

# Genetics of phenotypic variation

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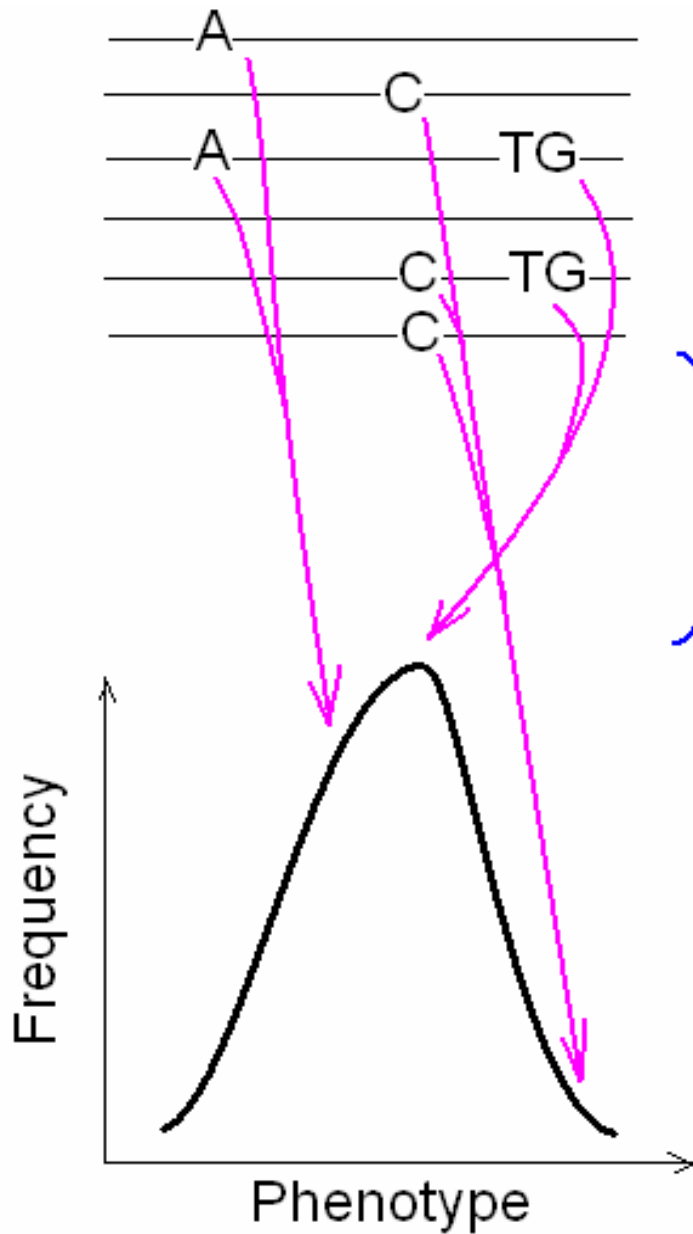
UFG

UNL

# Intro

- Genetics of local adaptation of *A. lyrata*, QTL → model selection;
- Is outcrosser (*A. arenosa*) better than selfer (*A. thaliana*)?
- Gene-level association studies, *bric-a-brack* in *D. melanogaster*?
- How variation in pathways scales into phenotypic variation, InR/TOR pathway in *D. melanogaster*.

# Intro



Molecular Evolution

Quantitative  
Genetics

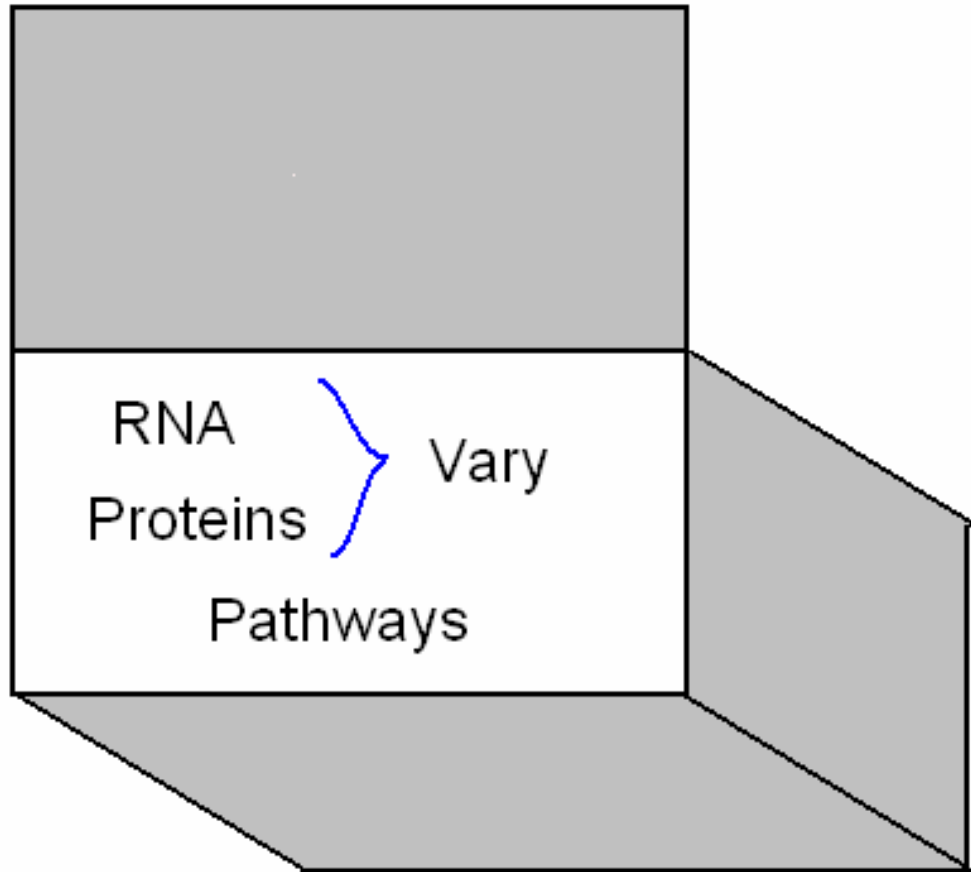
Evolution of Phenotype

# Intro

Genotype



Phenotype



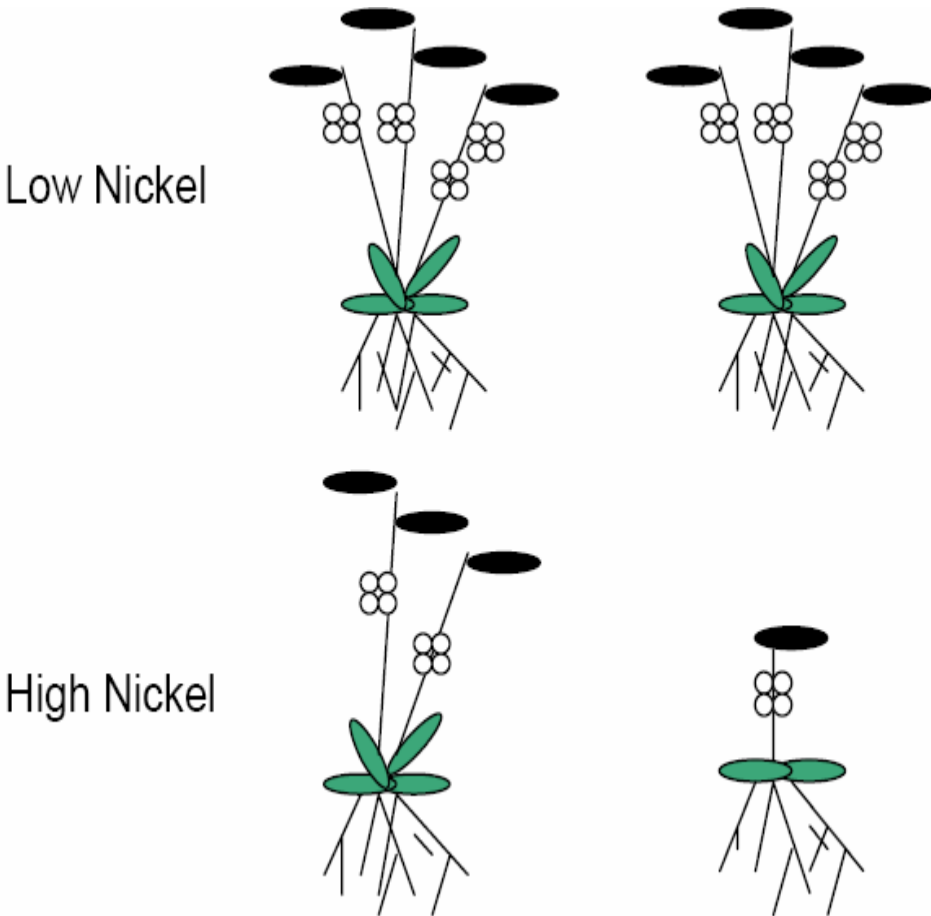
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# *Arabidopsis lyrata* adaptation to serpentine soils, how many genes?



# Sequence divergence, how much does it matter?



*Arabidopsis lyrata*

~95% seq. identity

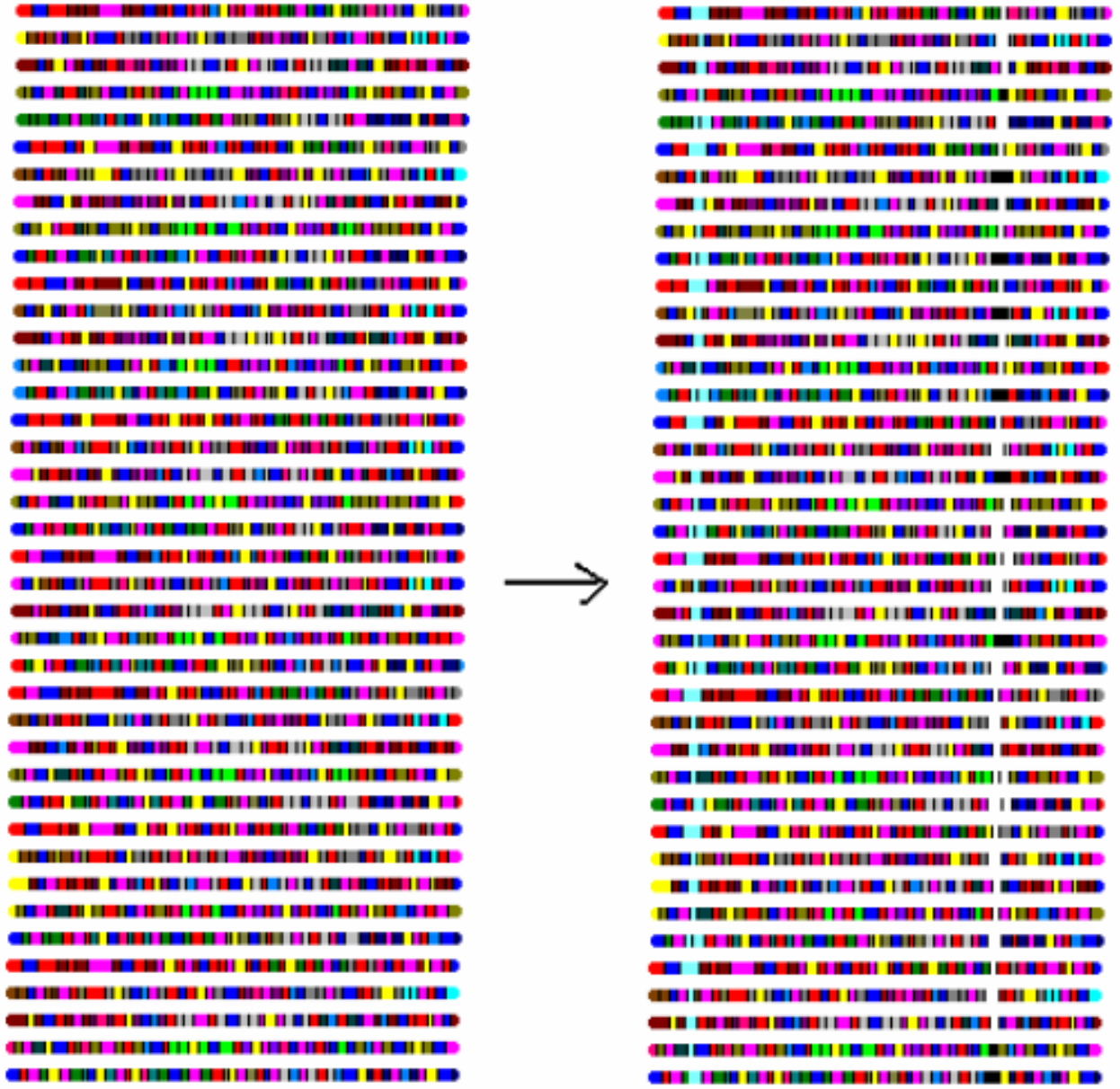
~tiling Arabidopsis array:

~3M PM/MM features

2 populations on normal

2 on serpentine X 3 chips

→ 12 arrays



Blue allele fixed due to selection, ↑  
black and white alleles both increased in frequency ↑



# Soil adaptation: steps

95% sequence identity →

→ 1 b out of 25 are divergent

2853369 PMs on the array →

→ 371642 are matches

28 ( $p < 0.05$  Bonferroni);

72 (FDR < 1%).

+ windows → a couple of hundreds significantly diverged genes!

# Reasonable? Overrepresented GO categories:

- Membrane proteins ( $p < 0.001$ )
- Transporters ( $p < 0.001$ )
- Potassium ion transport ( $p = 0.006$ )
- Cellular calcium ion homeostasis ( $p = 0.008$ )
- Cation transport ( $p = 0.03$ ).

*cax7* → 8/23 (70b) are polys in granitic  
/ fixed in serpentine soils.

→ At least a couple hundred genes are involved in adaptation to soil types.

- Genetics of local adaptation of *A. lyrata*, QTL  
→ model selection;

- 200 genes are divergent → ☹️
- 30000 – 200 are NOT diverged → 😊

(Solexa-resequenced to 60x, all holds up)

Imagine that we do RILs from 2 populations with  
KNOWN divergent genes →

Mapping exercises → Assigning effects and  
probabilities (priors: allele frequency  
differences, known gene functions etc.)

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# Is outcrosser (*A. arenosa*) better than selfer (*A. thaliana*)?

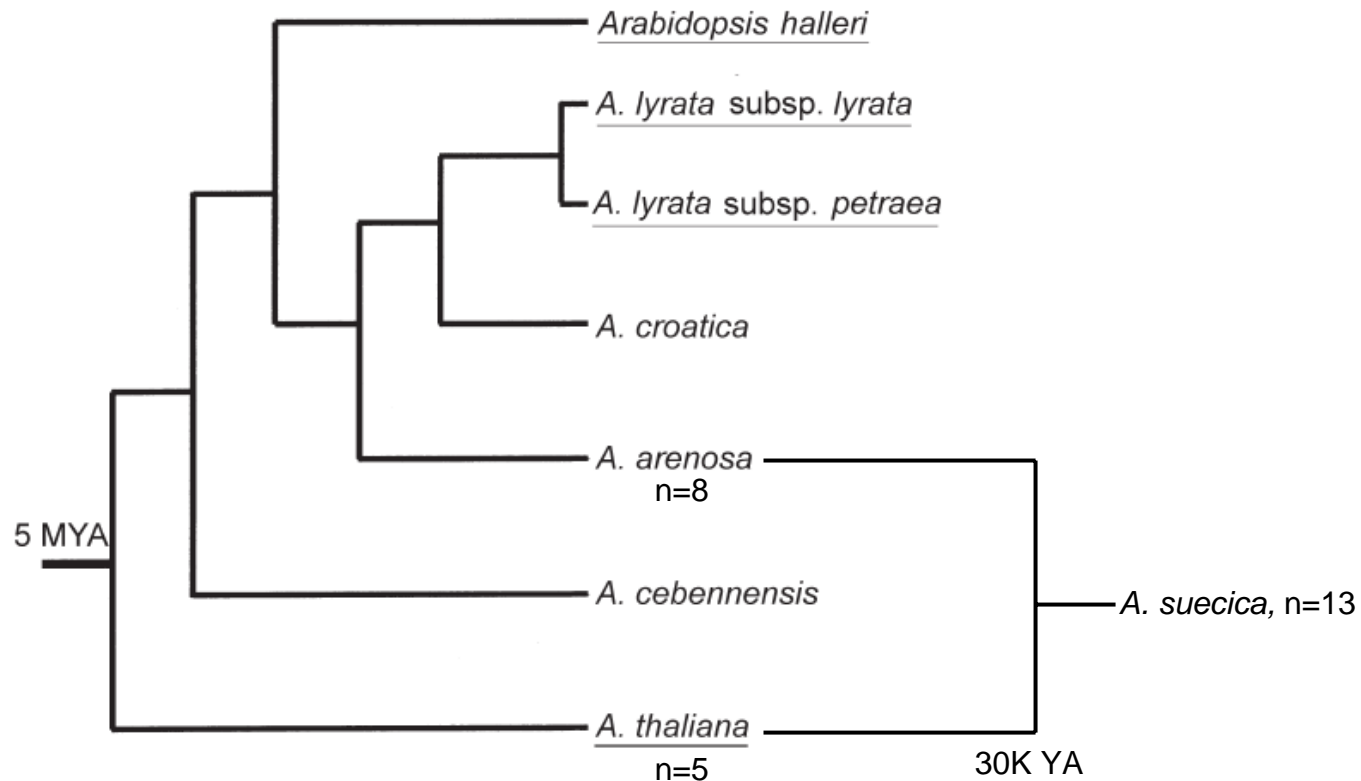
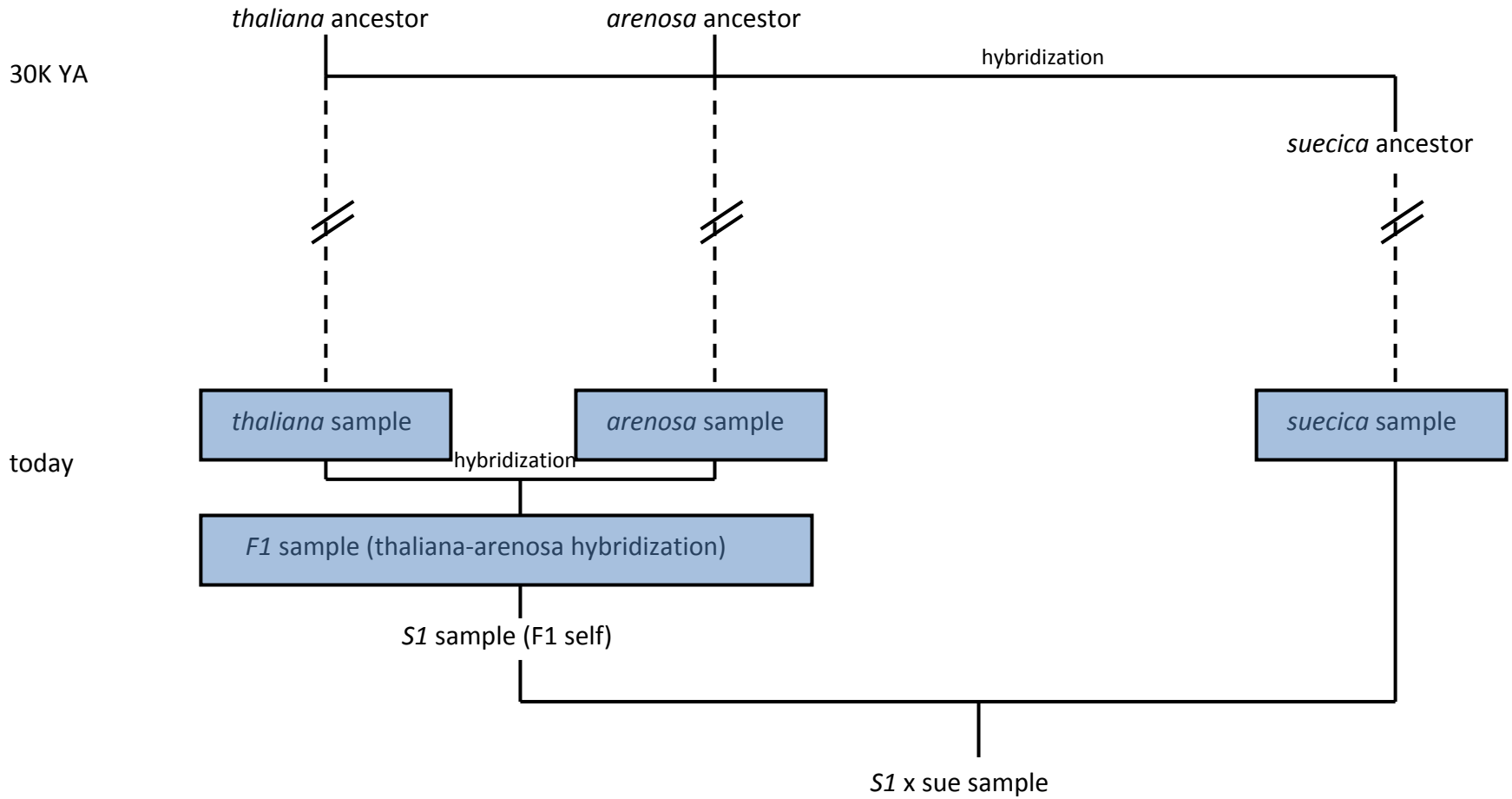


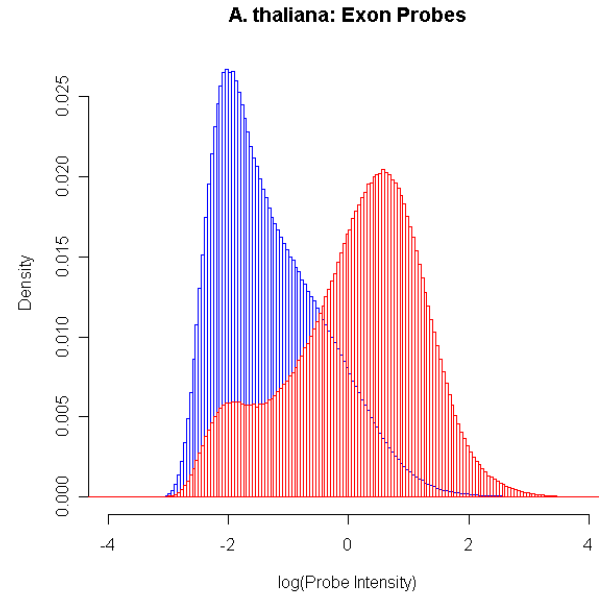
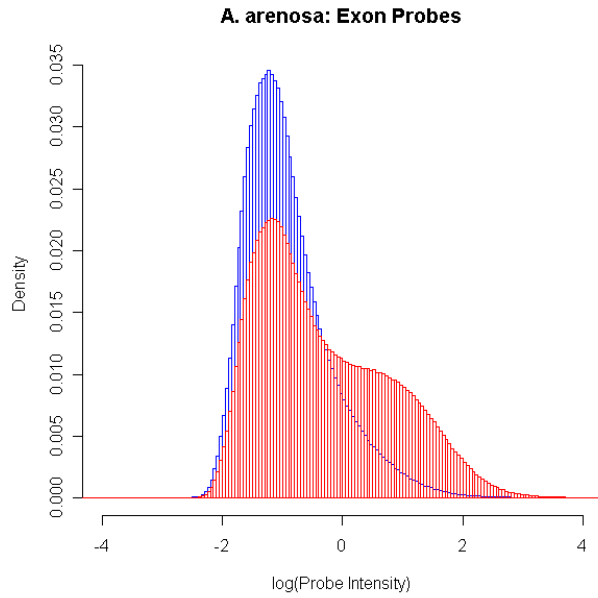
Figure 1. Phylogenetic relationships within *Arabidopsis*.



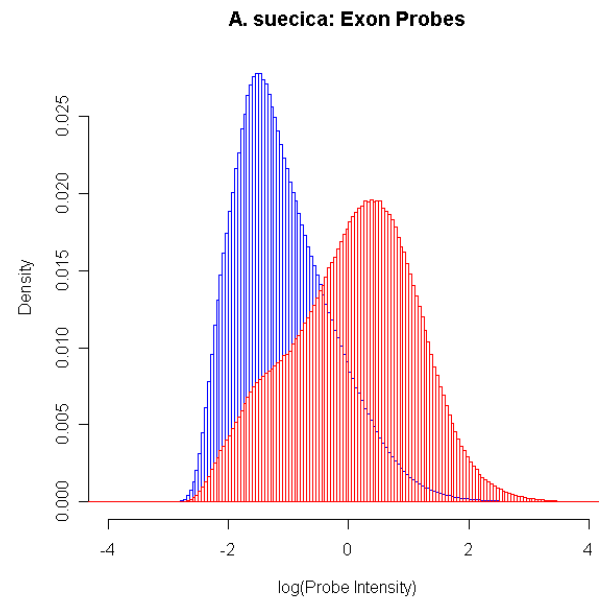
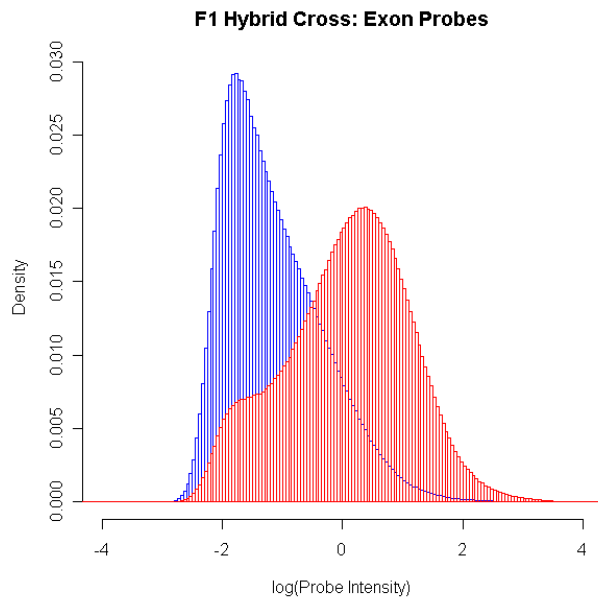
Question 1: Which allele has been maintained in *A. suecica*?

Question 2: Which allele has been silenced in *A. suecica*?

# *A. thaliana* chips hybed with:

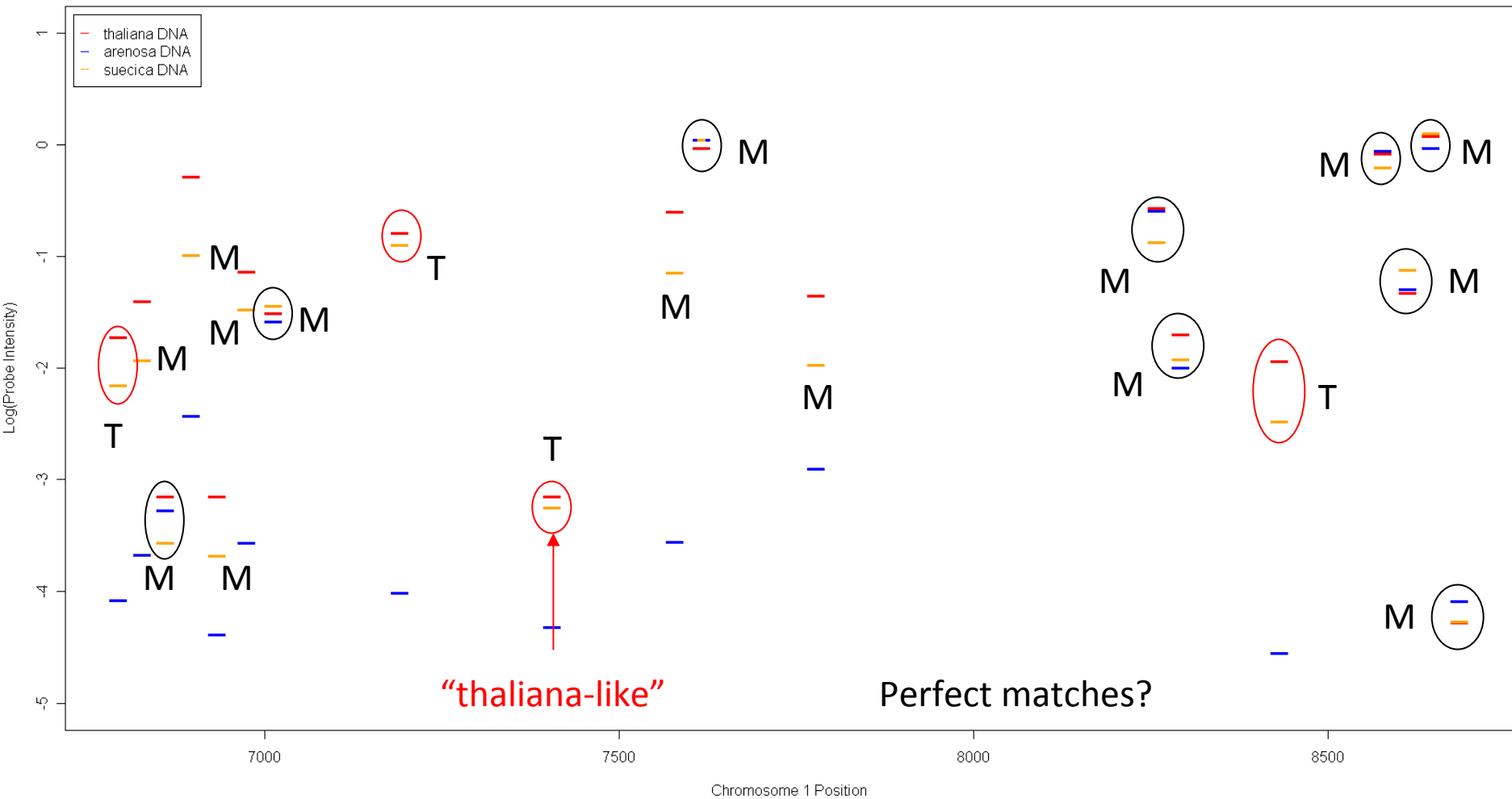


PM Probes  
MM Probes

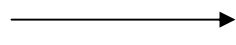


thaliana  
arenosa  
suecica

Gene: AT1G01020.1, 19 Probes



TMMMMMMTTMMMMMTMMMM



Probes Missing in arenosa: 4  
Probes Missing in thaliana: 0



# Probe Distribution

Arabidopsis Tiling Array At35b		Overall	
		Matches	Total
Number of Probes in Exons	F	601443	3039991
	R	611591	3039991
Number of Exons Covered	F	75448	78873
	R	76177	79197
Number of Genes Covered	F	15647	16018
	R	15519	15744
<b>Average Number of probes/exon</b>	<b>F</b>	<b>7.97</b>	
	<b>R</b>	<b>8.03</b>	
<b>Average Number of exons/gene</b>	<b>F</b>	<b>4.82</b>	
	<b>R</b>	<b>4.91</b>	

# What is retained?

<b>DNA</b>	<i>A. suecica</i>	Genes
<i>Deletions</i>	<i>arenosa</i> -biased	180
	<i>thaliana</i> -biased	45
<b>RNA</b>	<i>arenosa</i> -biased	939
<i>Silencing</i>	<i>thaliana</i> -biased	383

Is outcrosser (*A. arenosa*) better than selfer (*A. thaliana*)?

In F1 → silencing is unbiased and ‘random’  
(among progeny, tissues) Chen et al. 2006.

In *A. suecica* → *A. arenosa* alleles are:

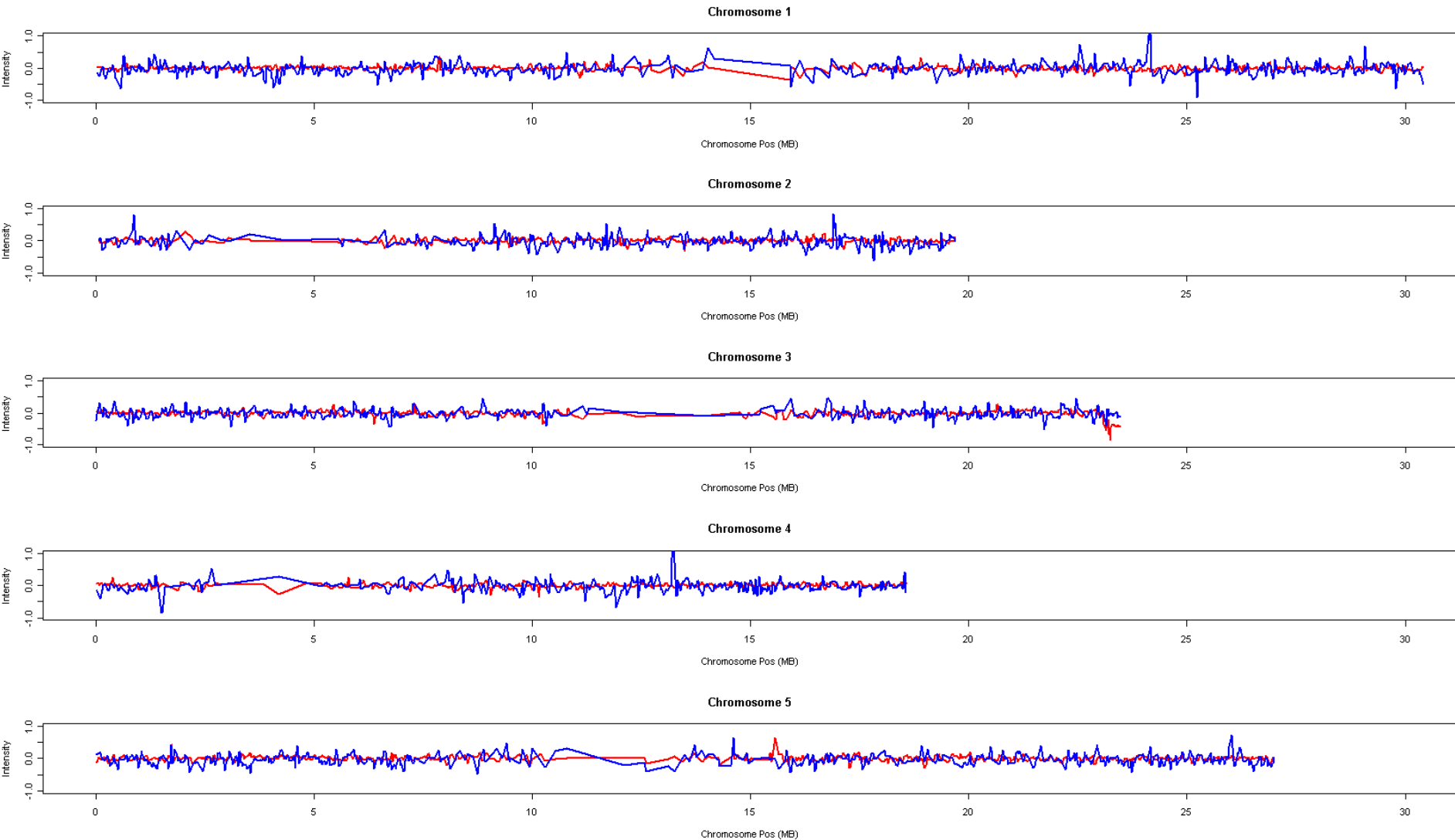
- more frequently retained;
- more frequently utilized.

Is silencing ‘region-specific’?

Is silencing ‘pathway-specific’?

Is silencing reflective of ‘population genetics’?

# Is outcrosser (*A. arenosa*) better than selfer (*A. thaliana*)?



# The *Drosophila* model for the genotype to phenotype map



wing song



dance

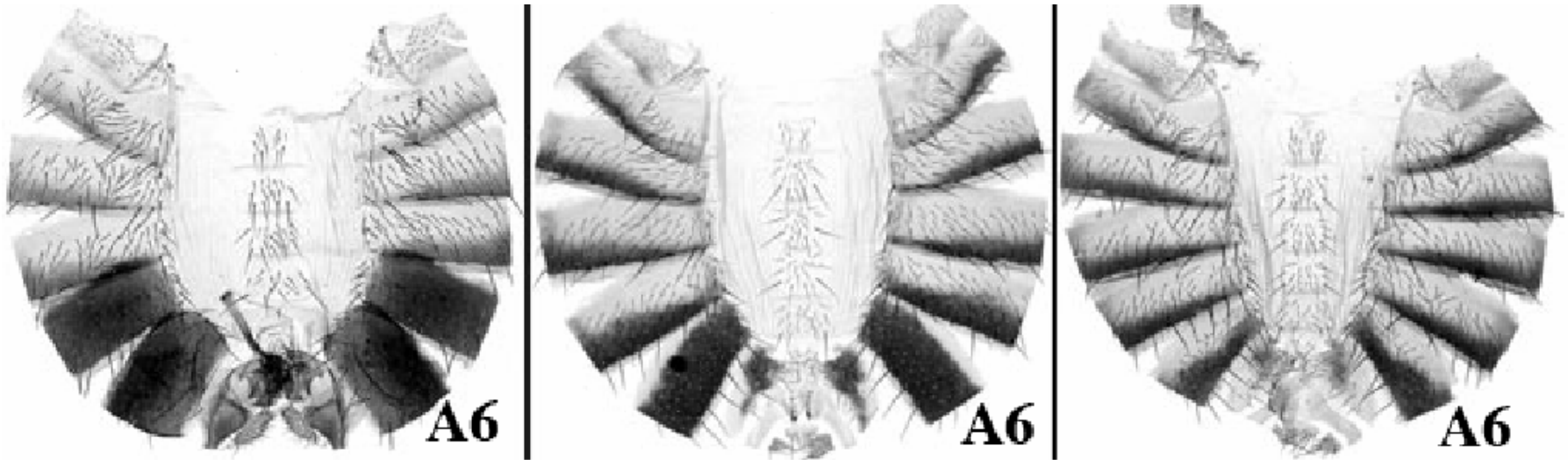


CHCs  
(pheromones)



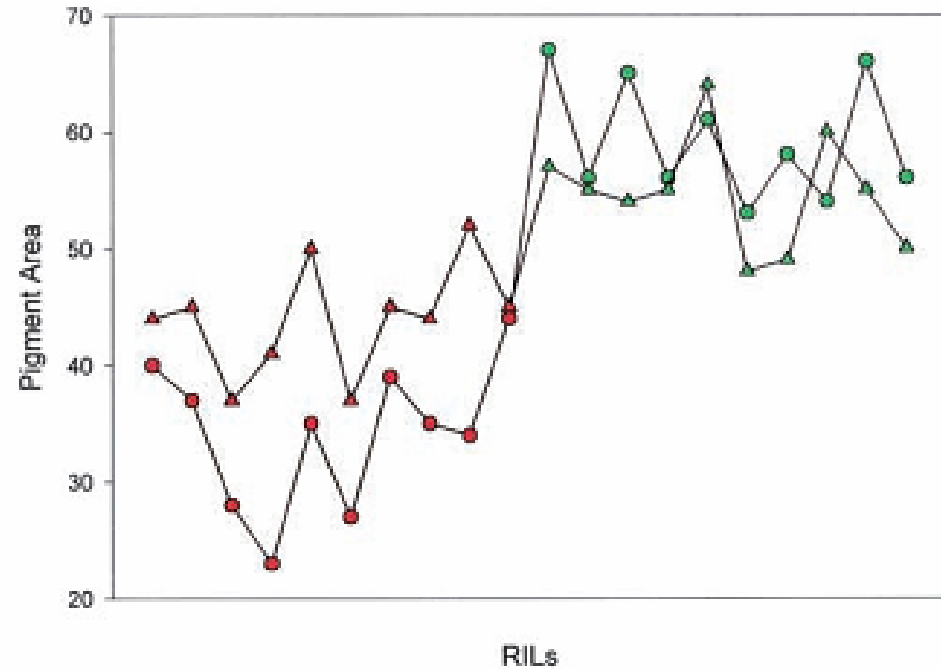
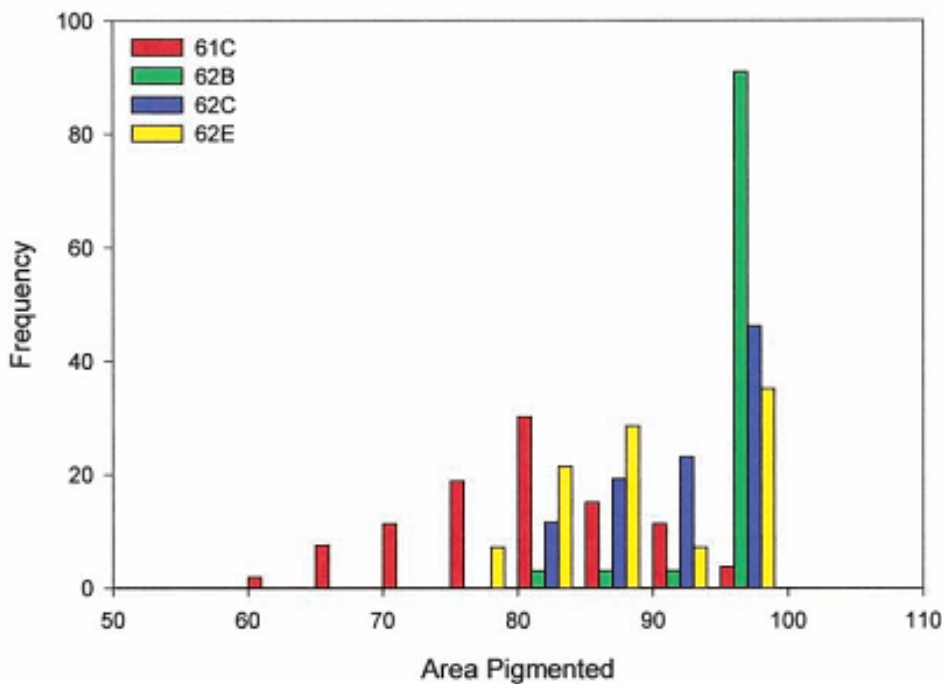


# Building a genotype to phenotype map

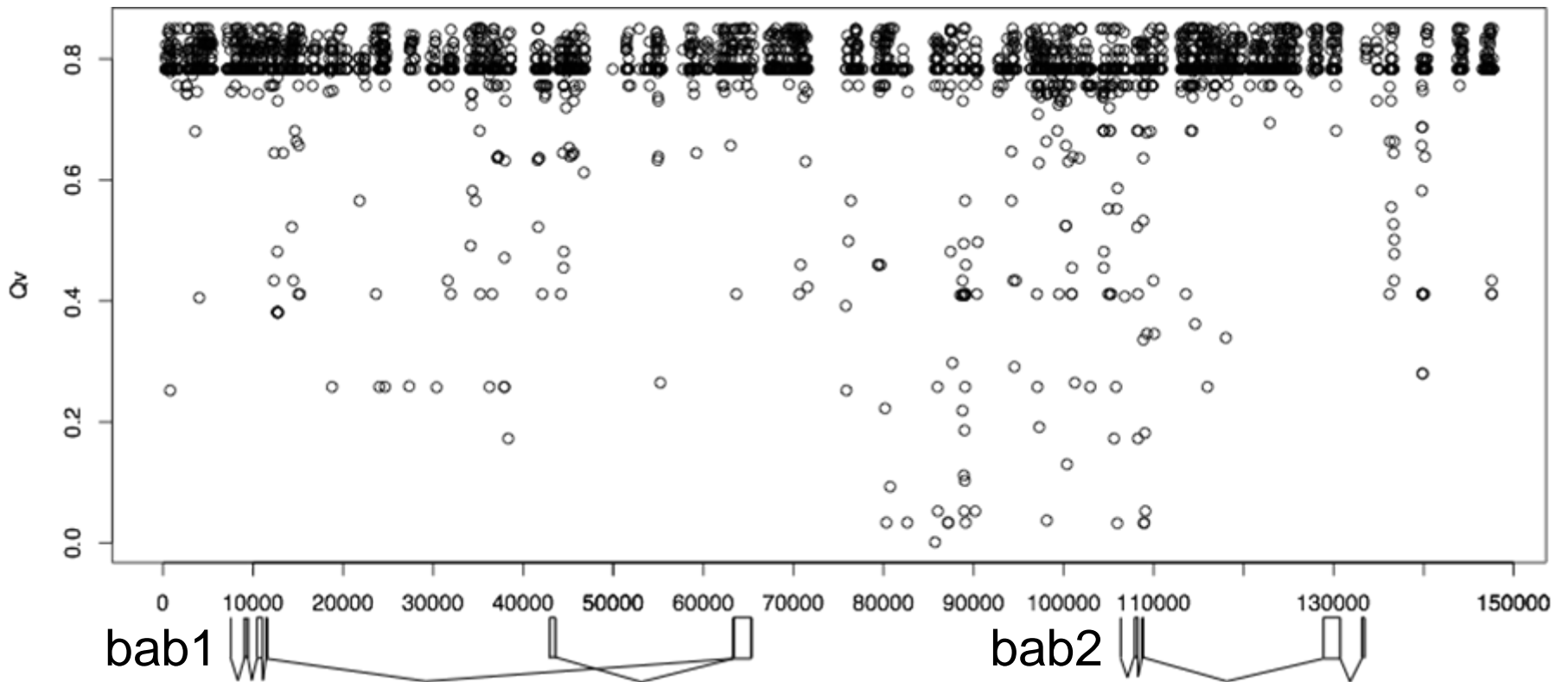
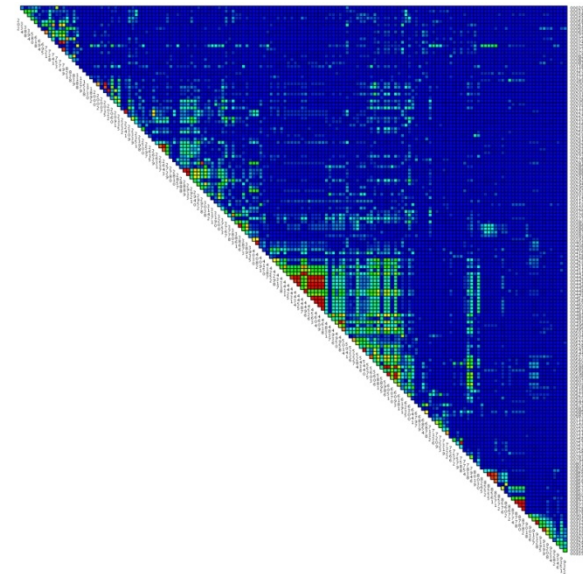


148 kb region → 40 SNPs / 6 indels per kb  
→ ~6000 SNPs and ~900 indels

# Effects of alleles are large, and the evidence for a gene is clear-cut

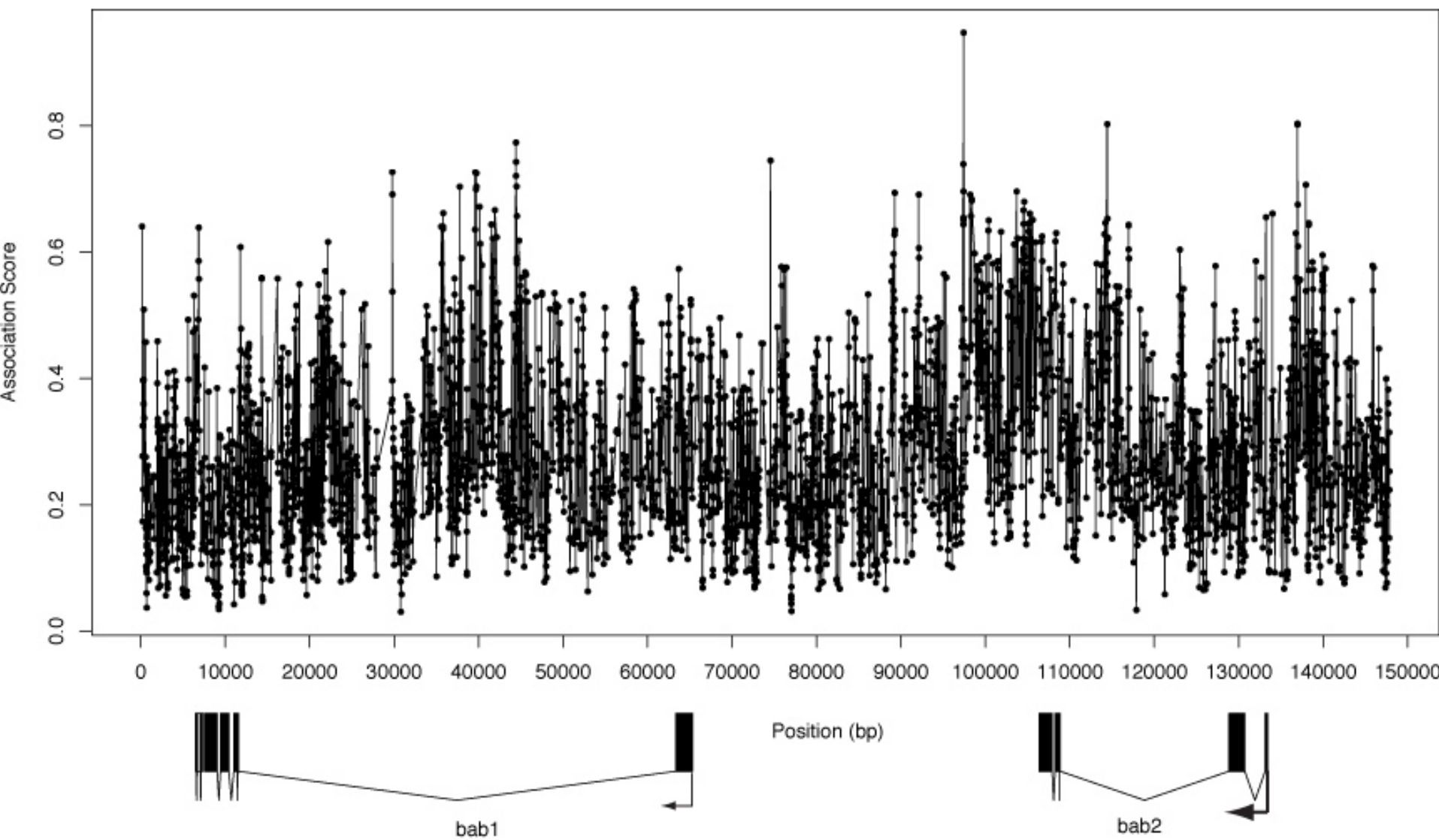


# Low LD (200 b) enables precise association inferences

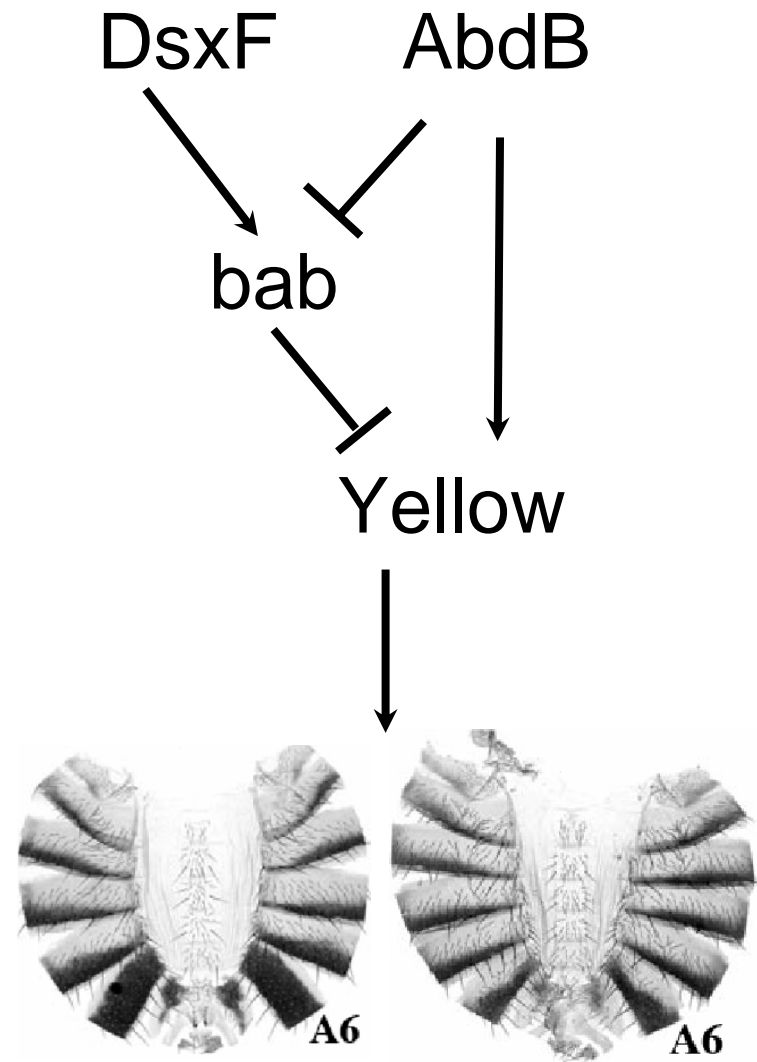
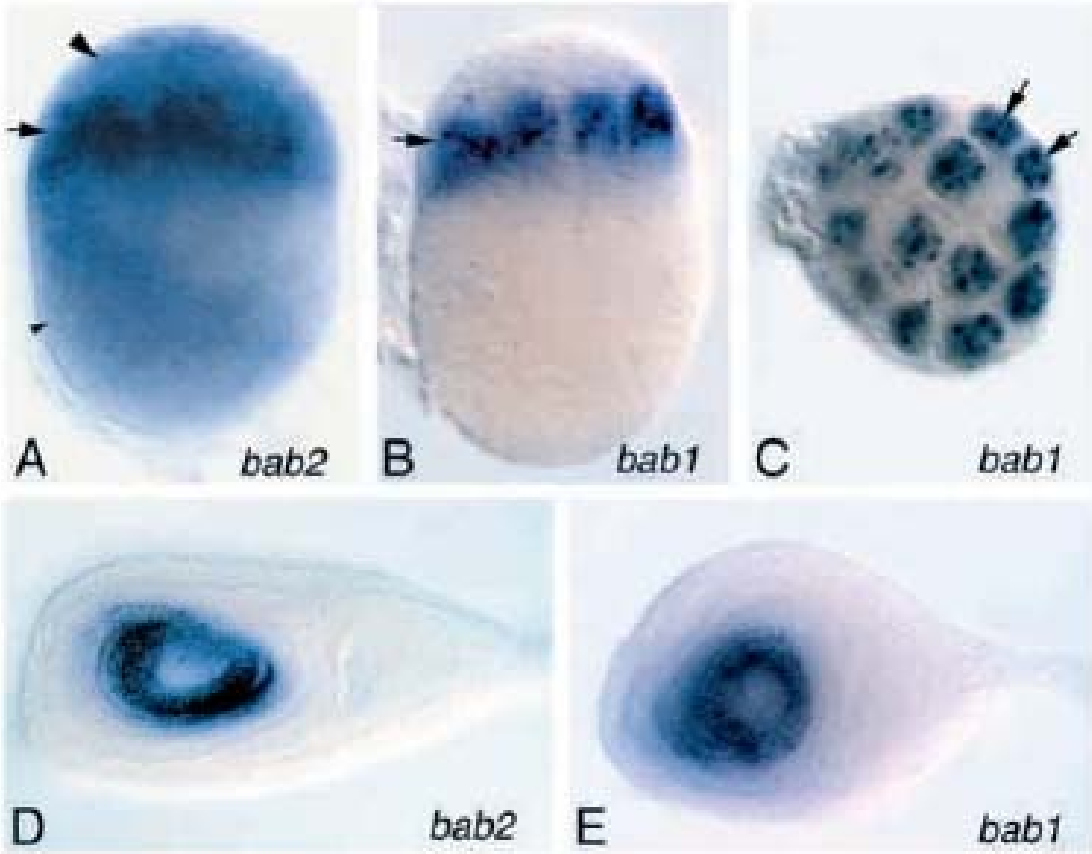




### HLD modeling of the bab region



Gene functioning might be assayed at the right time in the right tissues...



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# Quantitative genetics of transcript levels. Components of Variation.

- 9,863 genes ← no multiple splice products, probes higher than negative controls.

1,609 X linked genes

8,213 “autosomal” genes (2, 3 only)

35 4<sup>th</sup> chromosome, 6 Y chromosome

- Models run for sexes separately; evaluated significance at FDR = 0.20

$$\sigma^2_P = 2\sigma^2_{GCA} + \sigma^2_{SCA} + 2\sigma^2_{RGCA} + \sigma^2_{RSCA} + \sigma^2_{\varepsilon}$$

# Quantitative genetics of transcript levels.

Sires

Dams

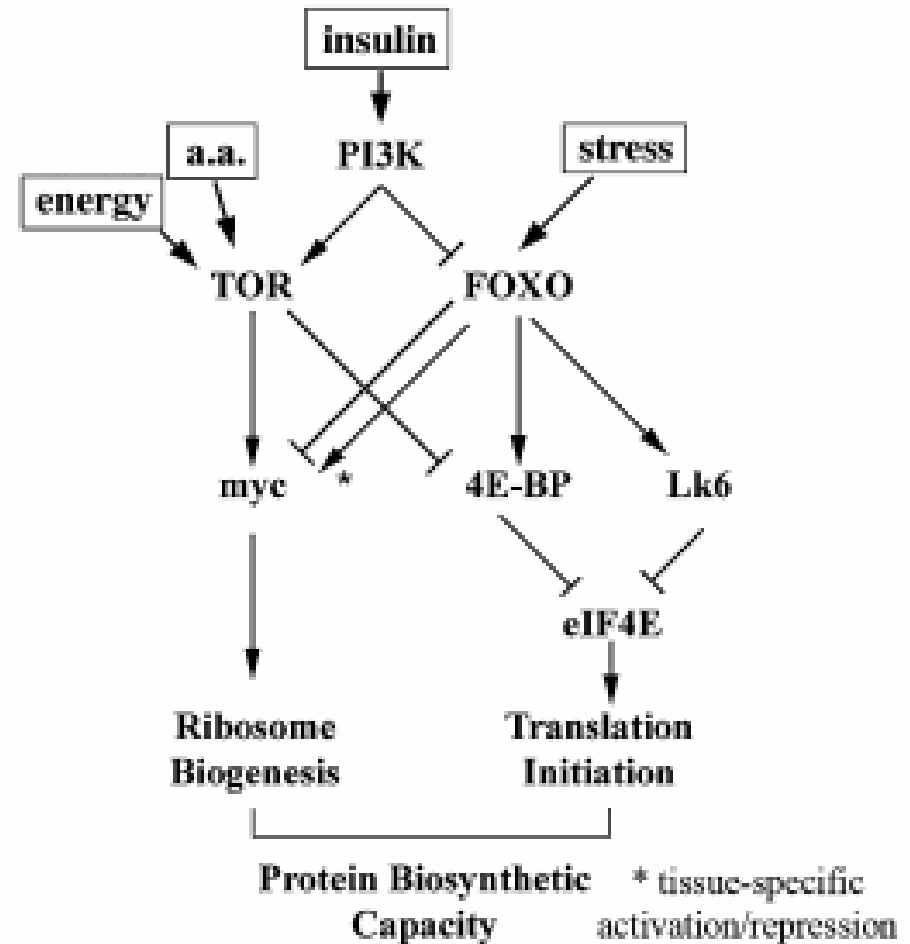
	1	2	3	4	5	6	7	8	9
1		1,2	1,3	1,4	1,5	1,6	1,7	1,8	1,9
2	2,1		2,3	2,4	2,5	2,6	2,7	2,8	2,9
3	3,1	3,2		3,4	3,5	3,6	3,7	3,8	3,9
4	4,1	4,2	4,3		4,5	4,6	4,7	4,8	4,9
5	5,1	5,2	5,3	5,4		5,6	5,7	5,8	5,9
6	6,1	6,2	6,3	6,4	6,5		6,7	6,8	6,9
7	7,1	7,2	7,3	7,4	7,5	7,6		7,8	7,9
8	8,1	8,2	8,3	8,4	8,5	8,6	8,7		8,9
9	9,1	9,2	9,3	9,4	9,5	9,6	9,7	9,8	

Conclusions: males possess ~no epistatic variation in transcript levels, while females are overwhelmed with it

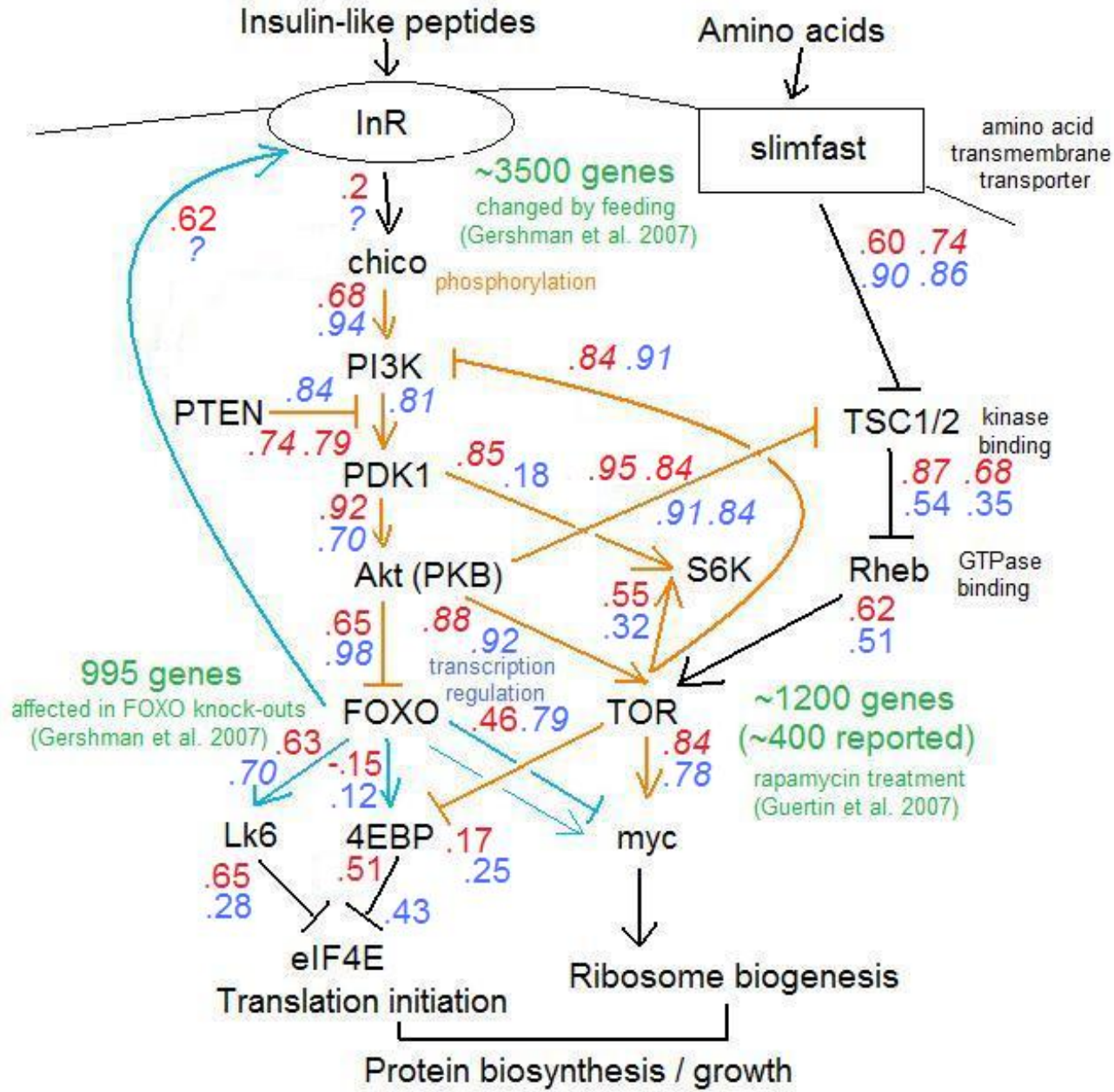
	Male		Female	
	X	Autosome	X	Autosome
Genetic	192	1337	604	2720
<b>GCA</b>	<b>185</b>	<b>1336</b>	<b>88</b>	<b>630</b>
<b>SCA</b>	<b>1</b>	<b>2</b>	<b>361</b>	<b>1284</b>
rGCA	44	23	1	1
rSCA	2	1	341	1380

# How variation in pathways scales into phenotypic variation, InR/TOR pathway in *D. melanogaster*.

- Does transcriptome variation shape with the networks inferred with major-effect mutations?
- Does genetic variation in networks map into phenotypic variation?



# Does transcriptome variation shape with the networks inferred with major-effect mutations?

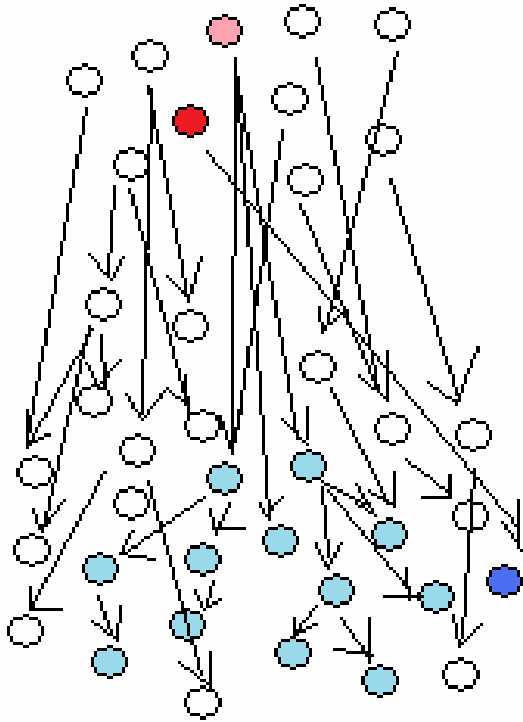




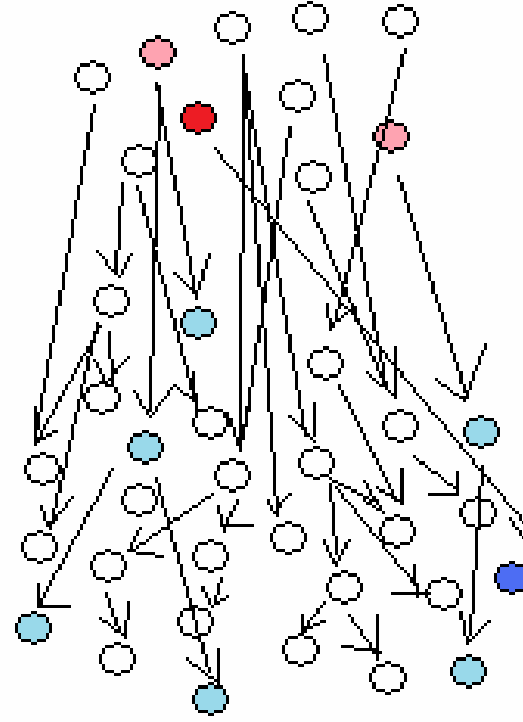
# Tons of correlations, naturally

Gene or factor	Dessicat.	Oxidat.	Starv.	Longev.	Develop.	Bodysize
Akt1	-.34	-.75, .02	-.43	-.46	-.01	-.37
InR	.10	-.63	-.22	-.11	.25	.24
Lk6	-.30	-.81, .005	-.30	-.47	-.06	-.13
PI3K	-.31	-.76, .01	-.44	-.58	.07	-.01
PDK1	-.41	-.67, .04	-.27	-.33	-.01	-.19
PTEN	-.29	-.72, .03	-.50	-.43	.02	-.59
Rheb	-.18	-.44	.11	-.24	.02	-.44
S6k	-.29	-.43	-.24	.28	.32	.03
4EBP	.27	-.04	.09	-.47	-.80, .008	-.71, .03
Tor	-.32	-.88, .0008	-.49	-.53	.05	-.01
Tsc1	-.33	-.69, .04	-.24	-.39	.07	-.20
chico	-.18	-.44	.14	-.17	.01	-.24
Myc	-.15	-.60	-.17	-.35	-.06	.31
eIF4E	-.43	-.53	-.21	-.45	-.02	-.46
foxo	-.42	-.93, $6.2 \times 10^{-5}$	-.79, .008	-.41	.23	.04
Tsc2	-.42	-.89, .0005	-.45	-.55	.03	.02
Slif	-.06	-.80, .008	-.37	-.48	-.15	-.47

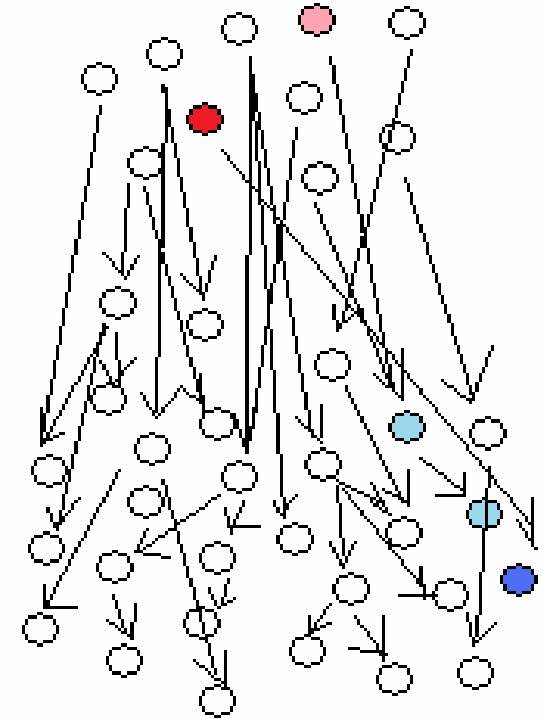
# What's causing what?



Genotype 1



Genotype 2



Genotype 3

- Allele specific expression  $\rightarrow$  *cis*;
- The rest are *trans*;
- 200 genotypes?

# Should work...., might be?

