Genetic determinants of maize phenotypic evolution





Maud Tenaillon

CNTS

L Ch



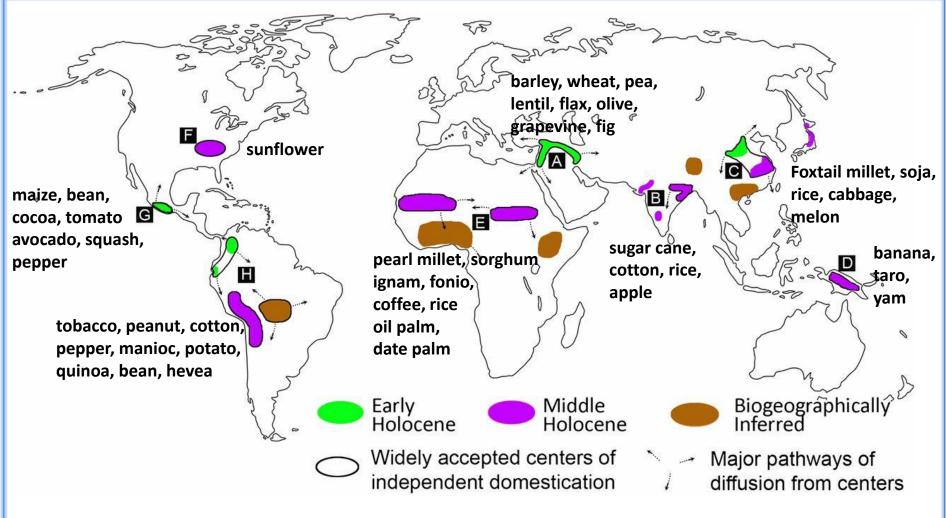
Adaptation to human-needs in domesticated species



Domesticated species are good models to study adaptation because

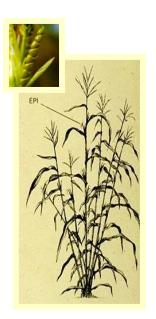
- Long term *in vivo* evolution (>8,000 years)
- The variation is particularly pronounced
- Cultivated range of domesticated species by far exceeds that of their wild "ancestors"
- Genomic data for most domesticated species are produced

Centers and epochs of plant domestication

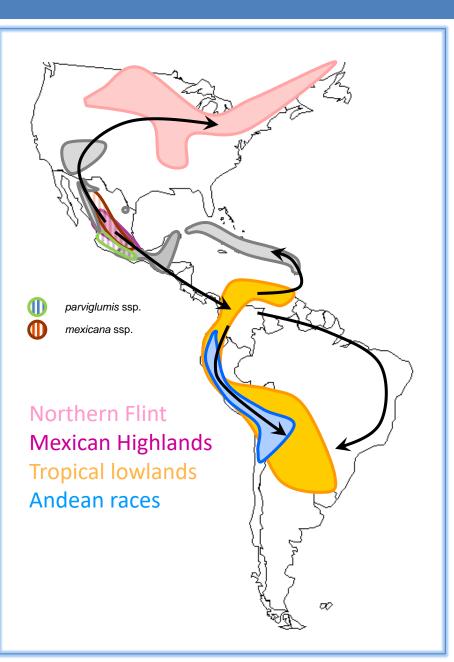


Early Holocene: -(12 000-8 200) years, Late Holocene: -(8 200-4 200) years

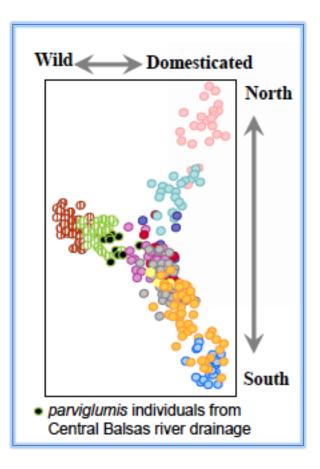
Origin and history of corn





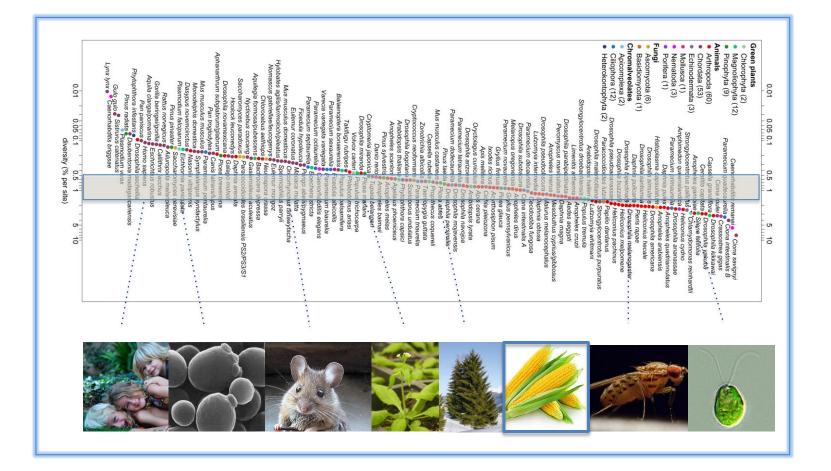


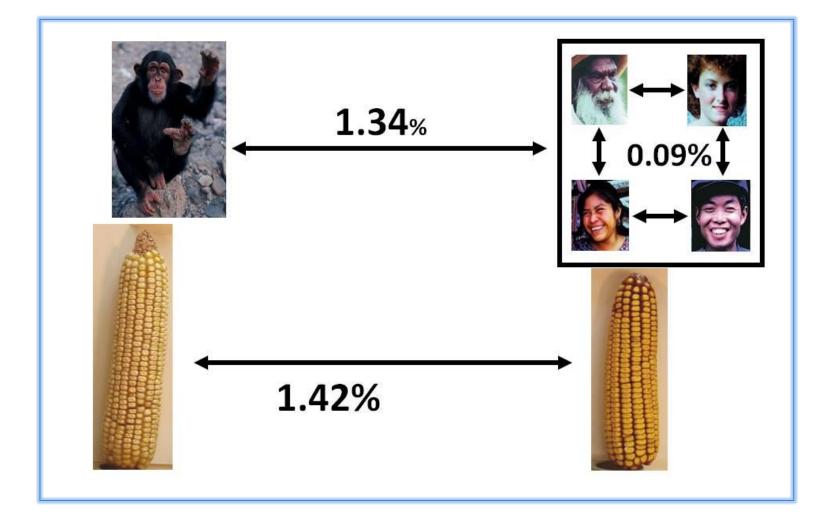
Domestication occurred in Mexico around 9000 years ago



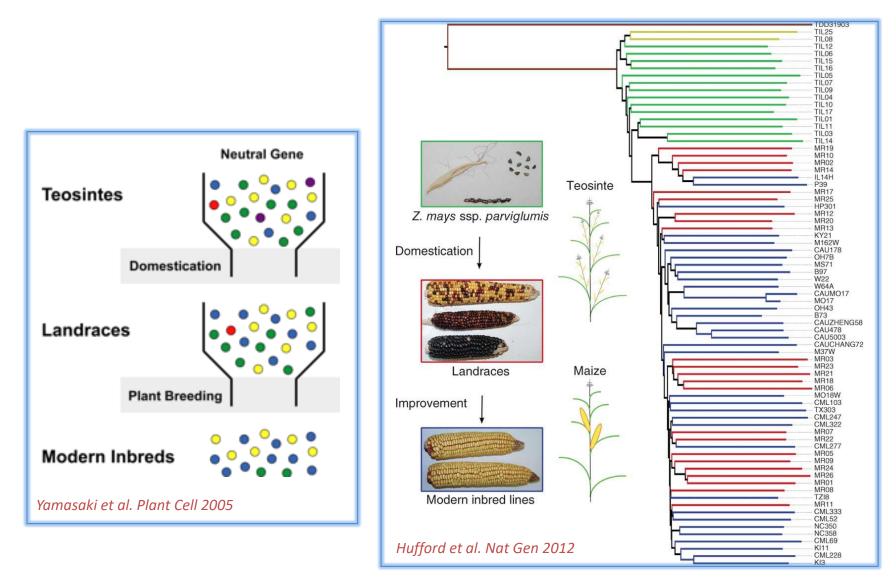
Matsuoka et al. PNAS 2002 Tenaillon and Manicacci Adv. In Maize 2011

Genetic diversity in maize



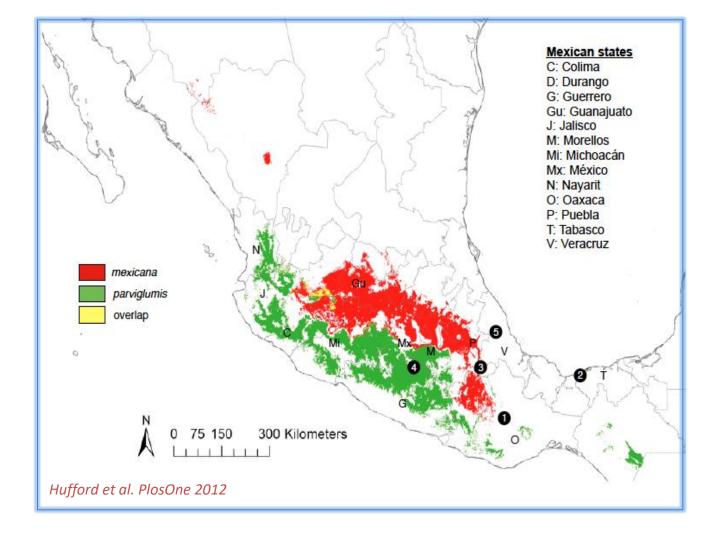


Maize underwent two bottlenecks

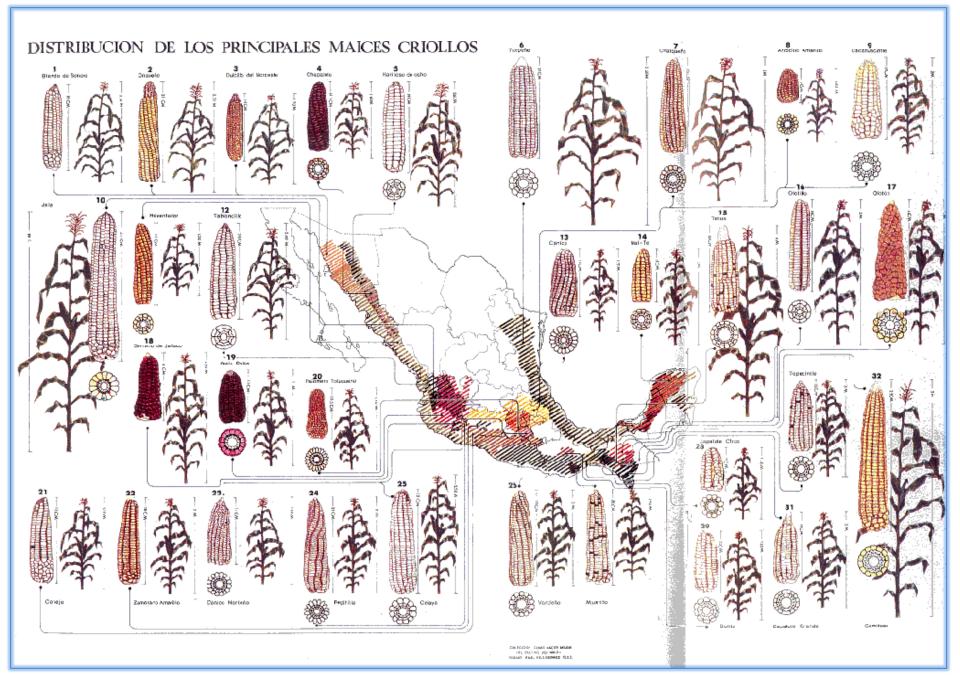


The reduction of diversity for the domestication bottleneck has been estimated to 20% and the breeding bottleneck to < 5%

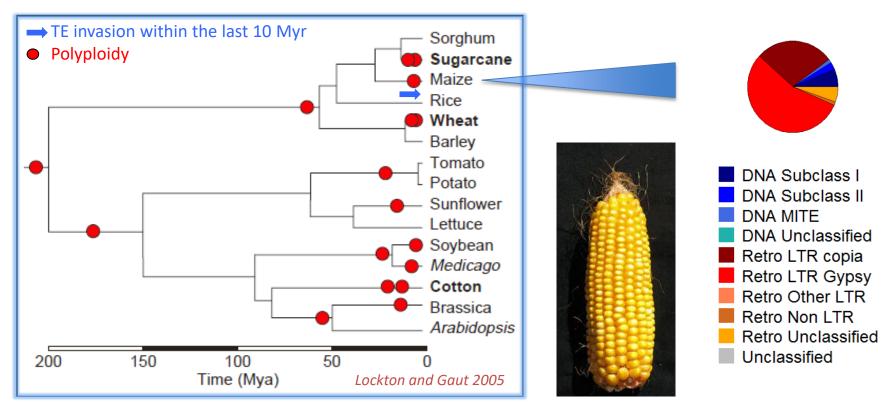
Niche modeling



Distinct ecological niches with little overlap Average altitude for *parviglumis* 1058 m, for *mexicana* 2105 m



Maize genome is about 2.5 Gb, is highly redundant



Vitte et al. Brief Funct Gen 2014

and contains 85% of transposable elements

• A bit more on the European history of maize

Drivers of maize adaptation:

- Short life cycle? Relation to genome size evolution
- Flowering time

• A bit more on the European history of maize

Jean-Tristan Brandenburg, GQE-Le Moulon, France Tristan Mary-Huard, GQE-Le Moulon, France Guillem Riguaill, IPS2, France Clémentine Vitte, GQE-Le Moulon, France Johann Joets, GQE-Le Moulon, France Stéphane Nicolas, GQE-Le Moulon, France Alain Charcosset, GQE-Le Moulon, France



J-T Brandenburg



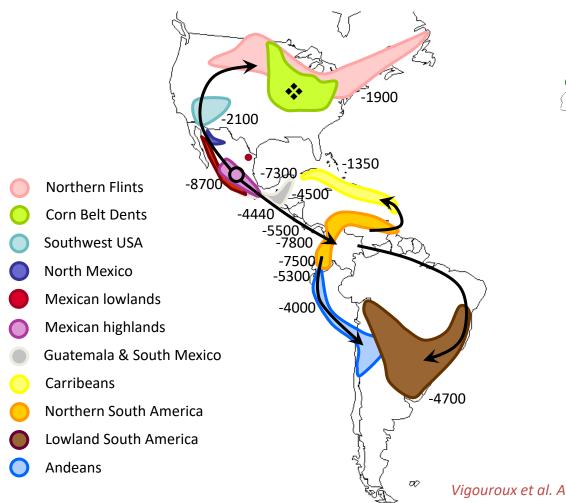
Origin and history of corn



Domestication center

Major expansions

Recent hybridization

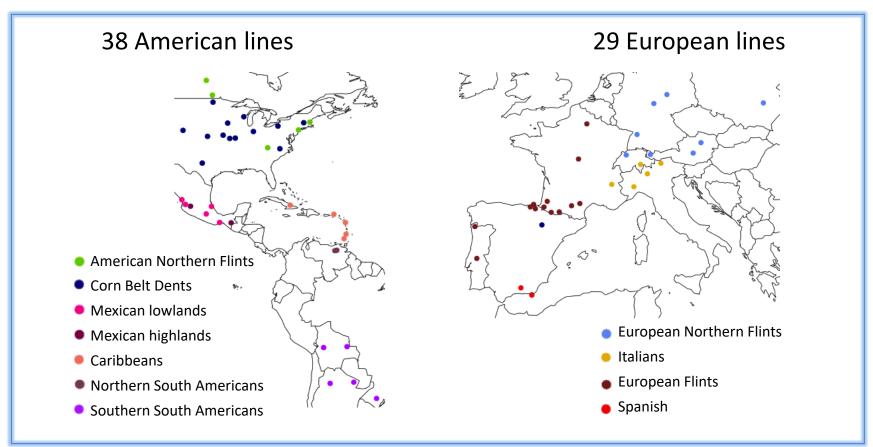




Vigouroux et al. Am J Bot 2008 Tenaillon and Charcosset CRAS 2011

Sampling

Favour inbred lines directly derived from landraces Ensure a correct representation of European genetic groups Include all possible American sources of European maize



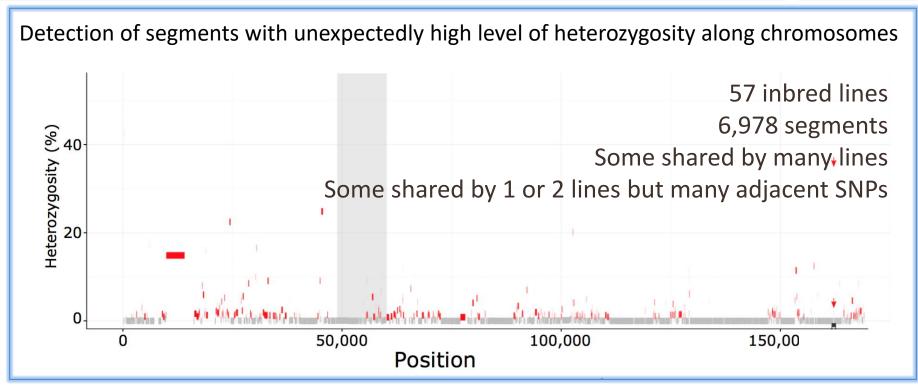
First-cycle inbreds + Single seed descents (57), Doubled haploids (10)

Sequencing:

67 mid-depth whole genome sequencing (18x) >22 million SNPs genotyped covering >85% of all maize genes Alignment to an outgroup: *Tripsacum dactyloides*

> False positive rate for homozygotes calls is 0.036% False positive rate for heterozygotes calls is 34%

Patterns of heterozygosity in inbreds

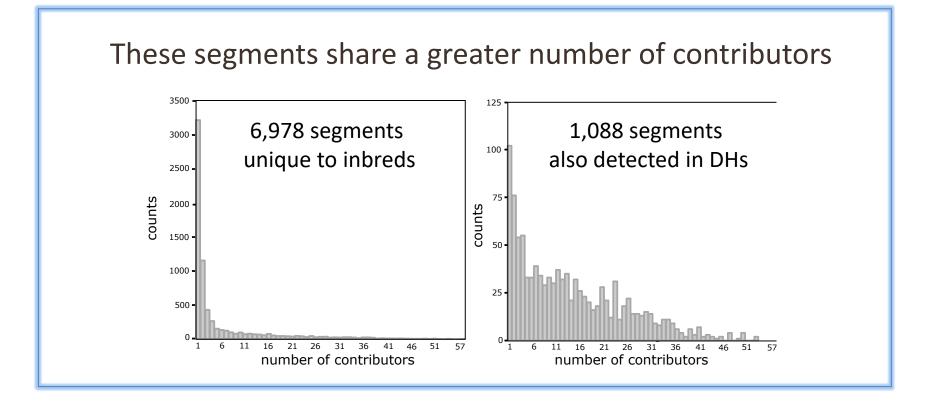


79% are unique to inbreds (not detected in Doubled Haploids) SIFT: heterozygous segments encompass significantly more deleterious variants than the rest of the genome

Selection against inbreeding depression contributes to shape patterns of residual heterozygosity

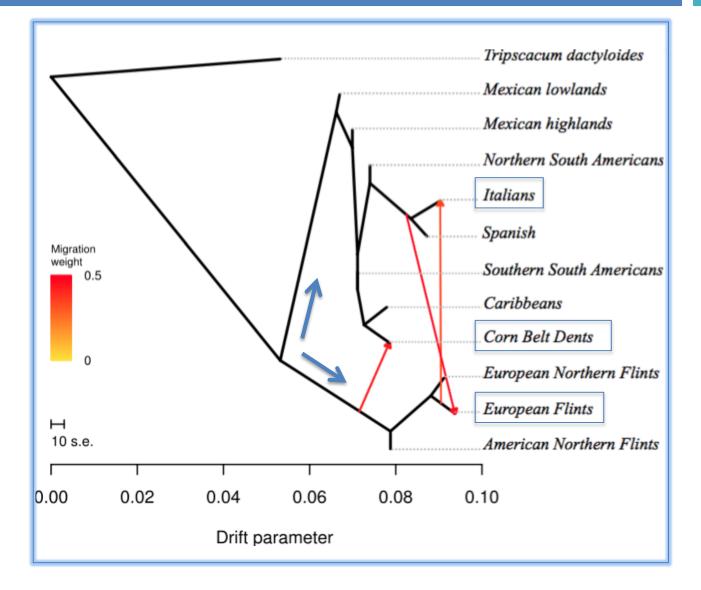
Patterns of heterozygosity in Doubled Haploids

21% of segments (1,088) are found in DHs = alignment artefacts due to structural variation



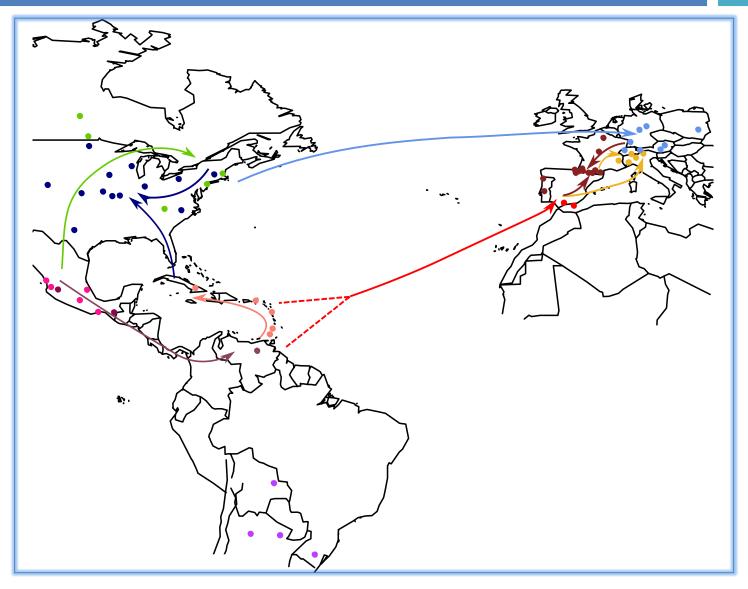
Substantial proportion of stretches of heterozygosity are caused by common structural variants

Genetic proximities and admixture



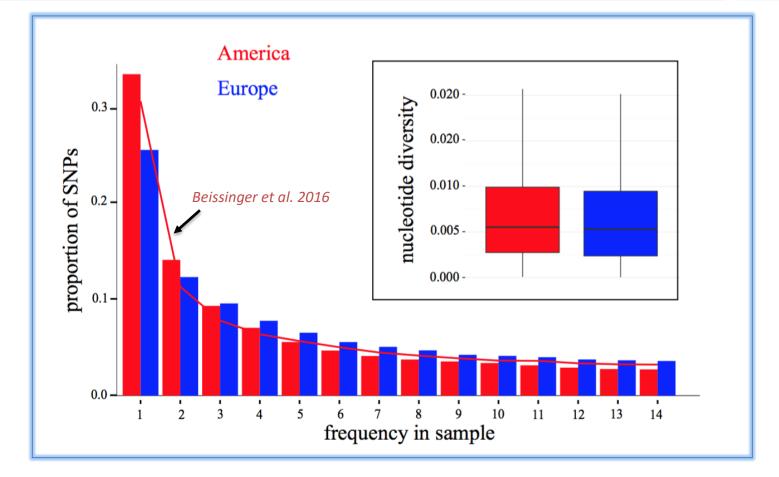
2 independent introductions, 3 admixed groups

Proposed scenario of European introduction



Admixture is an important contributor of adaptation to mid-latitudes

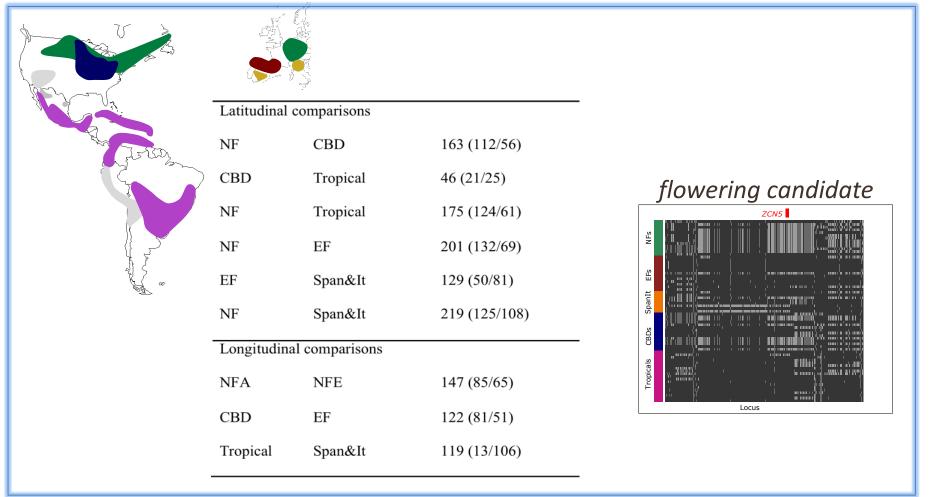
Footprints of European introduction



Modest footprints but: Significant loss of diversity (7%) Loss of rare variants

Targets of selection along latitudinal and longitudinal contrasts

Combination of 3 statistics in genes, using non-genic windows as controls



968 Differentially Selected candidates: more on average in latitudinal than in longitudinal contrasts, more in NFs than in the other groups

Adapted from *Daub et al. MBE 2013*:

294 networks Sum of normalized P-values among genes of a given network (P-values are computed from observed Fst and Fst of non-genic windows, network size is accounted for)

<u>Drought tolerance</u> tetrapyrrole synthesis ABA synthesis

<u>Cold tolerance</u> Putrescine pathway <u>Stress responses</u> cis- and trans-zeatin pathways

Defense against pathogens β-caryophylens

Early European corn



One of the earliest european drawing of maize can be found in the translation of the book Materia medica de Dioscorides edition 1543 (Frankfurt)

Illustration taken from *De historia stirpium* de Fuchs (1542) with notes on: the American origin of corn the 4 grain colours painted on a single plant

During the 18th century corn become a staple crop



Drivers of maize adaptation:

• Short life cycle? Relation to genome size evolution

We know very little:

- <5% in Hordeum maritimum Jakob et al. MBE 2004
- 16.6% among *Festuca pallens* populations *Smarda et al. Annals Bot 2008*
- about 10% among *Arabidopsis* ecotypes Long et al. Nat Gen 2013
- about 30% of variation in maize and its wild relatives

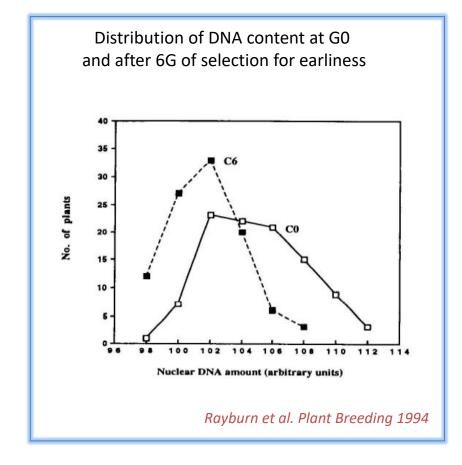
Maize intraspecific variation makes it an interesting model for studying factors driving GS variation



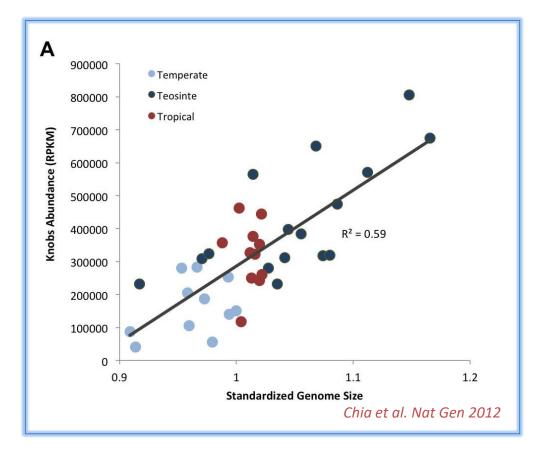








Genome size correlates with knob content

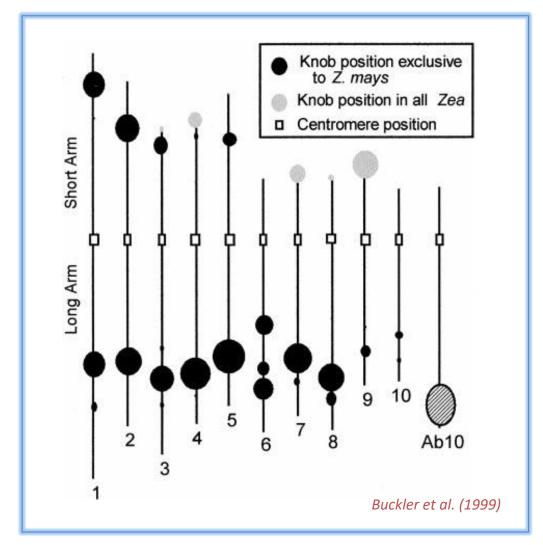


Genome size correlates with knob content (P-value=0.00046)

GS teosintes > GS maize inbred lines GS tropicals > GS temperate

→ GS may be adaptive (seems to correlate with environmental variables)

Knobs in Zea

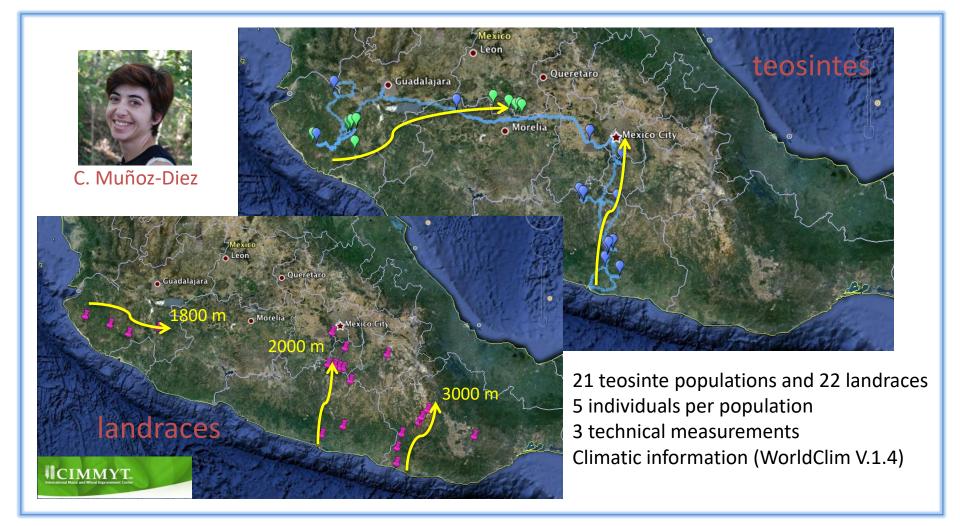


Area of circle is proportional to size and frequency of the knobs

Knobs

 vary in size and frequency

are constituted by 180-bp
360-bp tandem repeats
and transposable elements

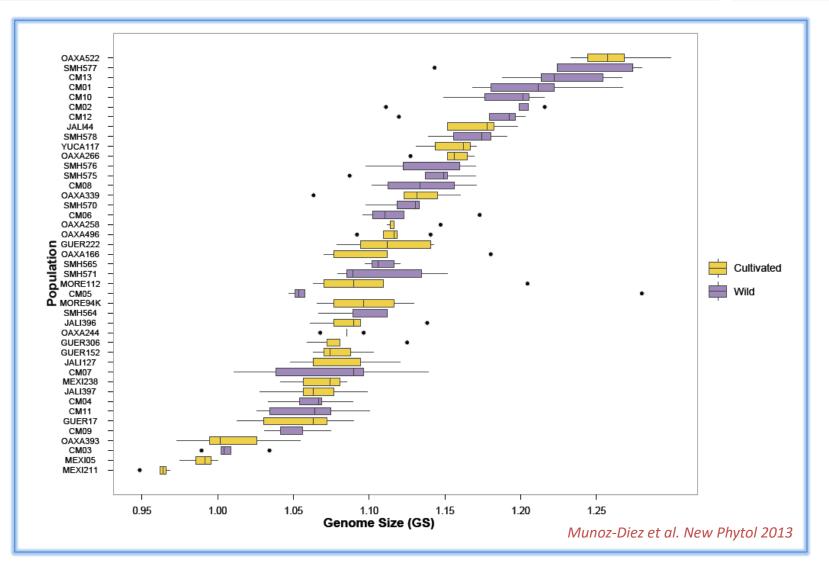


Conception Muñoz-Diez, Esteban Meca, Brandon Gaut, UC Irvine, USA Salvador Montes-Hernandez, INIFAP Celaya, Mexico Enrique Scheinvar, Luis Eguiarte, UNAM, Mexico



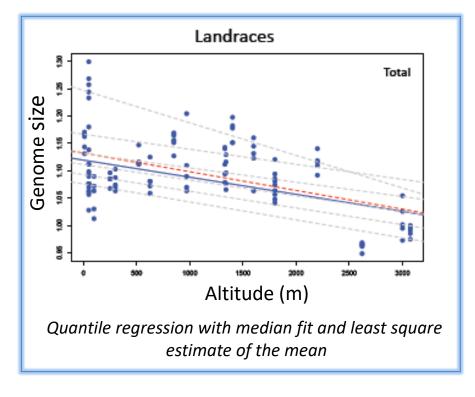


Estimating genome size in teosintes and landraces



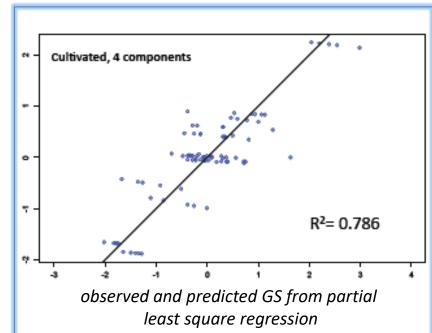
Genome size is significantly larger in teosintes than in maize (by few percent) Proportion of variance explained by variation among populations (\approx 71%) > within (\approx 20%) > subspecies (\approx 2%)

Is genome size variation adaptive in Zea mays ?



In maize, GS correlates negatively with altitude

And geographical components (altitude, longitude, latitude) are good Predictors of GS



Phenotypes that affect GS are likely complex, we investigated one of them: Leaf Elongation Rate

Measuring growth with LERmax

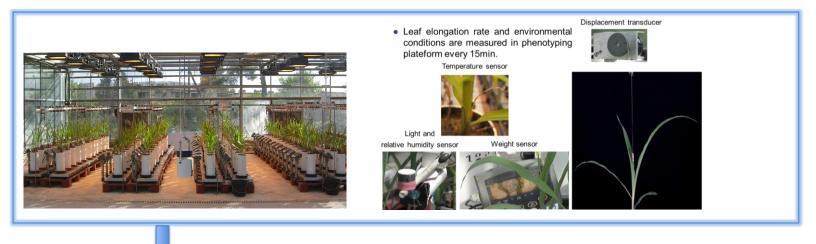


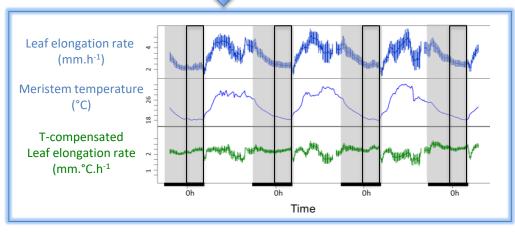
J. Melique

Claude Welcker, INRA Montpellier, France Francois Tardieu, INRA Montpellier, France Domenica Manicacci, Le Moulon, France



Measuring Leaf Elongation Rate at Phenodyn (http:montpellier.inra.fr/bioweb/phenodyn)

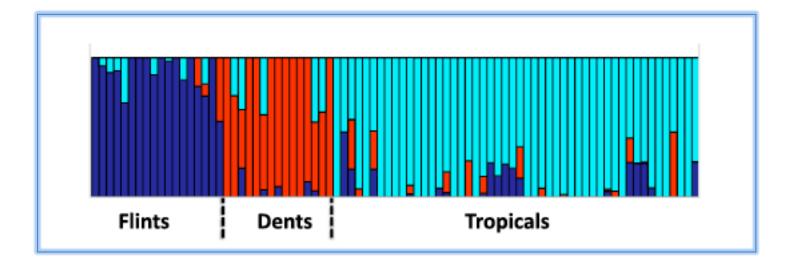




- Leaf growth fluctuates with °C
- In thermal time, a plateau is reached between 0-4h= LERmax
- LERmax measures the intrinsic ability of leaf to grow
- It is highly heritable

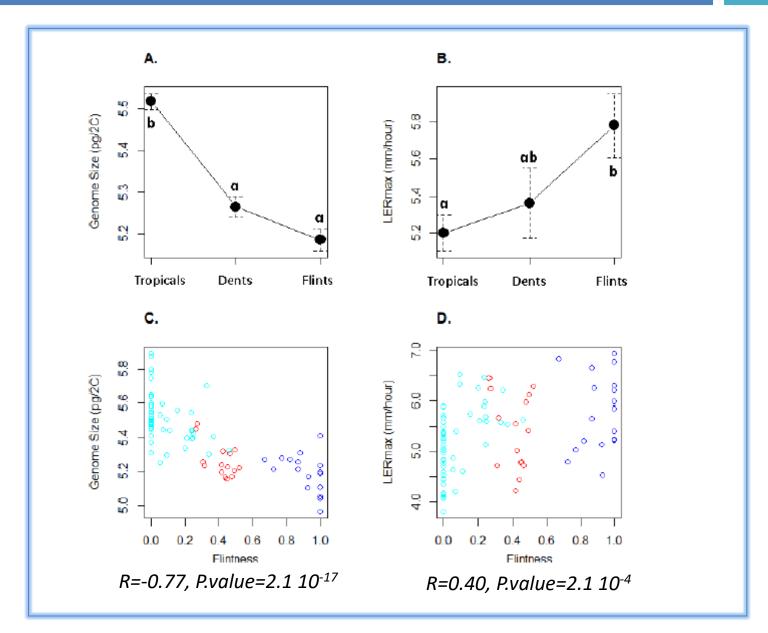
Sadok et al. Plant Cell Env 2007; Dignat et al. Plant Cell Env 2013

Sample: 83 maize inbred lines structured in 3 genetic groups

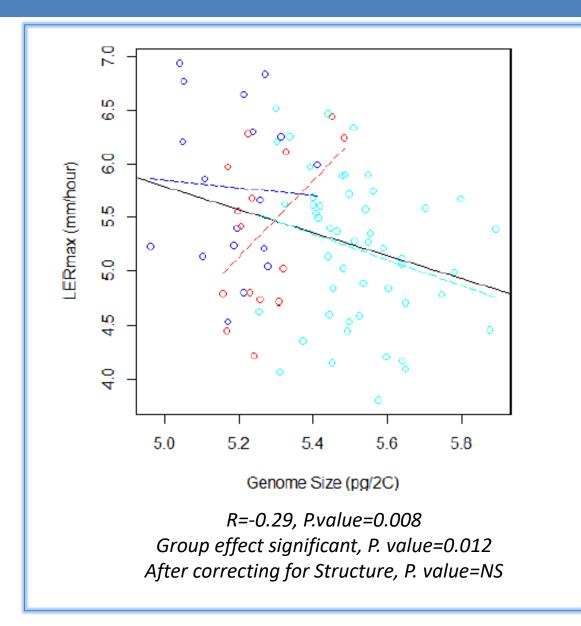




Genome size and LER max by genetic group



Relationship between genome size and LER max



Correlation mainly driven by among-group variation

Tenaillon et al. PeerJ 2016

Bilinski et al. bioRxiv, 2017 – Ross-Ibarra lab, UC Davis

Negative correlations between GS and altitude, knob repeat and TE content in Meso- and South-America (-108 and -154 Kb/meter) in maize landraces

Negative relationship between GS and cell production rate (inferred from LER) within a *mexicana* population,

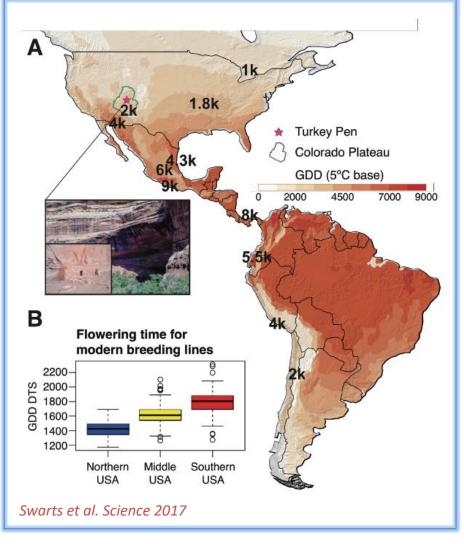
Negative relationship between flowering time and cell production in the meristem in maize lines Leiboff et al. Front Plant Science 2016 Drivers of maize adaptation:

• Flowering time

Flowering time adjustment: a major component of maize diffusion

Maize is originally a tropical plant sensitive to photoperiod flowers under short days (< 13hours of daylight). Variation in daylength indicates a transition between the dry and wet season in the Tropics.

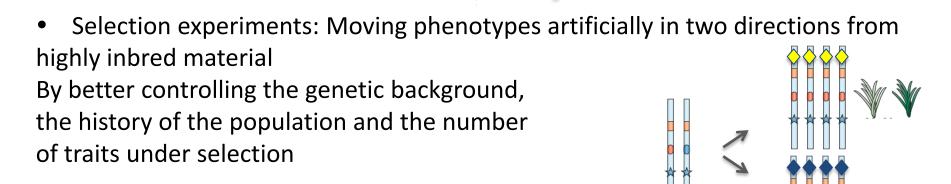
In temperate regions, the days are longer in the summer and temperatures lower. Selection of maize has triggered a loss of sensitivity to photoperiod, early flowering to lengthen the growing season.



Linking phenotypic to genotypic variation

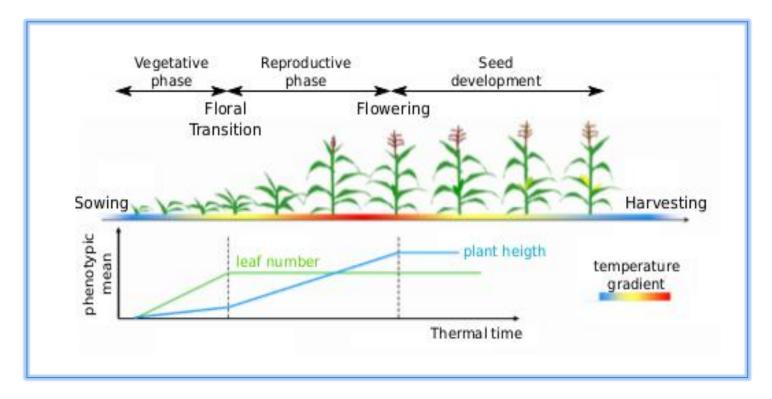
• QTL mapping: Assessing genetic differences in progenies of crosses between two (few) parents with contrasted phenotypes | AA BB

Genome Wide Association mapping (GWA): Statistically associating genotypic to phenotypic variation



Maize flowering time

Floral transition marks the transition between vegetative and reproductive phase, and contributes to determine flowering time







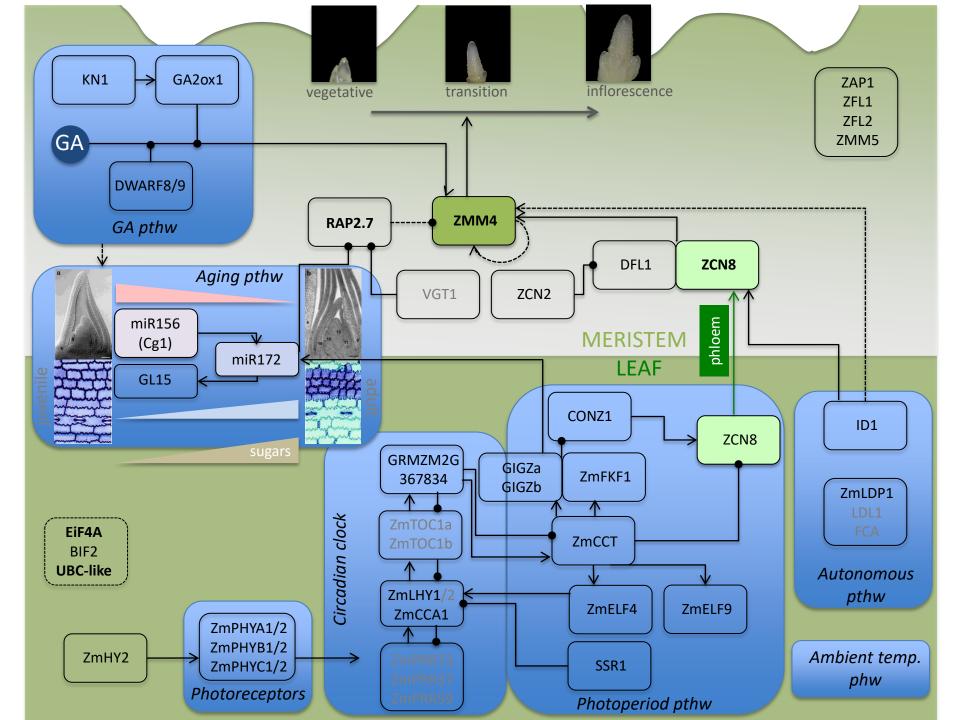
E. Durand



Christine Dillmann, GQE-Le Moulon, France

Adrienne Ressayre, GQE-Le Moulon, France Elodie Marchadier, IPS2, France Martine Le Guilloux, GQE-Le Moulon, France Aurélie Bourgais, GQE-Le Moulon, France Hélène Corti, GQE-Le Moulon, France





16 years of divergent selection for flowering time

Every generation:

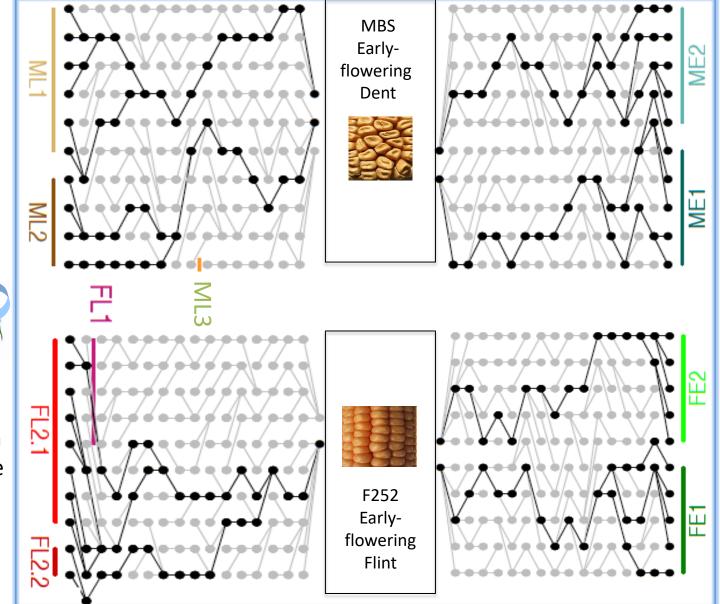
Selection of **10 Earliest** and **10 Latest** genotypes

Selfing

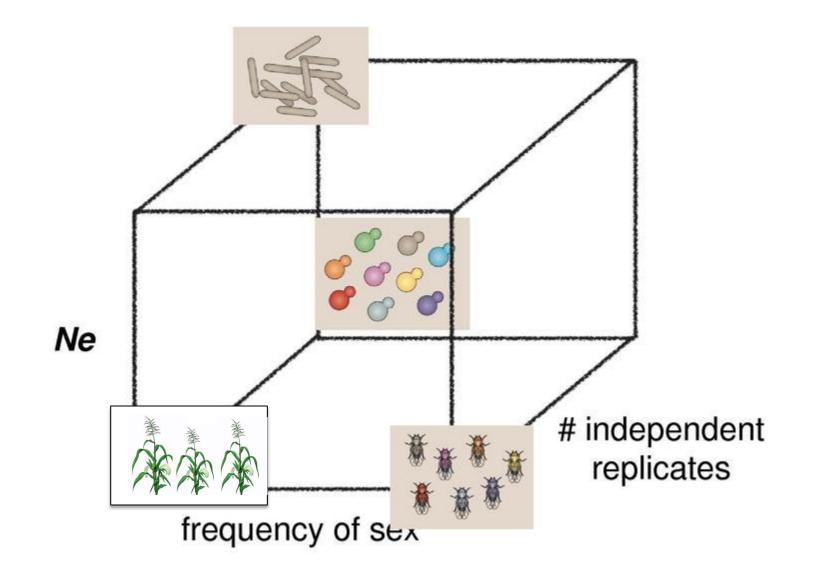


Phenotypic evaluation of 100 plants/genotype

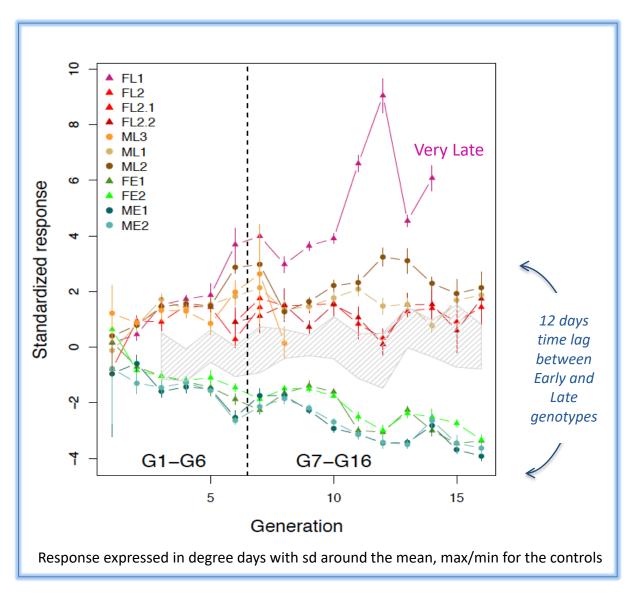
2000 plants total per selection experiment



Going back to Molly Burke slide



Characterization of the response to selection from G1 to G16

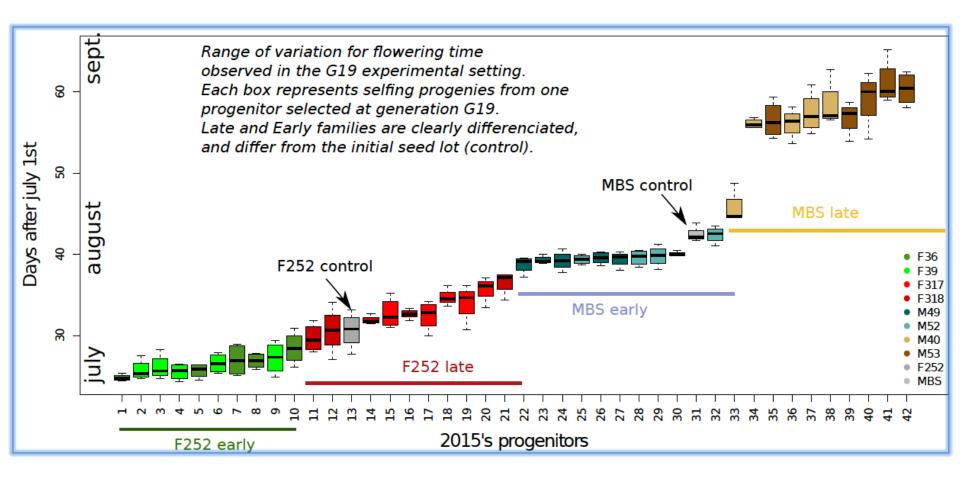


Sustainable, strong and significant response to selection from G 1-6 (stronger in Late)

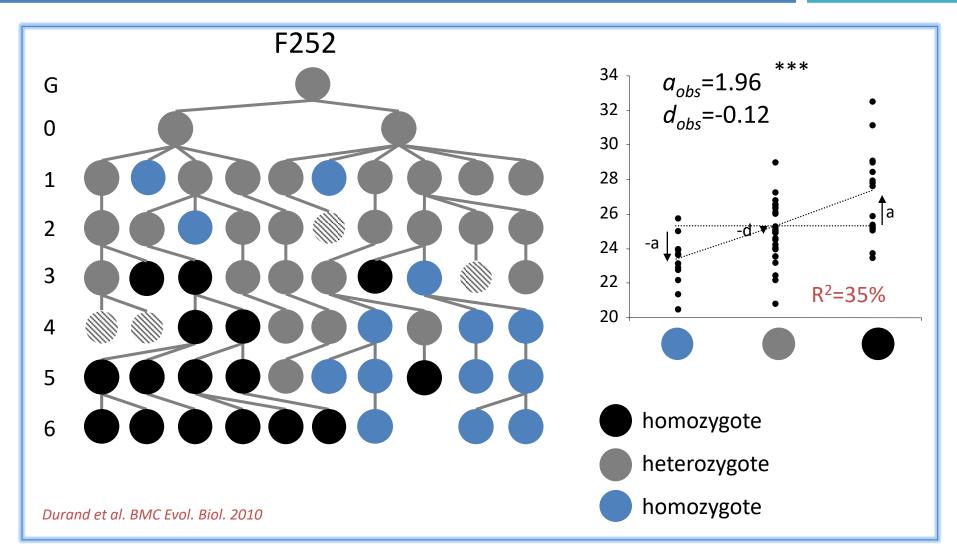
Significant in 6 over 8 families from generation 7-16, continuous in Early, much less so in Late

Durand et al. BMC Evol. Biol 2010

At G19: up to 3 weeks difference between Early and Late



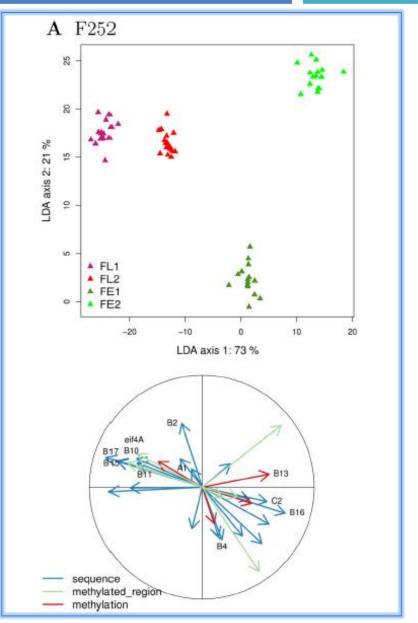
A major locus associated with flowering time variation in Late F252



Polymorphism present as standing variation Locus may explain up to 35% of the variation for flowering time in the Late F252 population

Other determinants: M-SAP and AFLP screening from G0-G6

- 24 markers total (highly inbred material) discriminating Early and Late subfamilies at G7
- 16 of these are sequence basedpolymorphisms8 are methylation-basedpolymorphisms
- All markers were segregating in the original seed lot, no *de novo* mutations



Durand et al. BMC Evol. Biol 2015

In progress: Samples preparation (G13 genotypes)

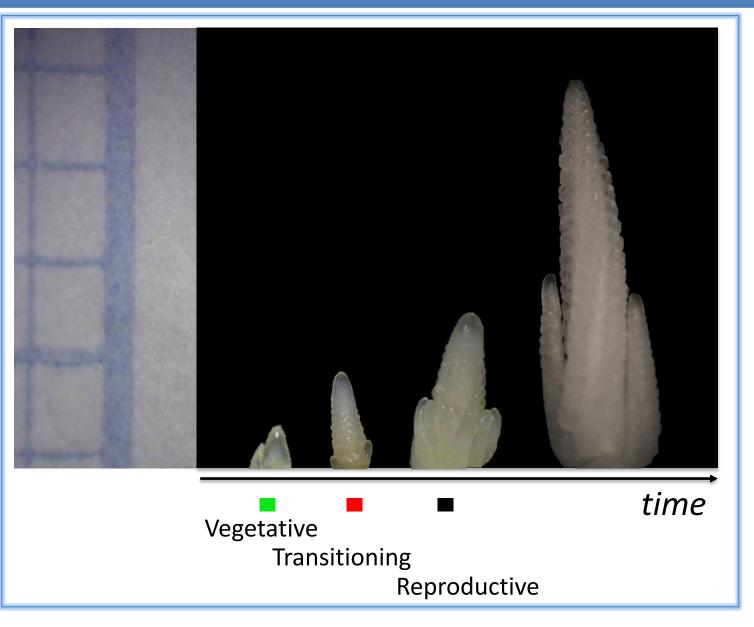


Sampling, leaf number and meristem stage determination

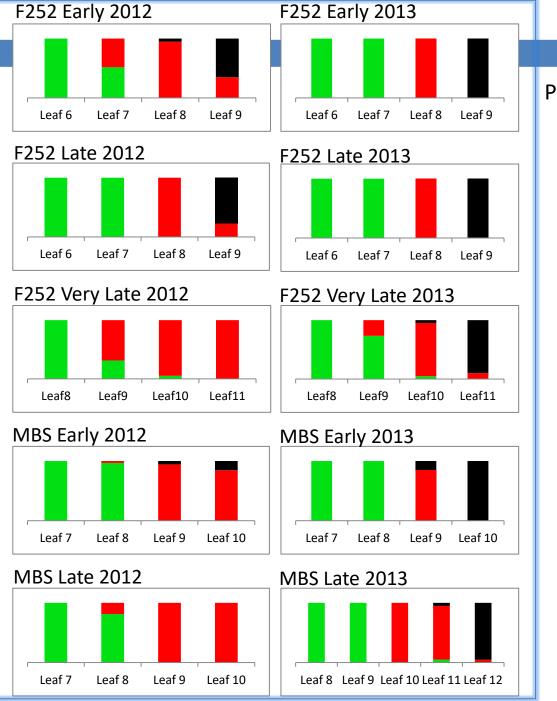




Meristem status



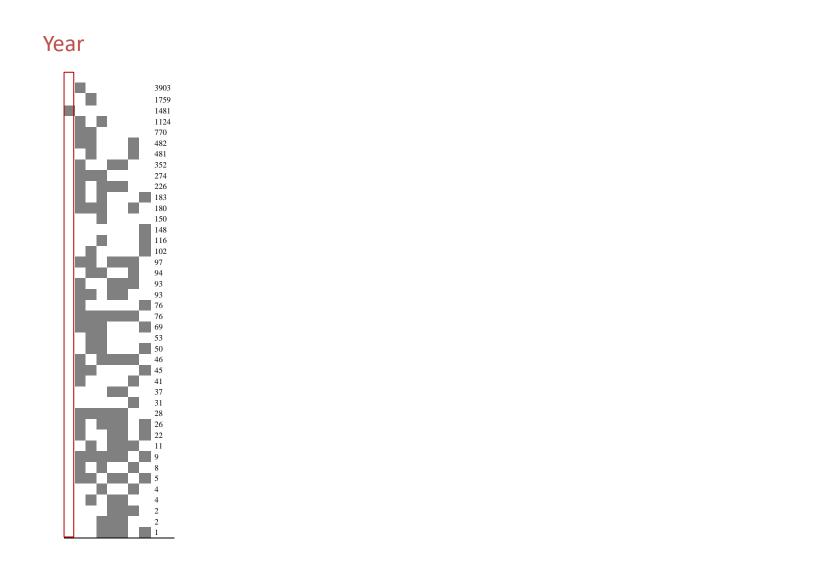




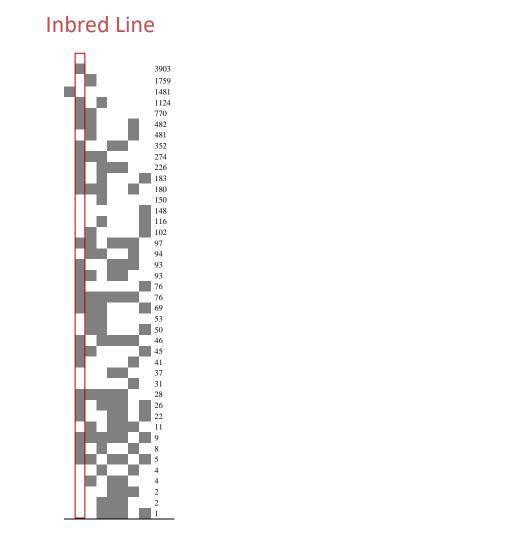
Collected samples

Proportion of meristems stages in pools of 20 to 35 meristems

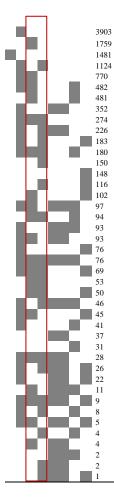
Defining DE genes from contrasts

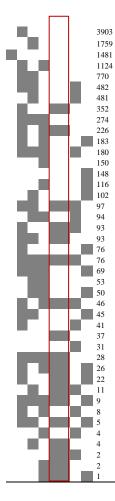


Defining DE genes from contrasts



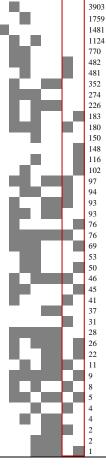
Status in F252 and MBS





Early vs Late/VeryLate in F252 and MBS=DE genes involved in the Early vs Late response

Status x Genotype and Status x Genotype differing in Early vs Late/VeryLate in F252 =DE genes involved in the Early vs Late response in F252



39,066 genes tested 12,754 DE genes (33%)

1481 with Year effect onlySELECTION2451 DE genes that differ between Early and Late genotypes (19%)

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2120 in F252
446 in MBS
115 common (P.value=9.14 10<sup>-6</sup>)
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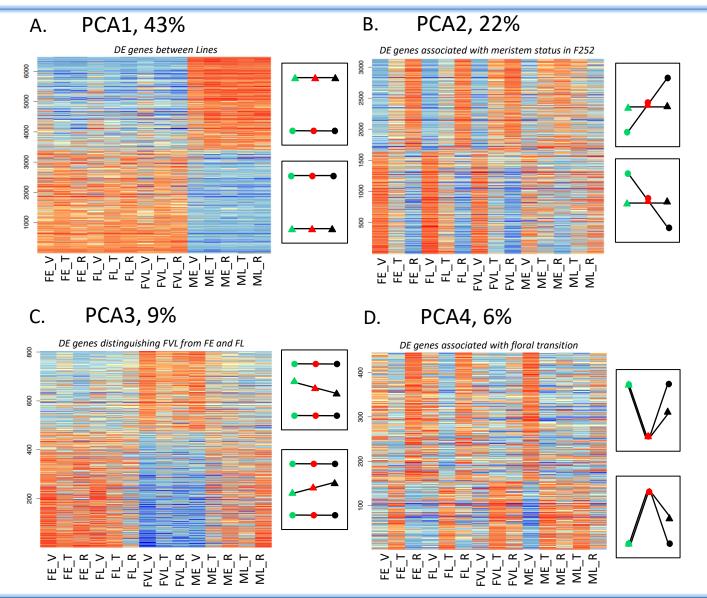
CONVERGENCE

157 candidates for flowering time95 are DE genes (60%)29 (18%)

ENRICHMENT

Heat maps of Differentially Expressed (DE) genes

DE genes with greatest correlation with the 4 first PCs (normalized average expression corrected for Year)

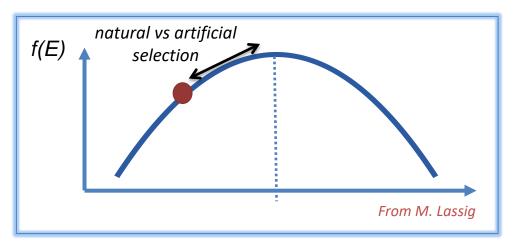


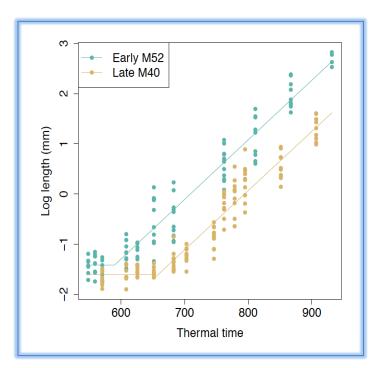
Perspectives

Evolution of allele frequencies, de novo vs existing variation

Integration of diverse sources of data Transcriptomic, proteomic, development

Modeling the response to selection to understand the steadiness of the response: few polymorphisms/many DE genes, role of epistasis?





Thank you to all collaborators and members of the DyGAP team











