

Analysis of immune receptor repertoires

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European Research Council
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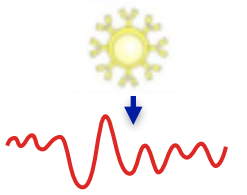
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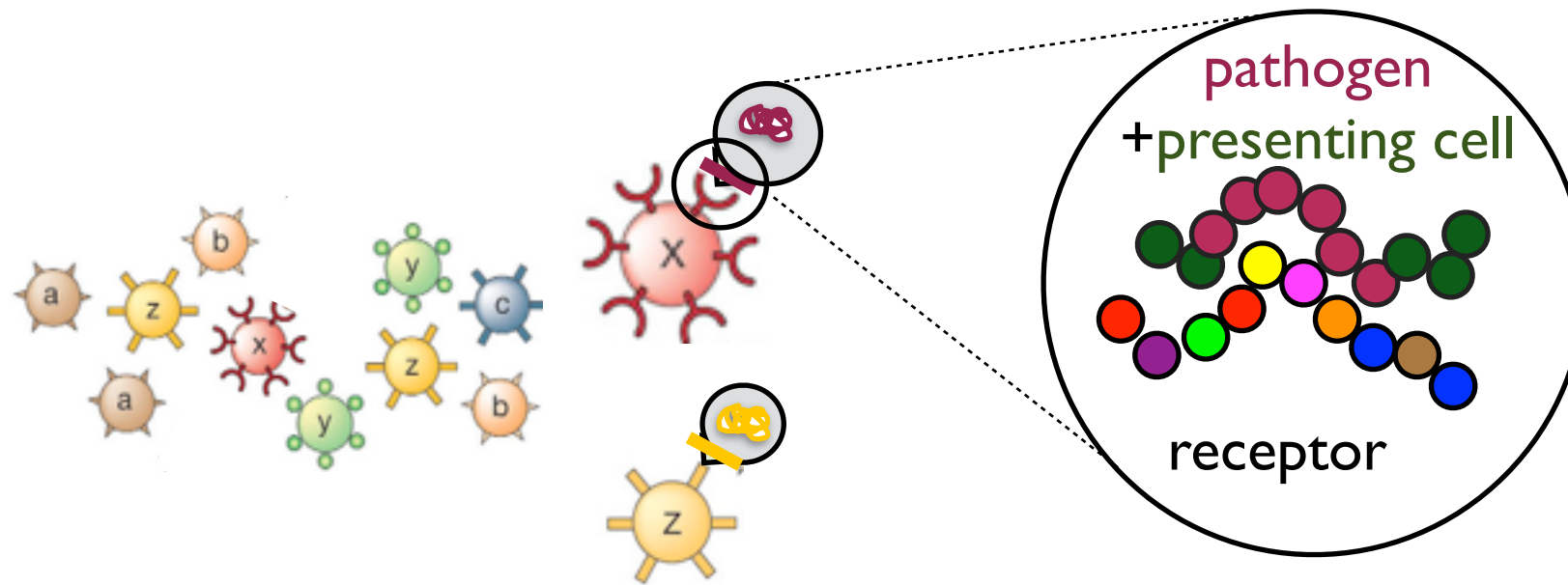
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Immune receptors

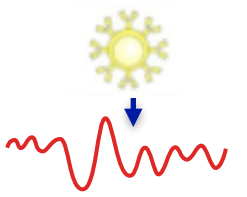


- T-cells important actors of immune system

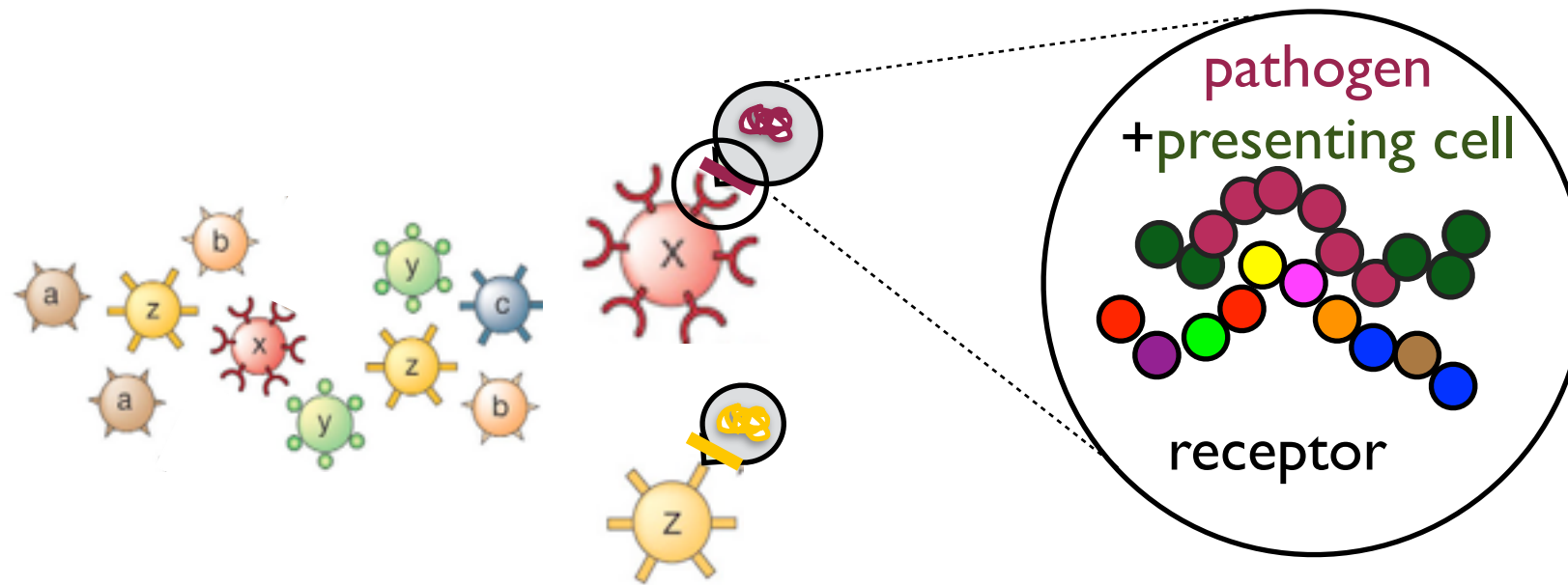


- triggers immune response

Immune receptors



- T-cells important actors of immune system



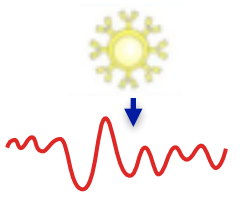
many unique receptors



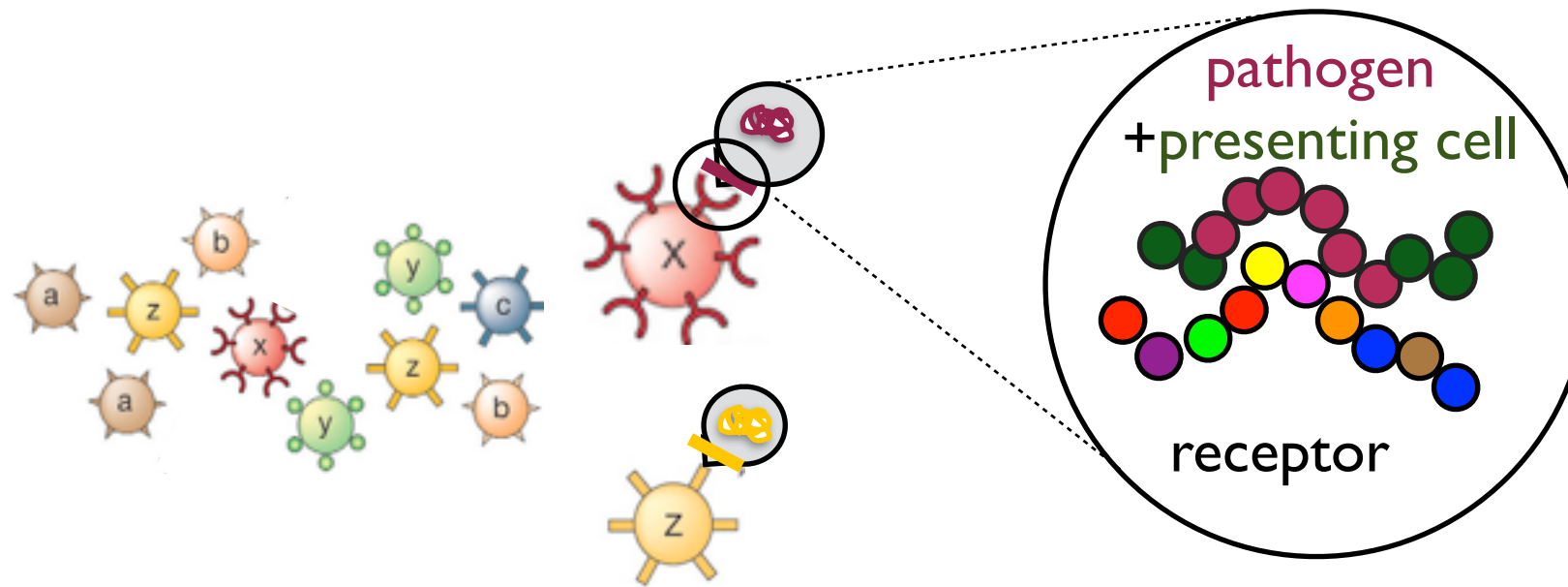
many different pathogens (virus...)

- triggers immune response

Immune receptors



- T-cells important actors of immune system



many unique receptors

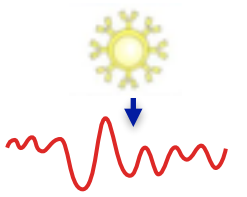


many different pathogens (virus...)

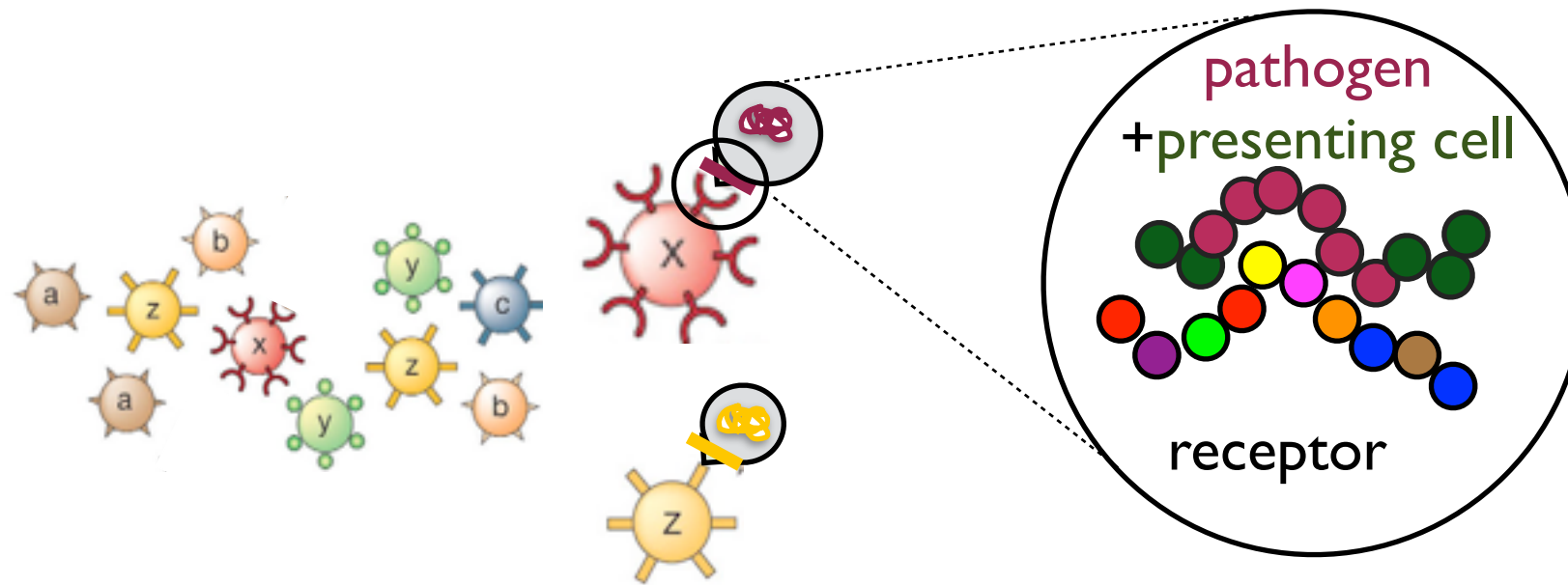
- triggers immune response

natural, healthy (=“normal”) diversity of immune receptors?

Immune receptors



- T-cells important actors of immune system



many unique receptors



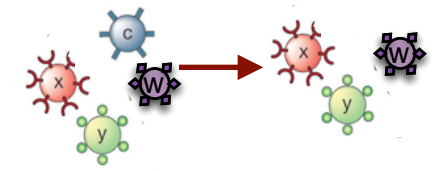
many different pathogens (virus...)

- triggers immune response

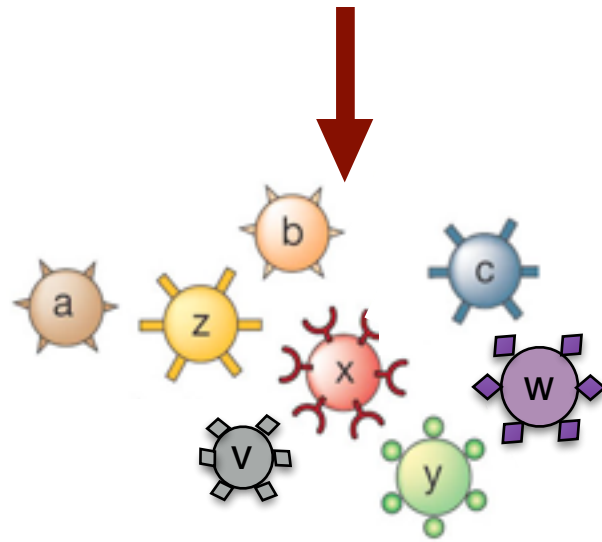
natural, healthy (=“normal”) diversity of immune receptors?

how diverse is it?
predicting repertoires ?

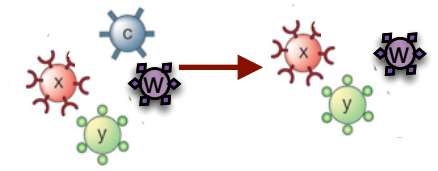
Repertoire evolution



RECEPTOR GENERATION

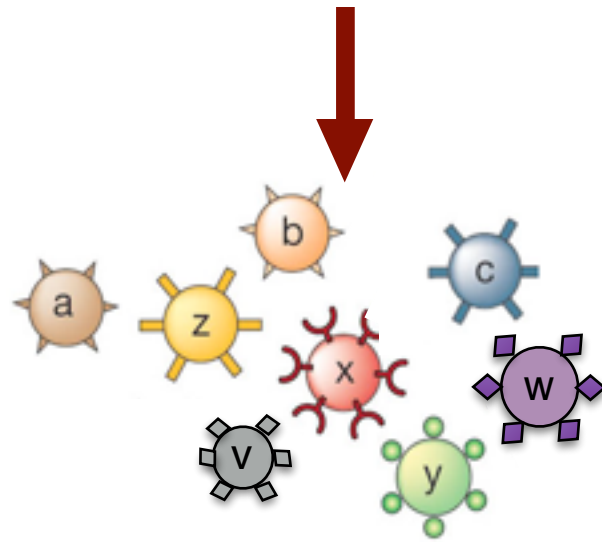


Repertoire evolution

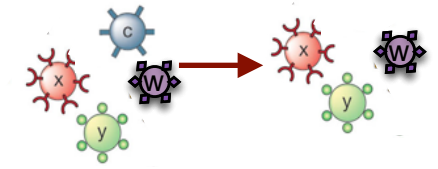


RECEPTOR GENERATION

combinatorics + *randomness* → diversity

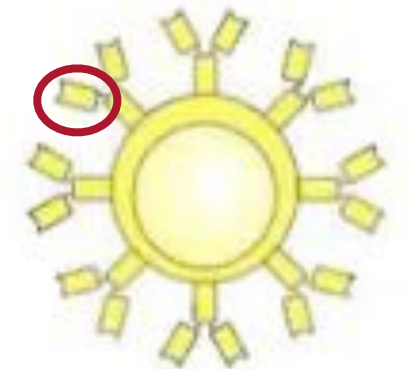
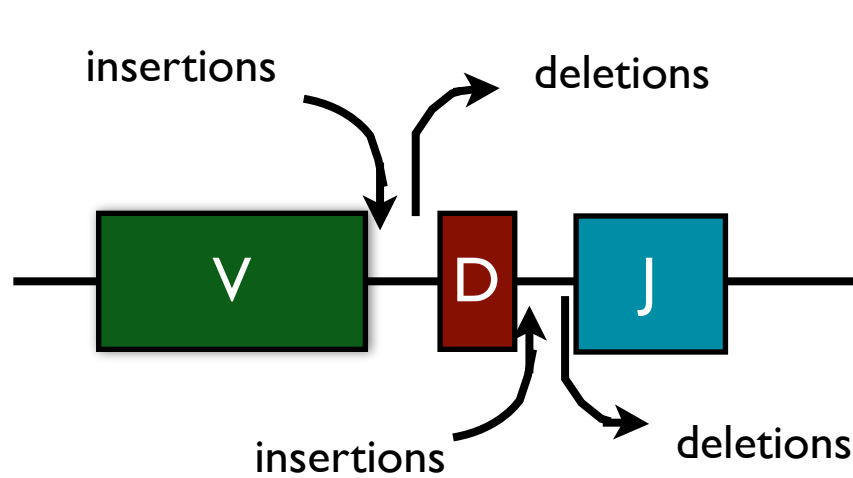
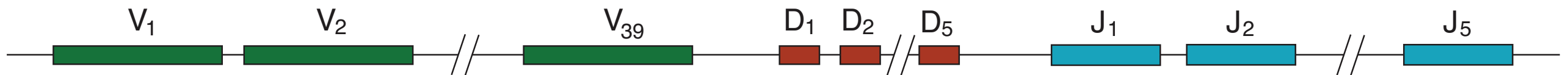
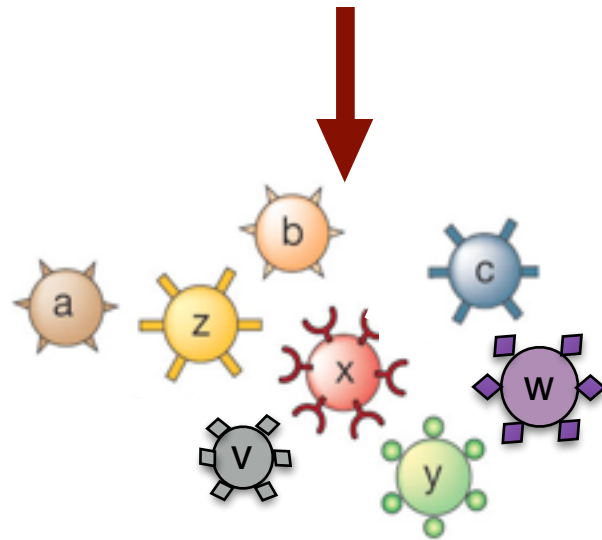


Repertoire evolution



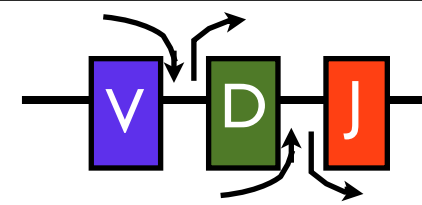
RECEPTOR GENERATION

combinatorics + *randomness* → diversity

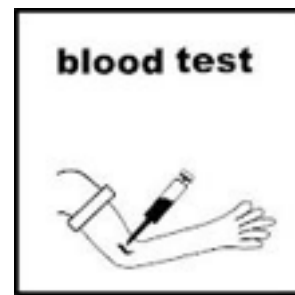


- two chromosomes per cell = two attempts

Sequence data



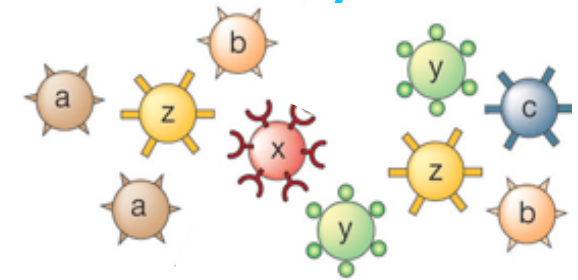
new data



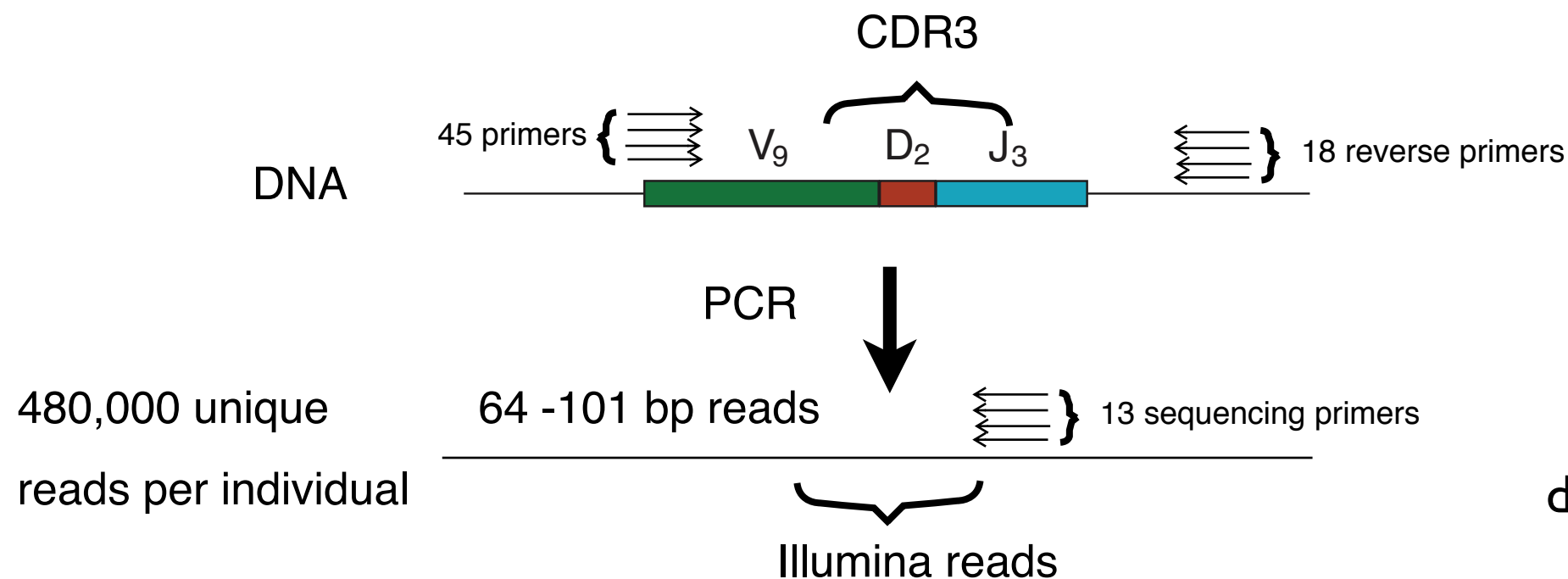
sequencing machine



natural diversity distribution

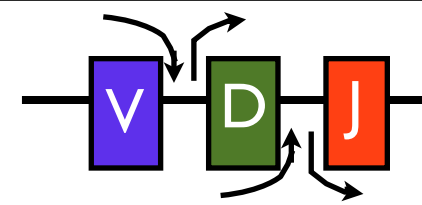


- human T-cell beta chain receptor sequences
- 9 people
- out of frame reads (~14% of each type of cells) → *generation*
- in frame reads (~235,000 unique reads) → *selection*



data from Robins lab and Chudakov lab

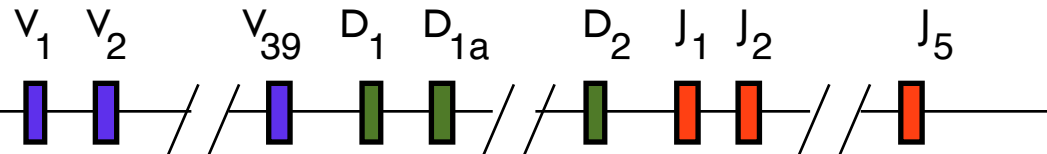
Probabilistic VDJ recombination annotation



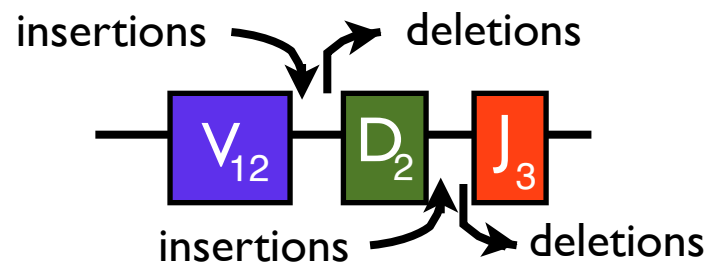
sequence generation:

combinatorics + randomness → diversity

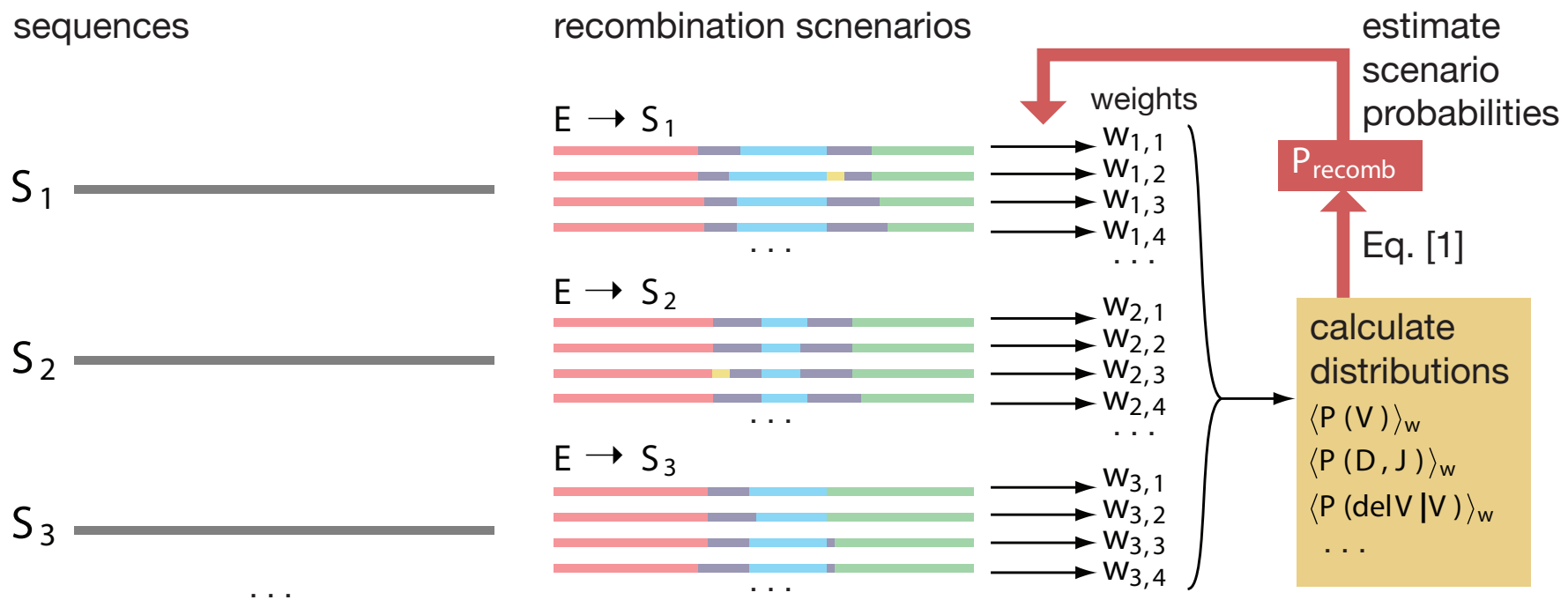
$\vec{\sigma}$ - receptor DNA sequence



```
TAGGACCTCGGAAACCTCTTCTGCTGGCGCAGAGATACAGGGCTGAGTCTTCTT
TATCTGCCGATGTCGCGAAGGCCCTCCCGCTAAGATCACTGGTGGCACAGAAGT
TAGGGAGAGGTGCCTAACTGCTGGCACAGAAGTACAGAGAGGTCTGGTTGGGGT
TCCGCCGCTAGTCCCTGAAACTACTGGCACAGAGATAGAAAGCTGTCGGGTTCT
CGAAACTGCTGGCACAGAAGTACACAGATGTTTGGGAGGGAGCAGCCGACTCCA
TCTTGGCCGCTAGTCCGAGAACTGCTGGCACAGAAGTACACAGATGTTTGGGA
TTGTAGGAGCCGGACCGGCCCTGTCCCTTGGCTGCTGGCGCAGAGATACAG
TCATTTAACGTGCGGCCCGCCTGGCACAGAAGTAAAGAGCTGTCTGGTTGTGGT
TAGTAACTCCGCTTCACTGCTGGCACAGAAGTACACAGATGTCTGGGAGGGAGC
TCCCTCCGGTTTGAAGGGTCTGCTGGCACAGAAGTACACAGATGTTTGGGAGGG
CCGGTGTTCGCACAGCCCTGGGGACCCTGGCGCAAACCCCGCTTCCCTCGAGGA
```



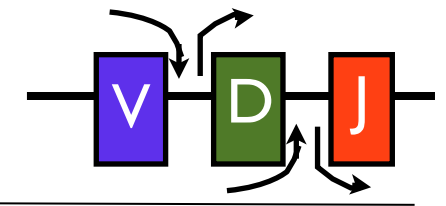
- probabilistic assignment of:



$$P^{\text{recomb}}(\text{scenario}) = P(V)P(D, J)P(\text{deletions } V|V)P(\text{insertions } DJ)...$$

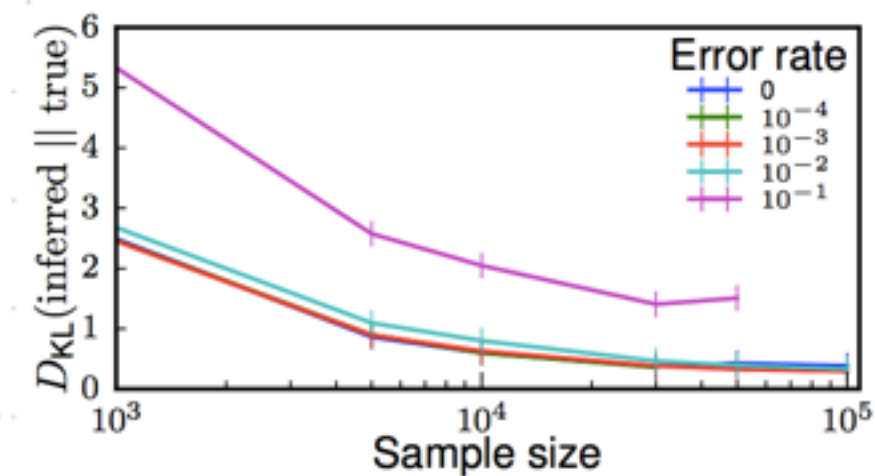
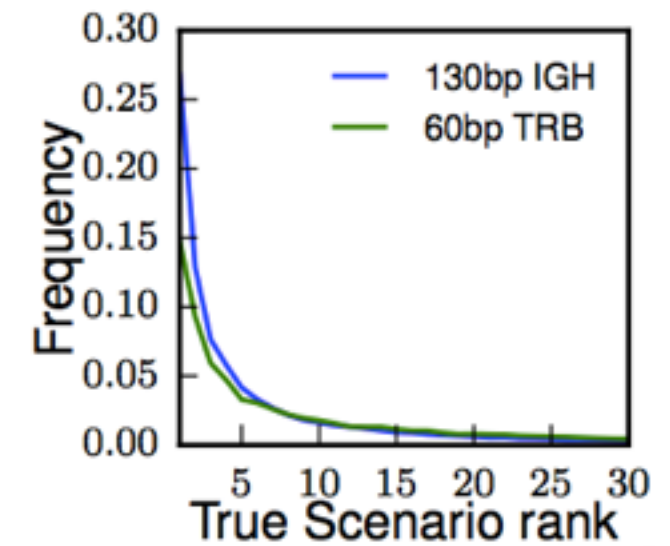
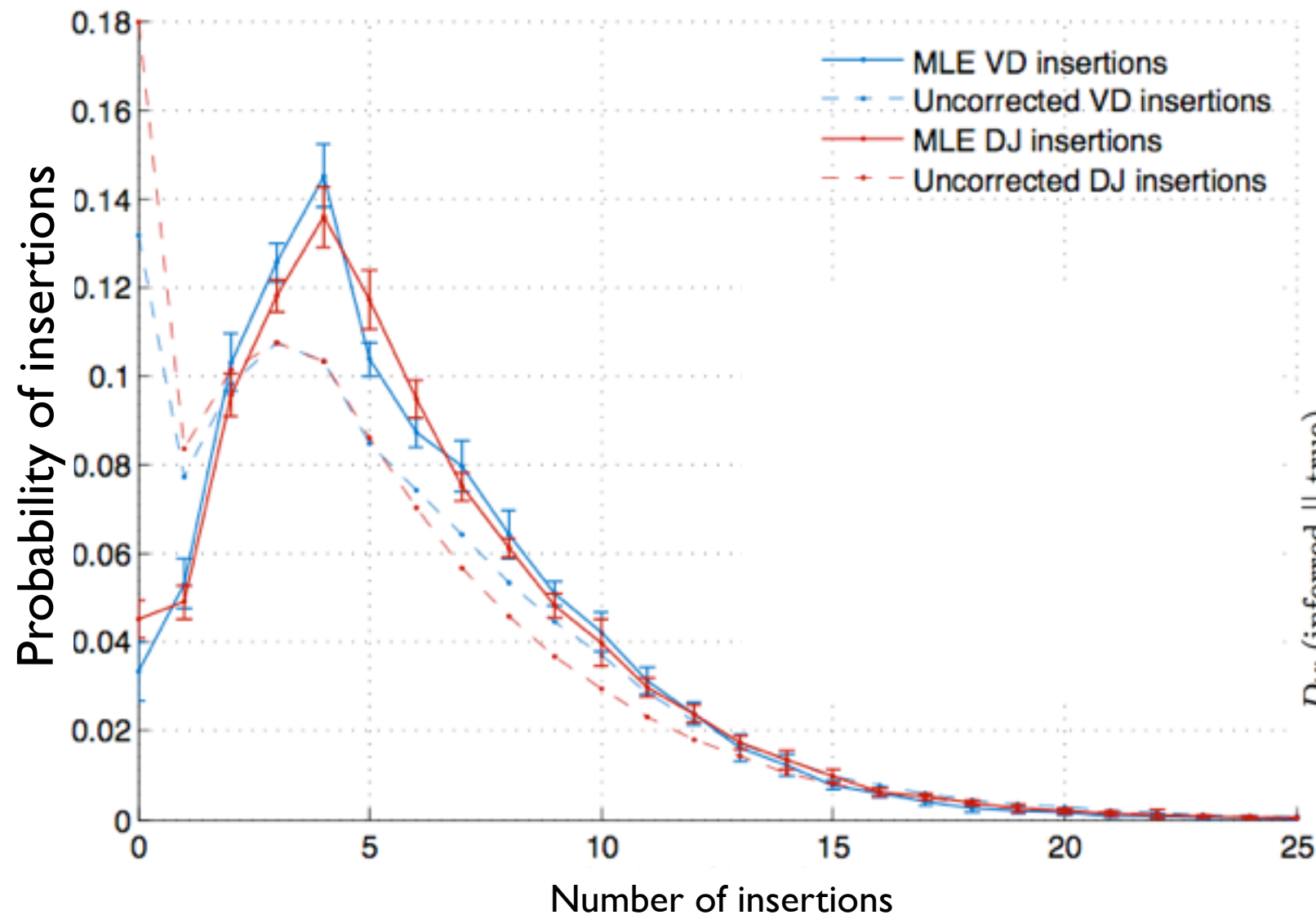
[1]

Universal insertion profiles

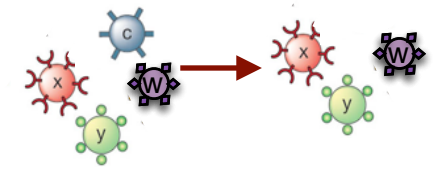


→ universal mechanism for generation

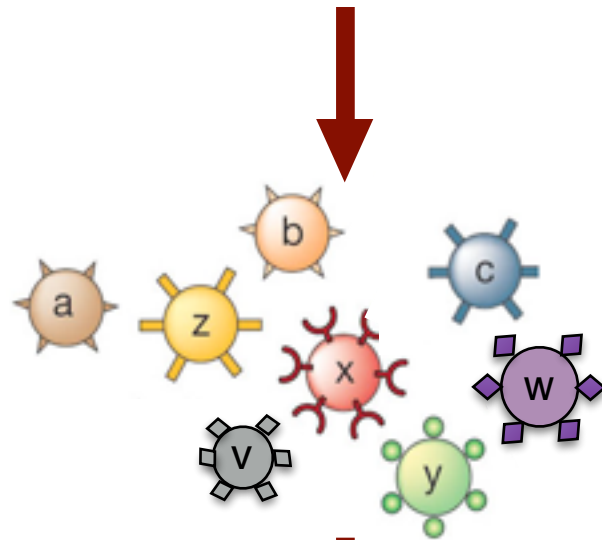
- VD and DJ insertion profiles are identical
- real scenario most likely in less than 30% of cases
- need < 2 ml of blood to learn model



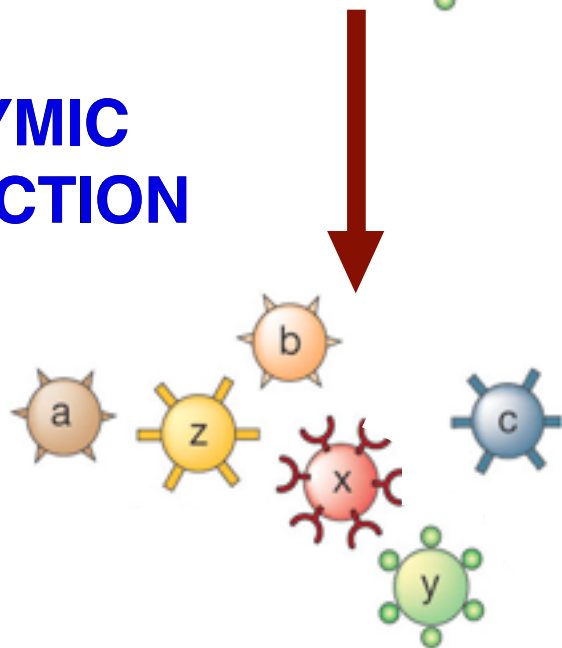
Next step: selection



RECEPTOR GENERATION



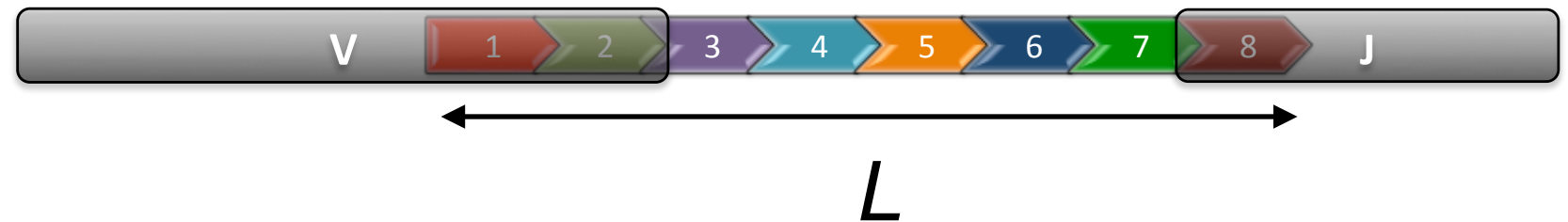
THYMIC SELECTION



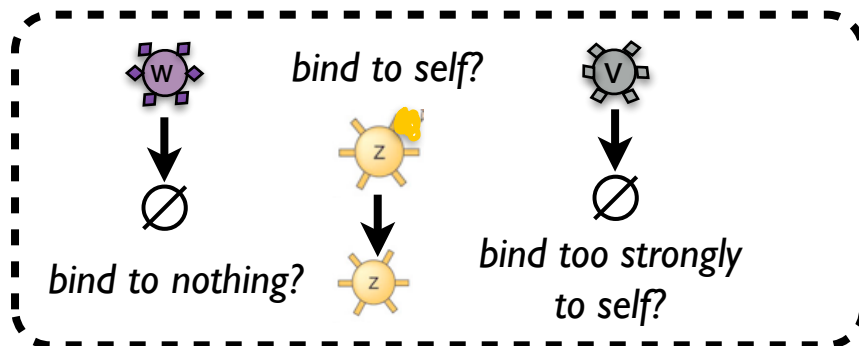
- quantify using selection factors

$$Q(\{\sigma\}) = \frac{P_{\text{post-sel}}(\{\sigma\})}{P_{\text{gen}}(\{\sigma\})}$$

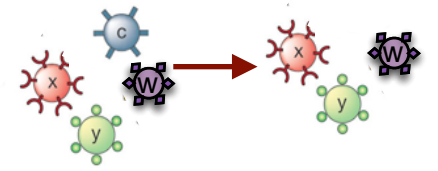
- a model for the observed probabilities



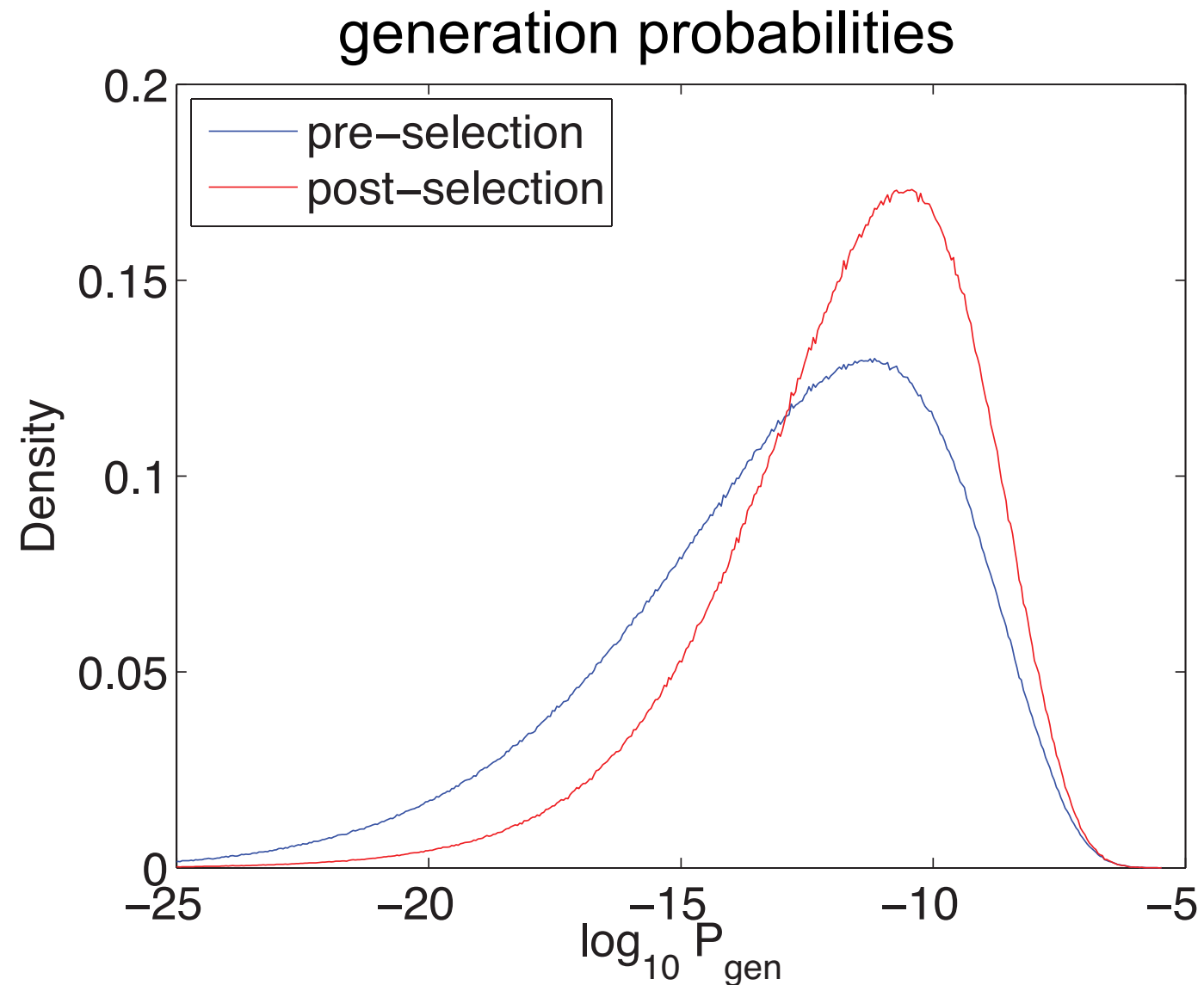
$$Q(\vec{\sigma}, V, J) = \frac{1}{Z} q_L q_{V,J|L} \prod_{i=1}^L q_{i;L}(a_i)$$



Natural selection anticipates somatic selection

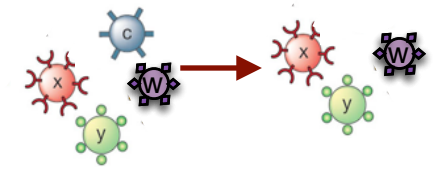


- sequences more likely to be *generated* → more likely to be *selected*



- true for all individuals independently

Receptor sharing

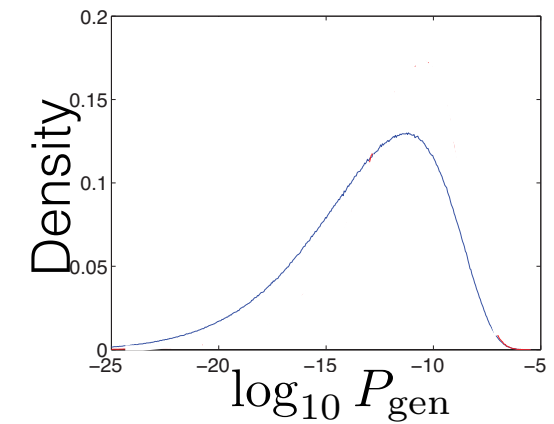


human beta TCR data from Robins lab

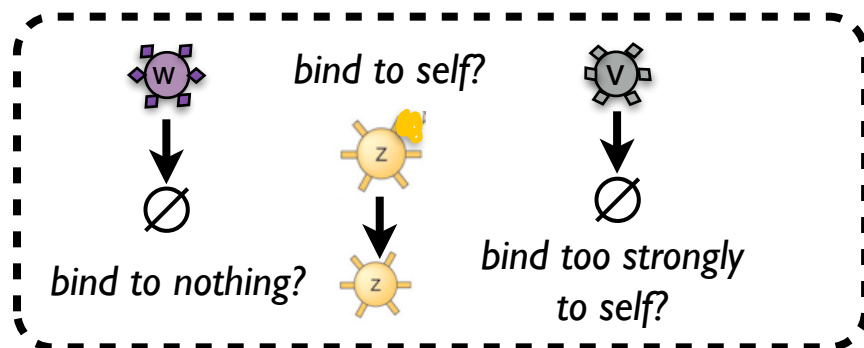
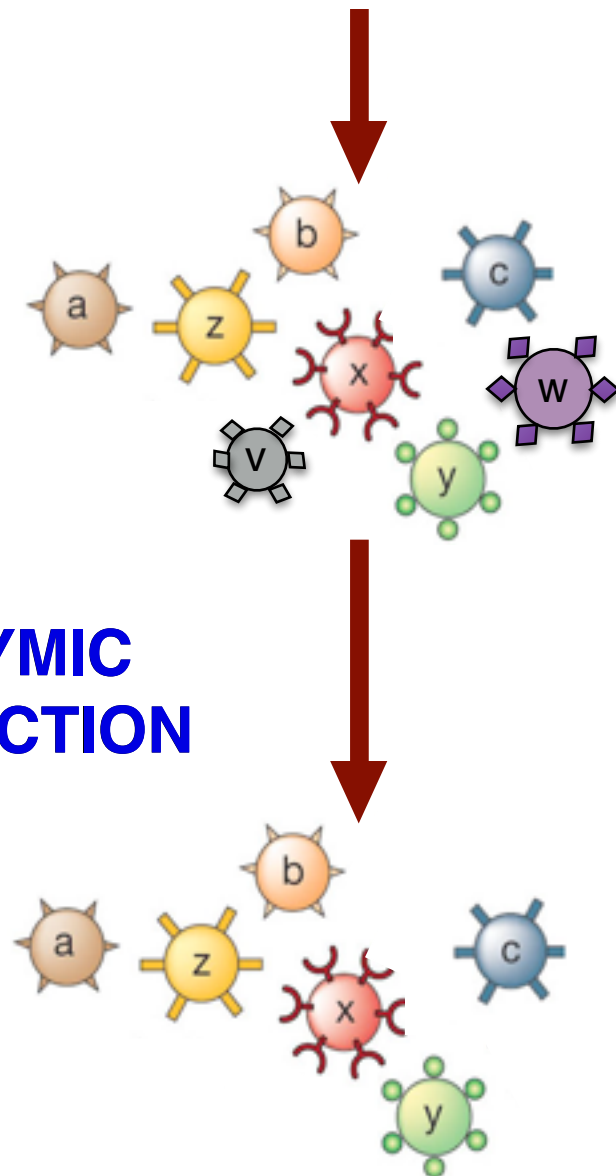
RECEPTOR GENERATION

- quantify using selection factors

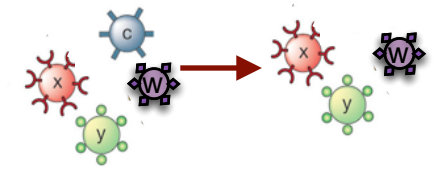
$$Q(\{\sigma\}) = \frac{P_{\text{post-sel}}(\{\sigma\})}{P_{\text{gen}}(\{\sigma\})}$$



THYMIC SELECTION

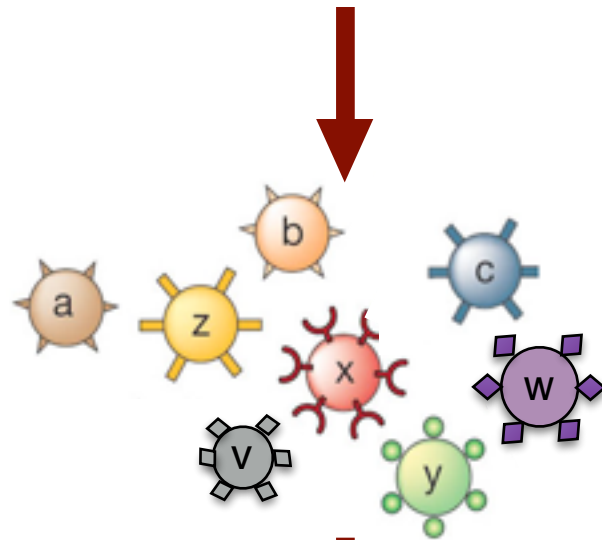


Receptor sharing



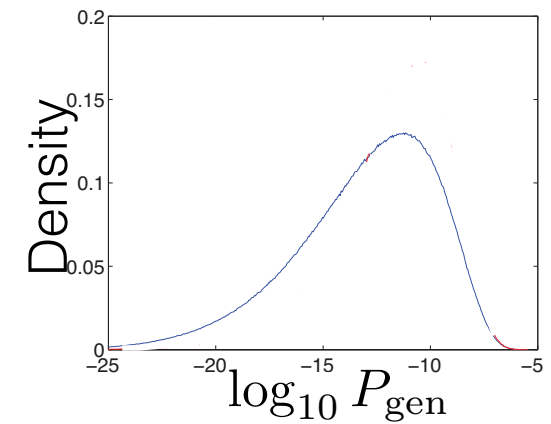
human beta TCR data from Robins lab

RECEPTOR GENERATION



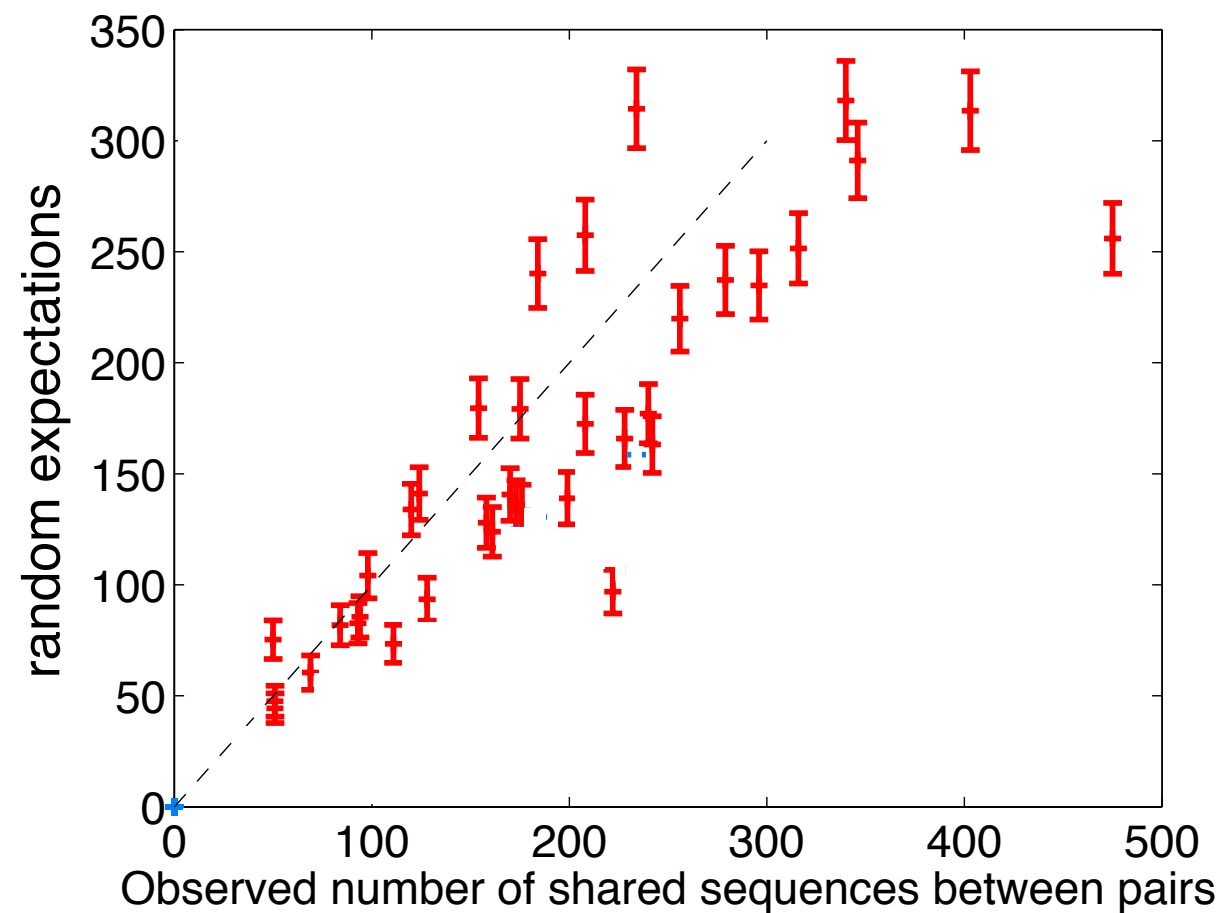
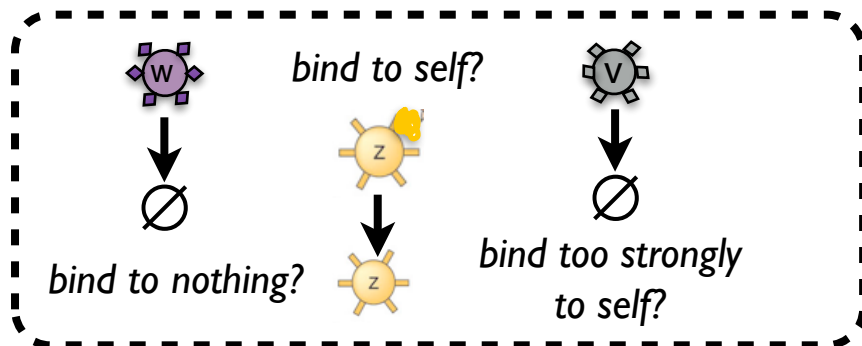
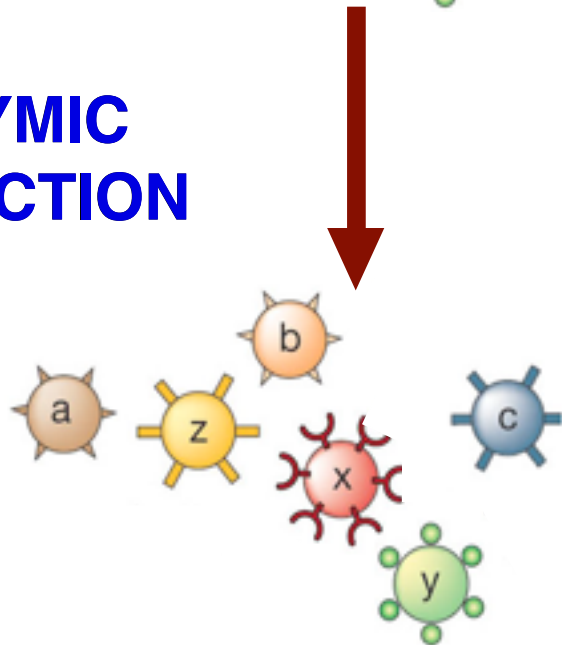
- quantify using selection factors

$$Q(\{\sigma\}) = \frac{P_{\text{post-sel}}(\{\sigma\})}{P_{\text{gen}}(\{\sigma\})}$$



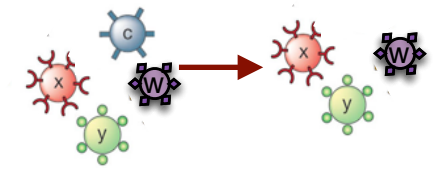
- how many shared receptors between 2 people?

THYMIC SELECTION

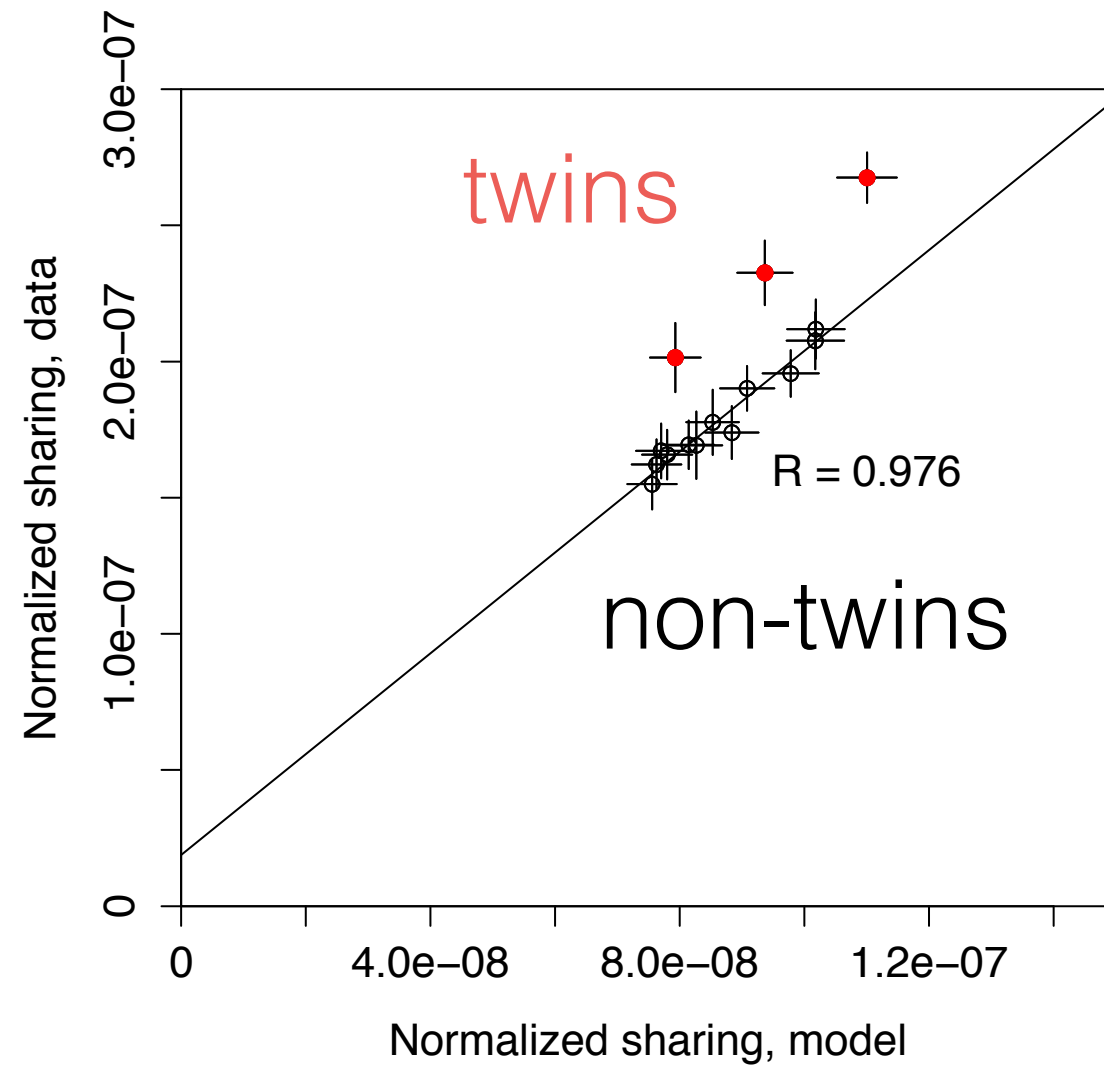


→ close to random expectations

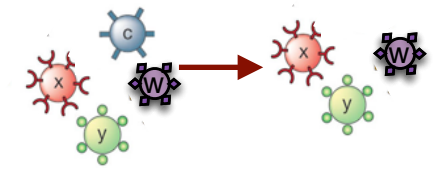
Is everyone the same?



at the level of generation



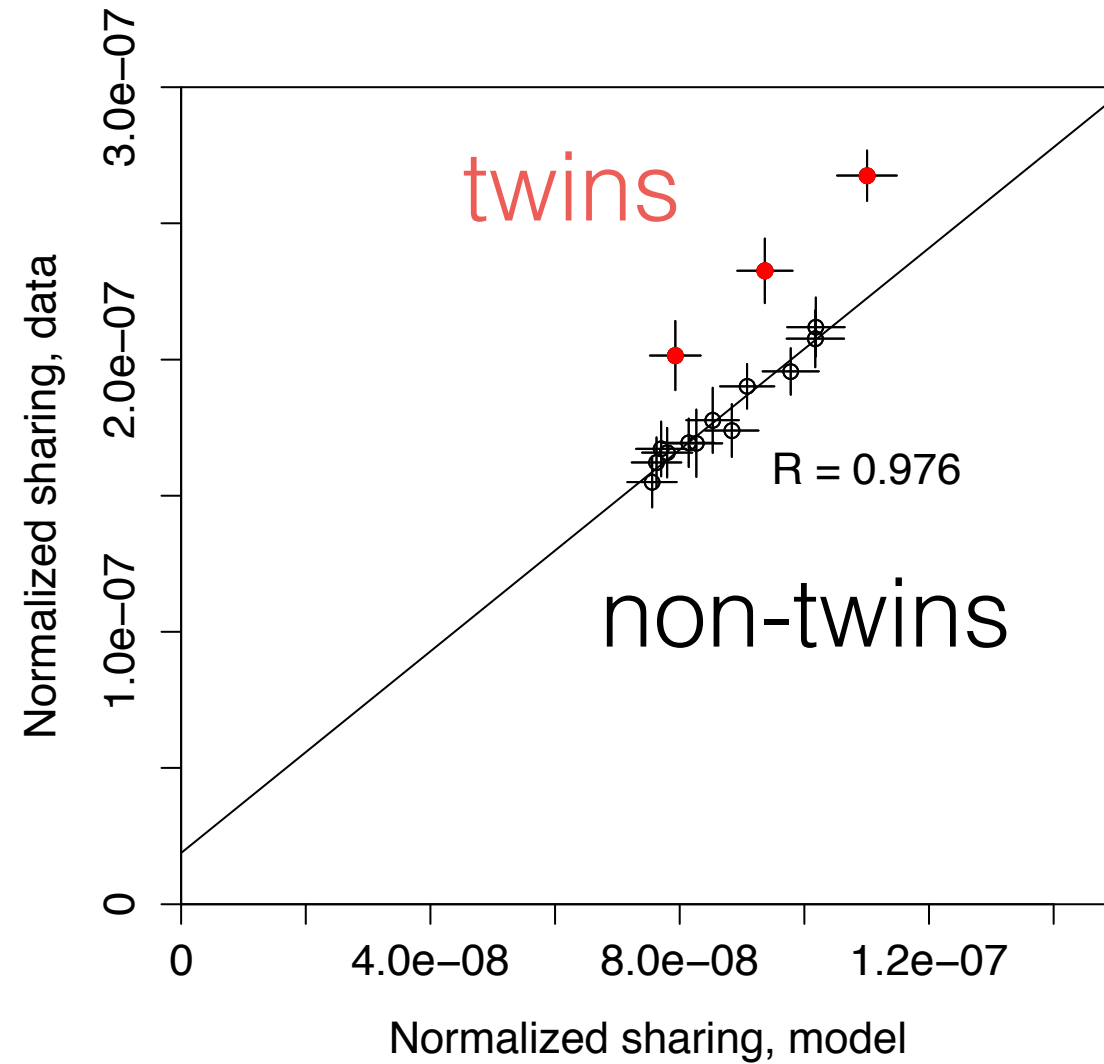
Is everyone the same?



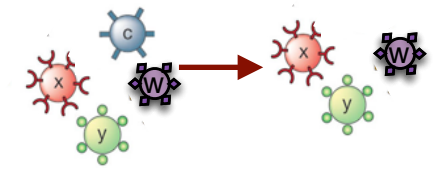
at the level of generation



twins are special



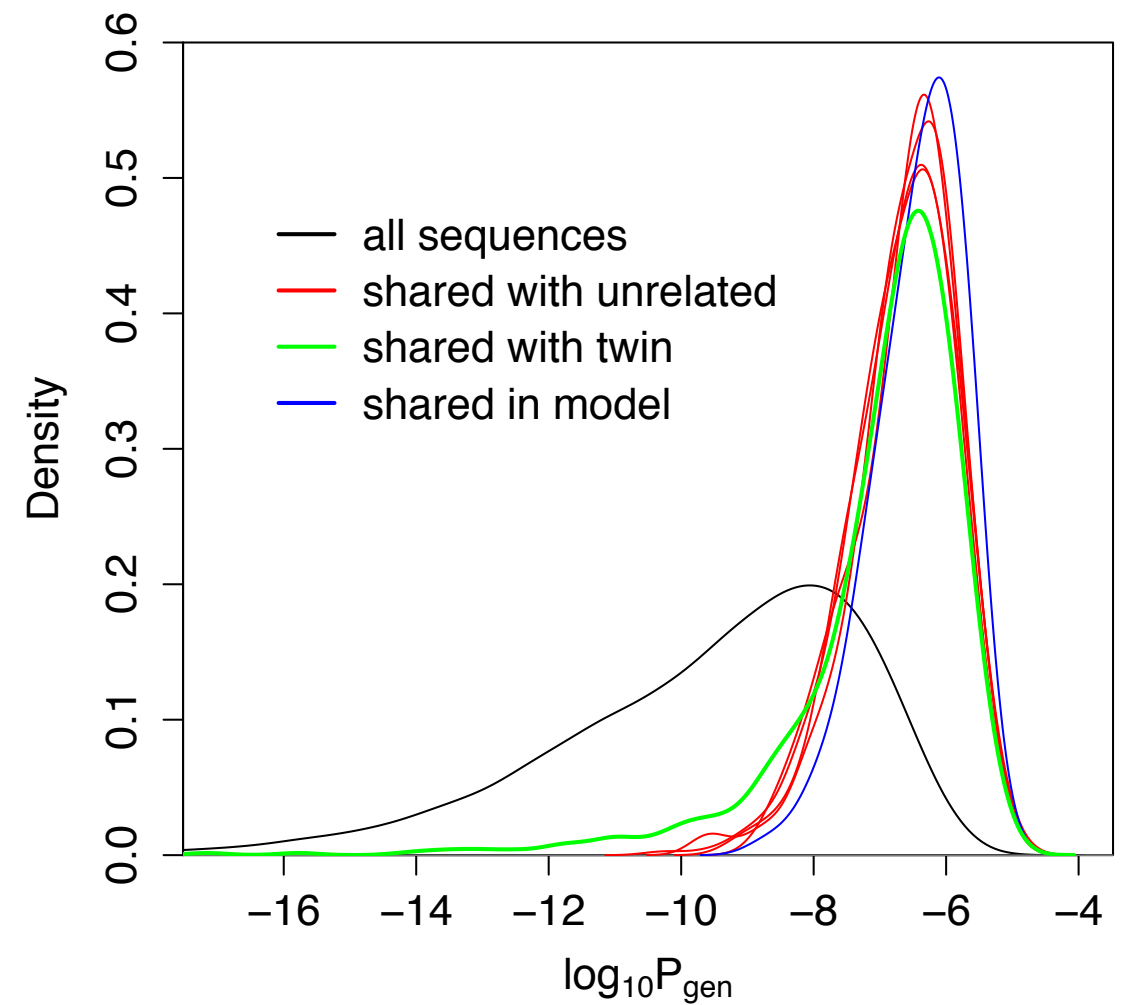
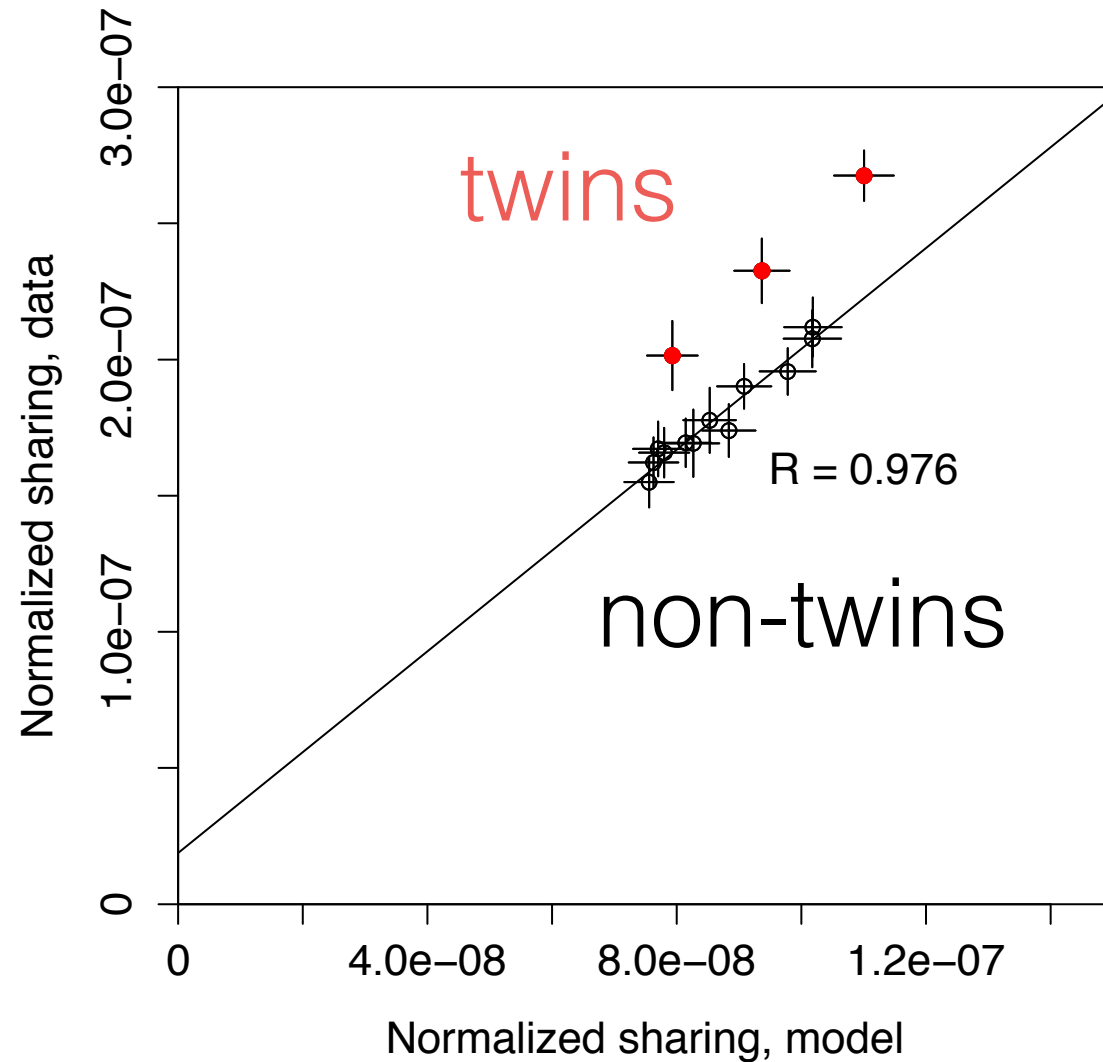
Is everyone the same?



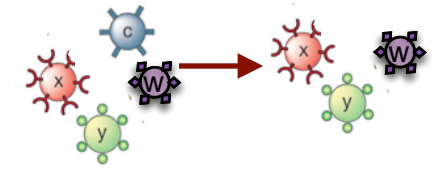
at the level of generation



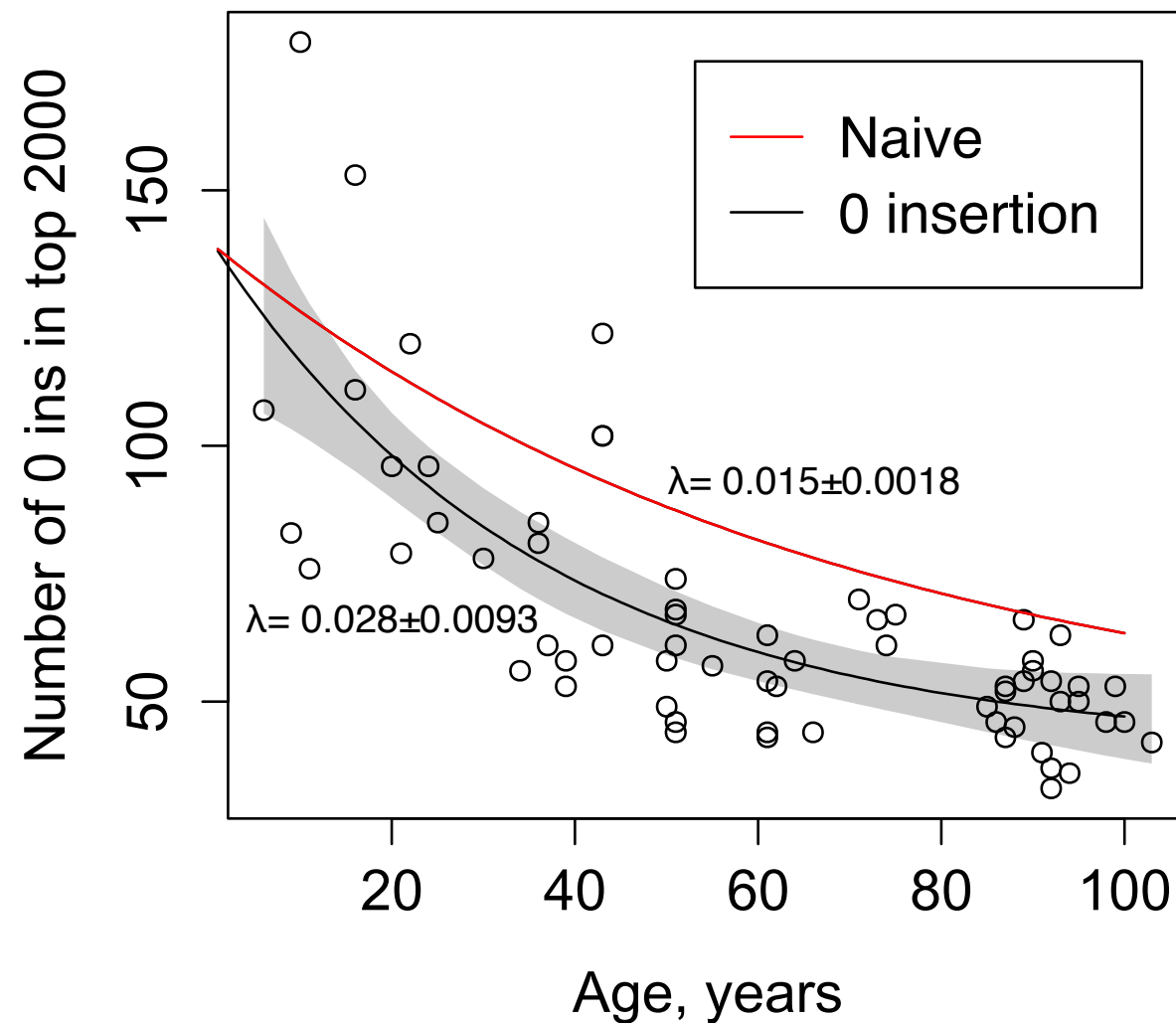
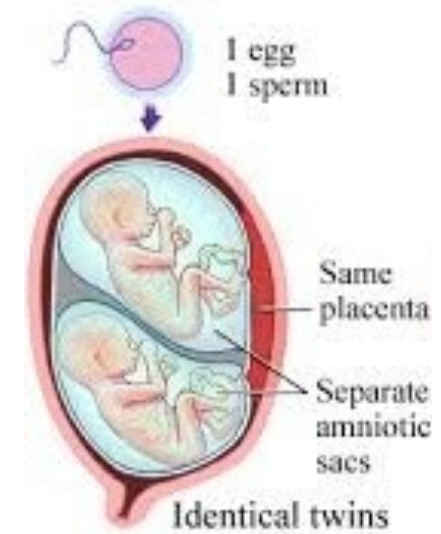
twins are special



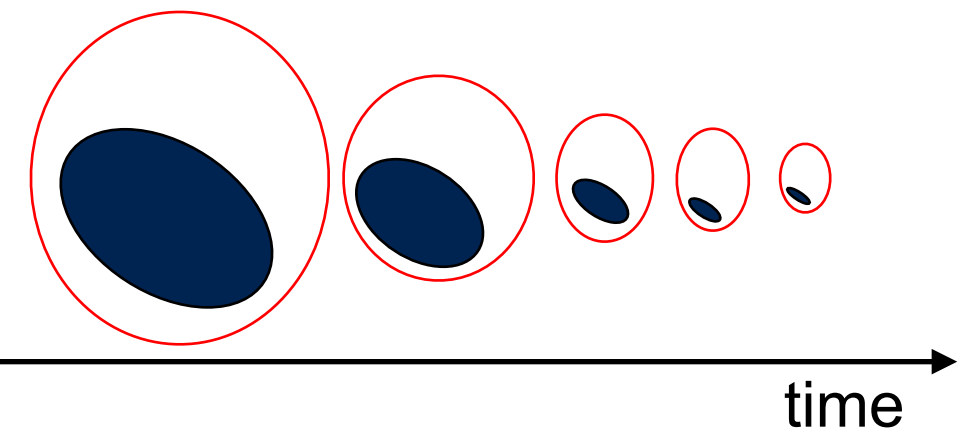
The source: long lived sequences



- last time twins shared blood: before birth
- insertions enzyme less active before birth



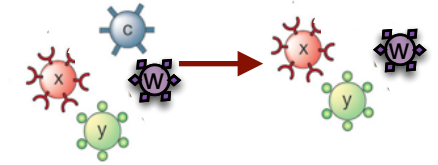
clone lifetime ~ 36 years



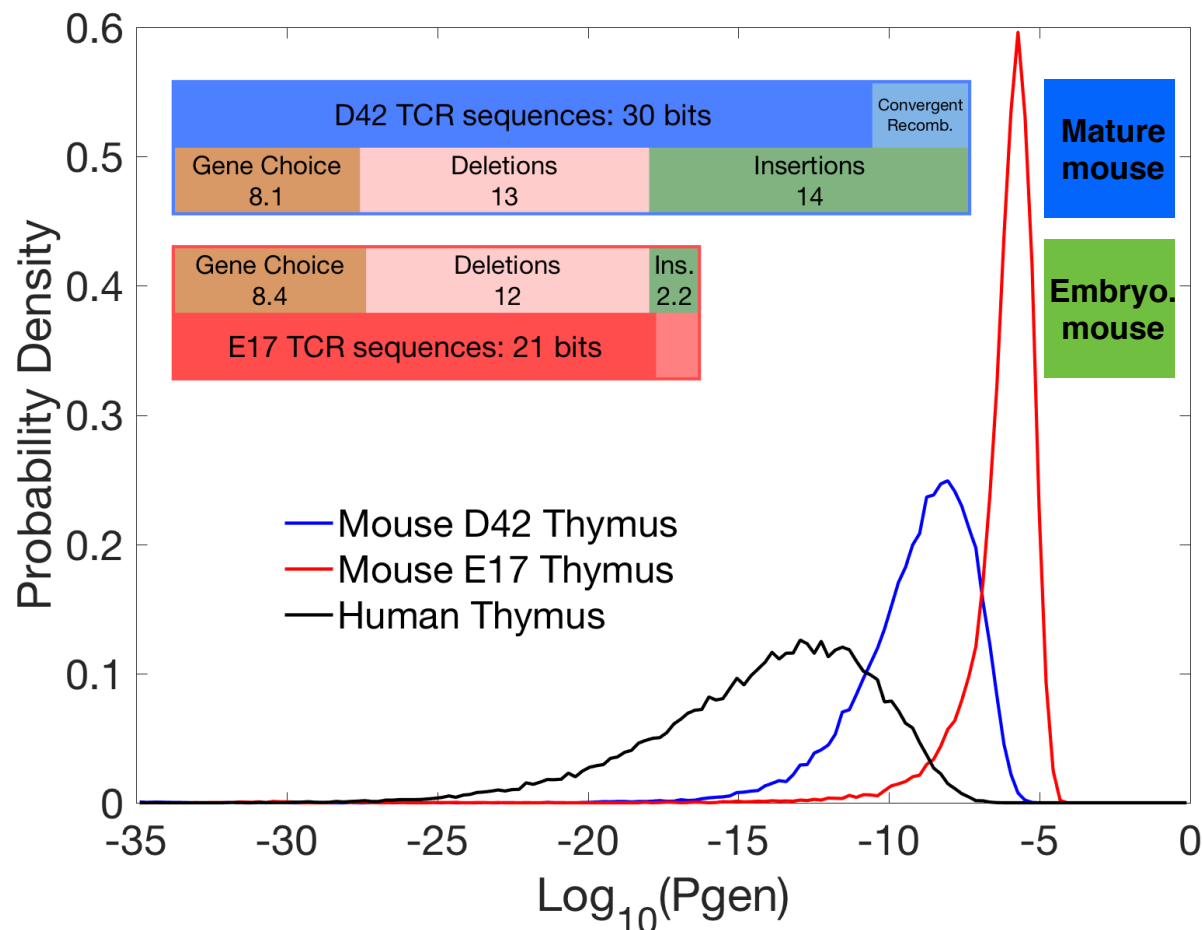
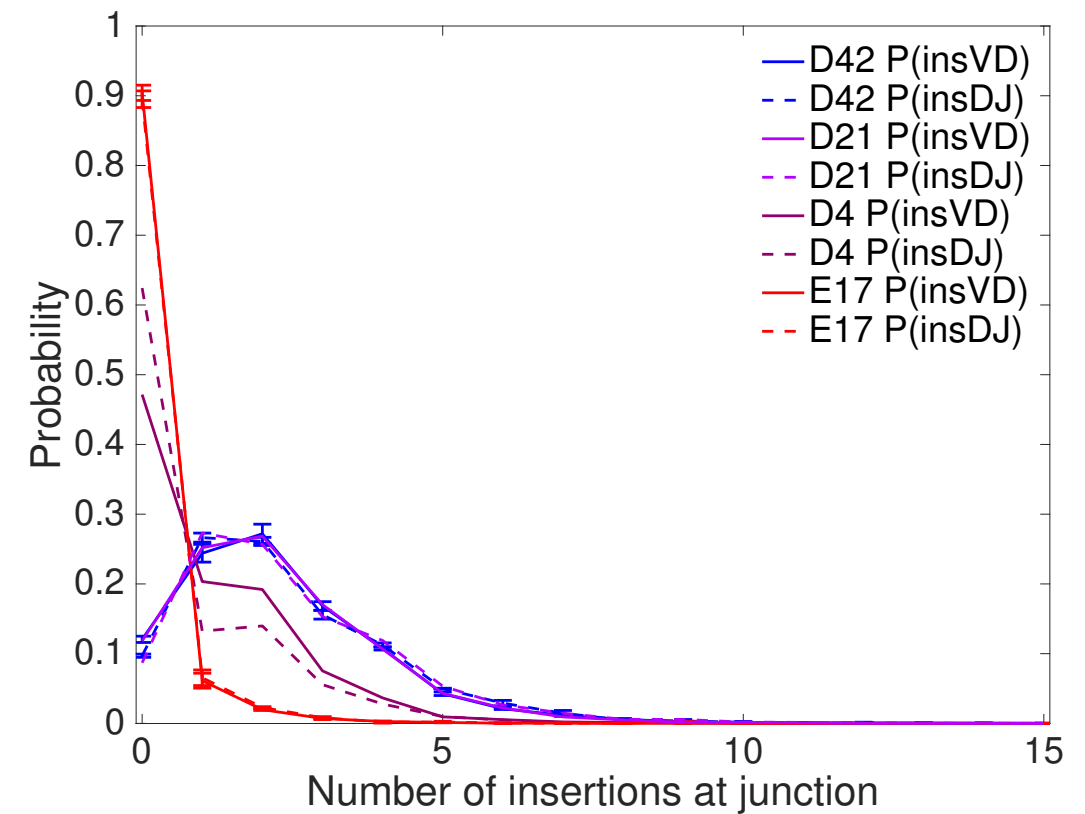
Decay of zero insertion clonotypes:

- zero insertion clonotypes within the naive pool
- size of the total naive pool

Insertions increase diversity

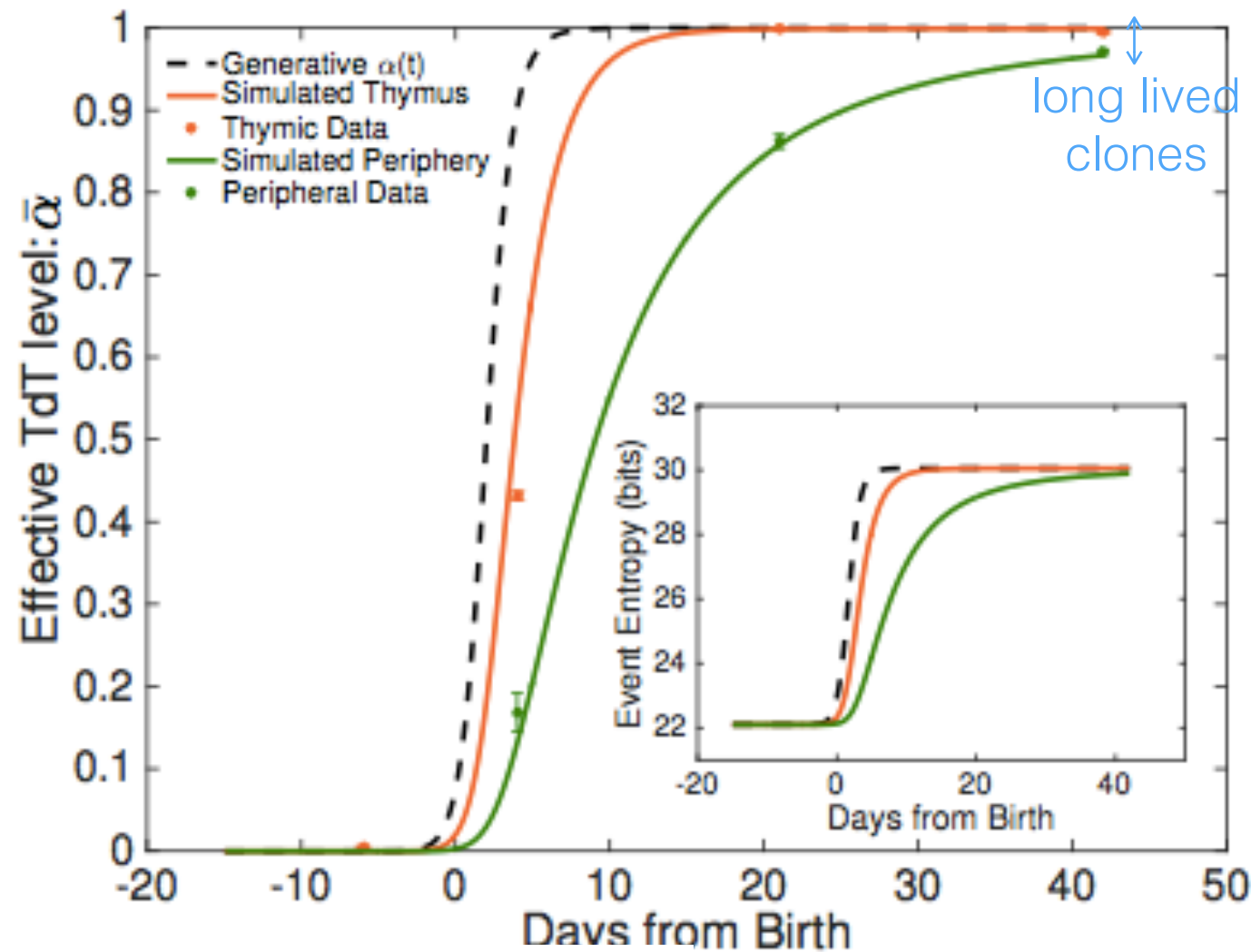
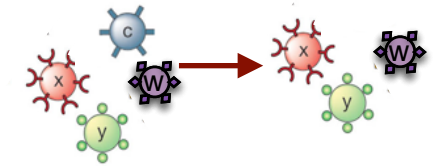


- mouse thymus T-cells
- insertions quickly increase during post-natal development
- diversity increase only due to insertions

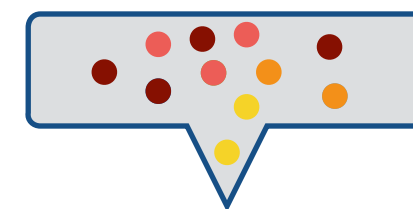


mice: Levine lab
sequencing: Robins lab

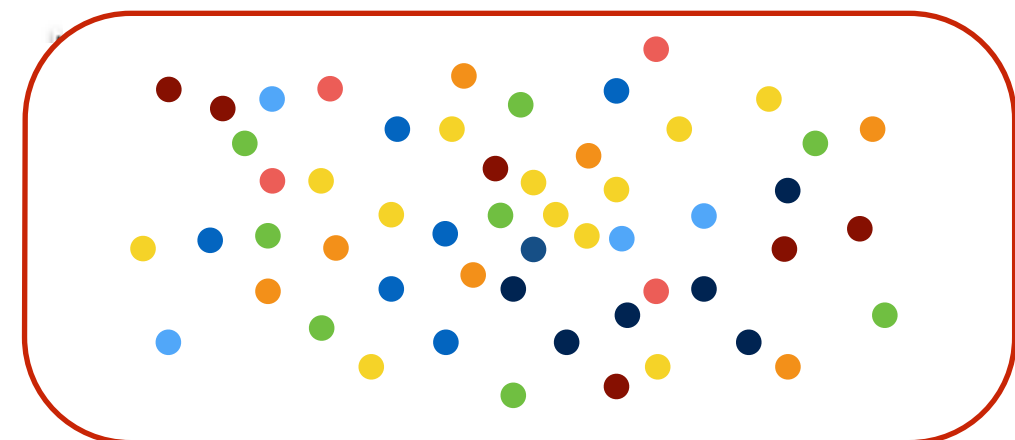
Blood tracks thymus



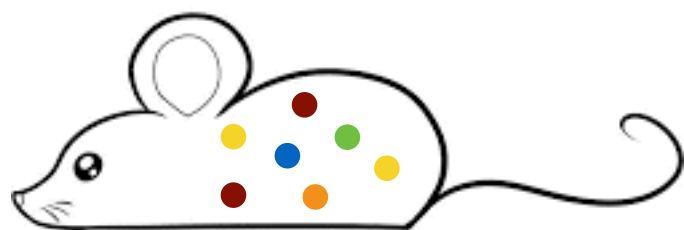
- blood is delayed with respect to thymus
- blood integrates different TdT levels



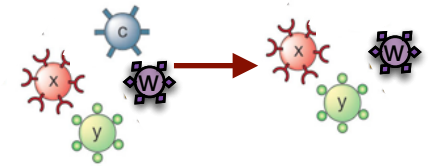
thymus



blood

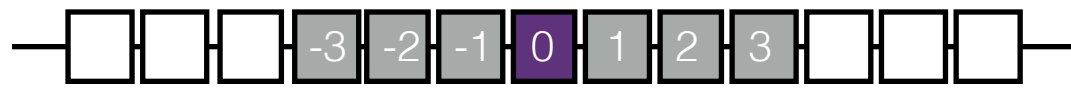


Somatic hypermutations



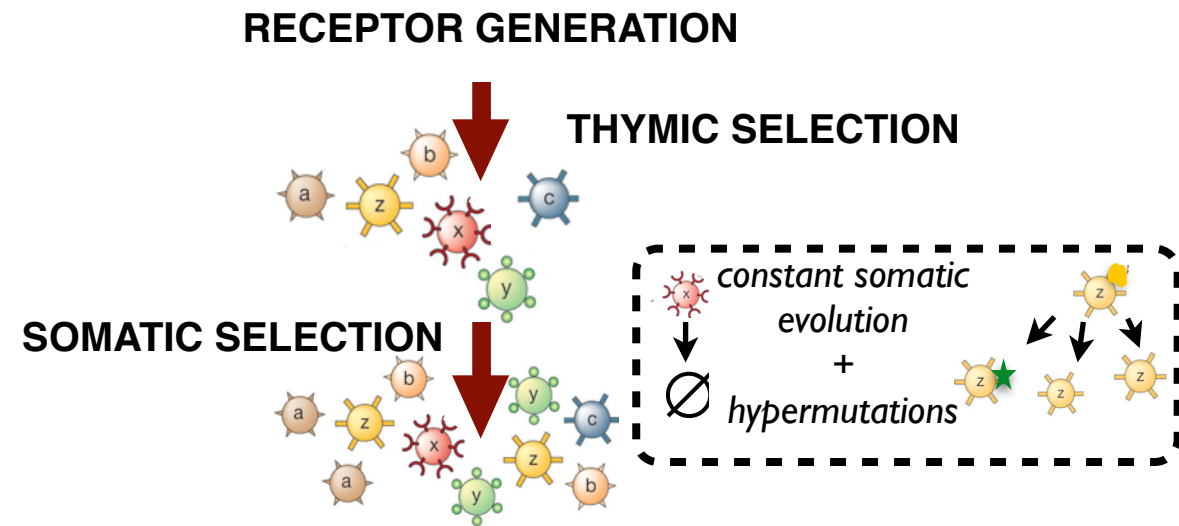
- identifying hypermutation hotspots

7-mer



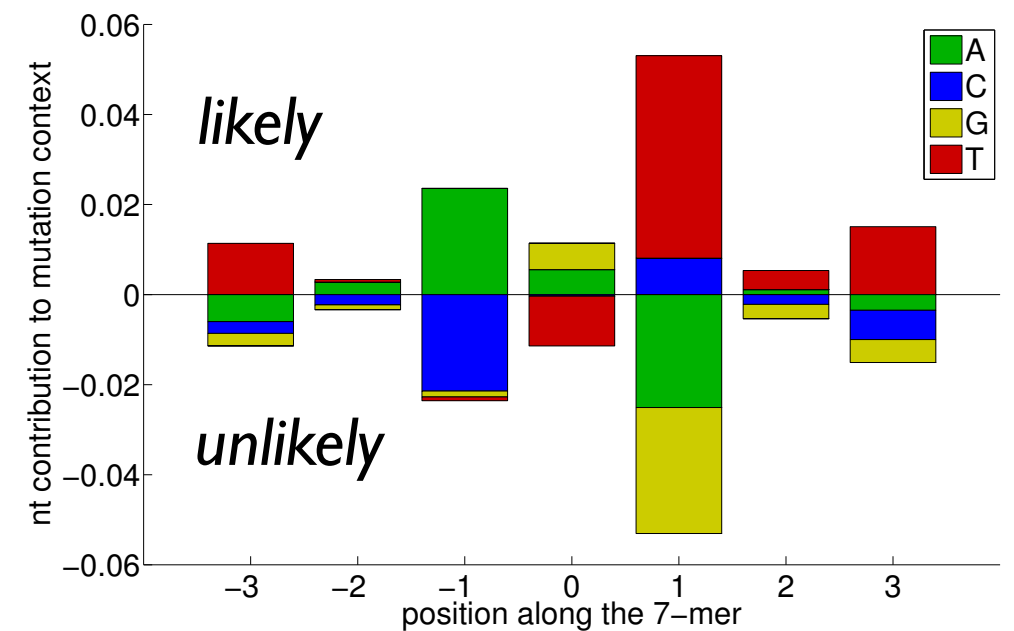
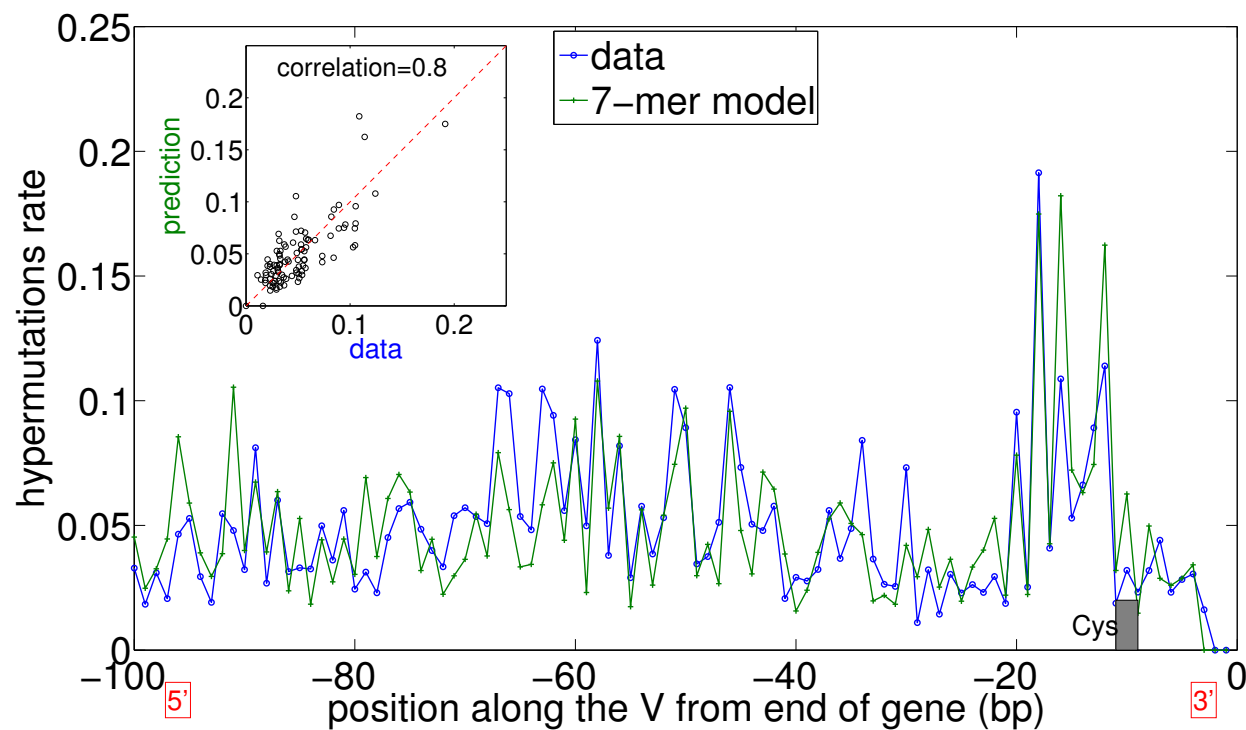
$$p_{\text{SHM}}(\sigma) \propto p_{\text{bg}}(\sigma) \exp\left[\sum_{i=-3,3} e_i(\sigma_i)\right]$$

background probability
of that 7-mer

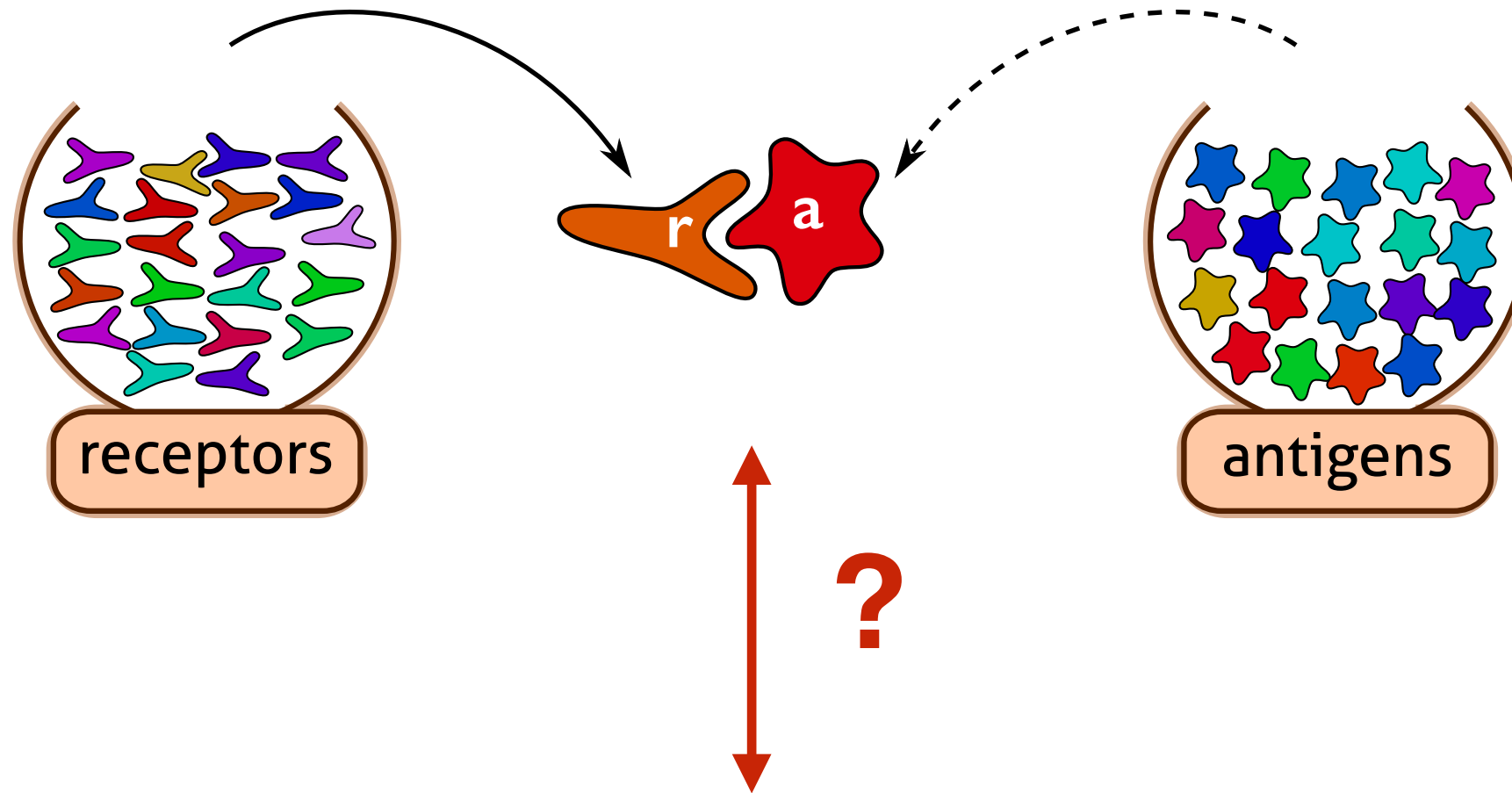


- nt that are likely to hypermutate

hypermutation hotspot 7-mer signature



Linking sequence to function

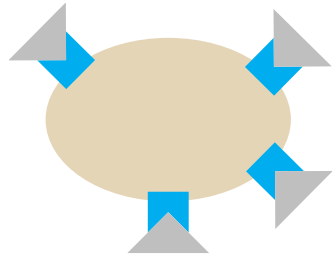


what matters

what we have

```
TAGGACCTCGGAAACCTCTTCTGCTGGCGCAGAGATACAGGGCTGAGTCTTCTT
TATCTGCCGATGTCGCGAAGGCCCTCCCGCTAAGATCACTGGTGGCACAGAAGT
TAGGGAGAGGTGCCTAACTGCTGGCACAGAAGTACAGAGAGGTCTGGTTGGGGT
TCCGCCGCTAGTCCCTGAAACTACTGGCACAGAGATAGAAAGCTGTCGGGTTCT
CGAAACTGCTGGCACAGAAGTACACAGATGTTTGGGAGGGAGCAGCCGACTCCA
TCTTGGCCGCTAGTCCGAGAAACTGCTGGCACAGAAGTACACAGATGTTTGGGA
TTGTAGGAGCCGGACCGGCCCTGTCCCCTTGGCTGCTGGCGCAGAGATACAG
TCATTTAACGTGCGGCCCGCCTGGCACAGAAGTAAAGAGCTGTCTGGTTGTGGT
TAGTAACTCCGCTTCACTGCTGGCACAGAAGTACACAGATGTCTGGGAGGGAGC
TCCCTCCGGTTTGAAGGGTCTGCTGGCACAGAAGTACACAGATGTTTGGGAGGG
CCGGTGTTCGCACAGCCCTGGGGACCCCTGGCGCAAACCCCGCTTCCCTCGAGGA
```

Tite-Seq



yeast display

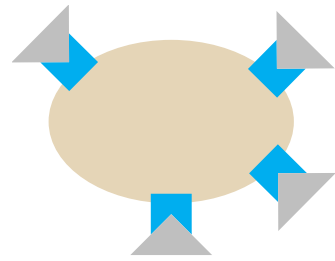
+

FACSort

+

sequence

Tite-Seq



yeast display

+

FACSort

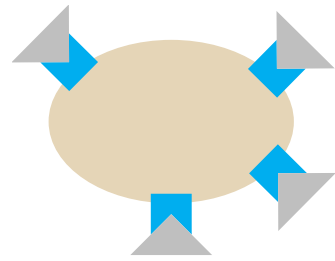
+

sequence

- proof of principle: antigen = fluorescein
antibody = scFv

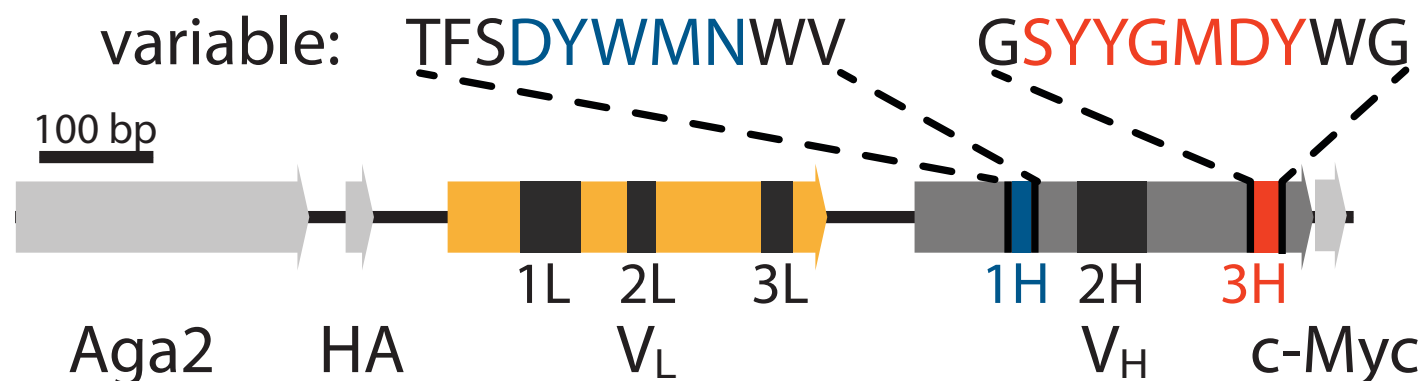
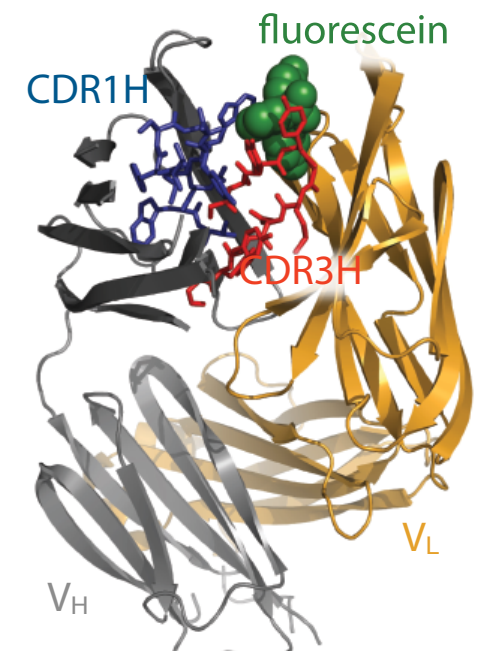


Tite-Seq



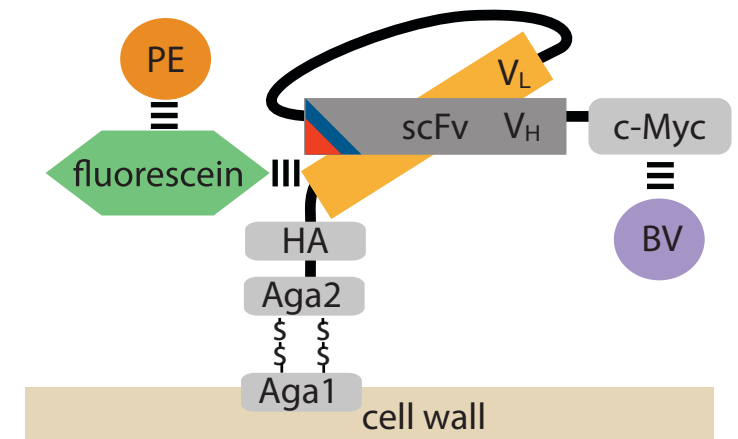
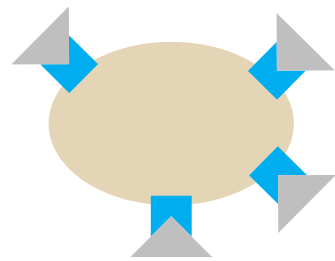
yeast display + FACSort + sequence

- proof of principle: antigen = fluorescein
antibody = scFv
- exhaustive single mutations in CDR1H and CDR3H



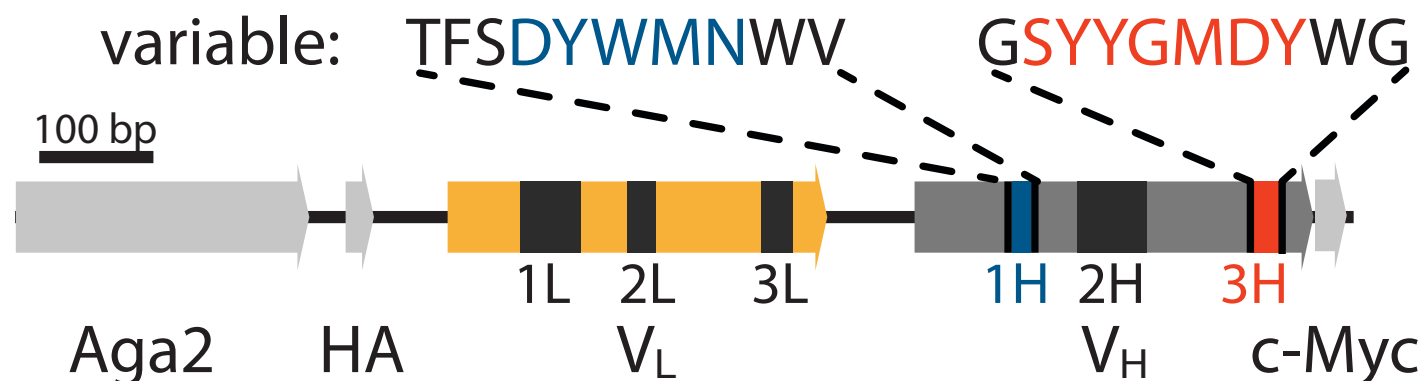
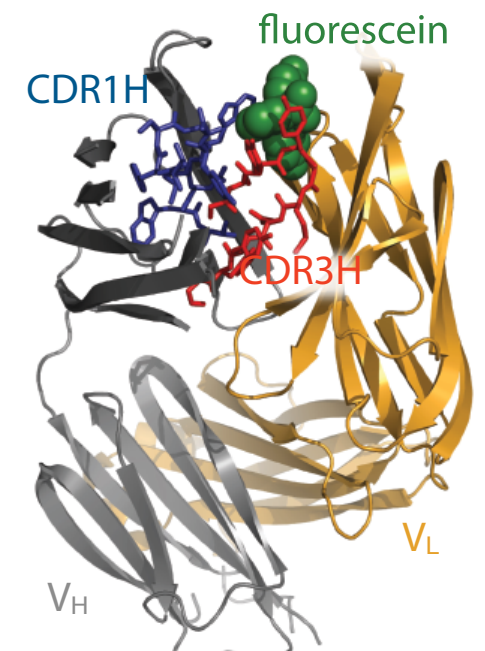
variation	1H	3H
1 codon	600	600
2 codon	1100	1100
3 codon	150	150

Tite-Seq



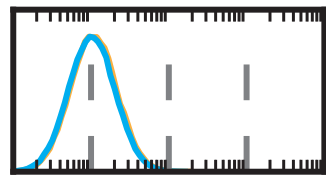
yeast display + FACSort + sequence

- proof of principle: antigen = fluorescein
antibody = scFv
- simultaneously measure affinity and expression
- exhaustive single mutations in CDR1H and CDR3H

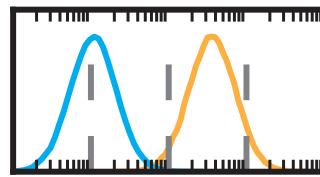


variation	1H	3H
1 codon	600	600
2 codon	1100	1100
3 codon	150	150

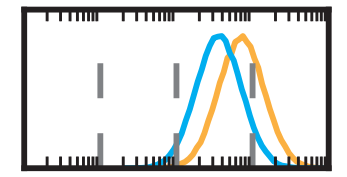
Tite-Seq



fluorescence

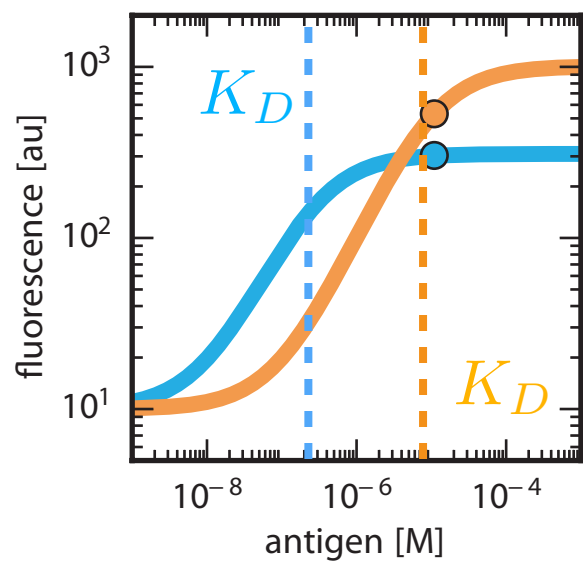


fluorescence

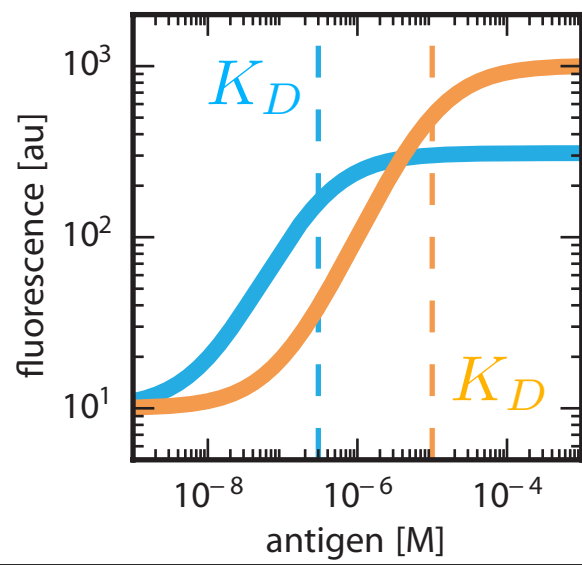
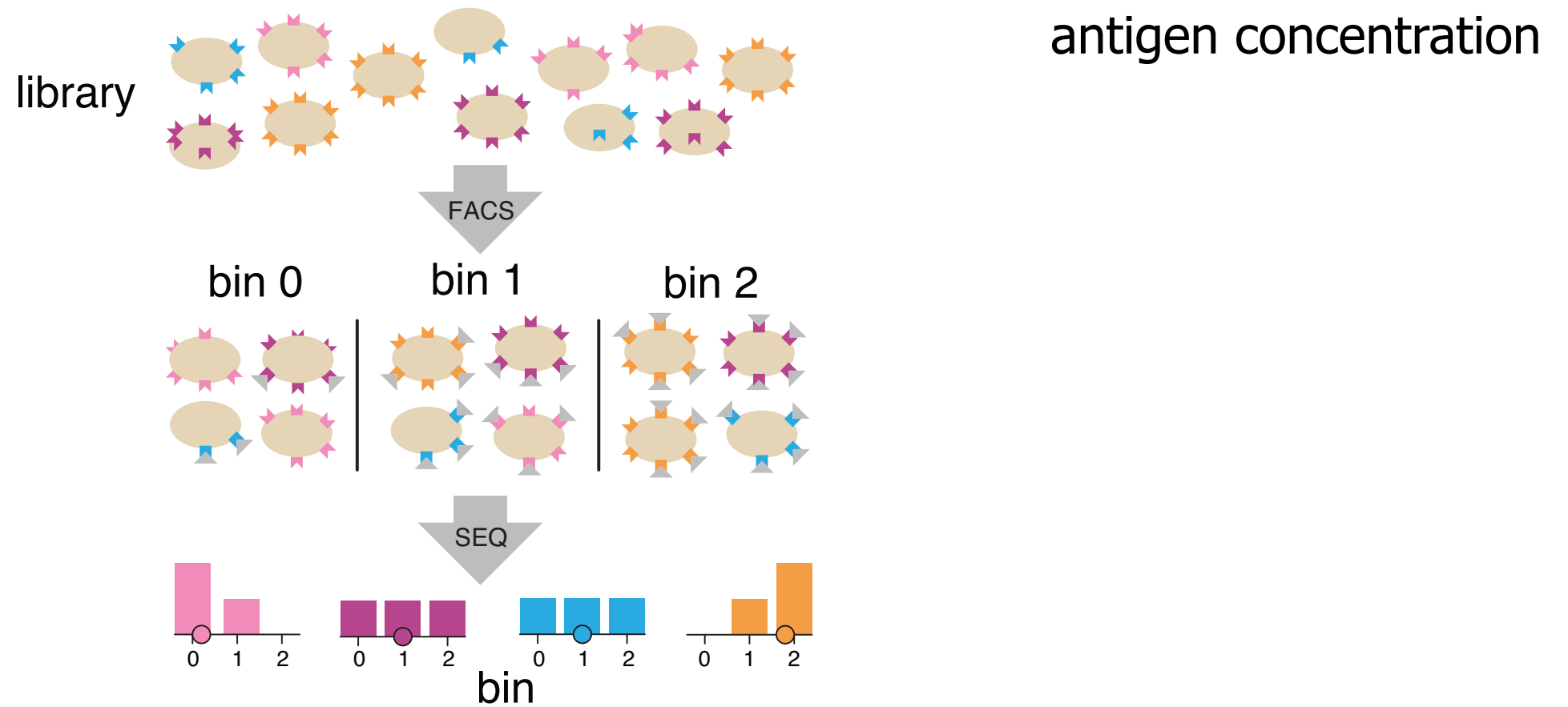
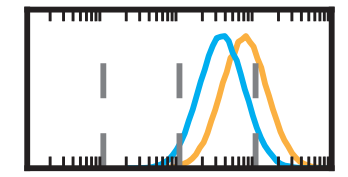
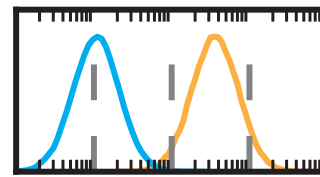
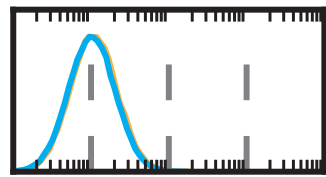


fluorescence

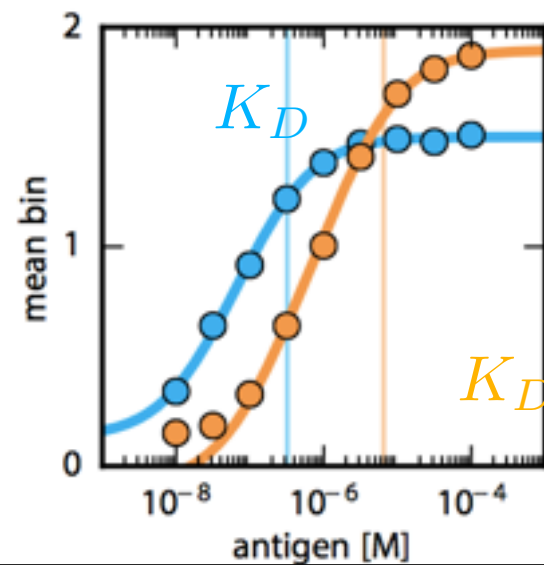
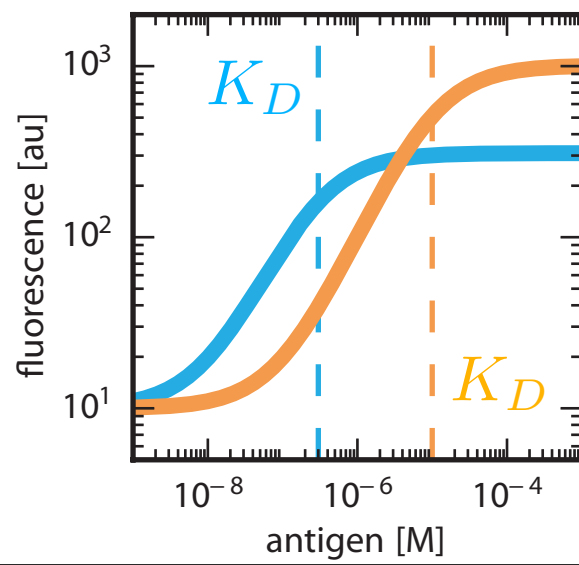
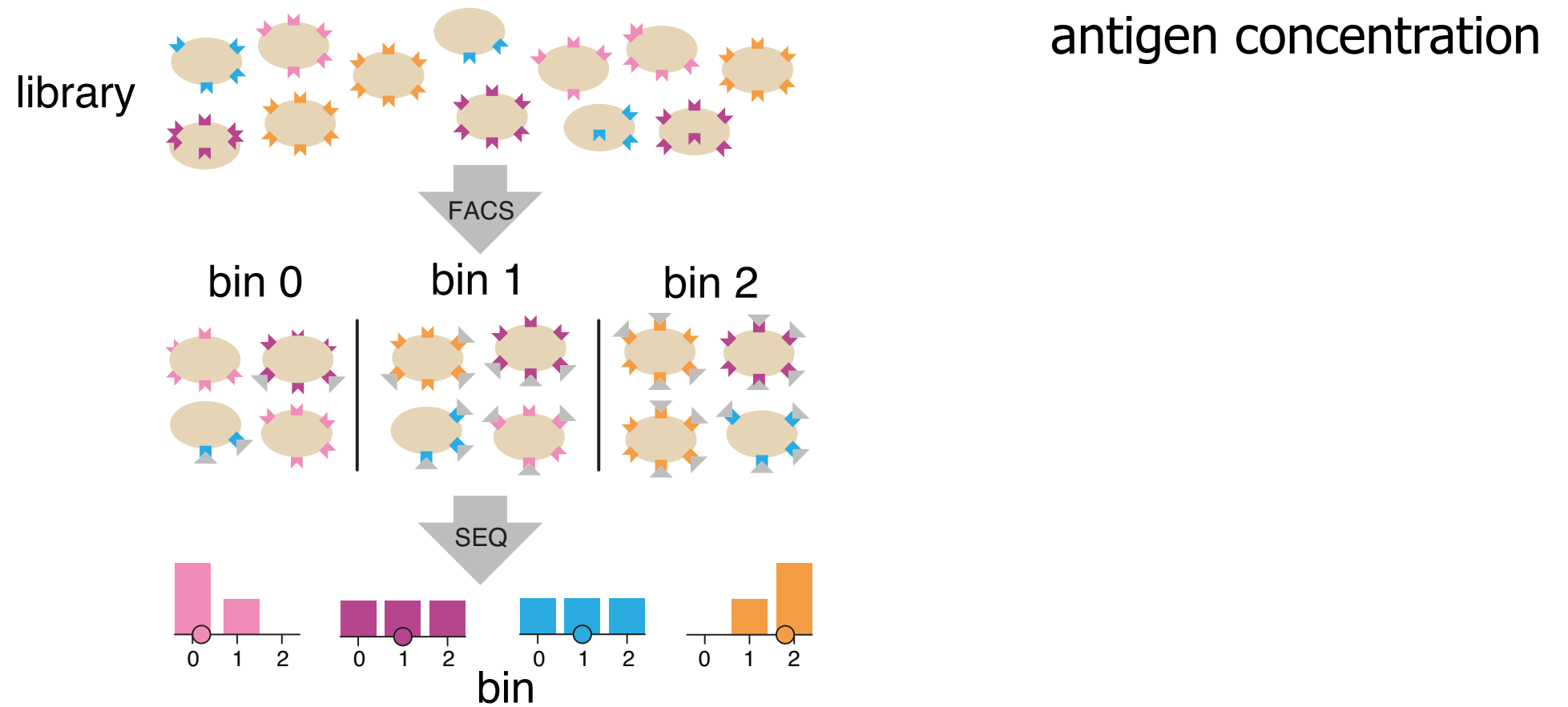
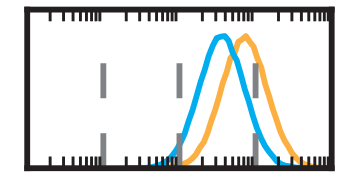
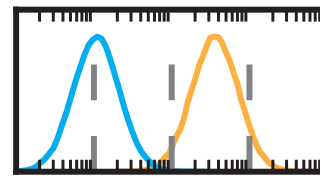
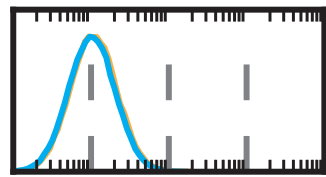
antigen concentration



Tite-Seq



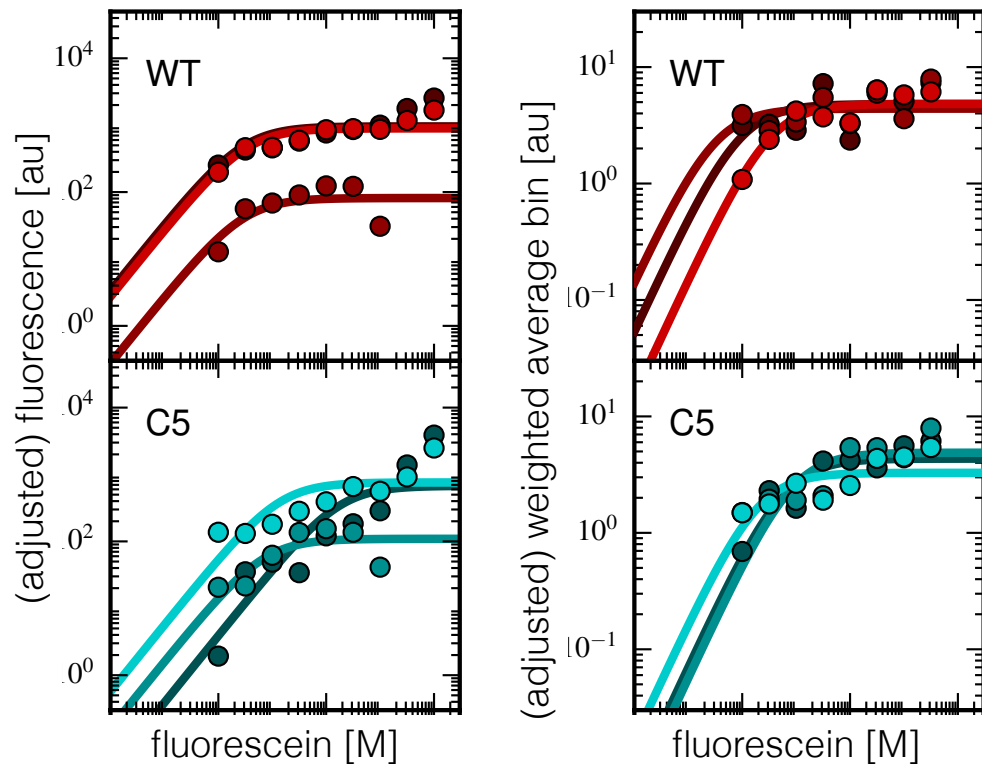
Tite-Seq



Comparison to fluorescence measurements

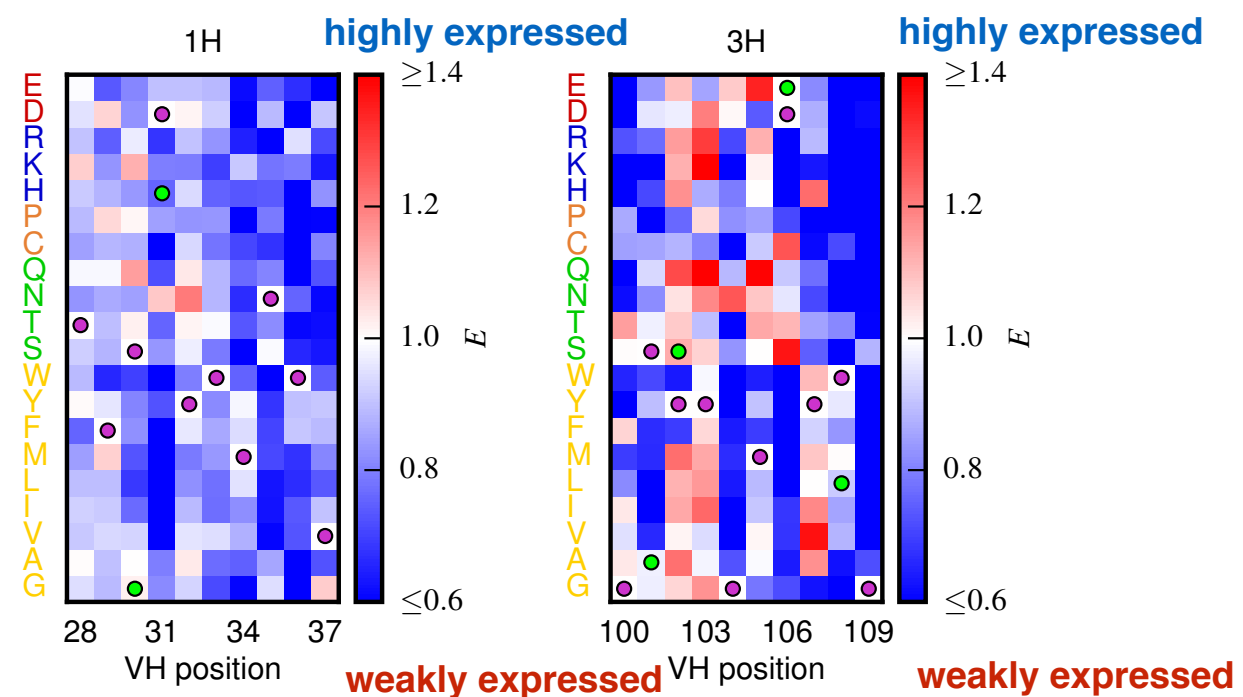
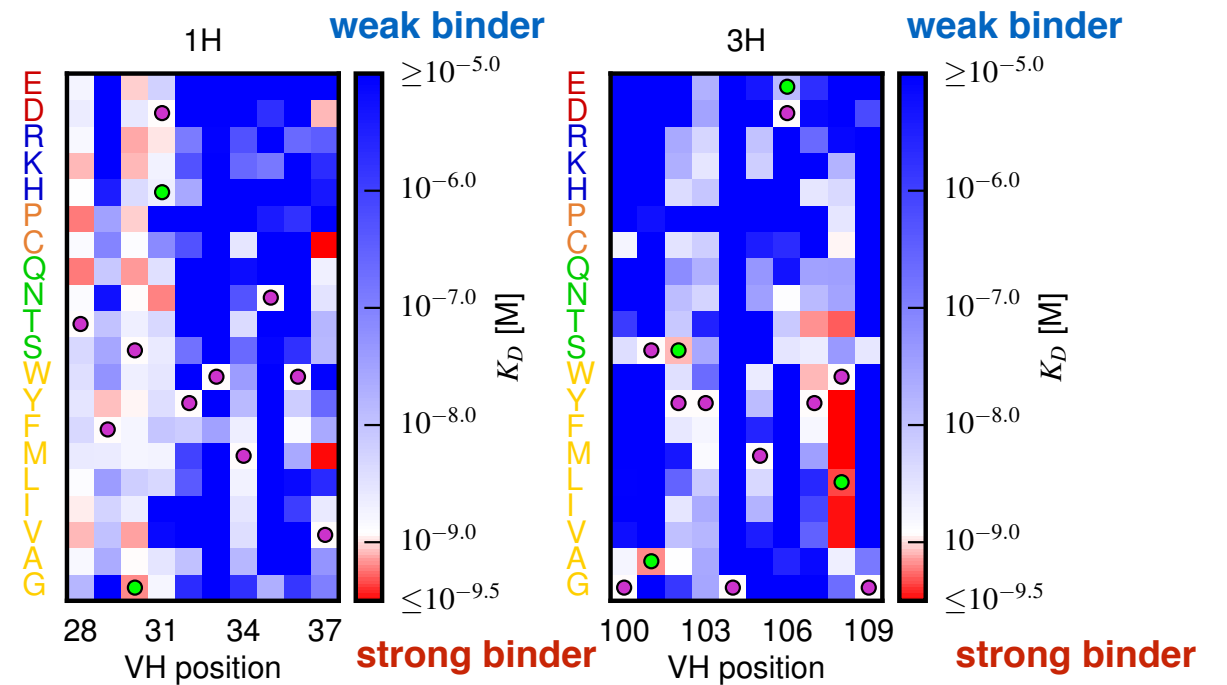
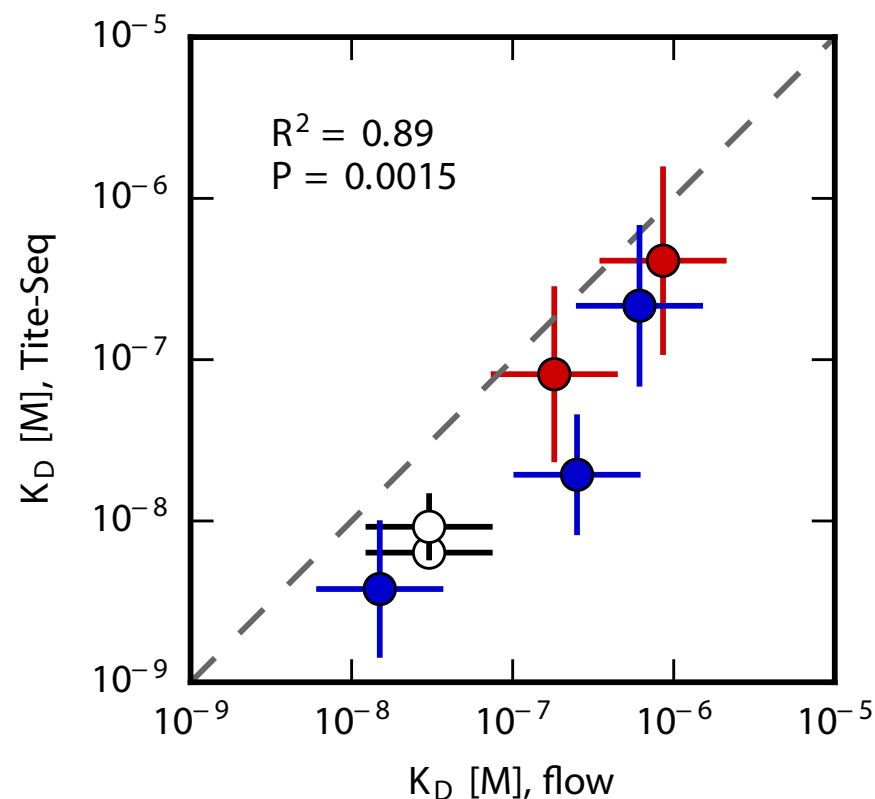
FLUORESCENCE

TITE-SEQ



- reproducible results from CDR1 and CDR3 mutants

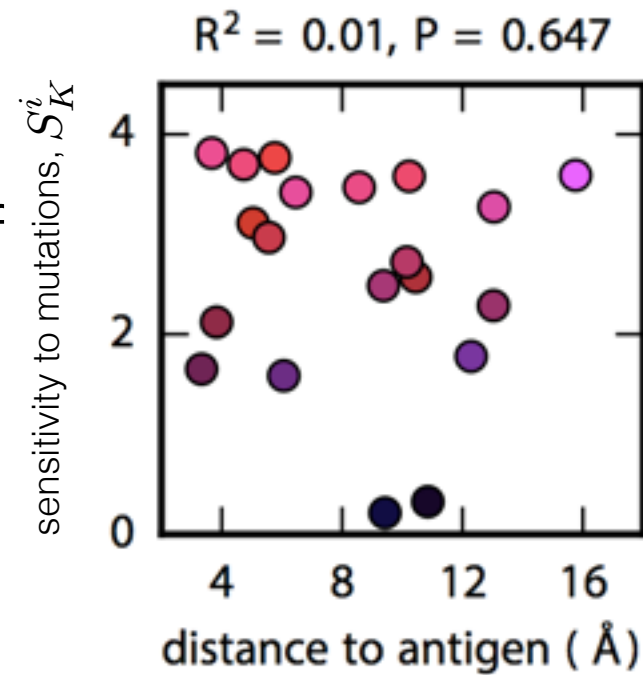
- wide range of affinities



Mutational effects by CDR

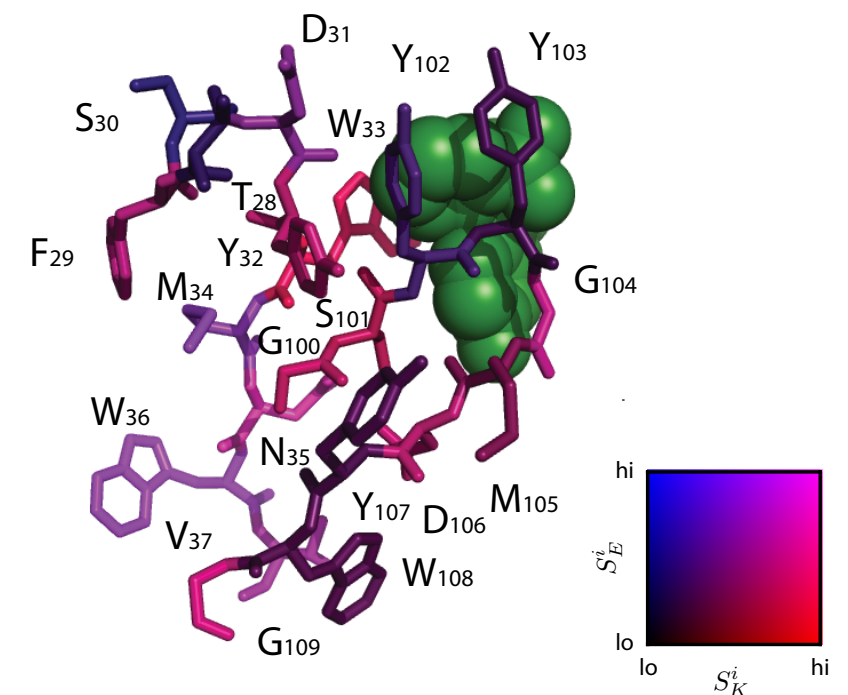
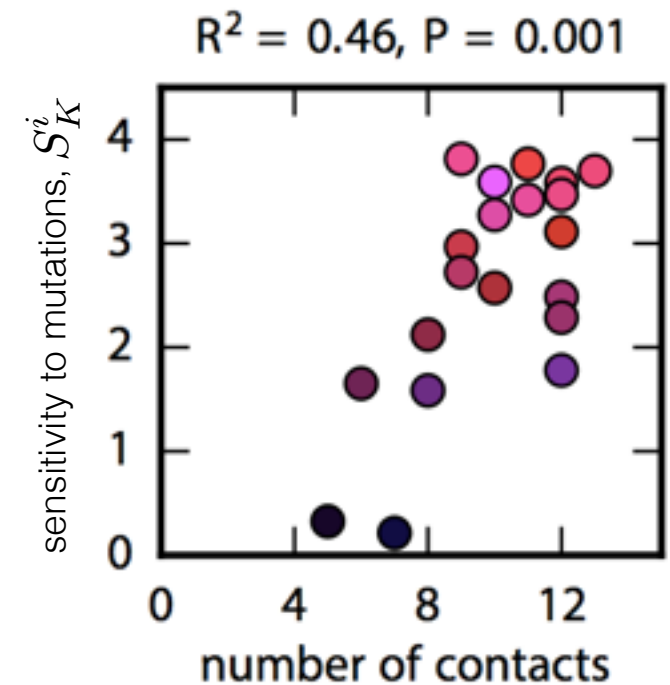
affinity:

- independent of distance of mutation to antigen



- depends on the number of contacts a residue makes in the receptor

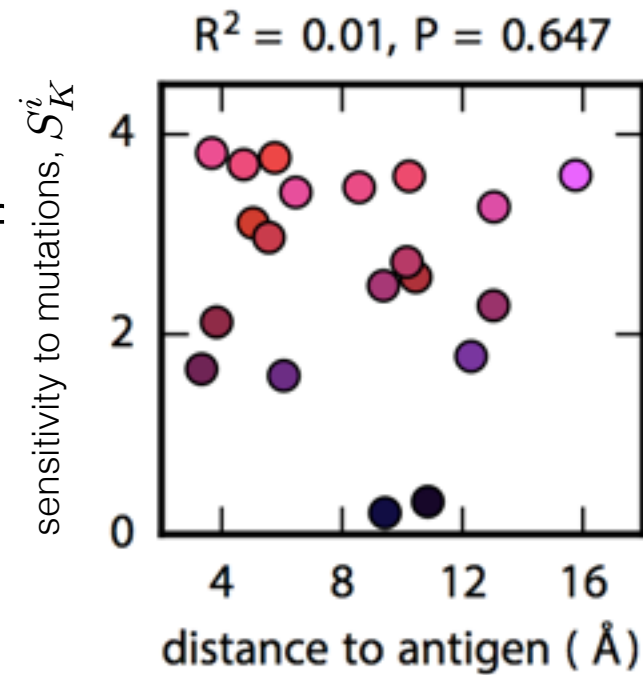
$$S_K^i = \sqrt{\langle (\log_{10} K_D^{ia} - \log_{10} K_D^{WT})^2 \rangle_a}$$



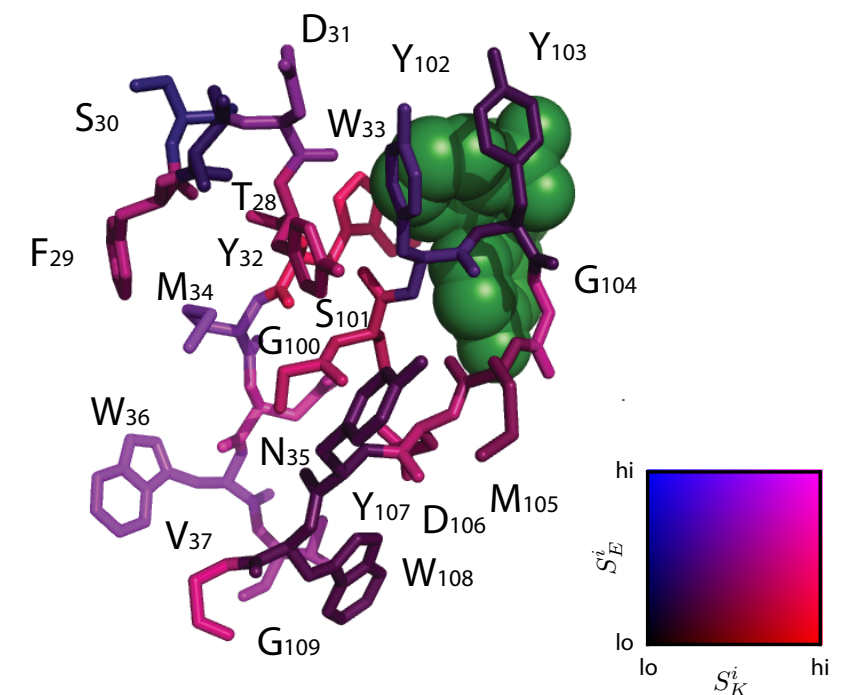
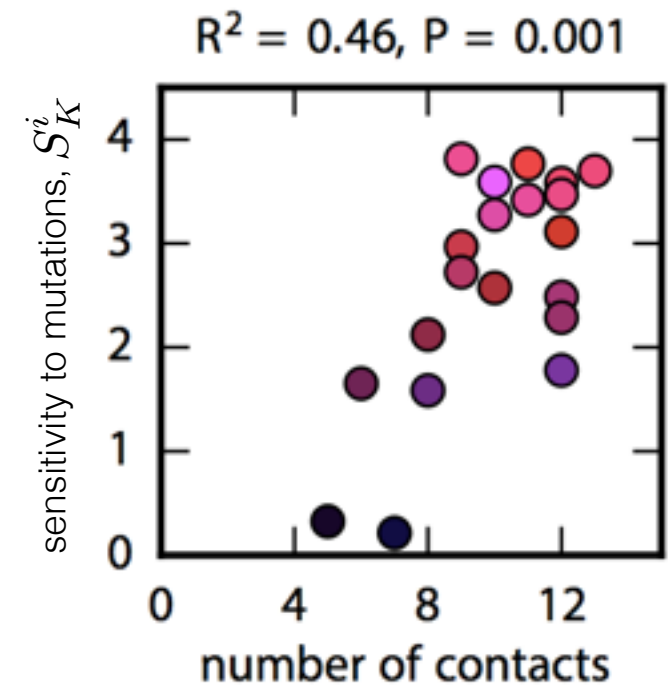
Mutational effects by CDR

affinity:

- independent of distance of mutation to antigen
- depends on the number of contacts a residue makes in the receptor
- non-local effects
- effect of interactions between receptor residues



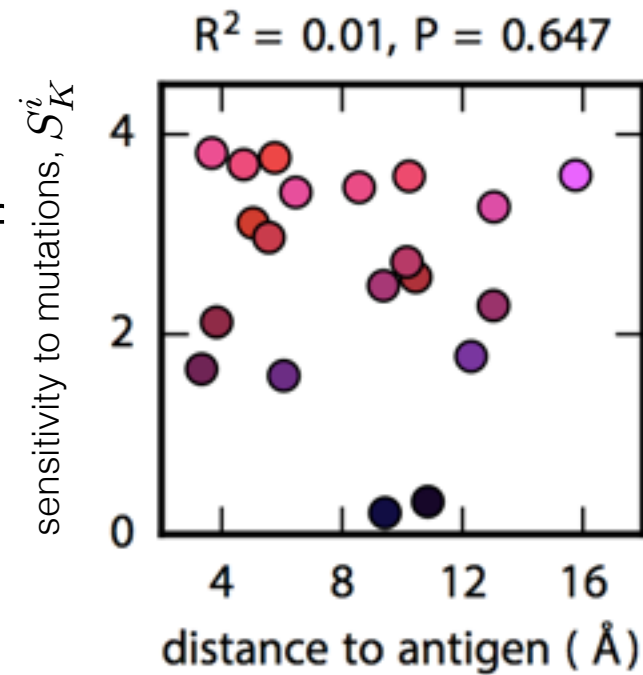
$$S_K^i = \sqrt{\langle (\log_{10} K_D^{ia} - \log_{10} K_D^{WT})^2 \rangle_a}$$



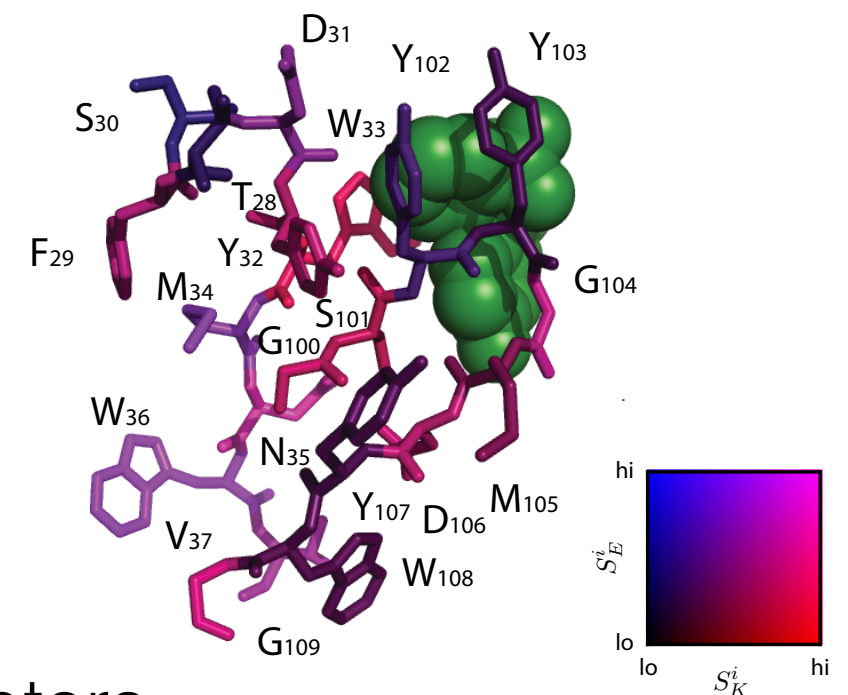
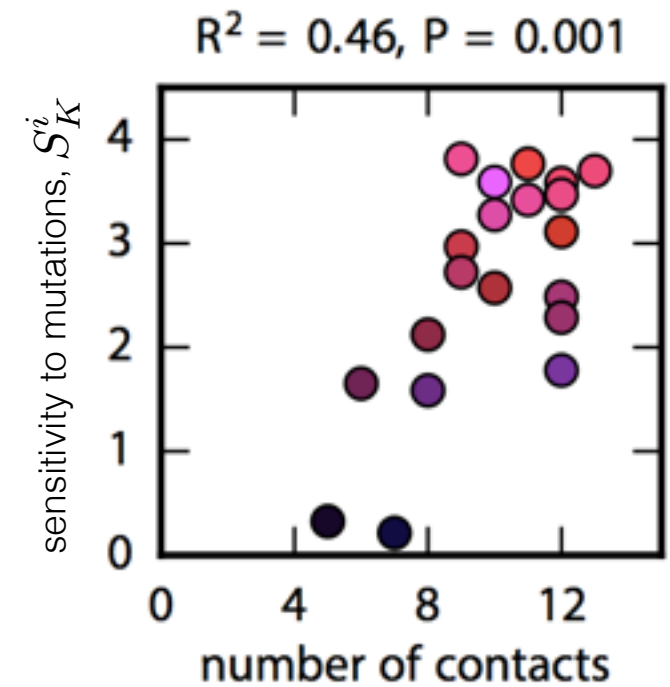
Mutational effects by CDR

affinity:

- independent of distance of mutation to antigen
- depends on the number of contacts a residue makes in the receptor
- non-local effects
 - effect of interactions between receptor residues
- CDR3 greater effect on affinity
 - more likely to be mutated in functional receptors



$$S_K^i = \sqrt{\langle (\log_{10} K_D^{ia} - \log_{10} K_D^{WT})^2 \rangle_a}$$





generating diversity and selection:

- random overlap between (most) individuals
- very long lived clones

response:

- identify responding clonotypes
- observe selection on standing variation

Tite-Seq:

- high-throughput affinity and expression assay
- interactions within receptors

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