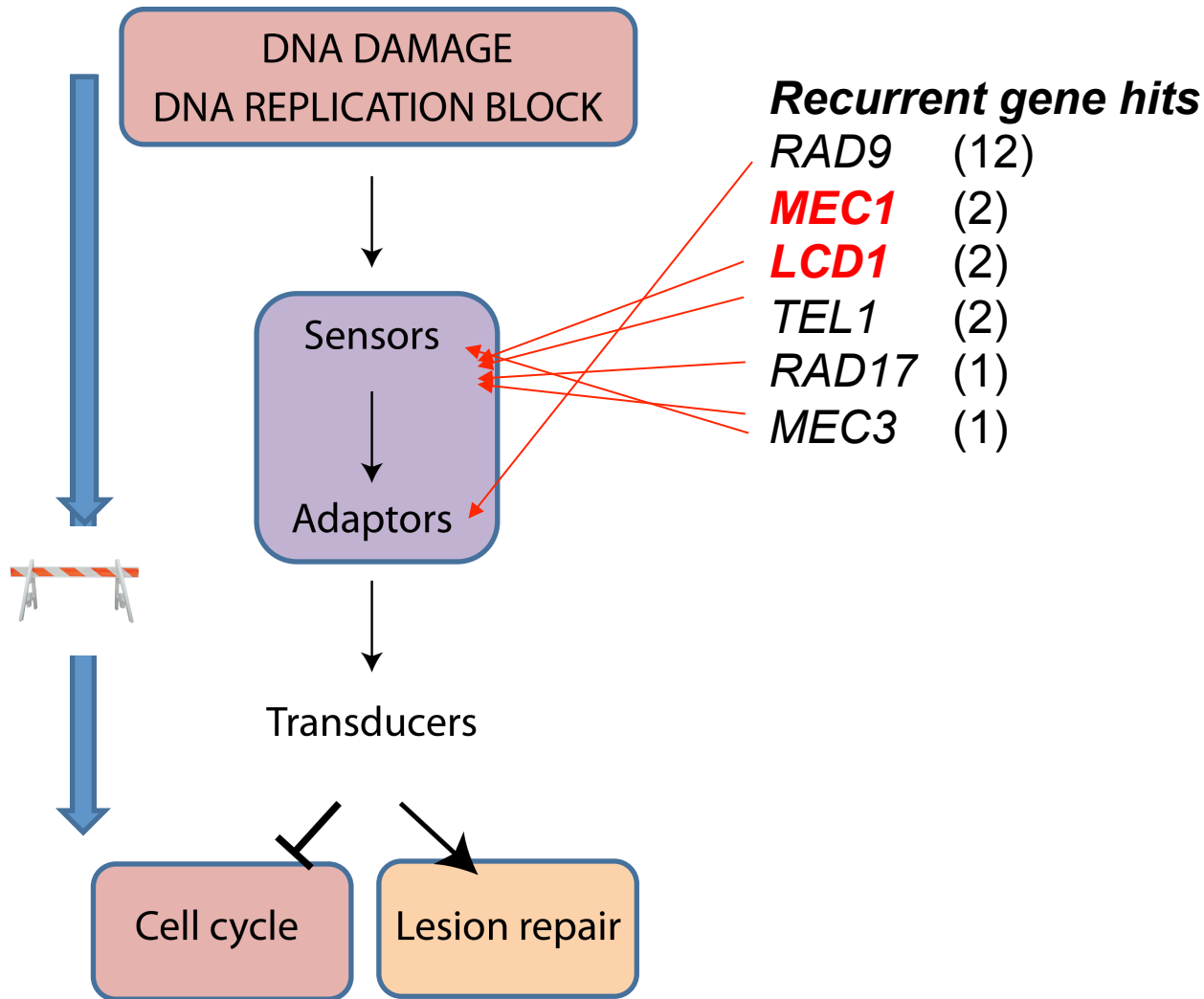
**DPB2** (2)

**TOP1** (2)

**POL32** (2)

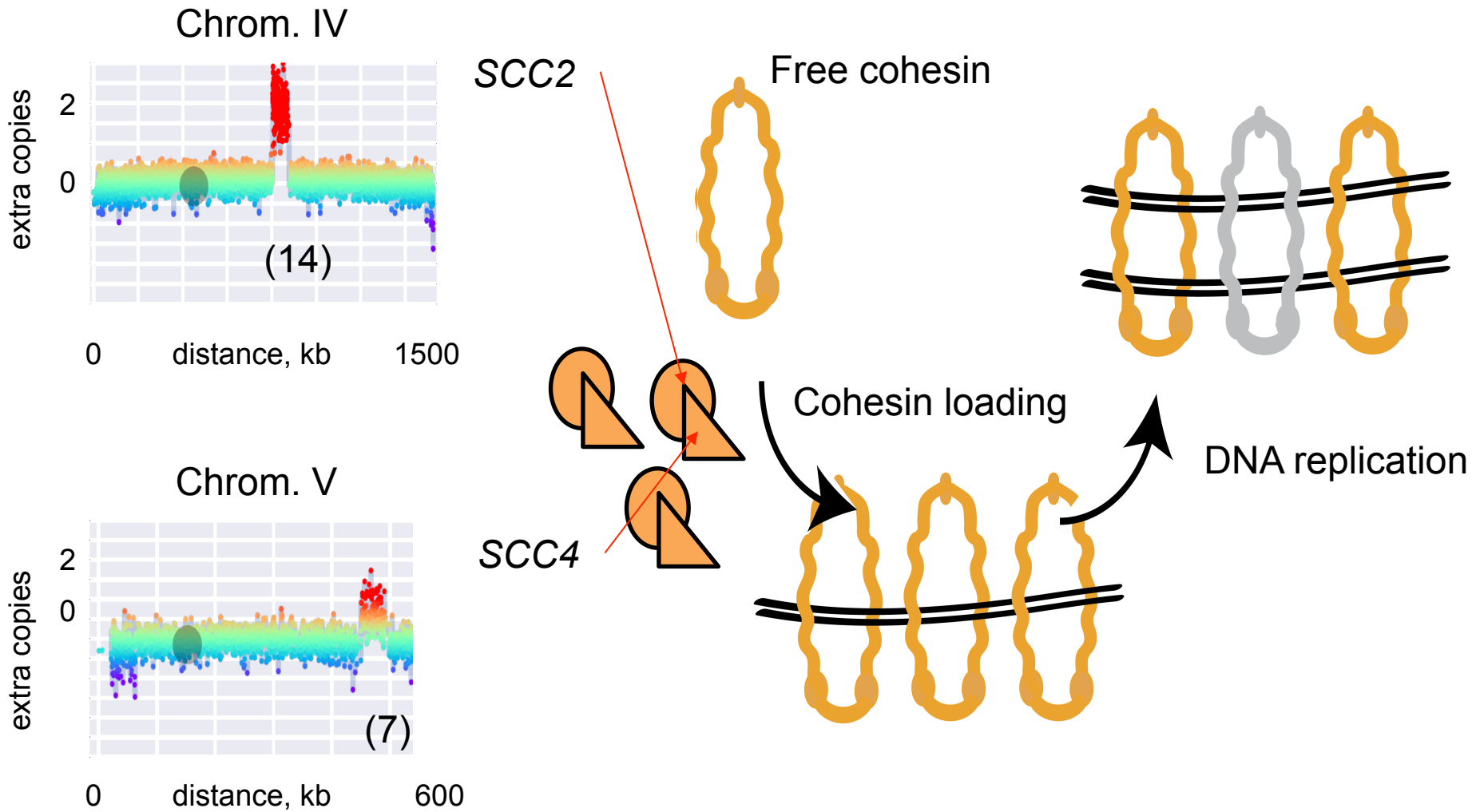


# Mutations break the DNA damage checkpoint



These mutations found only in haploids

# Recurrent chromosome rearrangements



These mutations found only in recombination<sup>+</sup> strains

# Summary

Cells adapt to perturbed DNA replication

The replisome can accept mutations in essential genes

Other conserved pathways have been turned up or down



# Are these straws in the wind?

Repair trajectories more reproducible?

Loss of function dominates causal mutations?

In nature, novelty produced by gene inactivation?