

From Genes to Growth and Form

KITP

08/23/16

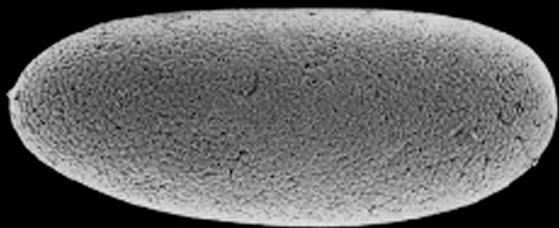
**Time-keeping mechanisms of embryonic  
cell cycles**

Stefano Di Talia

Department of Cell Biology

Duke University Medical Center

**Biological systems are 'noisy'. Yet,  
embryonic development is regulated so  
precisely!**

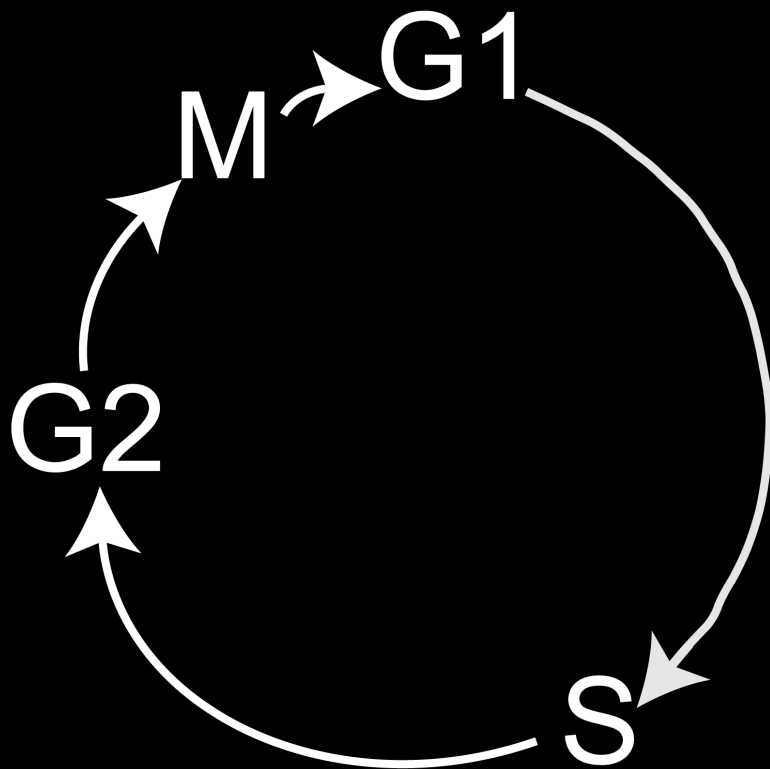


**How are developmental transition accurately  
controlled in time?**

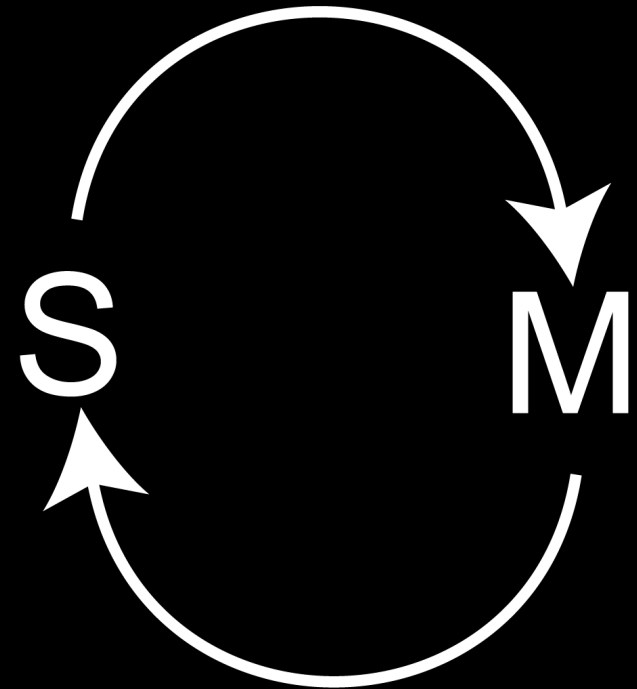


# The cell cycle as a model to study the precision of development

**Somatic** cell cycle



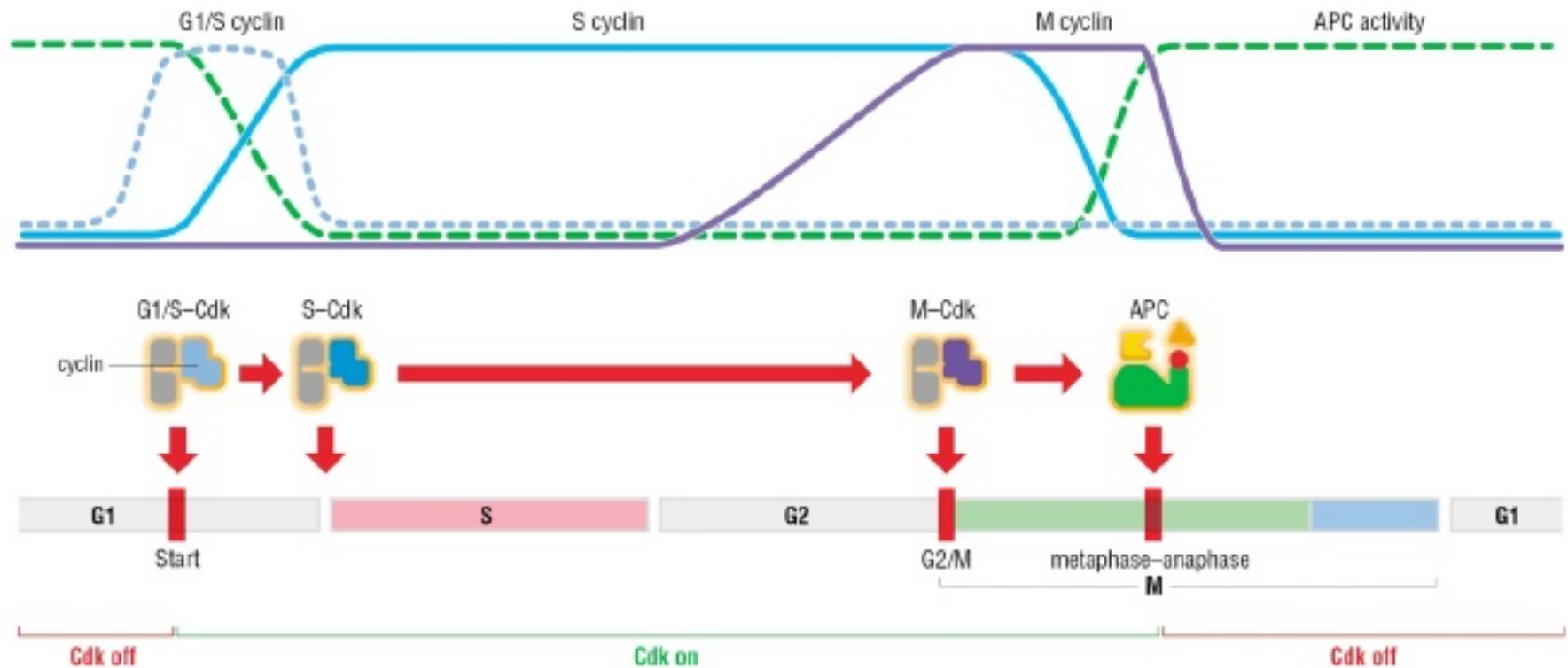
**Early embryonic** cell cycle



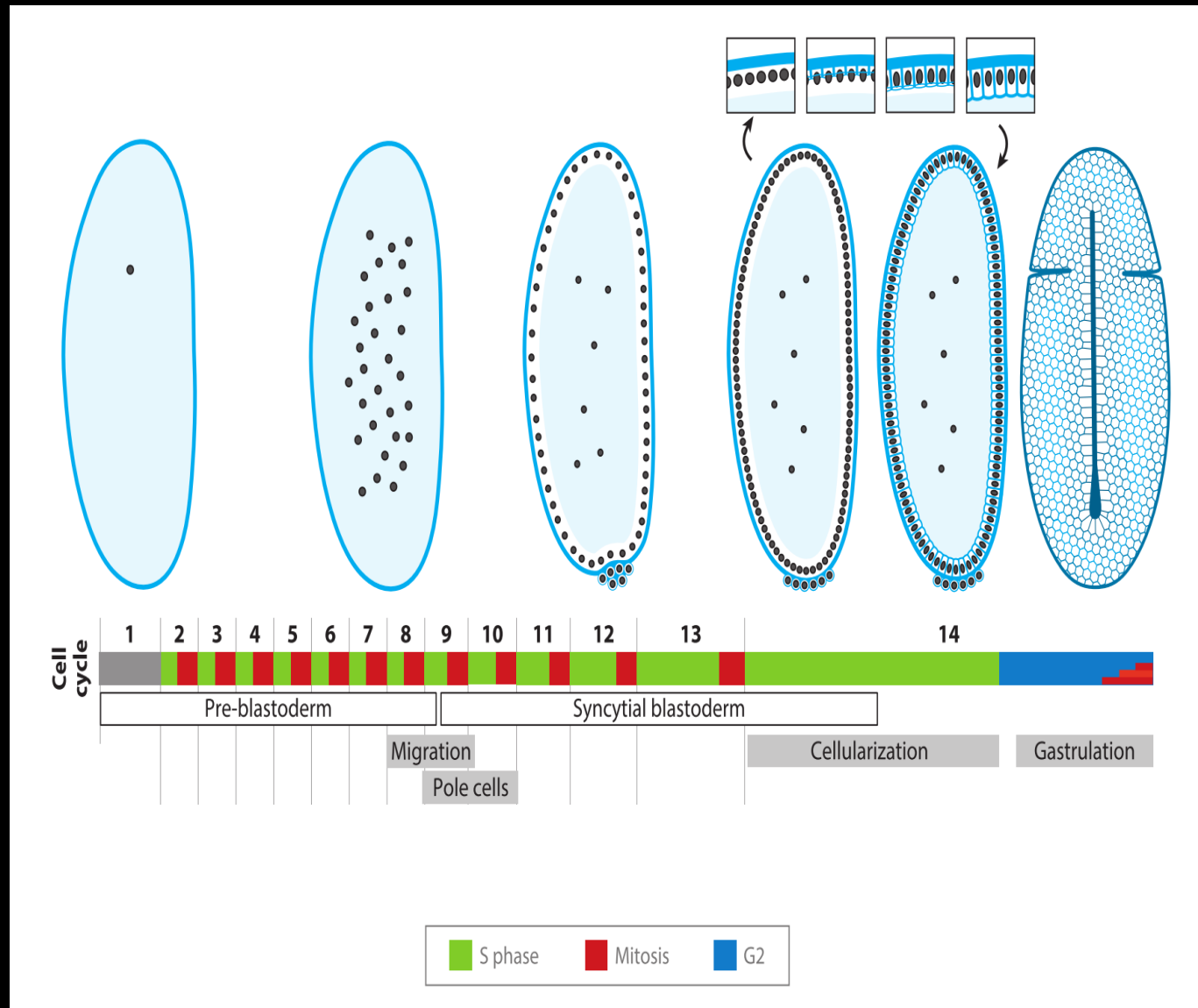
The major molecular pathways controlling the cell cycle have been identified

# The cell cycle is controlled by oscillations in Cdk activity

From **The Cell Cycle: Principles of Control** by David O Morgan

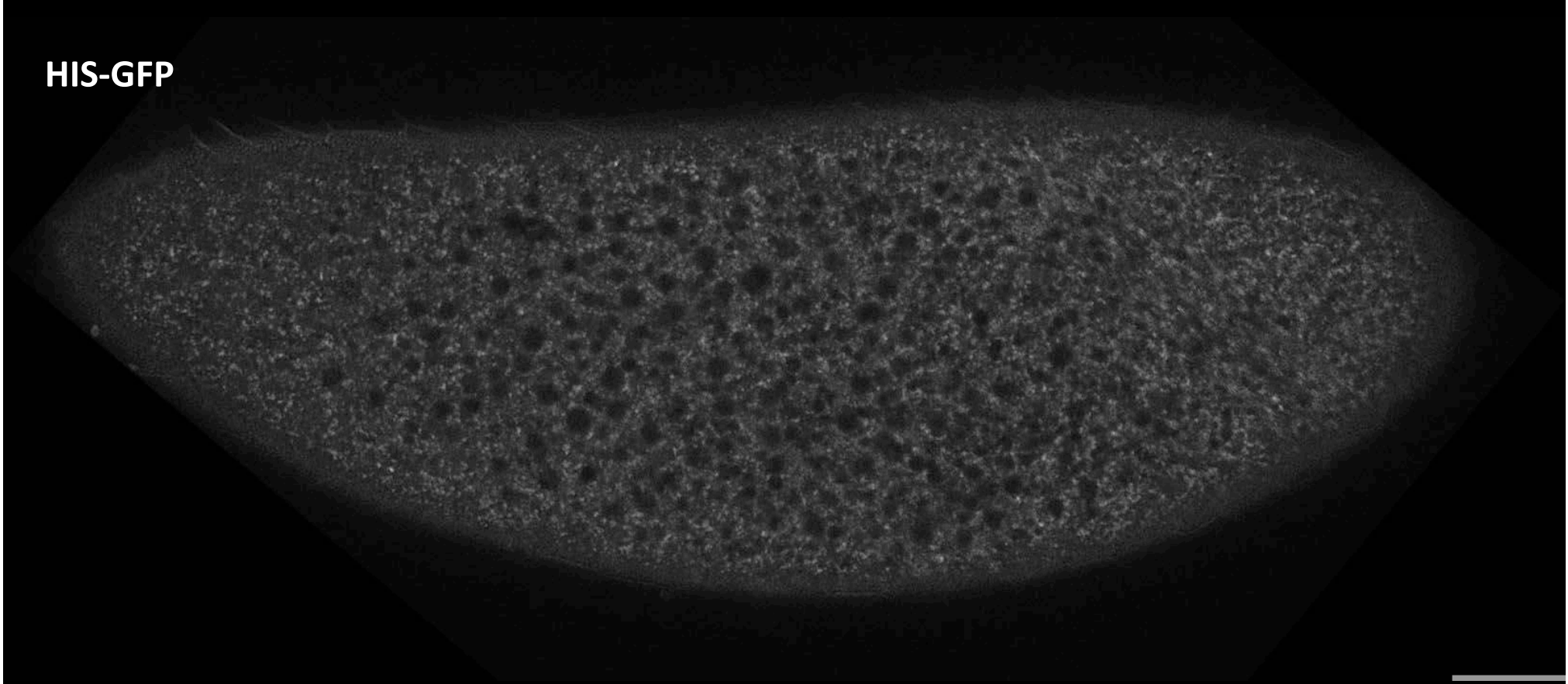


# Early development in *Drosophila*



# Visualization of *Drosophila* early cell cycles

HIS-GFP



# **Three remarkable examples of regulation**

- 1. Synchrony of the early mitosis**
- 2. Cell cycle pause at the mid-blastula transition**
- 3. Cell cycle re-entry in a spatiotemporal accurate pattern**

# Synchronization of mitosis across large scale ( $\sim 0.5$ mm)



**Victoria Deneke**



Anna Melbinger



Massimo Vergassola

UCSD, Physics

# How are mitotic events synchronized across large spatial scales?

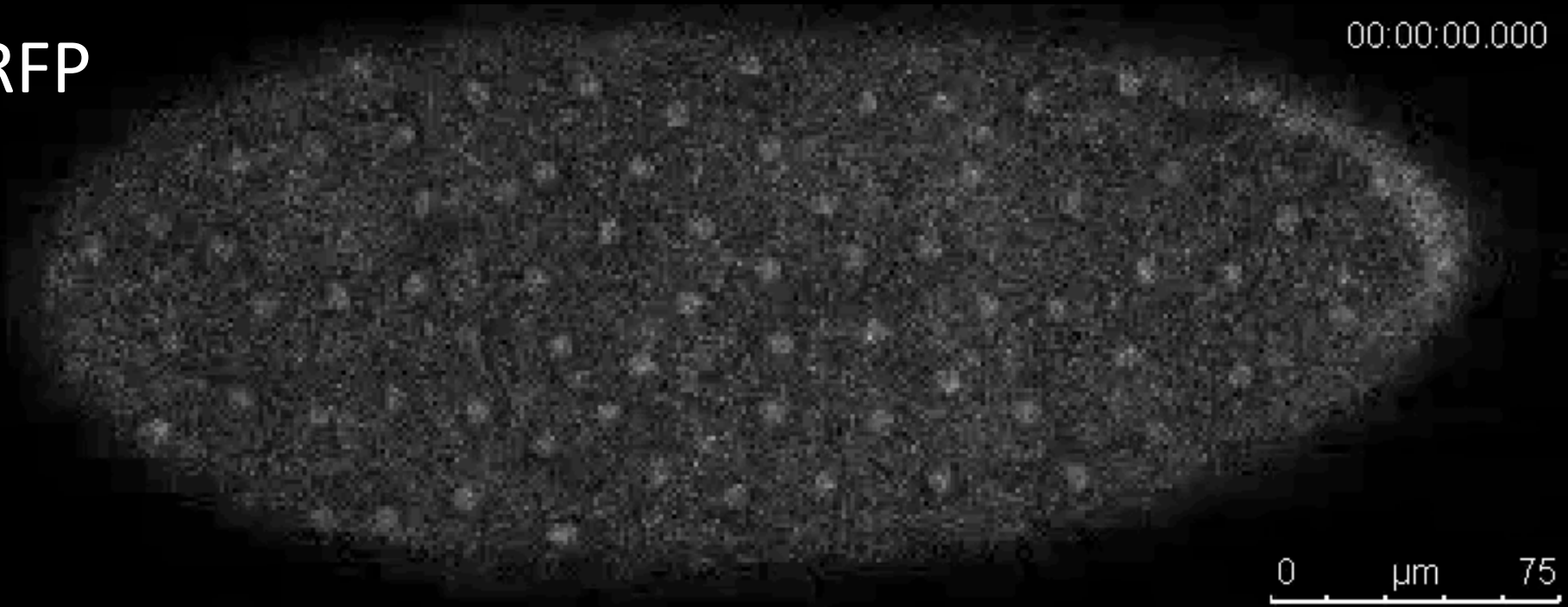
His-RFP

00:00:00.000

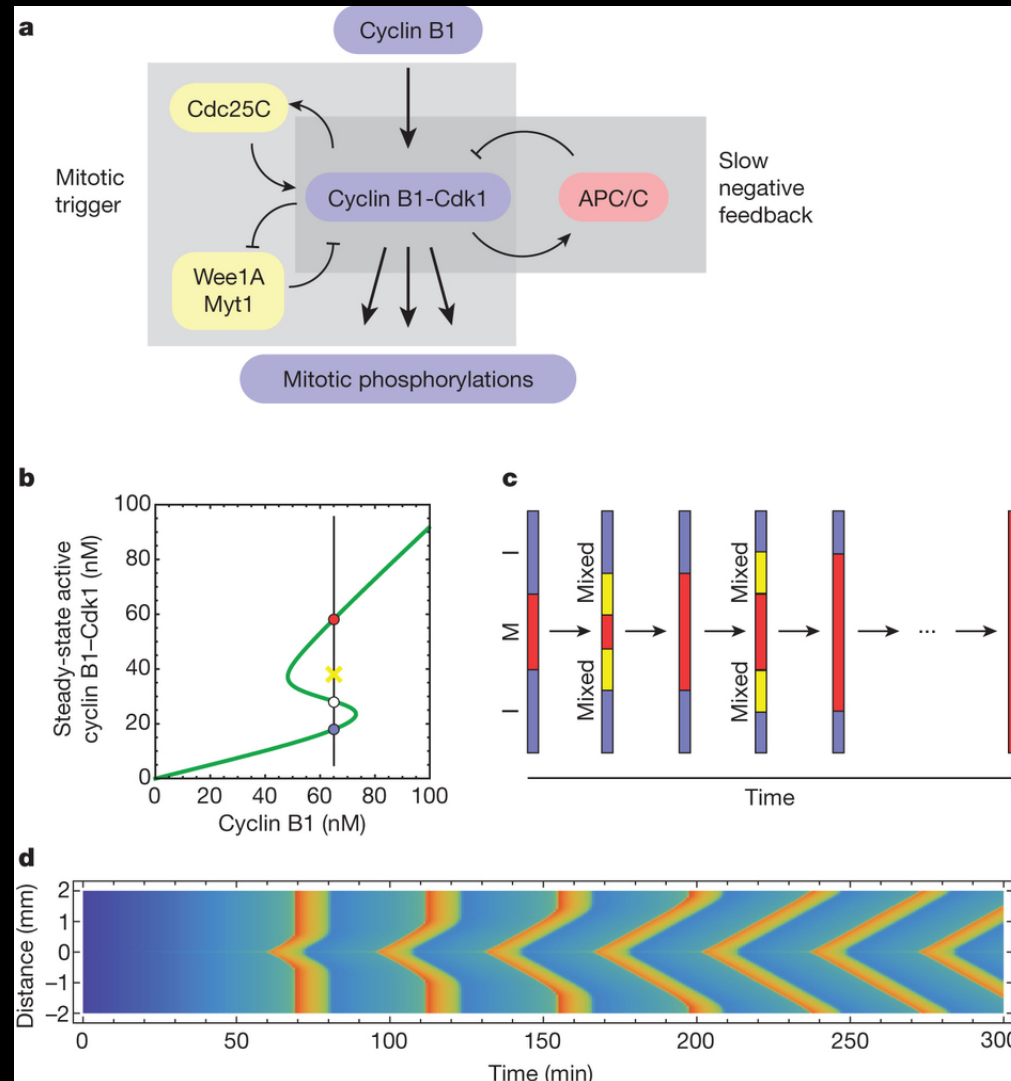
0  $\mu\text{m}$  75

$\sim 500\mu\text{m}$

Victoria Deneke



# Cdk1 trigger waves

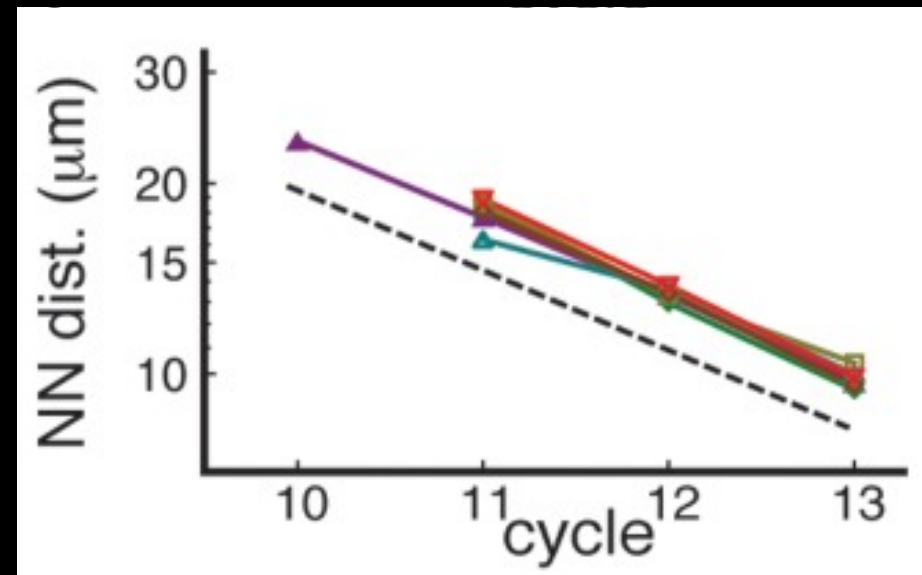
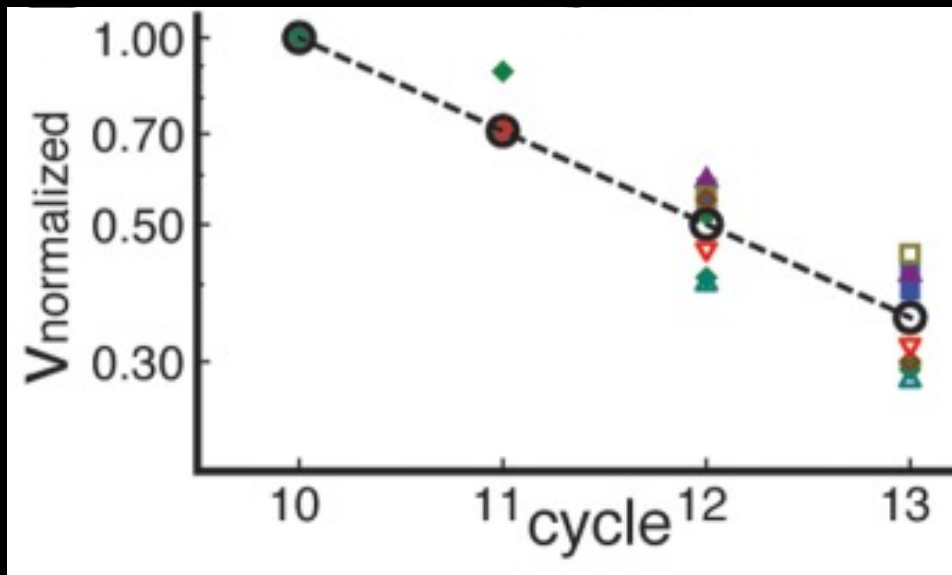


Novak and Tyson *J Cell Sc and J Theor Biol* 1993  
Chang and Ferrell *Nature* 2013



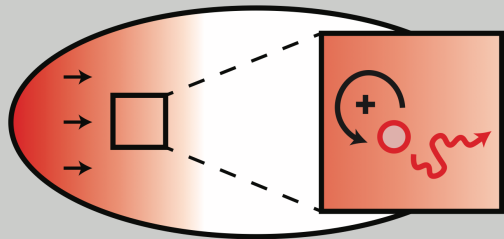
# Why do waves slow down during development?

Scaling of the mitotic wave speed with nucleus-to-nucleus distance in *Drosophila* has been proposed as a signature of mechanical waves propagating through the embryo

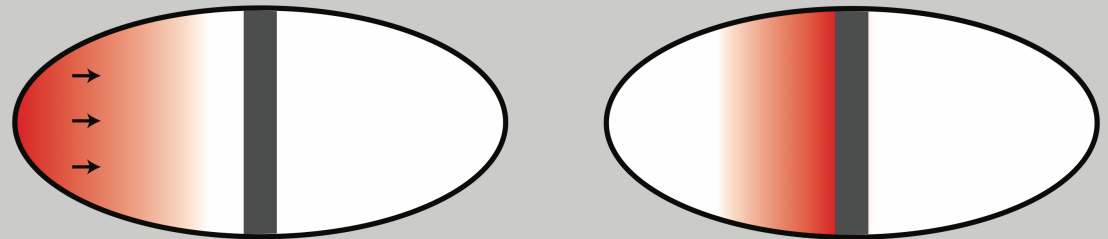


# Physical mechanisms of chemical waves

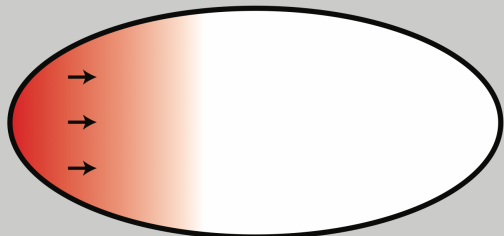
Trigger wave



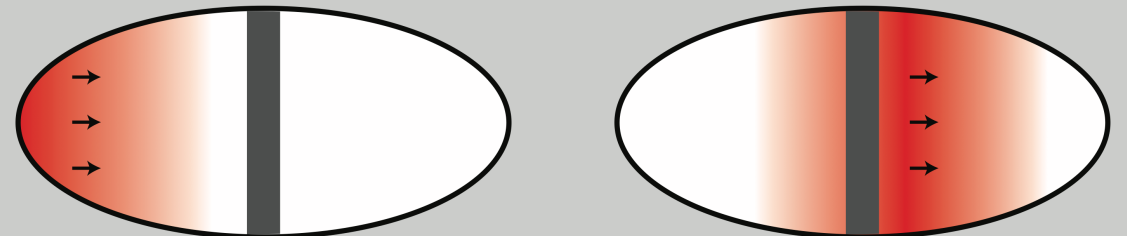
Trigger wave with barrier



Phase wave

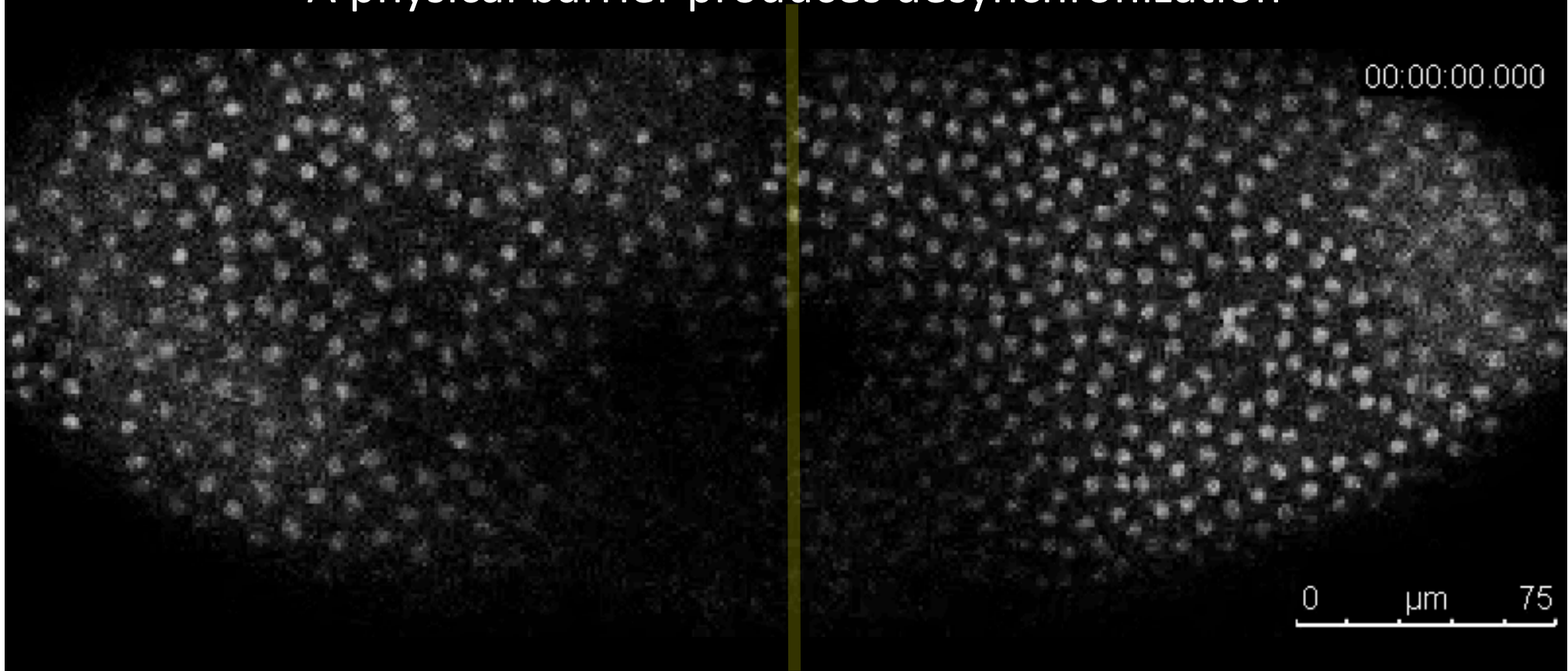


Phase wave with barrier



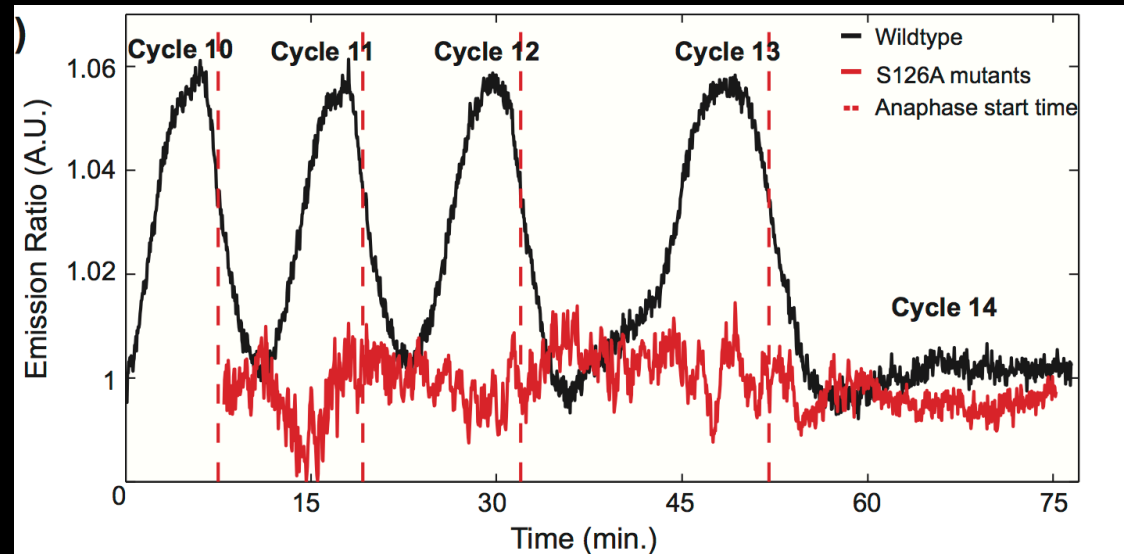
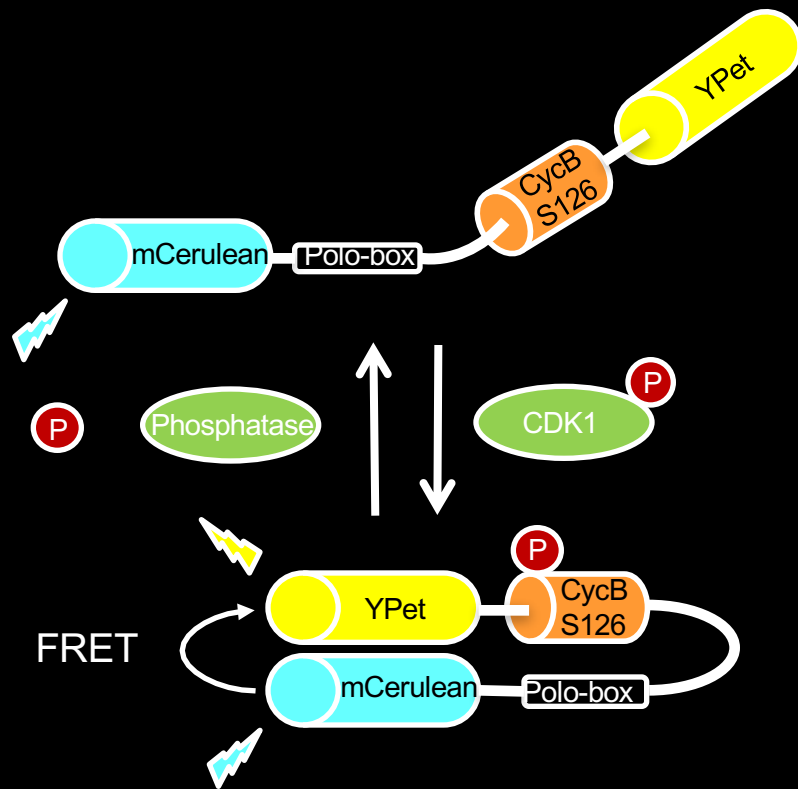
# Is it really collective or just well synchronized clocks?

A physical barrier produces desynchronization



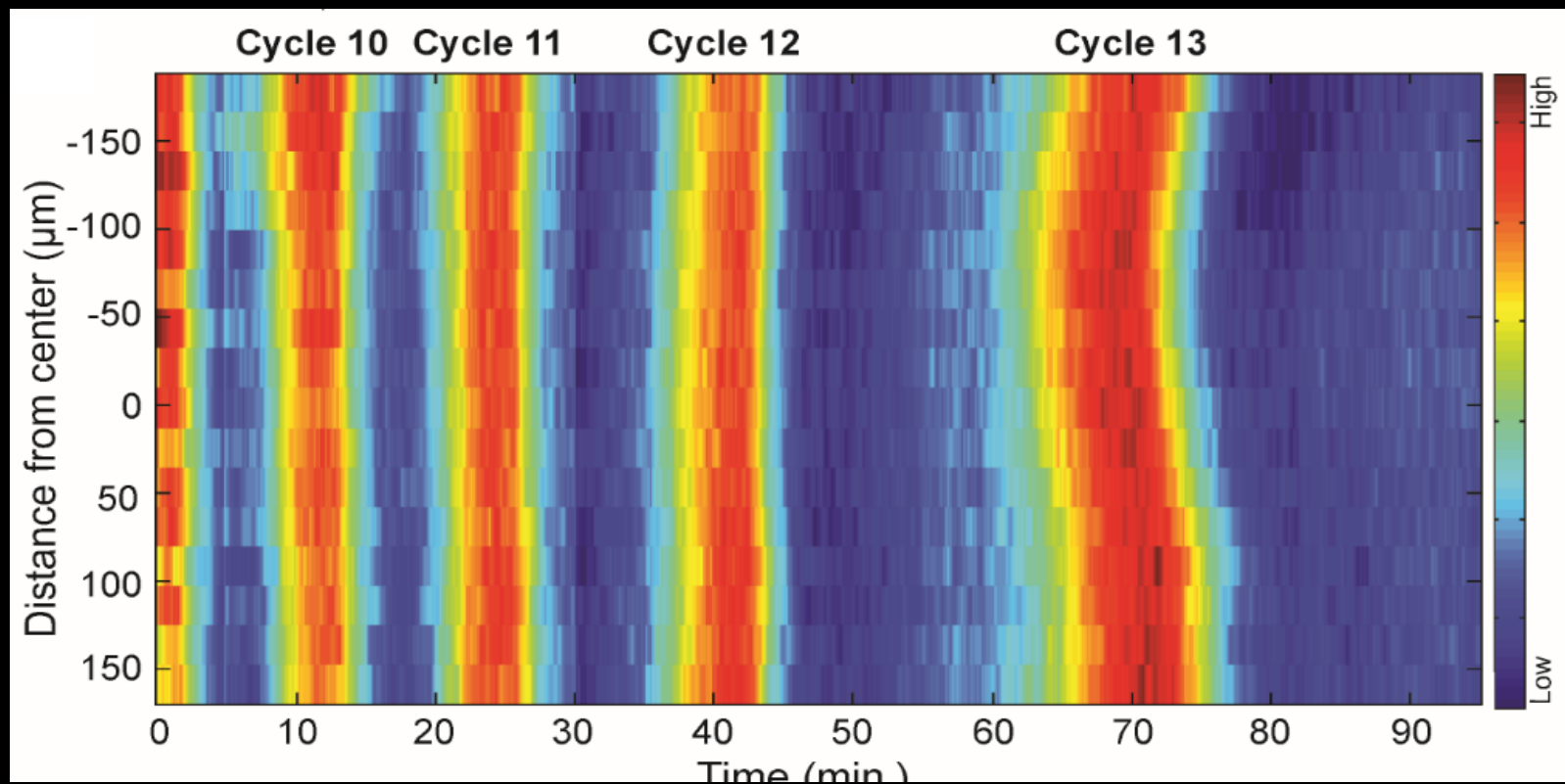
**Can we understand the molecular and physical mechanisms controlling the wave?**

# Cdk1 FRET biosensor specifically measures Cdk1 activity

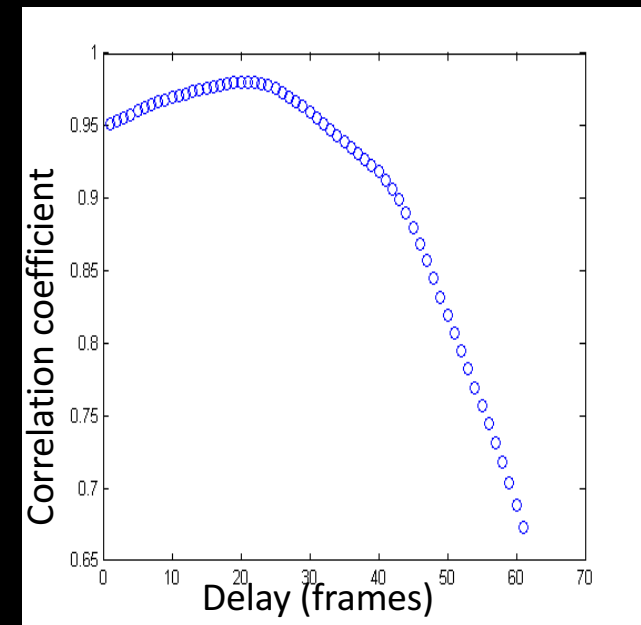
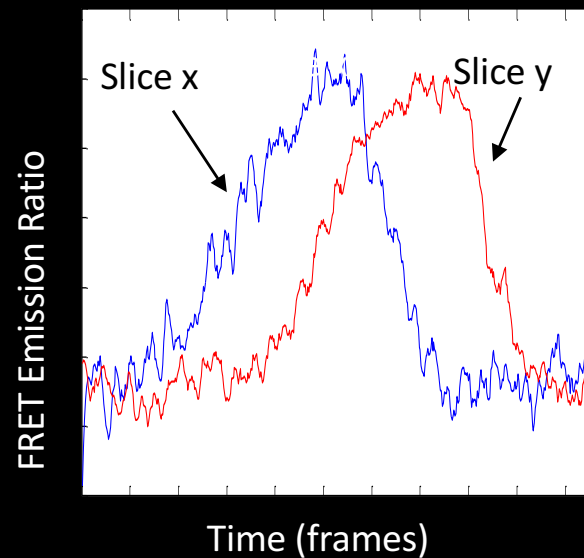
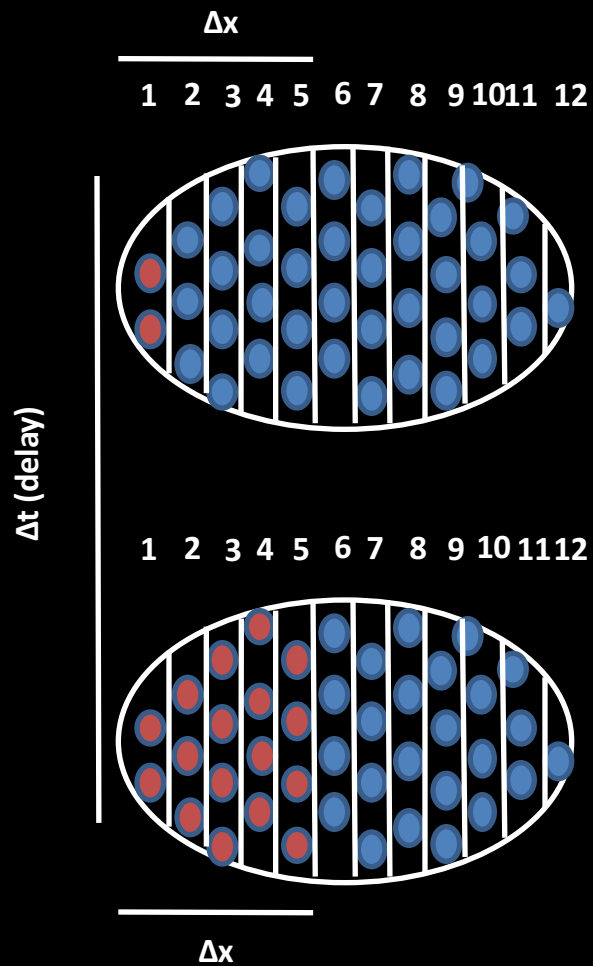


Gavet and Pines Dev Cell and JCB 2010

# Measuring Cdk1 activity over multiple cell cycles with seconds resolution

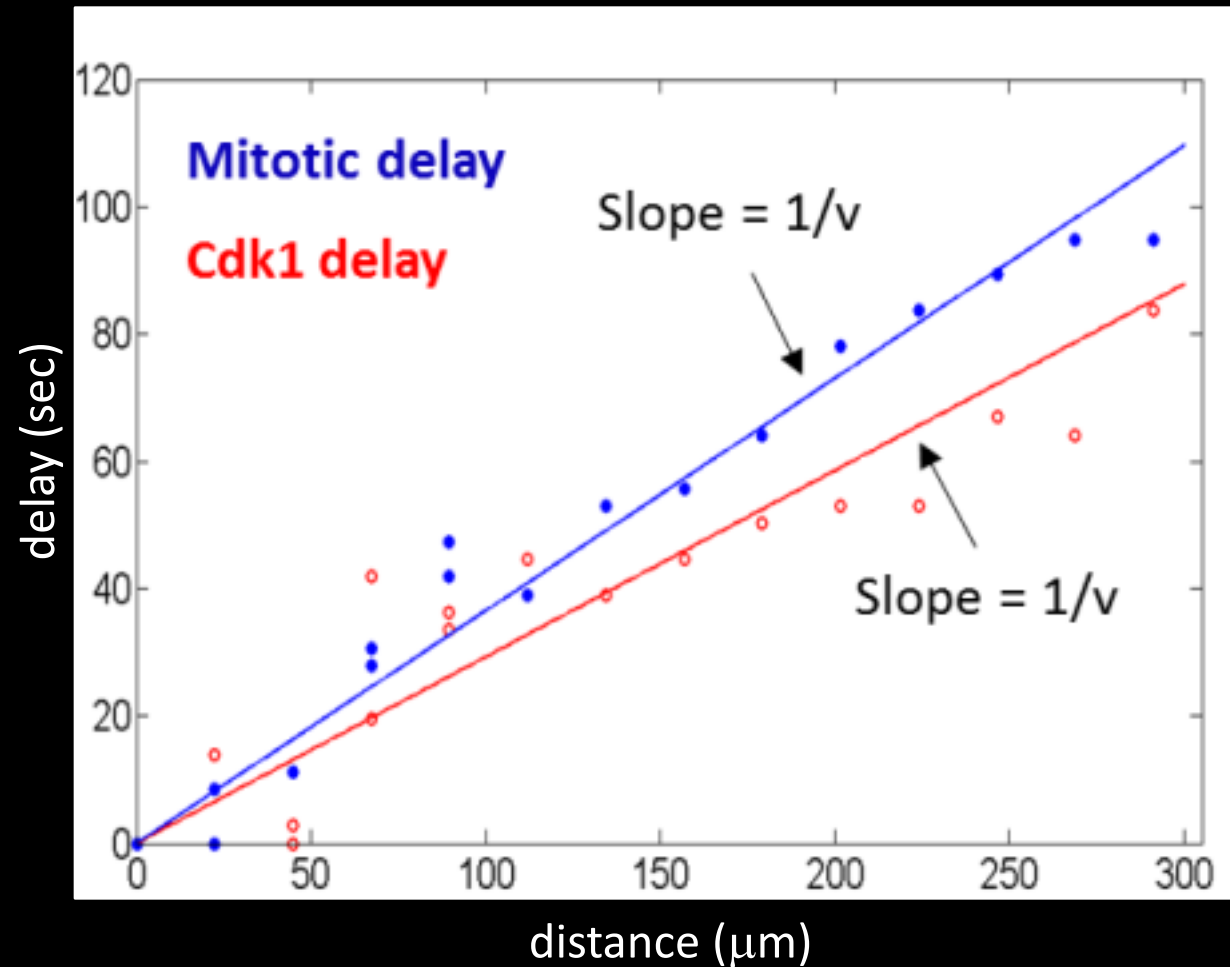
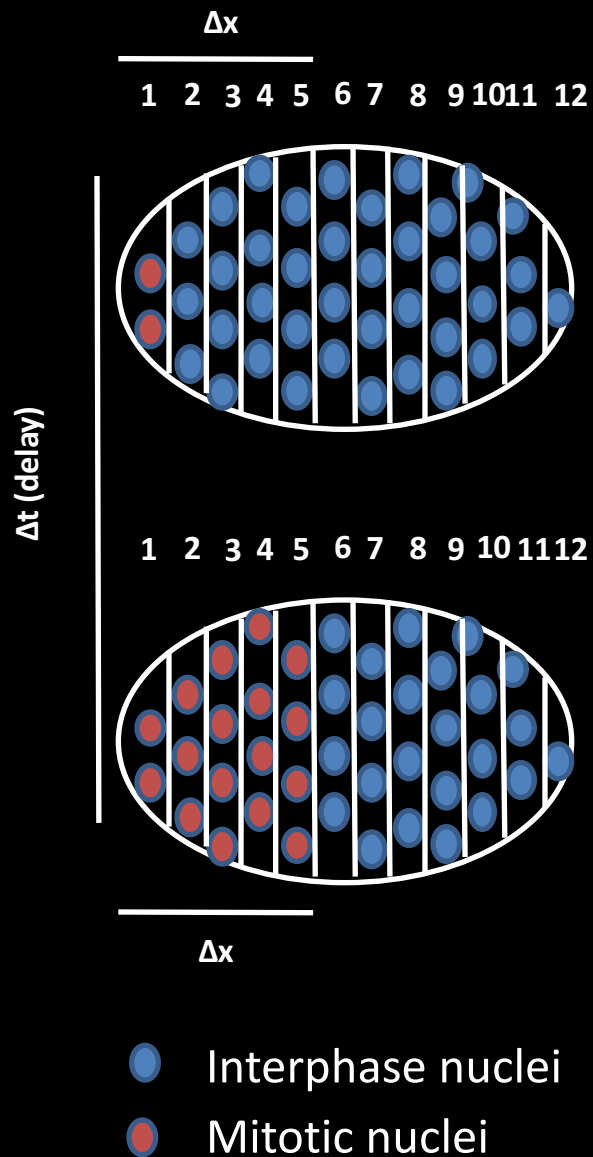


# Wave speed can be determined from time delays in vertical slices

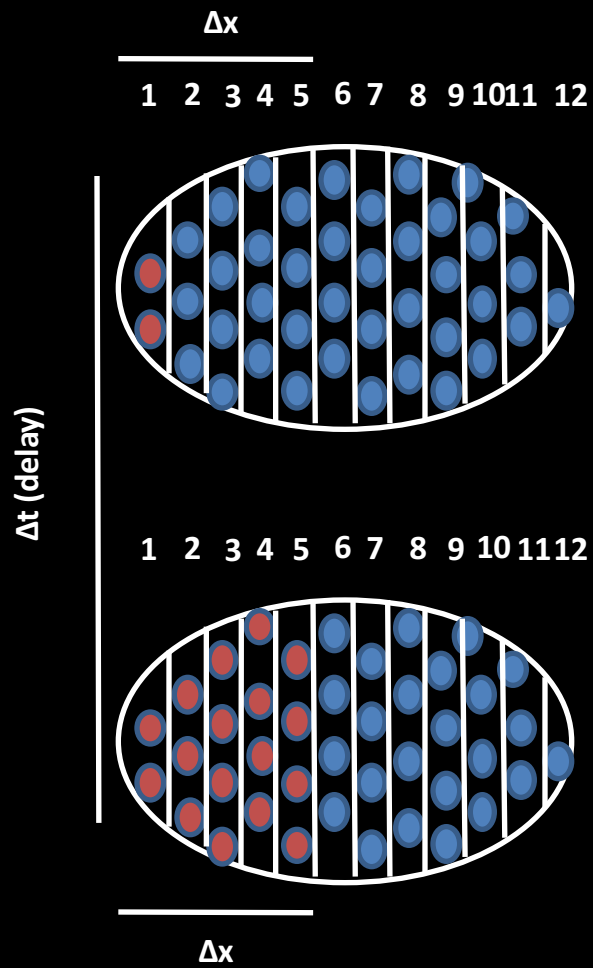


- Interphase nuclei
- Mitotic nuclei

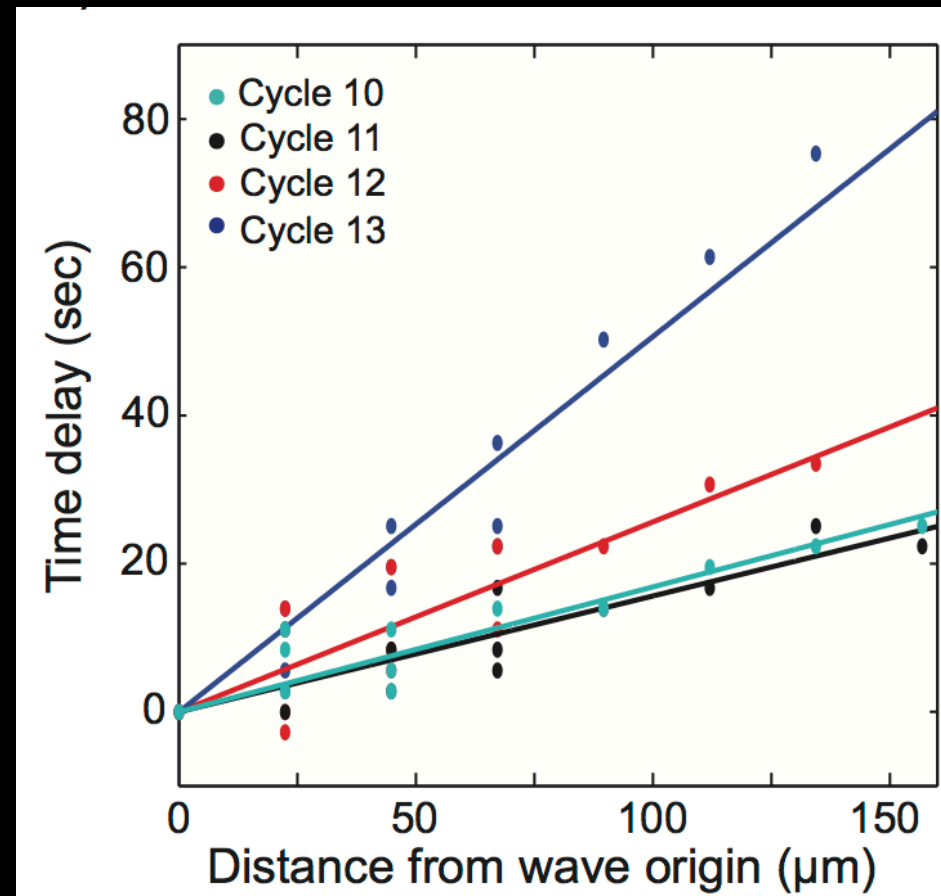
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# Wave speed can be determined from time delays in vertical slices

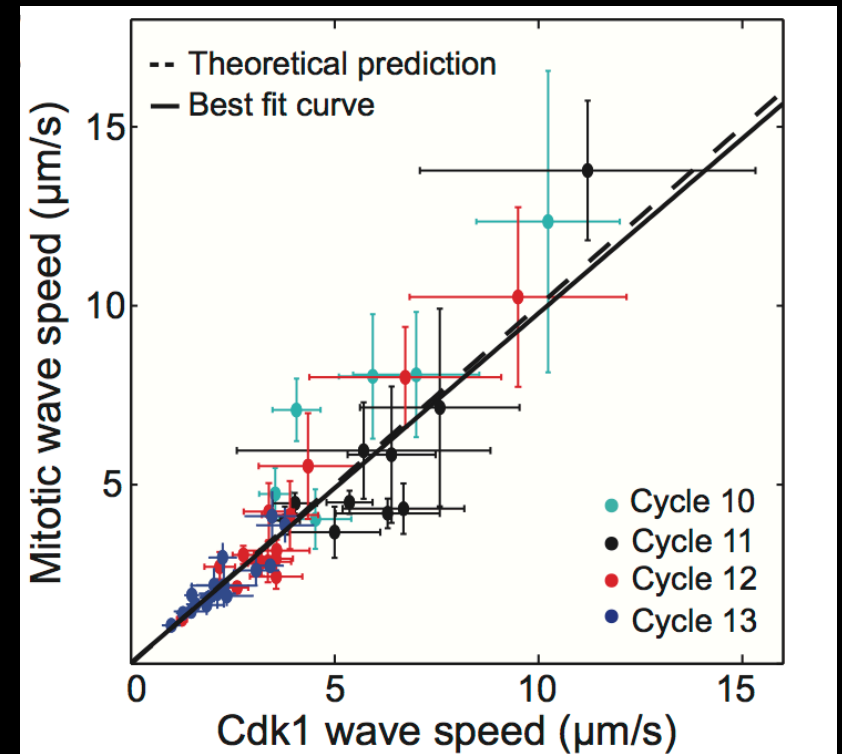
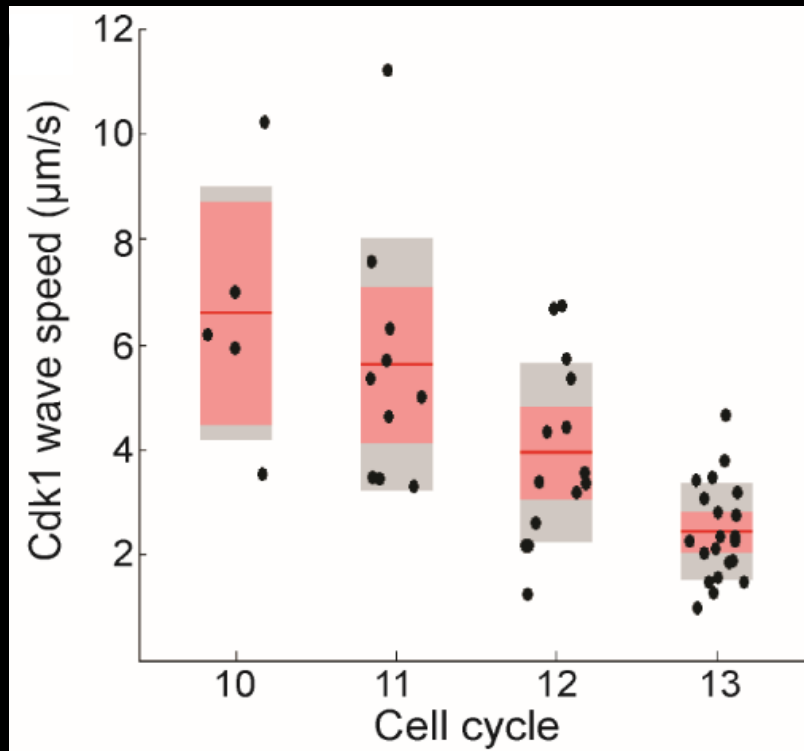


- Interphase nuclei
- Mitotic nuclei





# Cdk1 waves control mitotic waves



Can we understand what controls the physical properties of the waves?

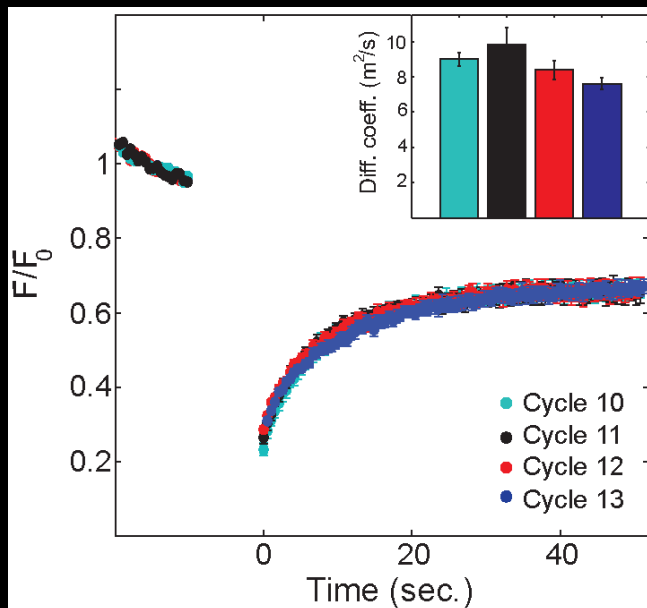
# Back of the envelope

Dimensional analysis suggests that the speed should scale as:

$$v \approx \sqrt{\frac{D}{\tau}}$$

where  $D$  is the diffusion coefficient and  $\tau$  is the relevant timescale.

FRAP measurements of the diffusion coefficient of Cdk1-YFP



**Diffusion cannot explain  
why the waves slow down**

# Back of the envelope

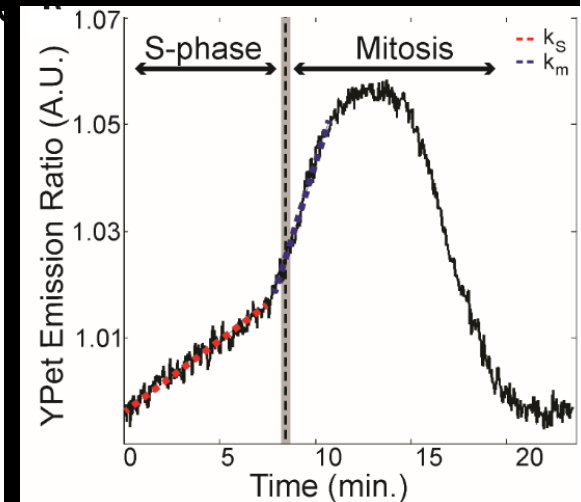
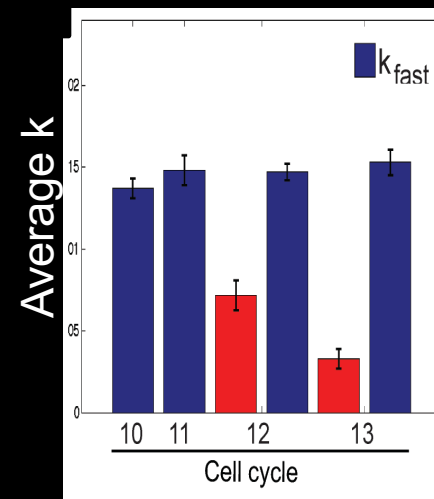
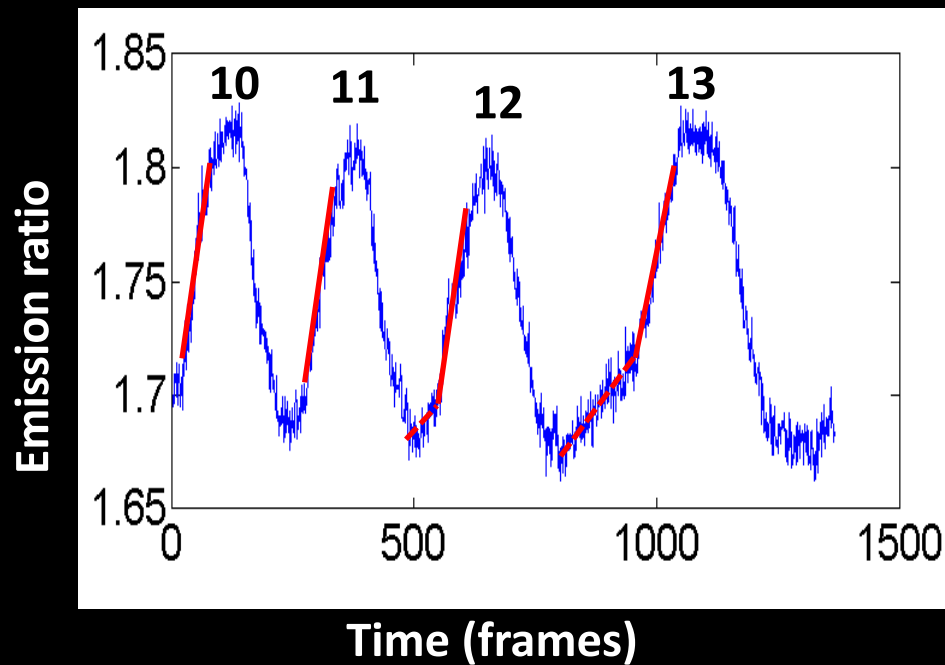
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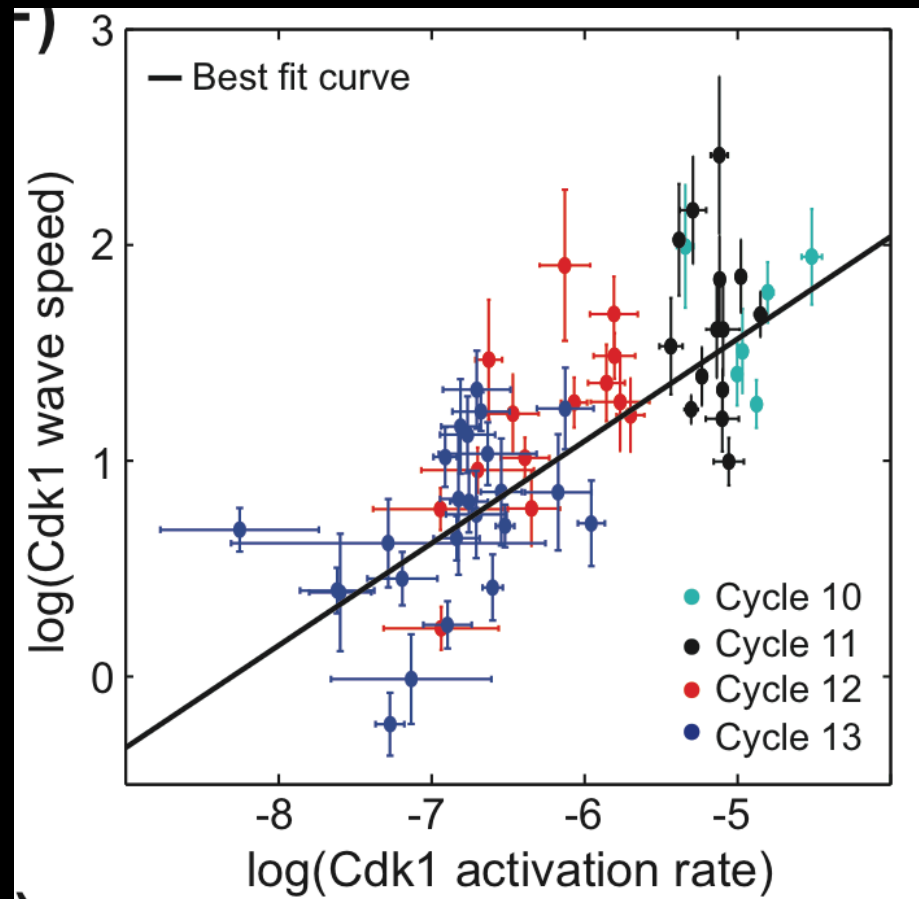
**Which changes in the Cdk1 reaction dynamics can explain the change in speed?**

# The activity of Cdk1 during mitosis does not regulate the speed of Cdk1 waves

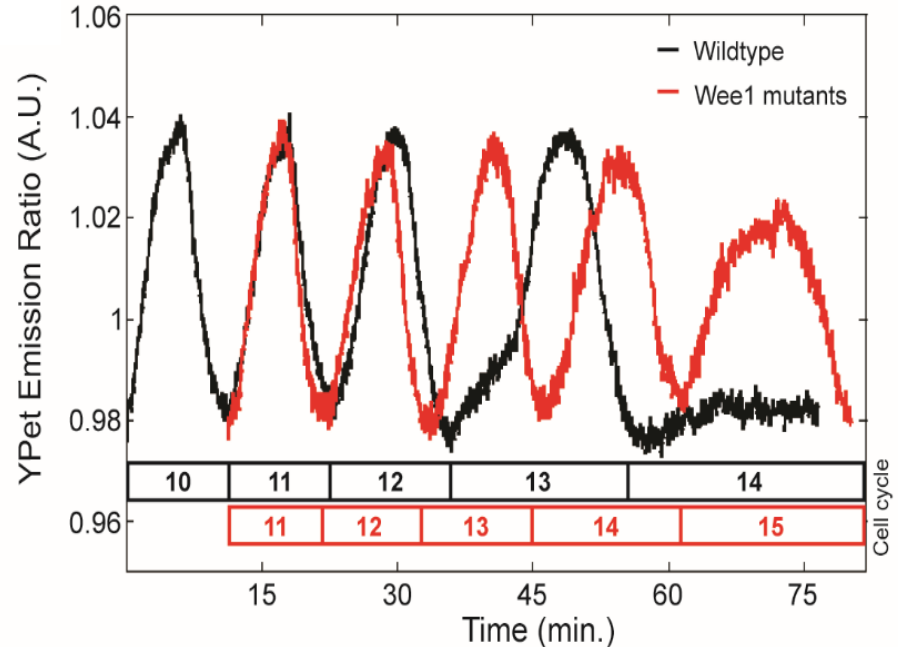
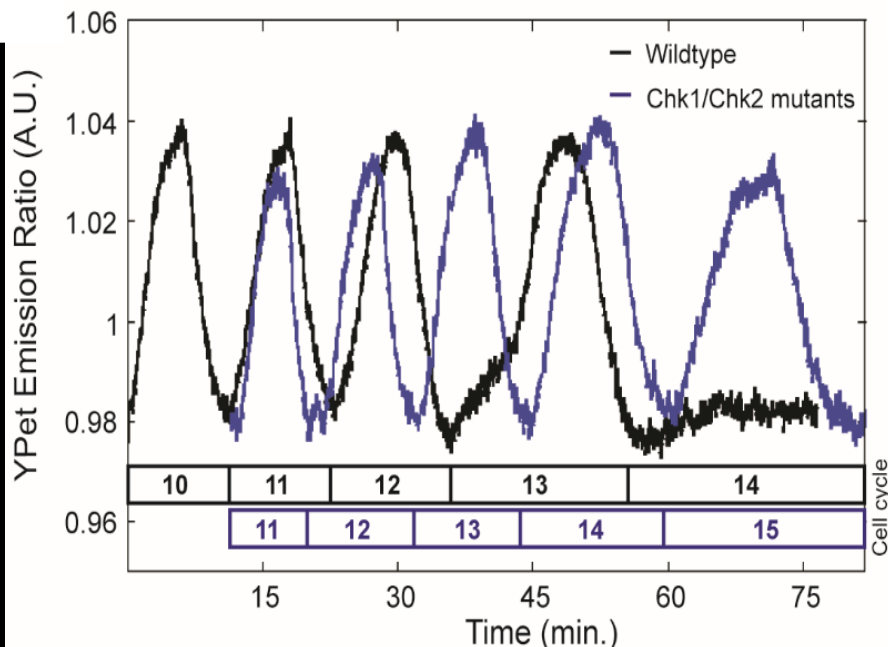
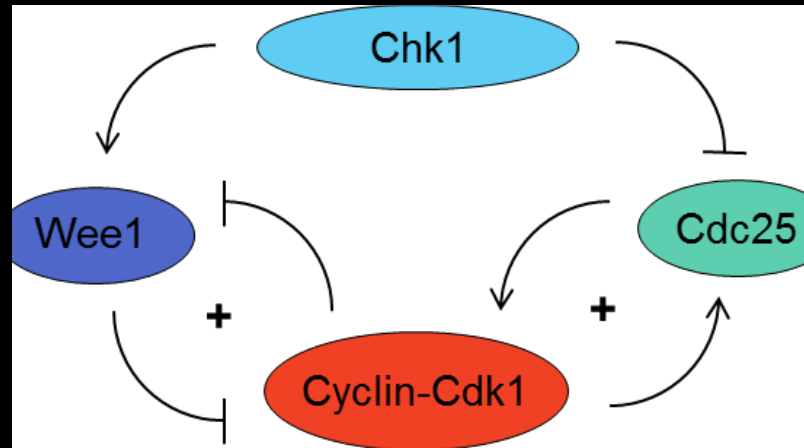


# Cdk1 wave speed can be predicted by the rate of activation of Cdk1 in S-phase

$$v \sim (Dk_S)^{1/2}; \log(v) = 0.5 * \log(k_S) + C$$

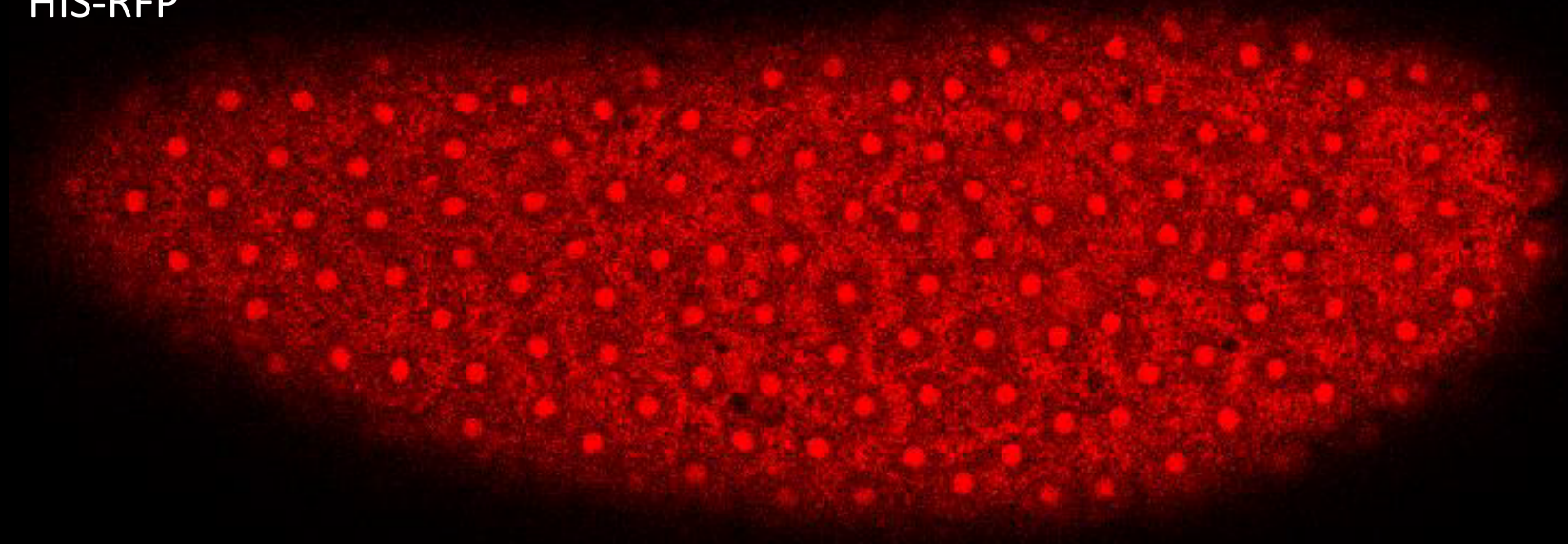


# The dynamics of Cdk1 is regulated by the DNA replication checkpoint via Chk1/Wee1

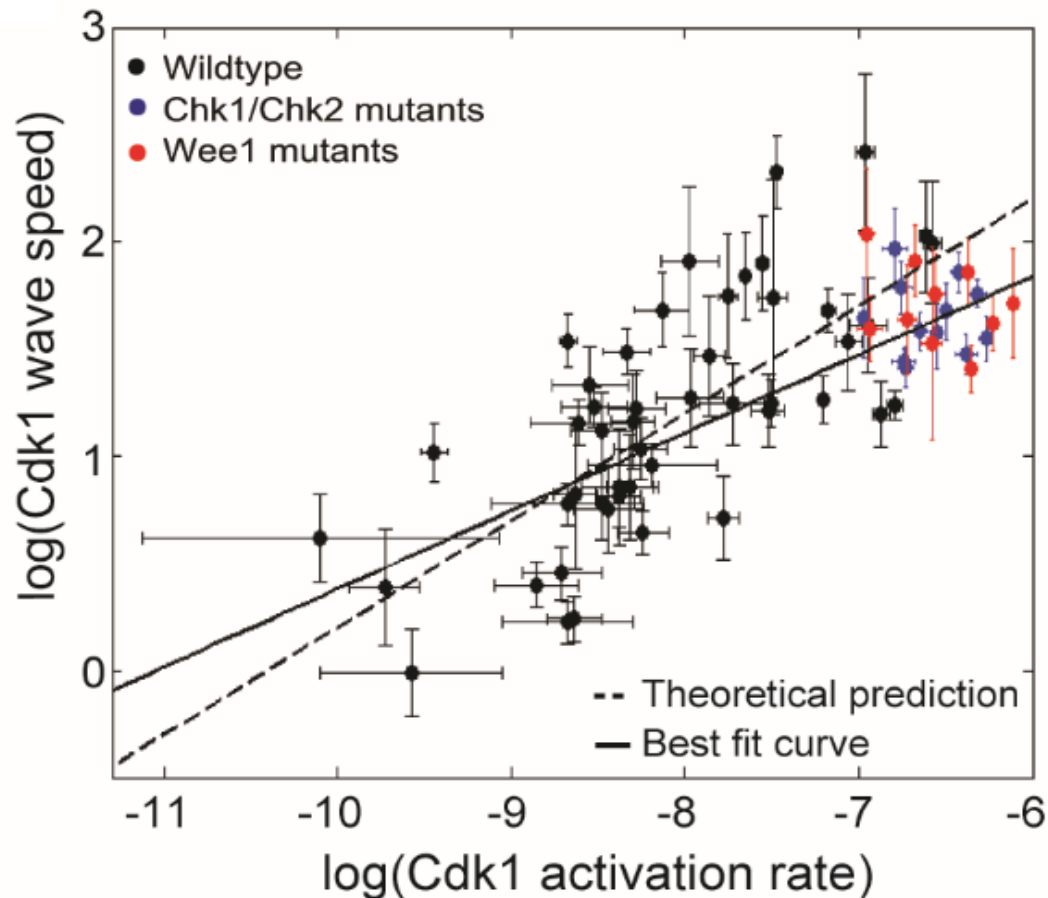


# The mitotic waves do not slow down in *chk1 chk2* mutants

HIS-RFP

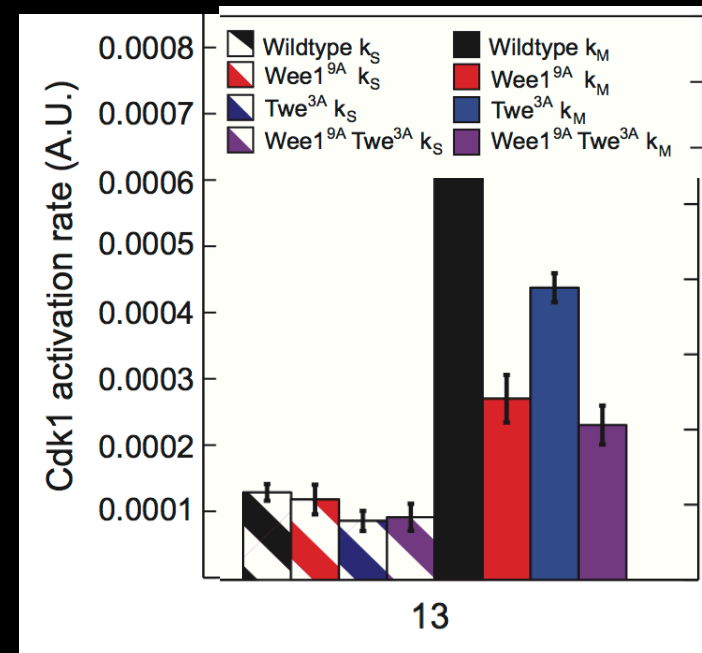
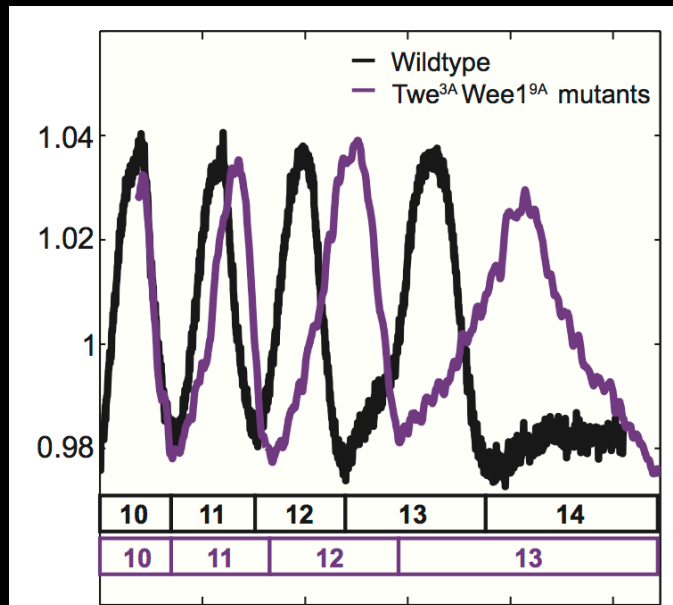
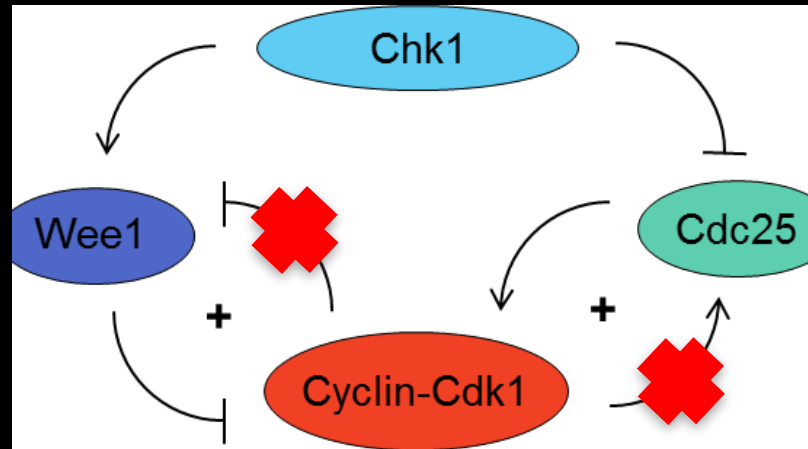


# The slowdown of the Cdk1 waves requires the Chk1/Wee1 pathway

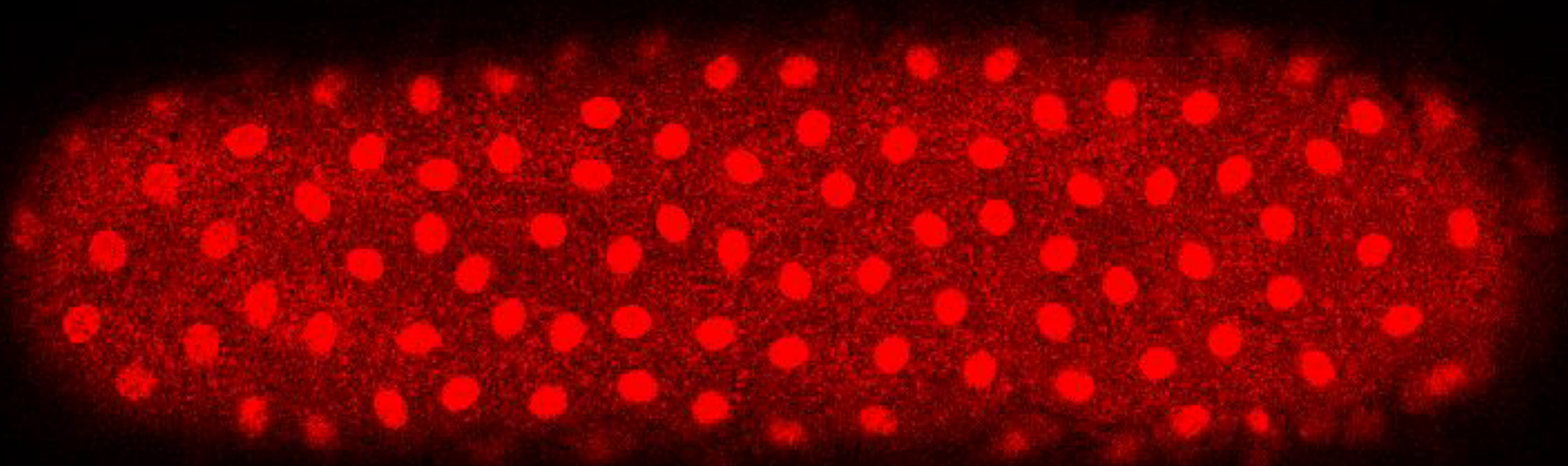




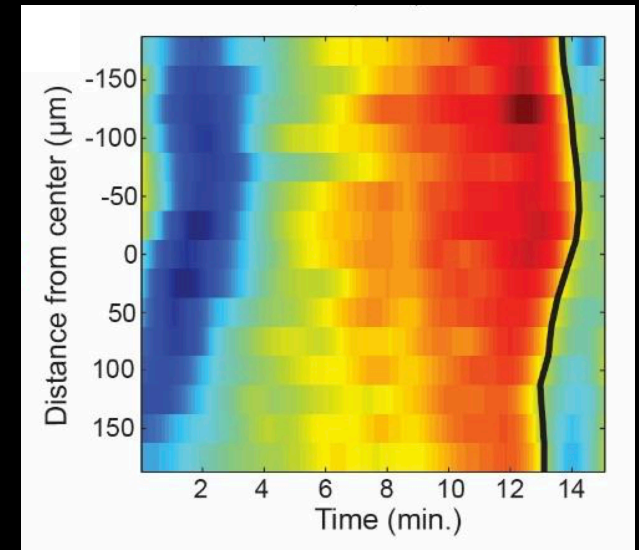
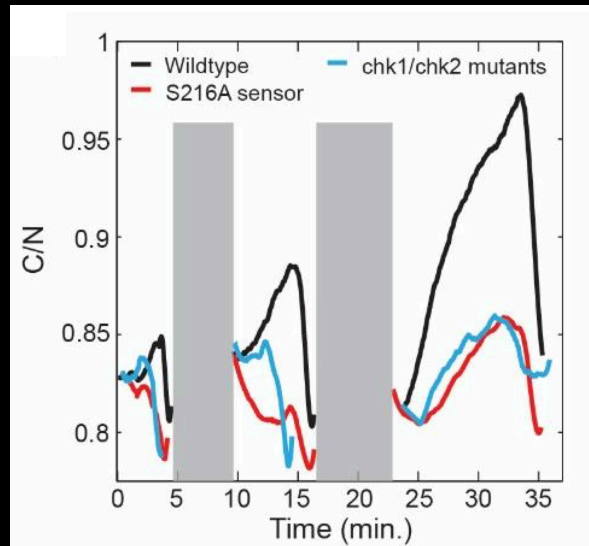
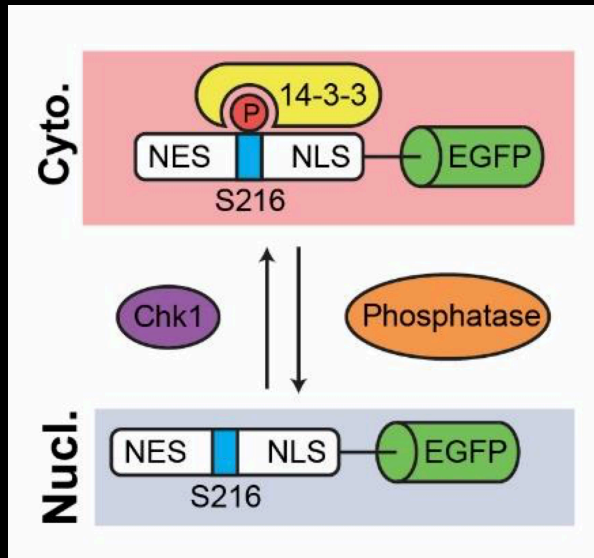
# Disabling the Cdk1/Wee1/Cdc25 feedbacks alters the rapid phase of Cdk1 activation



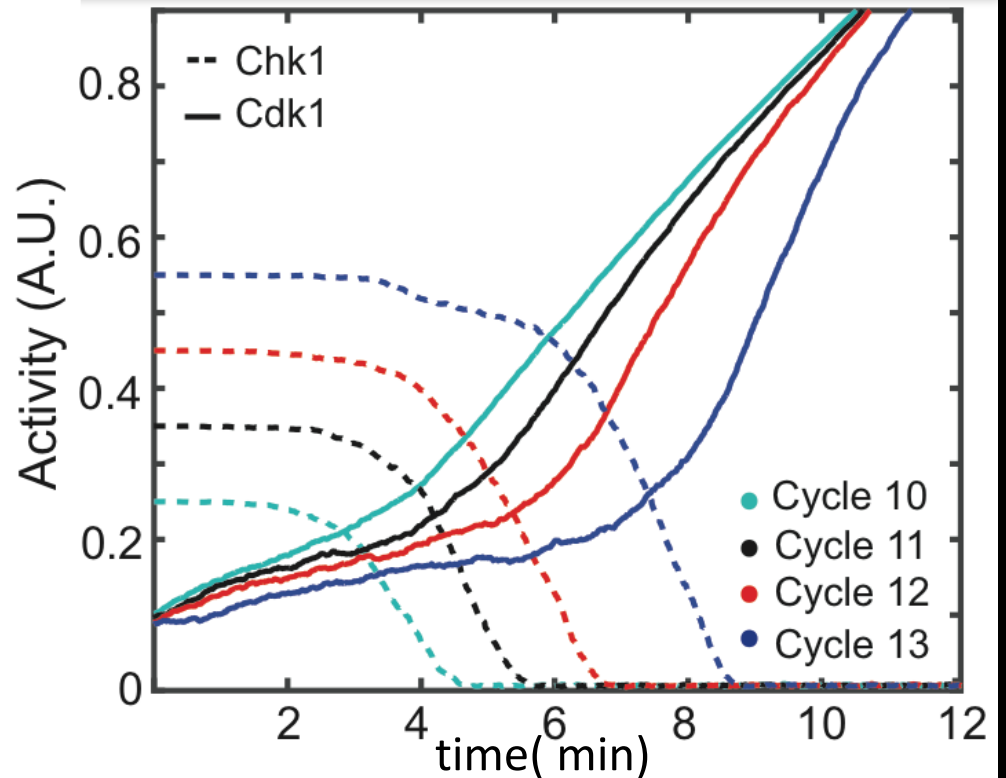
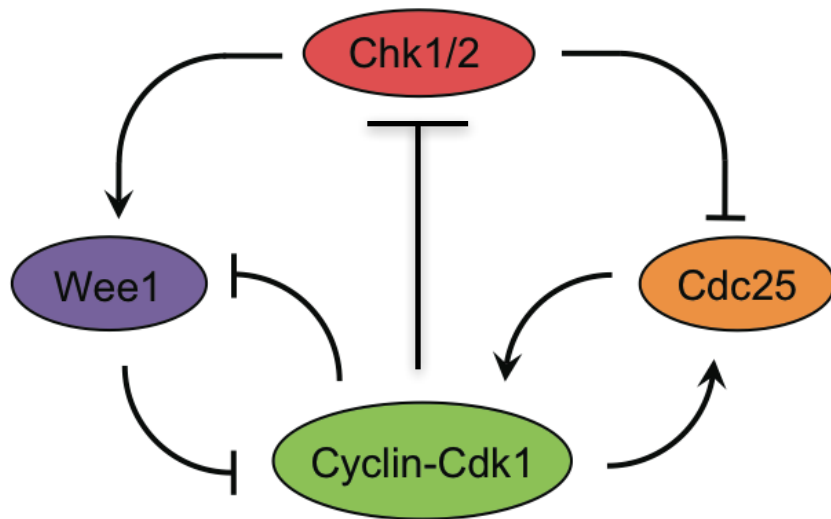
**Disabling the Cdk1/Wee1/Cdc25 feedbacks does not prevent the propagation of mitotic waves**



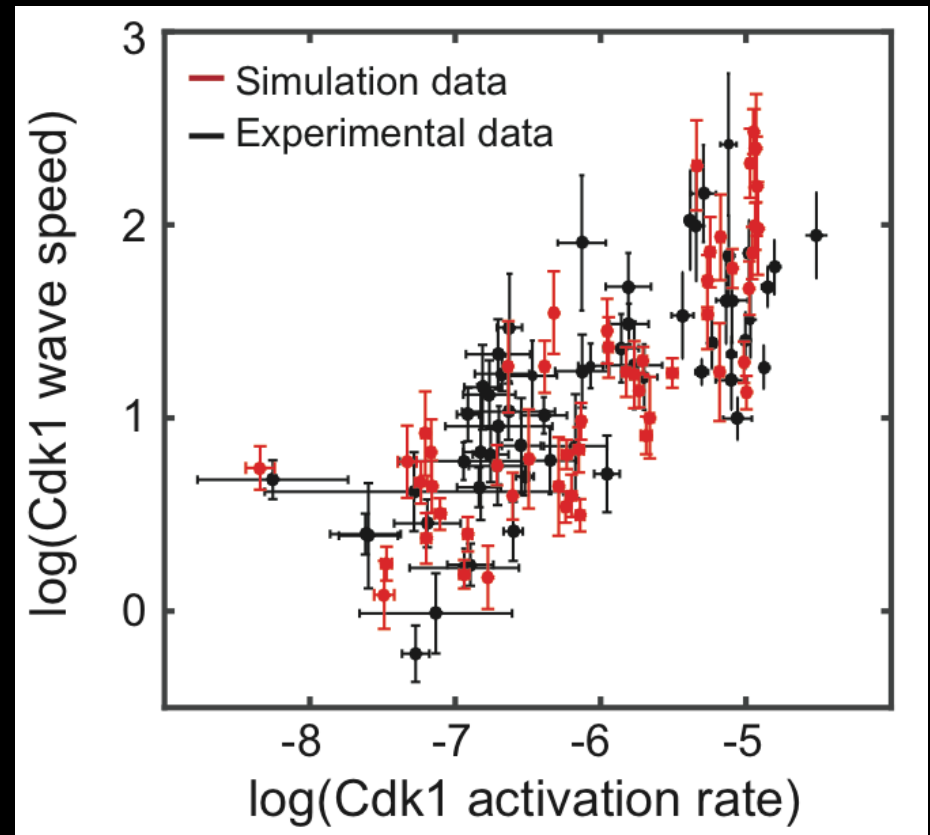
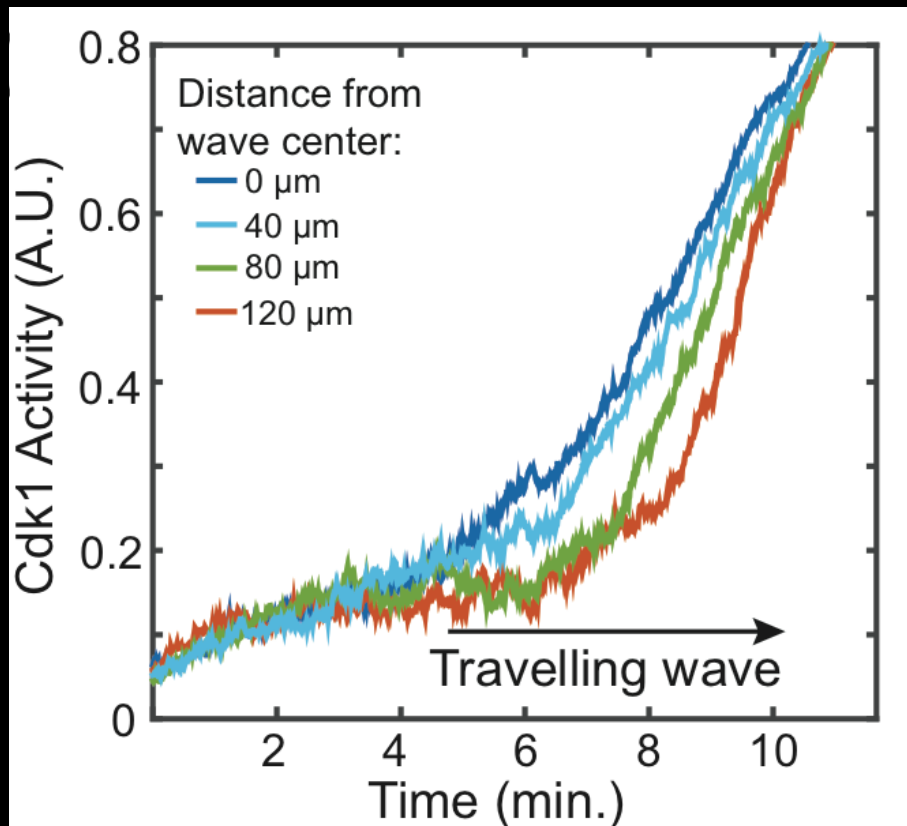
# Chk1 activity is turned off in a wave-like pattern



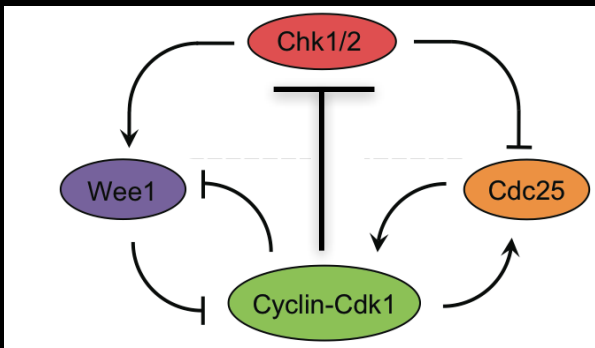
# A mathematical model of Cdk1 waves



# A simple model can reproduce the scaling of the speed of Cdk1 waves



# Reduction to an effective one-species dynamics



$$\frac{\partial f}{\partial t} = D_{Chk1} \frac{\partial^2 f}{\partial x^2} - \frac{a^\sigma}{K_{Chk1}^\sigma + a^\sigma} r_0 f + \xi_f(x, t)$$

$$\frac{\partial a}{\partial t} = D_{Cdk1} \frac{\partial^2 a}{\partial x^2} + \alpha + r_+(a, f)(c(x, t) - a) - r_-(a, f)a + \xi_c(x, t) + \xi_r(x, t)$$

$$\frac{\partial c}{\partial t} = D_{Cdk1} \frac{\partial^2 c}{\partial x^2} + \alpha + \xi_c(x, t)$$

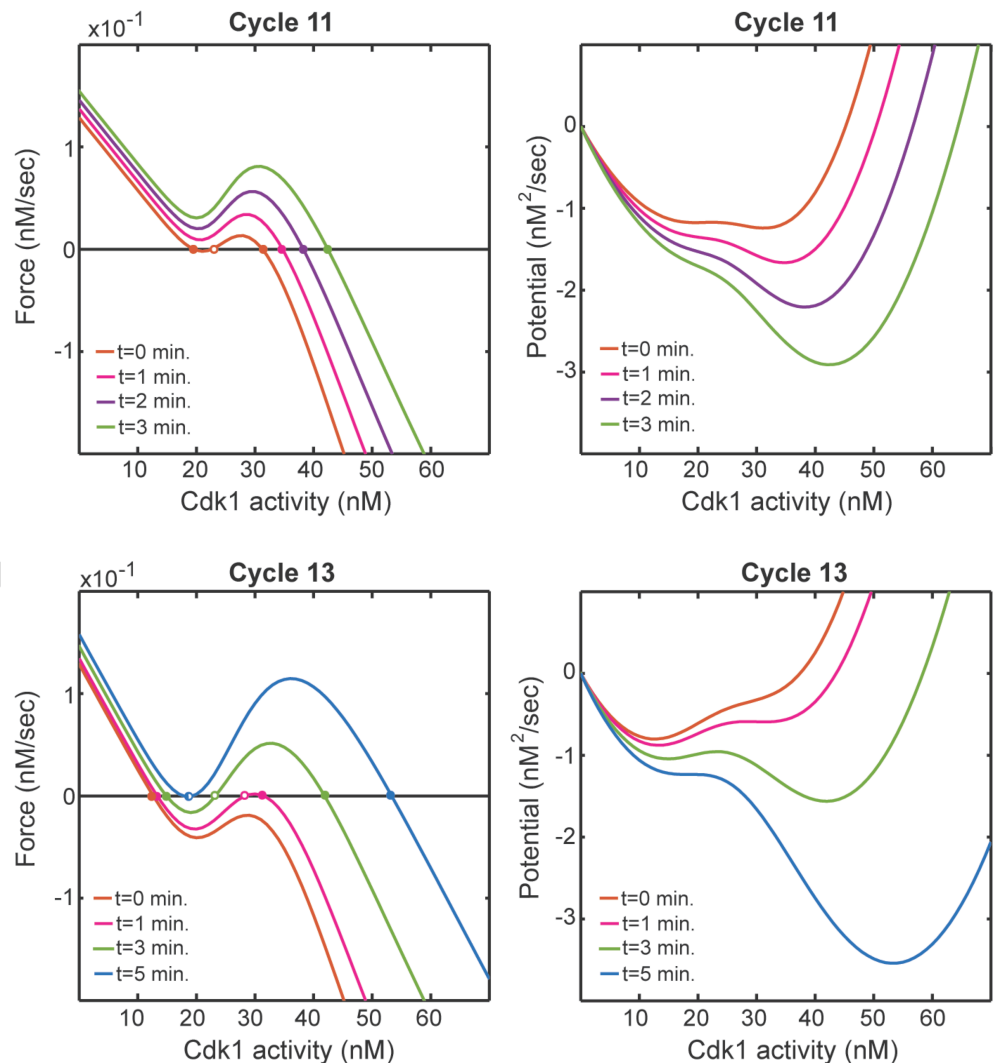
1. The dynamics of  $f$  is fast and effectively slaved to the dynamics of Cdk1
2. The dynamics of  $c$  (total Cdk1) is correlated over scales  $(4Dt)^{1/2} \approx 70 \mu\text{m}$  large compared to  $D/v \approx 1 \mu\text{m}$  so safely  $c(x, t) \approx c(t)$
3. Curvature of the embryo safely ignored by same reasoning

# Effective one-species dynamics

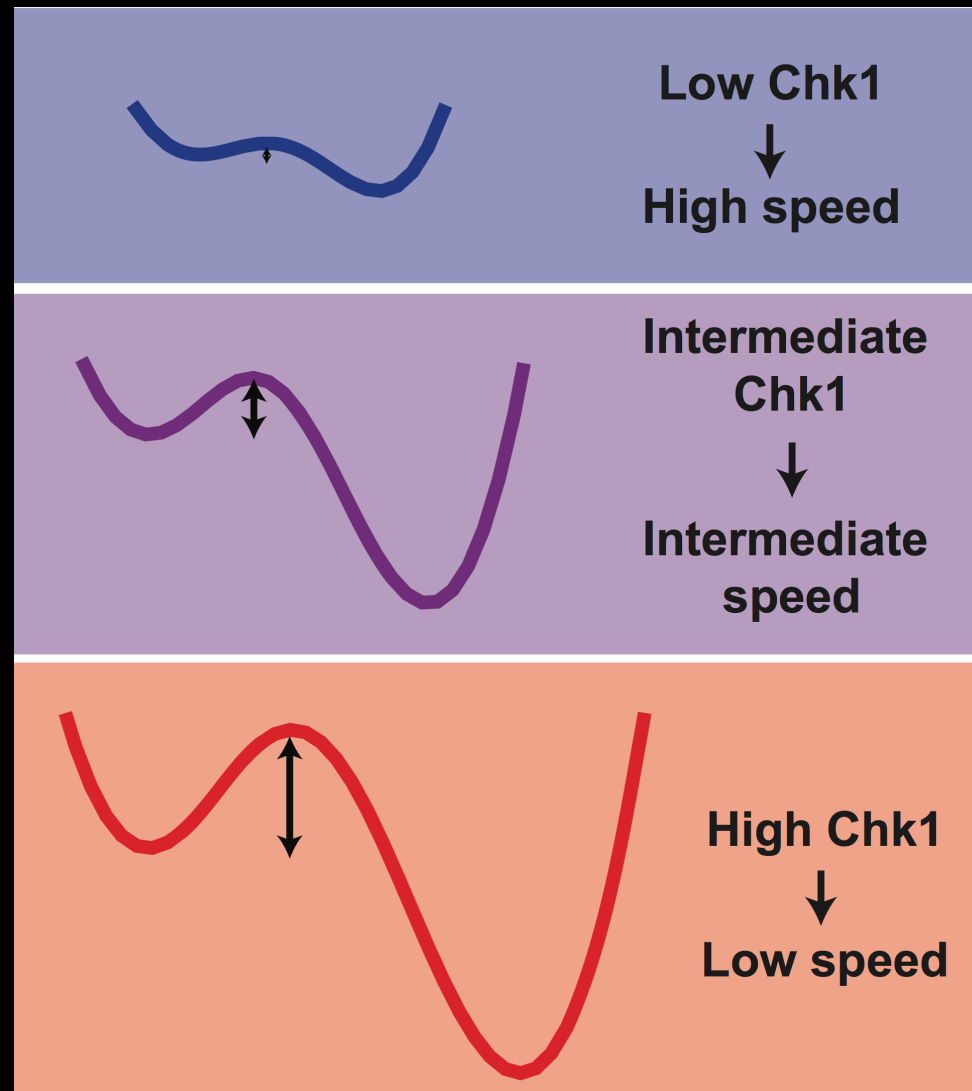
$$\frac{\partial a}{\partial t} = D_{Cdk1} \nabla^2 a - \frac{\partial V(a)}{\partial a} + \xi_{eff}(x, t)$$

The waves are noise triggered: noise comparable to barrier

1. Noise controls the time of the jump and as a consequence the speed of the wave as potential depends on time
2. The spreading is not a perfect wave



# An intuitive explanation for the wave slowdown



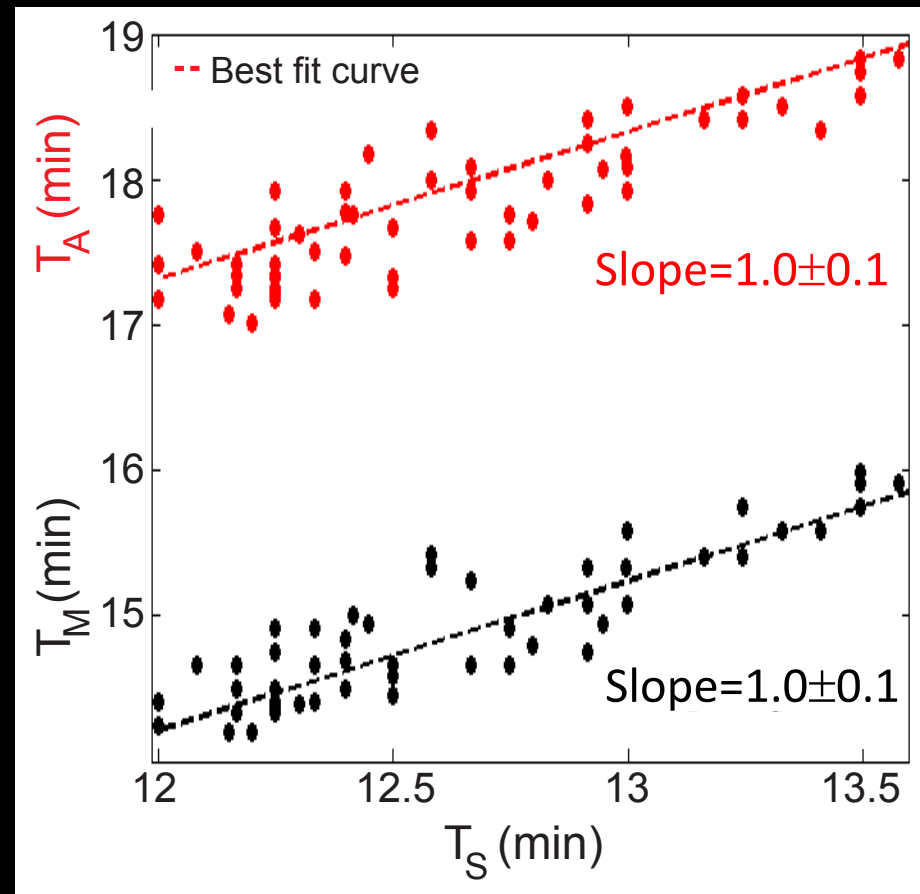


## **Important prediction:**

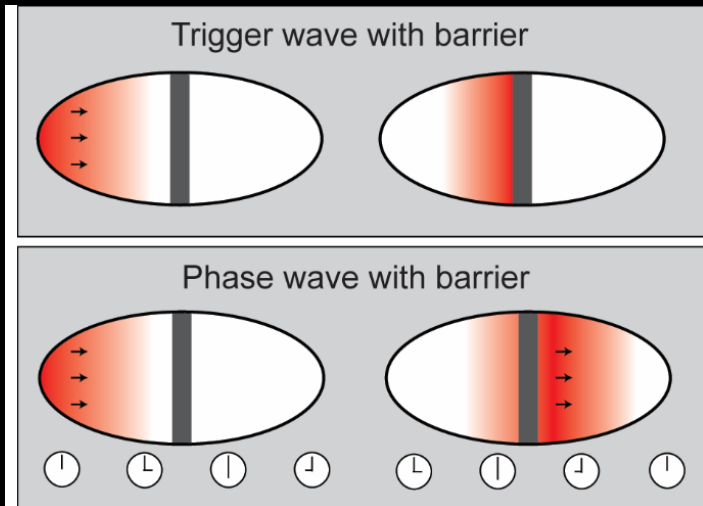
**Once the wave has passed and M-phase begins, diffusion is slow and cannot alter the delays set by the S-phase Cdk1 trigger wave: the system is effectively on a clock**

# S-phase waves predict the mitotic and anaphase waves

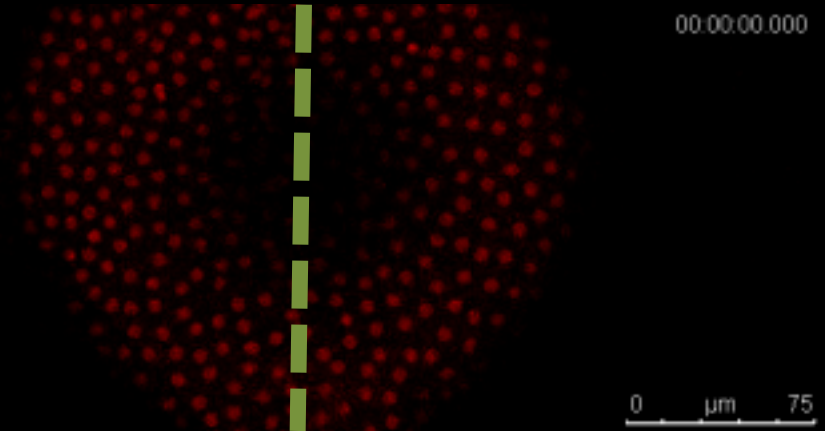
Phase wave prediction:  $T_M = T_S + \Delta_M$ ;  $T_A = T_S + \Delta_A$



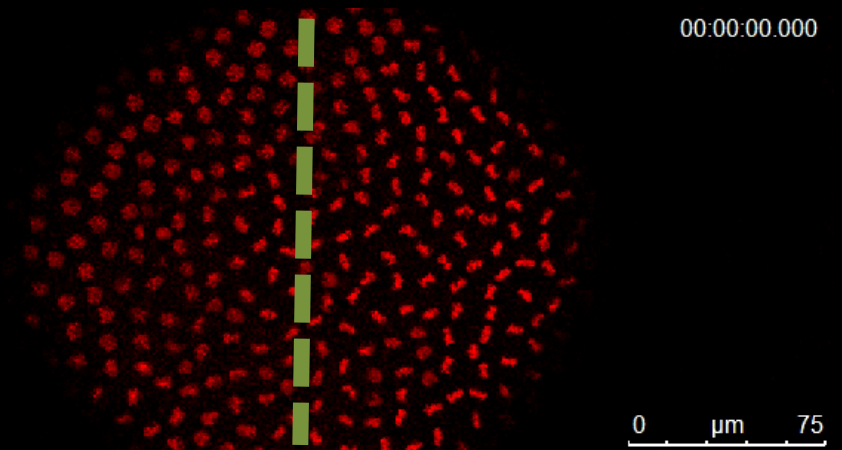
# Causality: timed ligation experiments



S-phase ligation

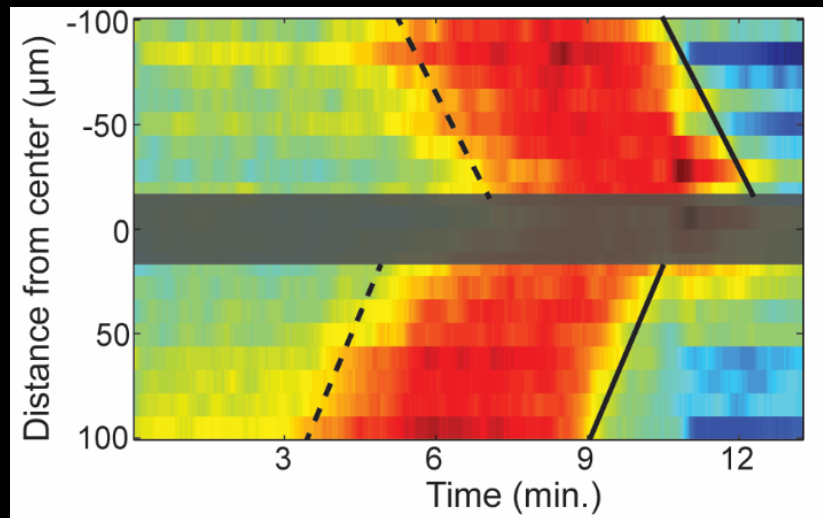


M-phase ligation

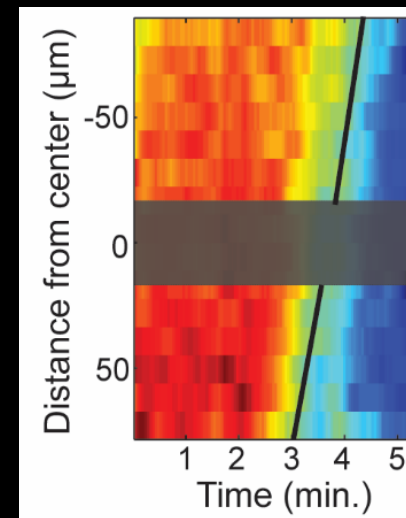


# The wave of anaphase is a phase wave, which reflects the delays set by the trigger wave during S-phase

S-phase ligation



M-phase ligation

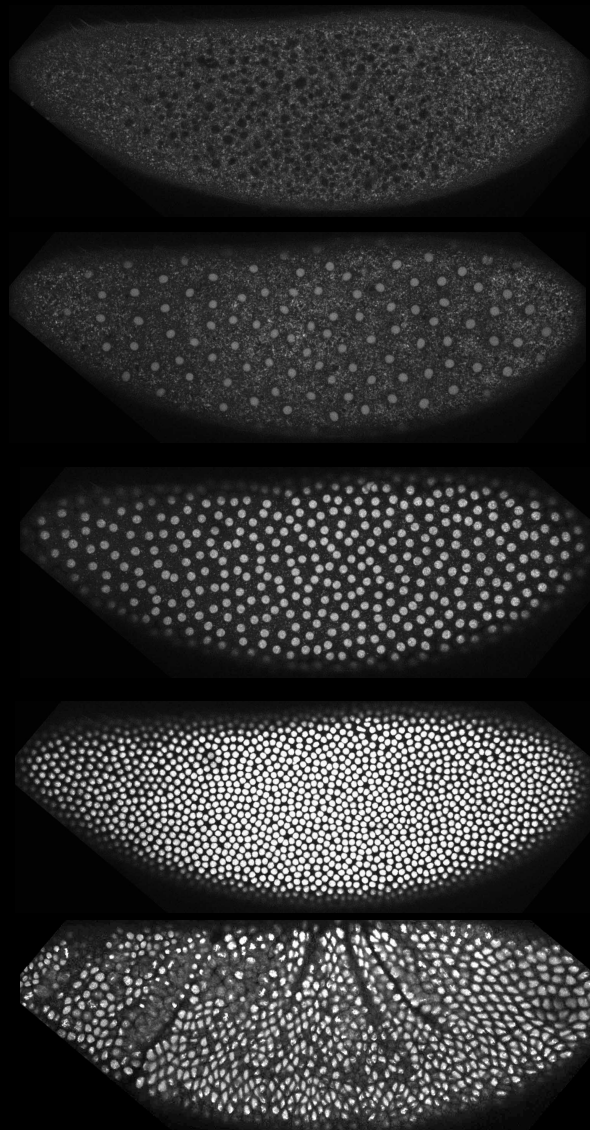


# Summary

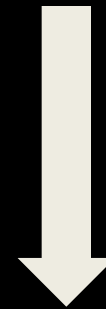
- The speed of Cdk1 waves depends on the S-phase DNA replication checkpoint
- Cdk1 positive feedback serves primarily to ensure the rapid onset of mitosis and is dispensable for wave propagation in the *Drosophila* early embryo
- The Cdk1 wave during S-phase is a trigger wave and the mitotic wave is a phase wave that follows the delays set by the trigger wave

## Part II

# Cell cycle control during gastrulation



**Synchronous  
Nuclear  
Replication**

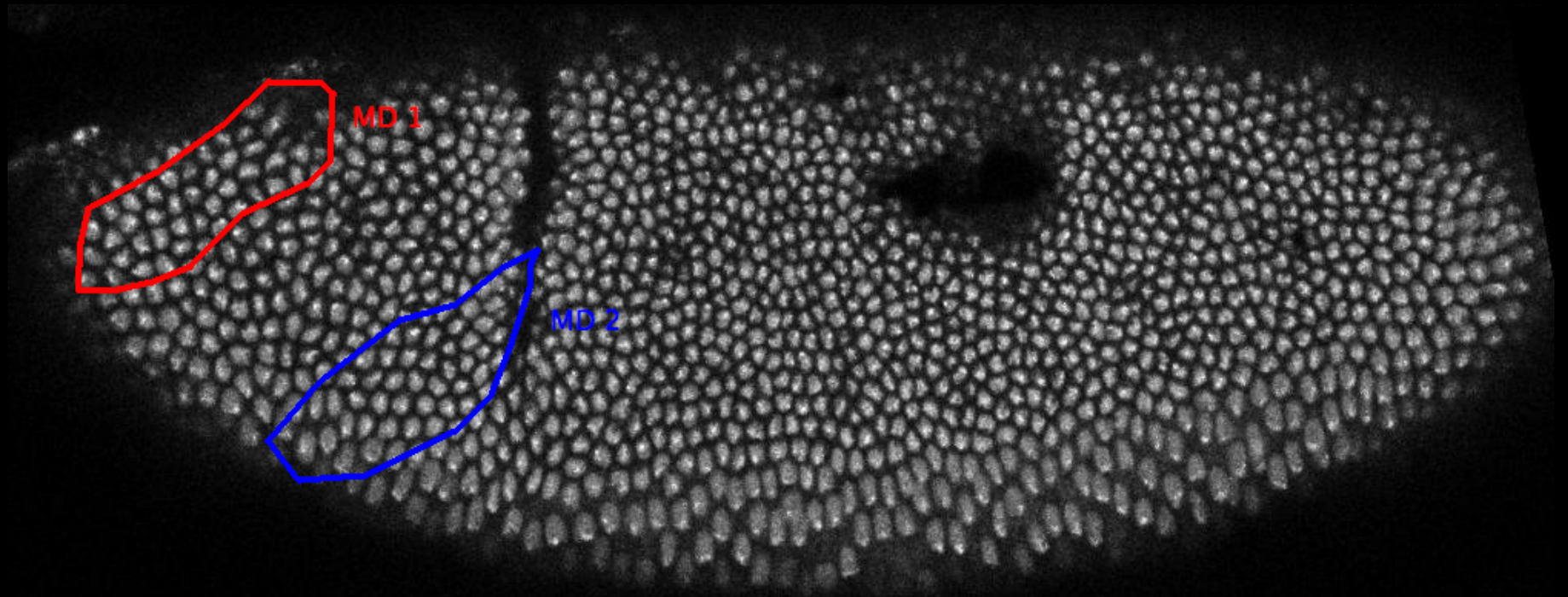


**Cell Cycle Pause**  
maternal gradients  
establish cell fate



**Gastrulation**  
cells in different  
regions behave  
differently

# *Drosophila* Embryo Shows a Distinct Temporal Patterning

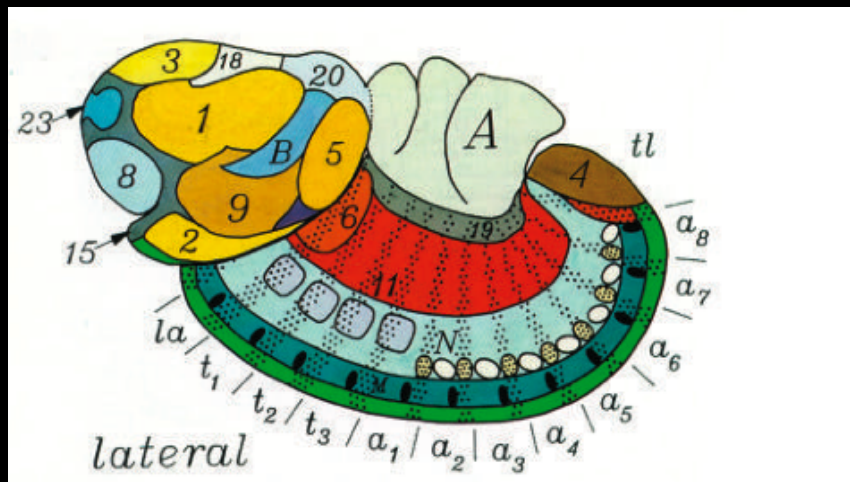




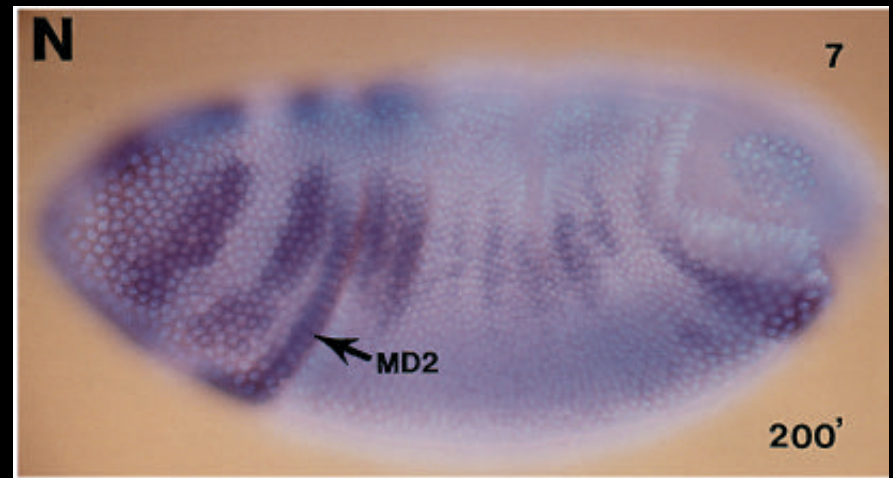
# Pattern of cell division correlates with the activation of transcription of *string*

Following depletion of String and Twine and the MBT, maternal gradients establish cell fates. Cell cycle re-entry correlates with transcription of *string* in specific mitotic domains.

## Mitotic Domains



## *string* mRNA expression





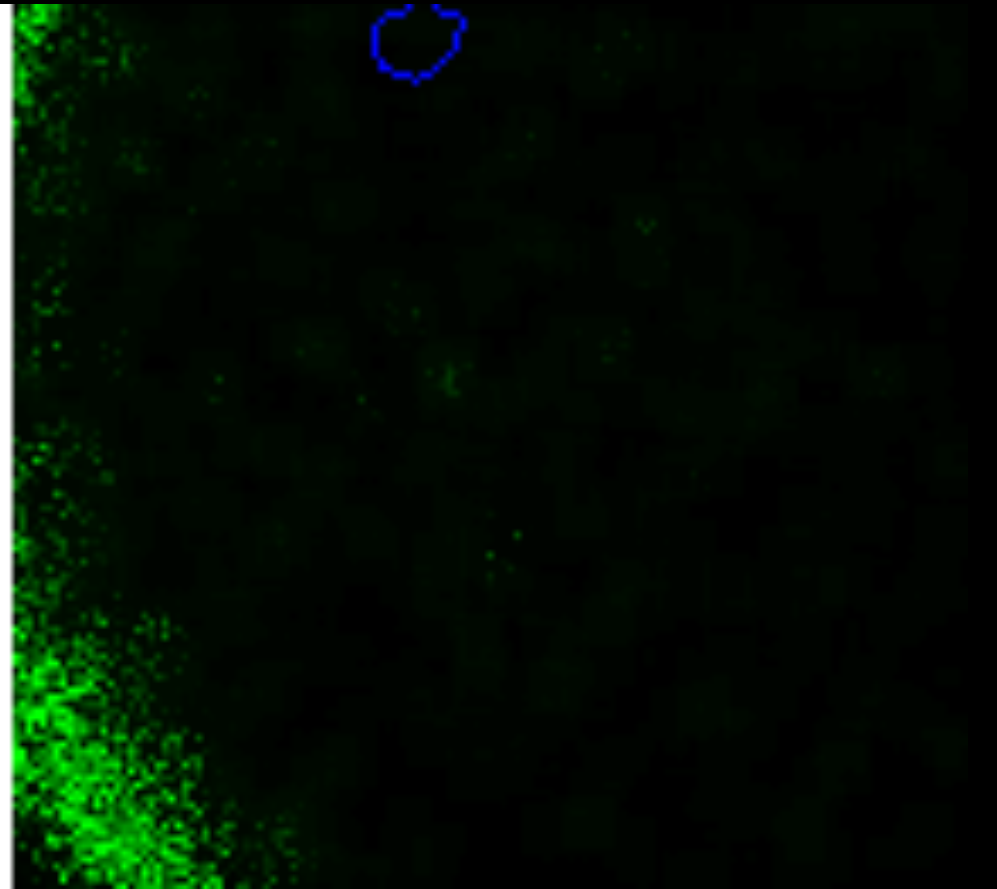
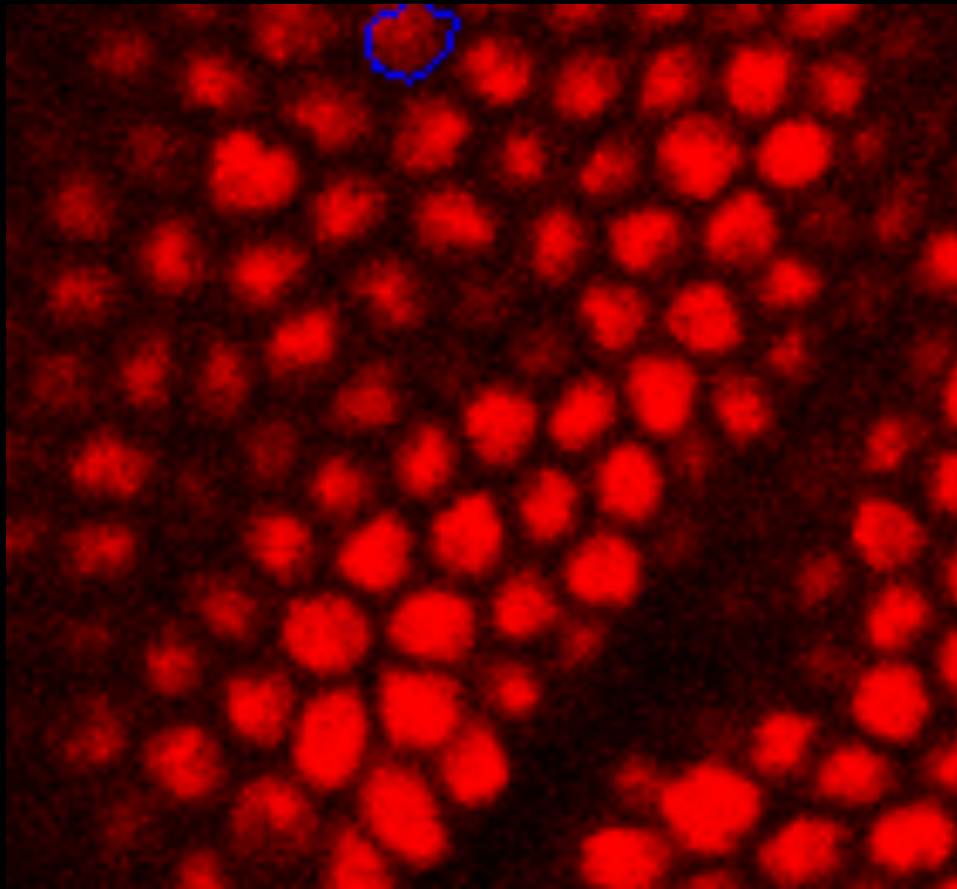
Is the time of mitosis controlled solely by *string* transcription?

Or is there some other process that controls entry into mitosis once adequate levels of String are expressed?

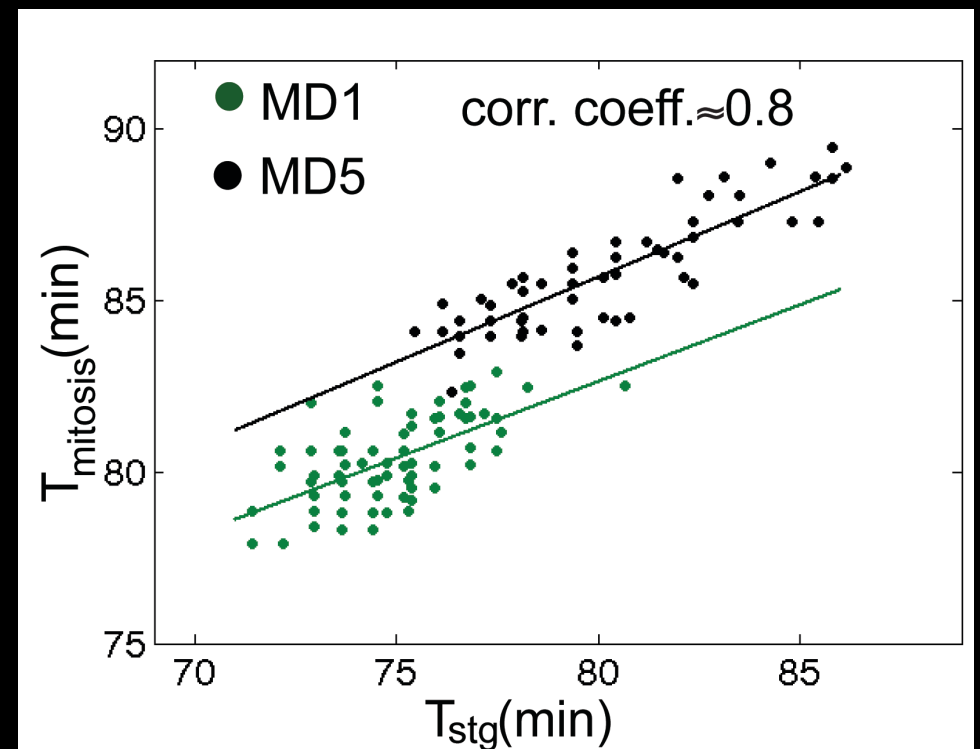
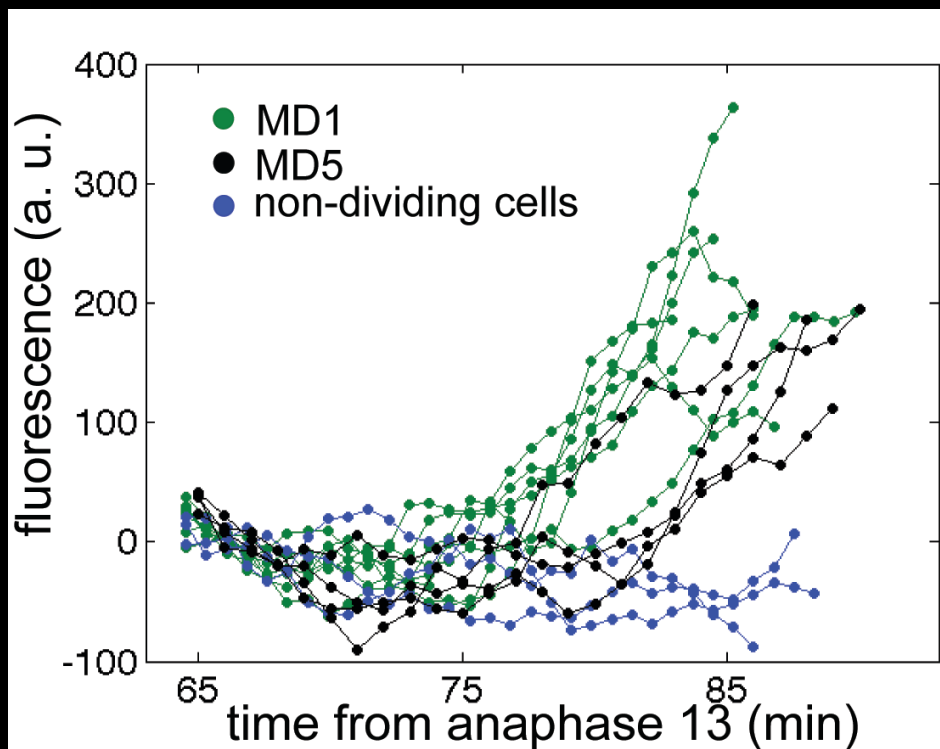
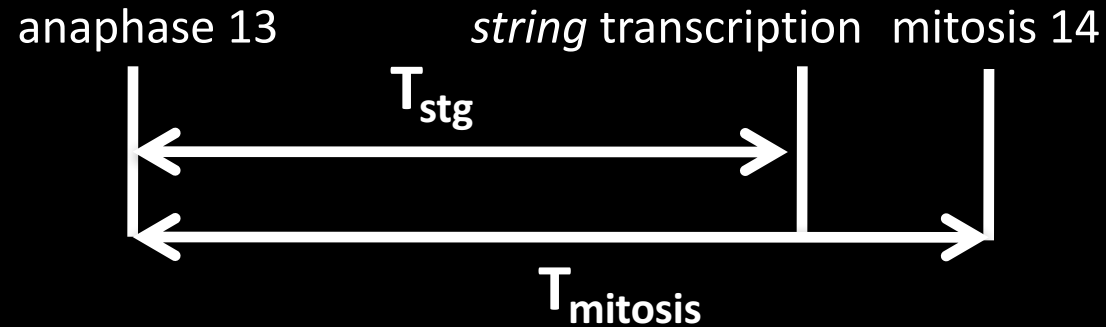
# Dynamical analysis of *string* transcription

HIS-RFP

*string* enhancer-GFPNLS



# Activation of *string* transcription controls the timing of mitosis

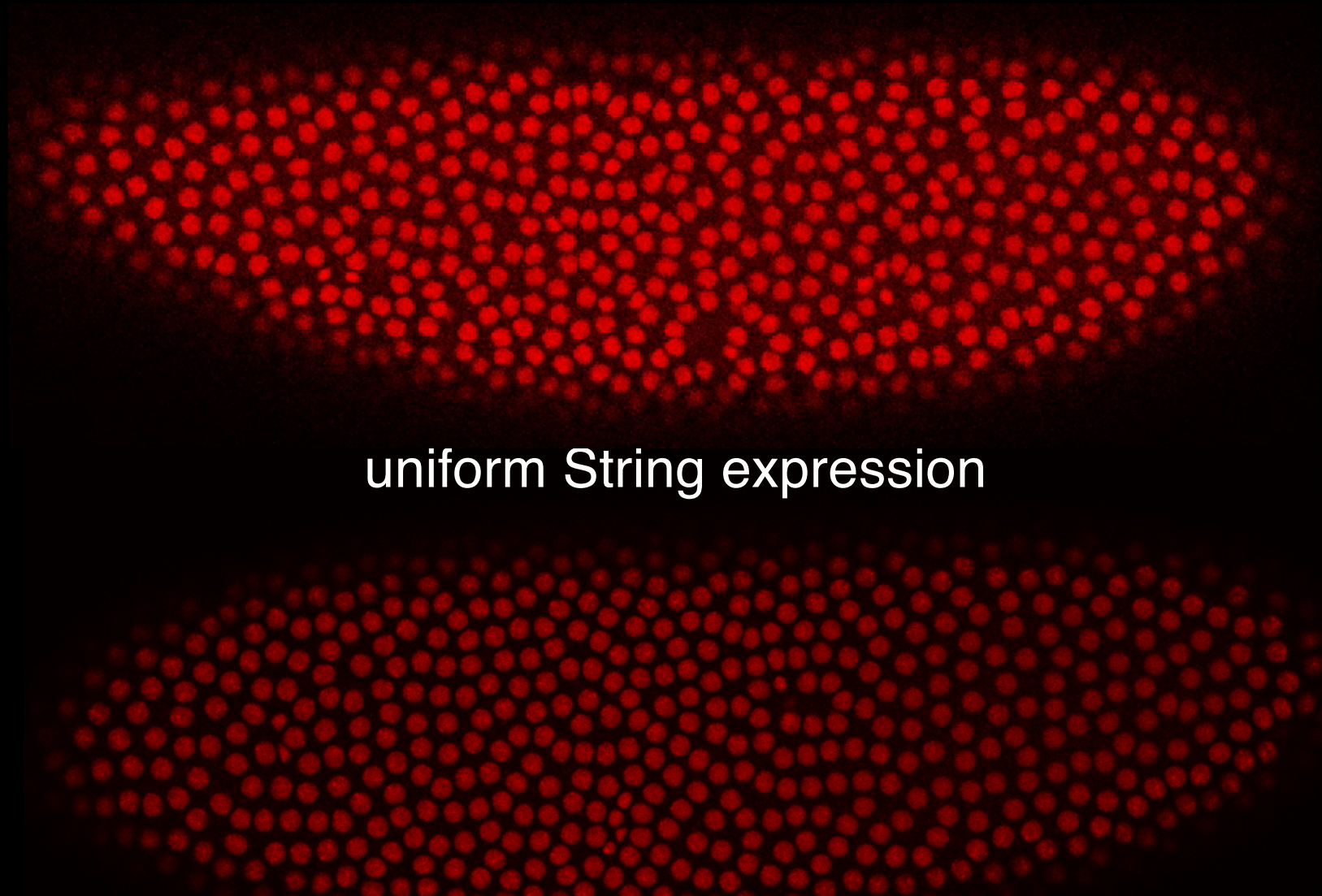


# How is the expression of *string* translated into reliable control of mitosis?

Is String the only input in the decision of entering mitosis? Eliminate transcriptional variability (in space and time) to study how the levels of String control mitosis

# Uniform expression of String induces a synchronous mitosis

wild type



uniform String expression

# Identifying Dosage-Dependent Regulators of Mitosis

A genetic screen for trans-activating factors

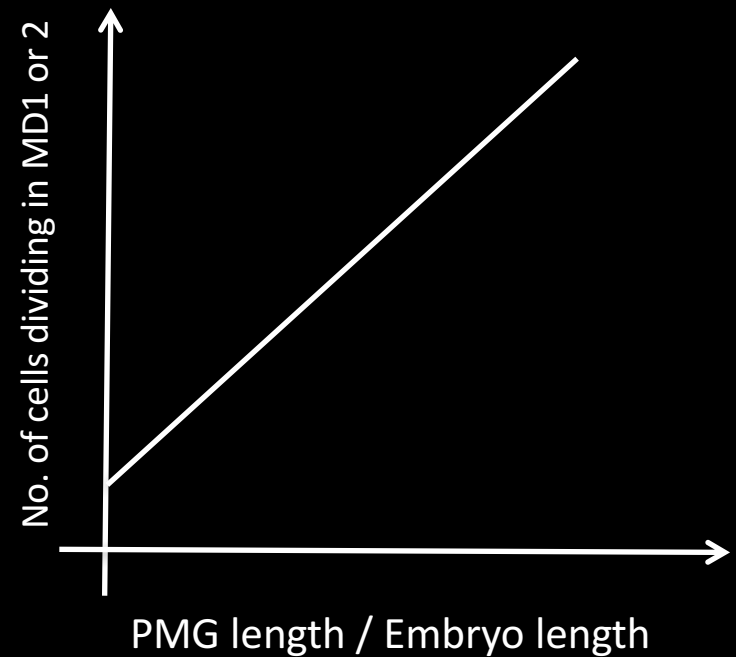
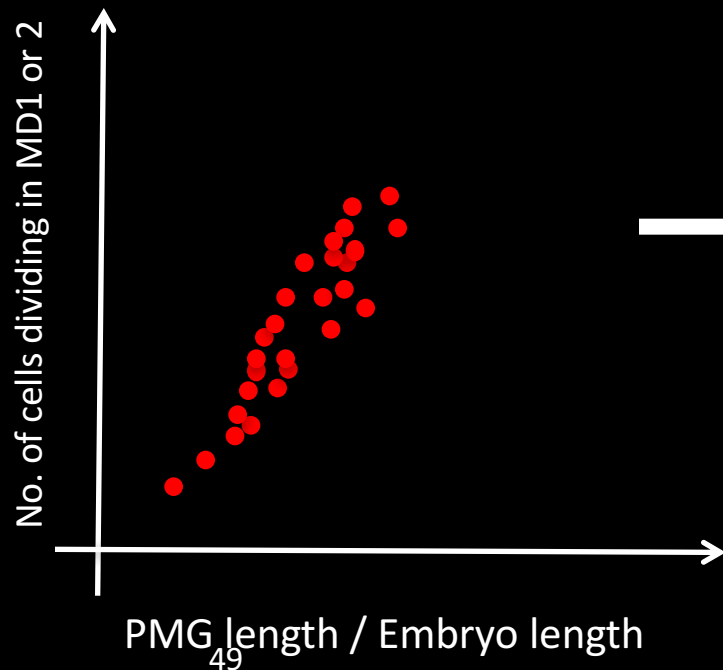
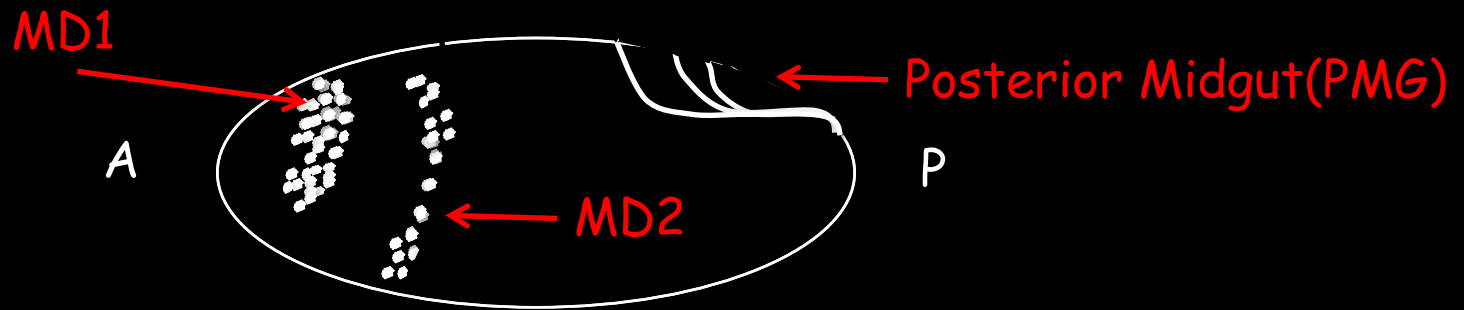


Amir Momen Roknabadi  
Princeton

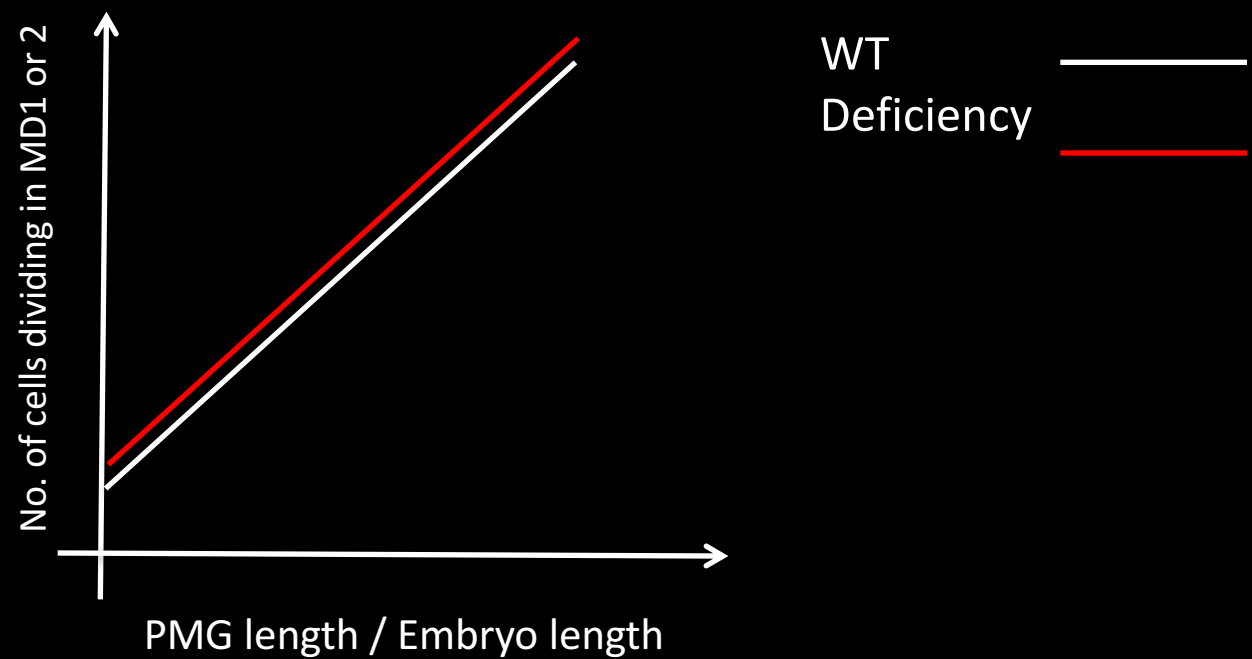


Eric Wieschaus

# Identifying Dosage-Dependent Regulators of Mitosis

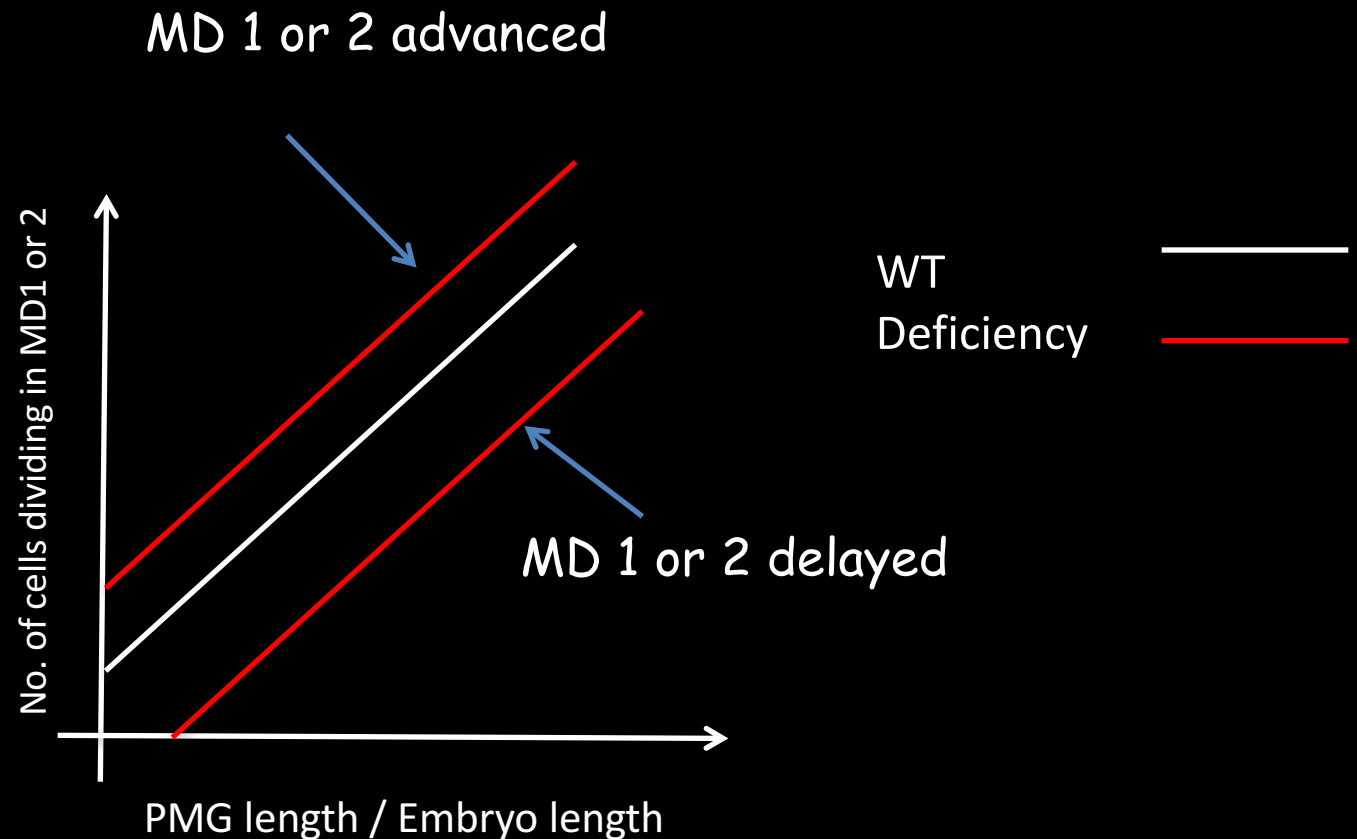


# Deficiency Does Not Affect MD1 or MD2 Timing





# Deficiency Affects MD1 or MD2 Timing: Change in the Intercept

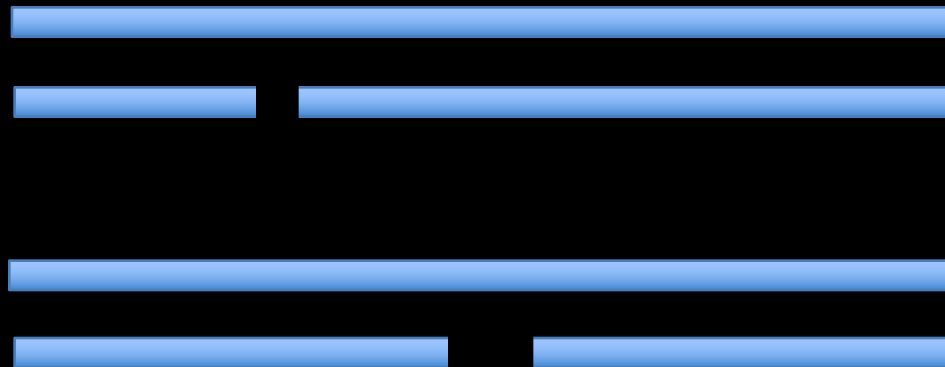


# Temporal control by a two-step process

A genetic screen for trans-activating factors

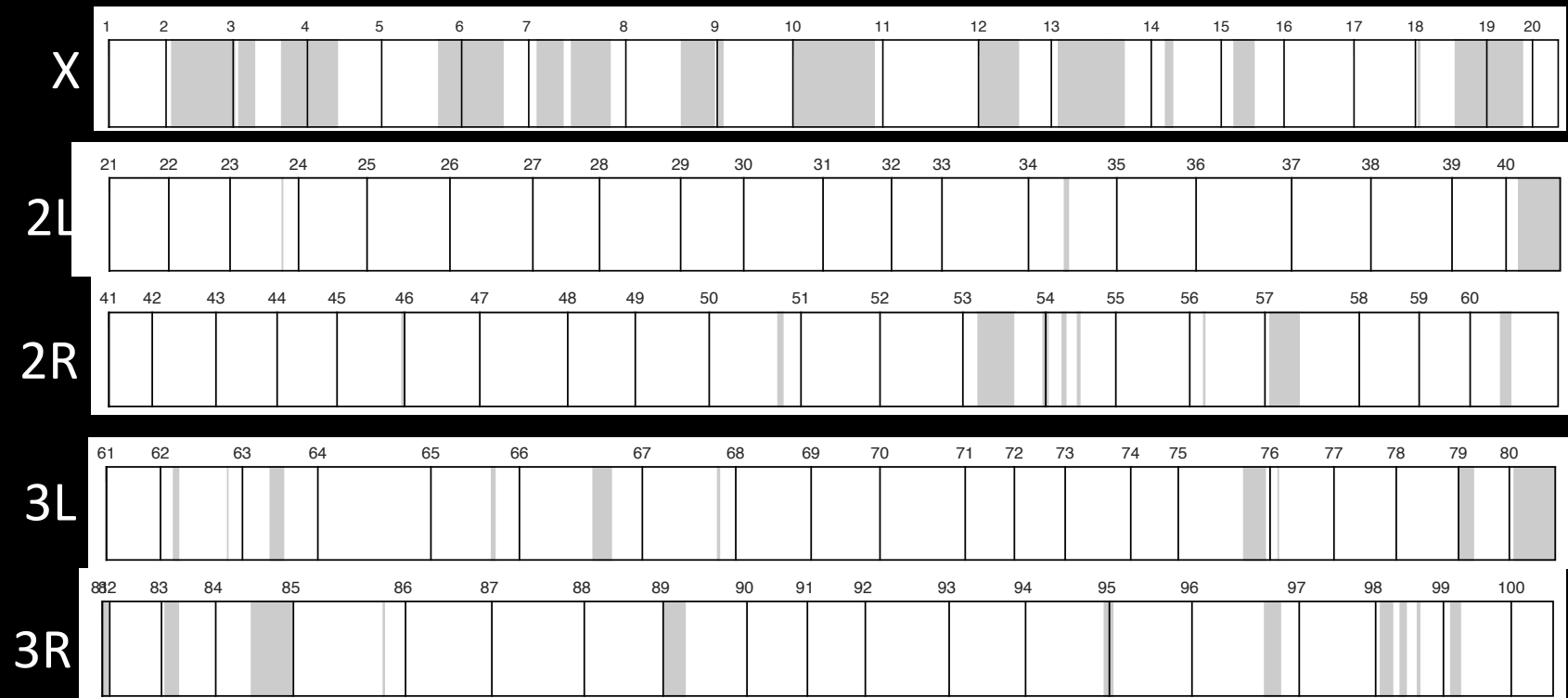
**Chromosomal deficiencies:**

**Flies can survive carrying large genomic deletion on one chromosome**



# Identifying Dosage-Dependent Regulators of Mitosis

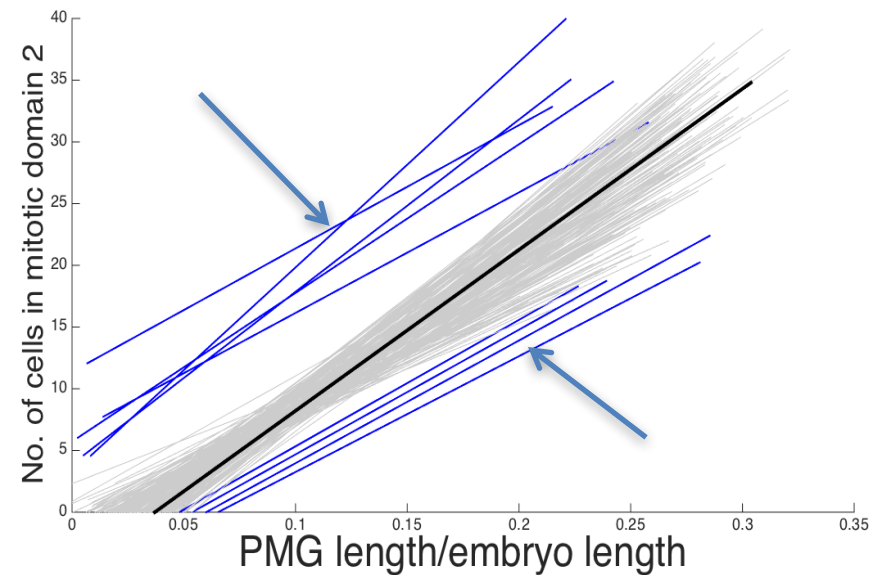
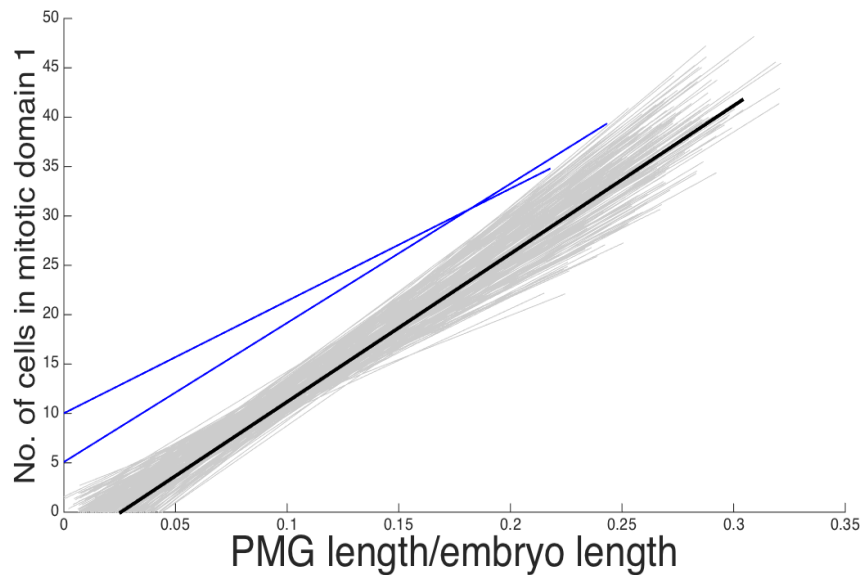
Genome-wide screen using heterozygote deficiencies, coverage of 85% of the genome

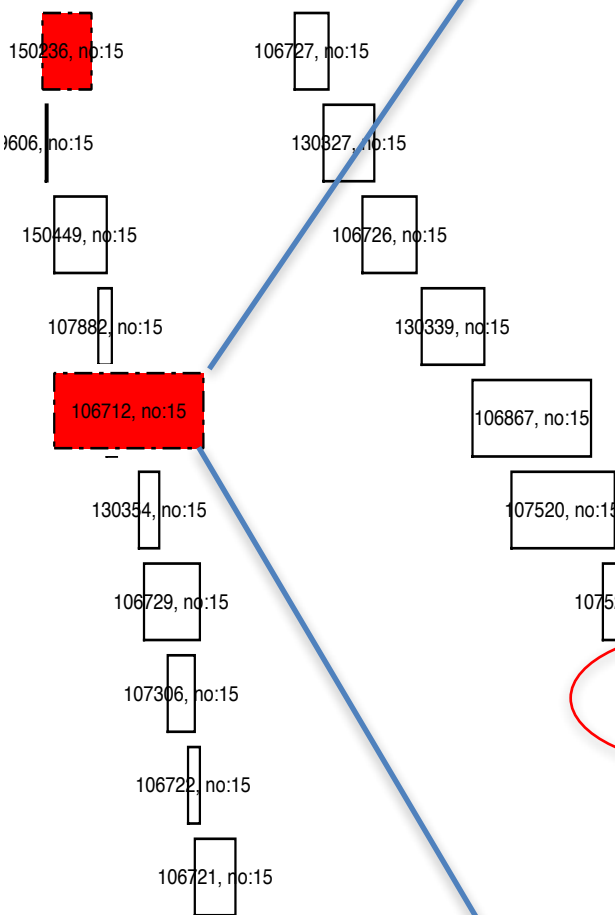
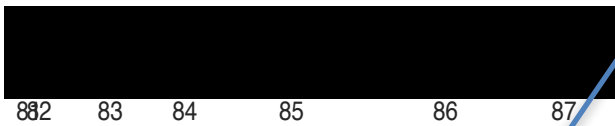


# 11 Regions Affect Either MD1 or MD2

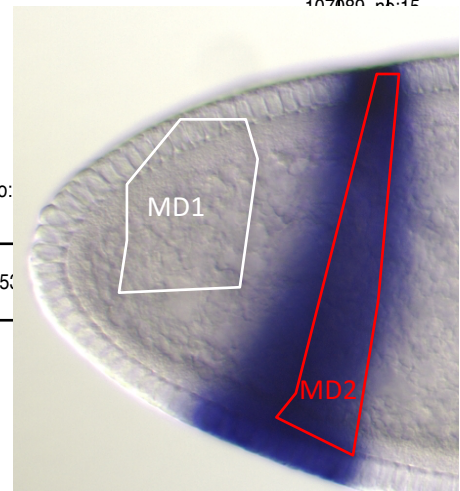
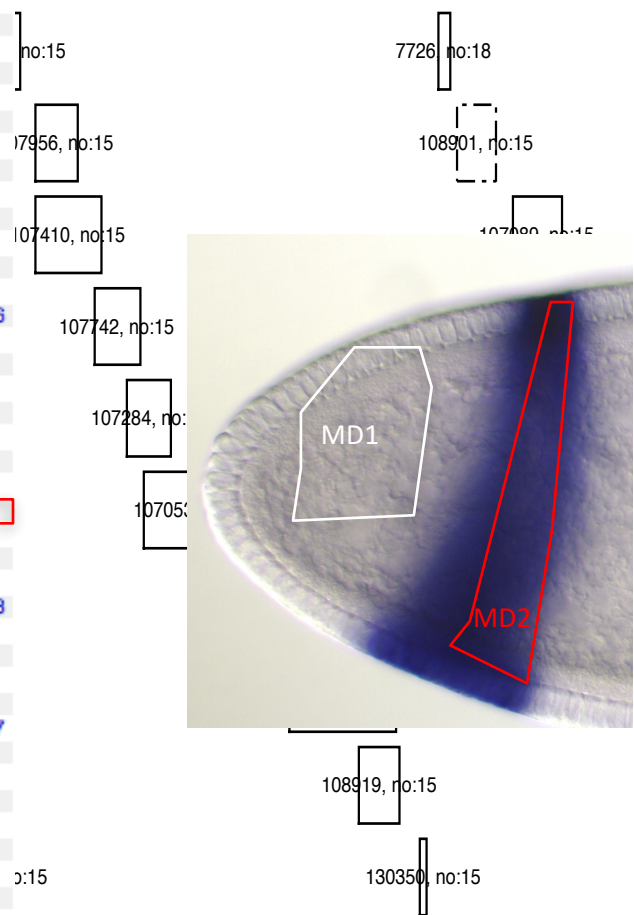
Two deficiencies advance MD1

Four deficiencies delay MD2  
Five deficiencies advance MD2





87F11-87F11	3R:13700565..13703164	ninaB	4 stocks
87F11-87F11	3R:13703084..13705402	Adgf-D	5 stocks
87F12-87F12	3R:13706757..13709129	Adgf-C	2 stocks
87F12-87F12	3R:13709159..13709939	CG31469	5 stocks
87F12-87F12	3R:13710017..13711835	primo-2	4 stocks
87F12-87F12	3R:13710017..13711835	primo-1	5 stocks
87F12-87F12	3R:13712469..13714204	CR34044	
87F13-87F13	3R:13715207..13764667	CG34383	13 stocks
87F12-87F12	3R:13717454..13718176	Ubc87F	5 stocks
87F13-87F13	3R:13765756..13767103	CG9312	3 stocks
87F13-87F13	3R:13767549..13770732	Chf5	7 stocks
87F13-87F13	3R:13771495..13779317	CG9297	3 stocks
87F13-87F13	3R:13779624..13780795	CG42375	3 stocks
87F13-87F13	3R:13779624..13780795	CG9288	4 stocks
87F13-87F13	3R:13780964..13782082	CG9286	4 stocks
87F13-87F13	3R:13782288..13785676	Dip-B	5 stocks
87F13-87F13	3R:13785806..13786583	CR45590	
87F14-87F14	3R:13797953..13798025	tRNA:CR31588	
87F14-87F14	3R:13798198..13798270	tRNA:CR31331	
87F14-87F14	3R:13813109..13814648	tal-2A	
87F14-87F14	3R:13813109..13814648	tal-3A	
87F14-87F14	3R:13813109..13814648	tal-1A	
87F14-87F14	3R:13813109..13814648	tal-AA	
87F14-87F14	3R:13815279..13815778	CR43641	
87F14-87F14	3R:13816231..13817127	CR43642	FBst0059486
87F14-87F14	3R:13827151..13828008	Mst87F	3 stocks
87F14-87F14	3R:13827968..13828427	CR43300	
87F15-87F15	3R:13833497..13837991	Nsf2	7 stocks
87F15-87F15	3R:13838240..13839934	CG31495	2 stocks
88A1-88A1	3R:13854599..13855454	CG14362	2 stocks
88A1-88A1	3R:13864597..13865095	CR46239	
88A1-88A1	3R:13867345..13874916	E5	5 stocks
88A2-88A2	3R:13901853..13904620	ems	9 stocks
88A2-88A2	3R:13942238..13943460	Art9	3 stocks
88A2-88A2	3R:13944654..13945812	Art6	4 stocks
88A3-88A3	3R:13948420..13949742	CG31517	2 stocks
88A3-88A3	3R:13949454..13951138	CG9926	FBst0472368
88A3-88A3	3R:13958789..13960474	lpp	3 stocks
88A3-88A3	3R:13960712..13964174	CG9925	8 stocks
88A3-88A3	3R:13964885..13967075	Orc2	12 stocks
88A3-88A5	3R:13967061..14032057	rdx	52 stocks
88A3-88A3	3R:13971733..13974675	CR43460	FBst0053207
88A4-88A4	3R:14029228..14031483	Cyp6d5	3 stocks
88A4-88A4	3R:14032371..14035041	CG3061	3 stocks
88A4-88A4	3R:14035018..14036398	CG9922	4 stocks
88A4-88A4	3R:14044525..14044606	tRNA:S2b:88A	
88A5-88A8	3R:14056945..14087978	foxo	29 stocks
88A8-88A8	3R:14089517..14092425	Npc2b	9 stocks
88A8-88A8	3R:14092504..14096835	CCHa1	5 stocks
88A8-88A8	3R:14097293..14098080	CG9920	6 stocks
88A8-88A8	3R:14099832..14103122	PK1-R	4 stocks
88A8-88A8	3R:14104488..14108321	CG12402	4 stocks
88A8-88A8	3R:14109286..14110726	CR44237	
88A8-88A8	3R:14109445..14110823	Or88a	3 stocks
88A9-88A9	3R:14111151..14112166	CG14357	6 stocks
88A9-88A9	3R:14112451..14118450	Kif19A	7 stocks
88A9-88A9	3R:14119262..14122420	Abi	7 stocks
88A9-88A9	3R:14122654..14124474	twf	10 stocks
88A9-88A9	3R:14124163..14125548	140up	6 stocks
88A9-88A9	3R:14125791..14130111	Rpl140	15 stocks
88A9-88A9	3R:14130680..14132072	CG14356	2 stocks
88A9-88A10	3R:14148508..14151940	CG14355	4 stocks



Momen-Roknabadi, Di Talia\* and Wieschaus\* to appear in Cell Reports

# Conclusions

- The same transcription factors that control the spatial pattern of mitosis encode the temporal pattern
- The precise timing of mitosis is encoded by a combination of transcriptional activators and repressors.

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Members:

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Patrick Ferree  
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**Victoria Deneke**

Collaborators:



**Anna Melbinger**

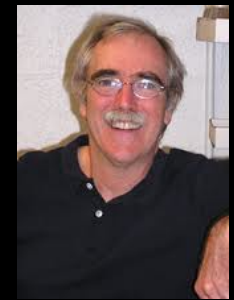


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