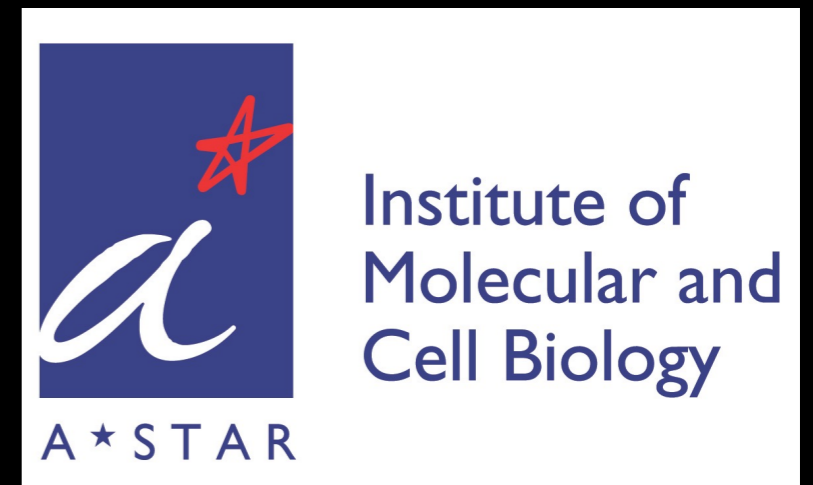


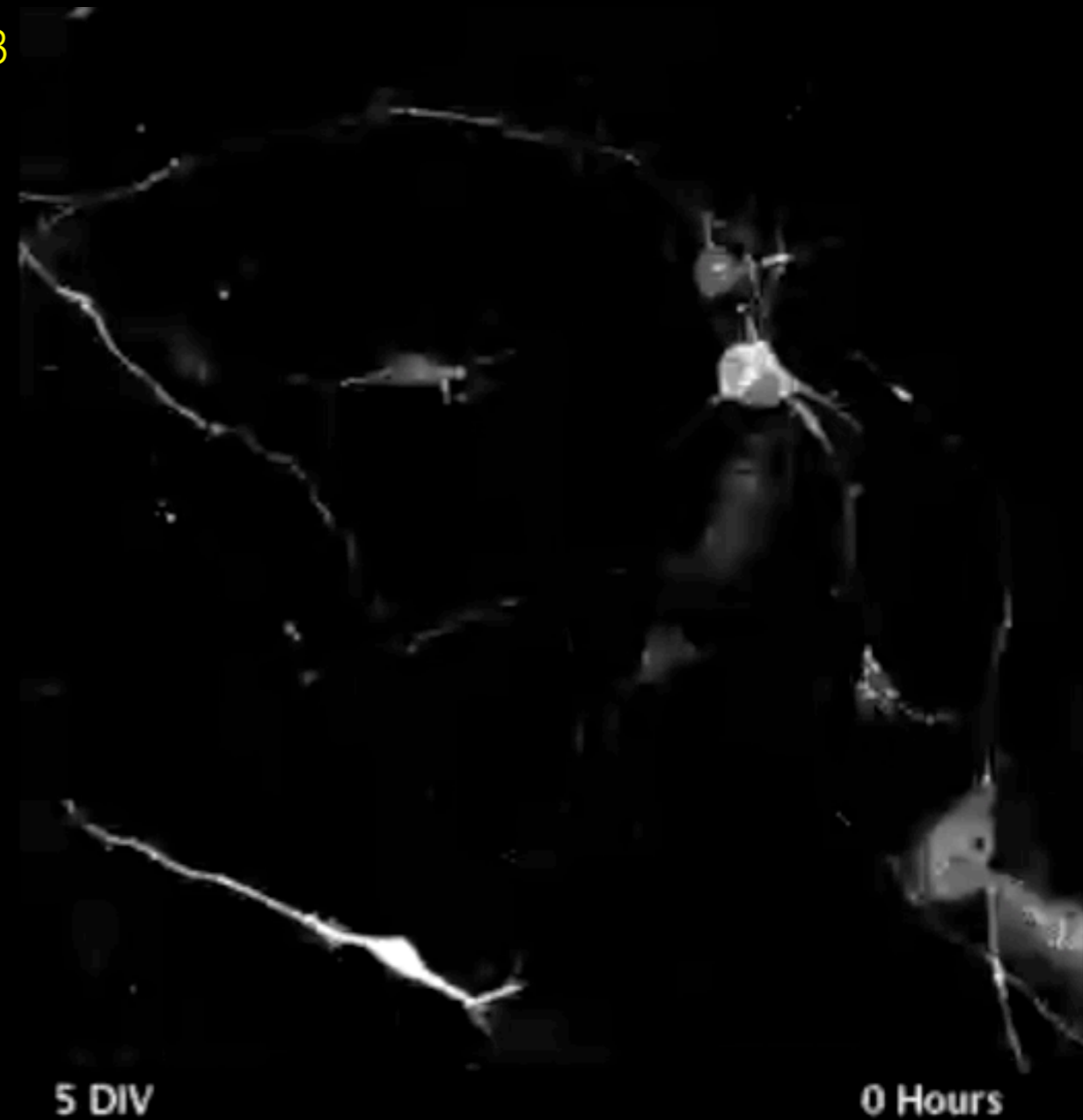
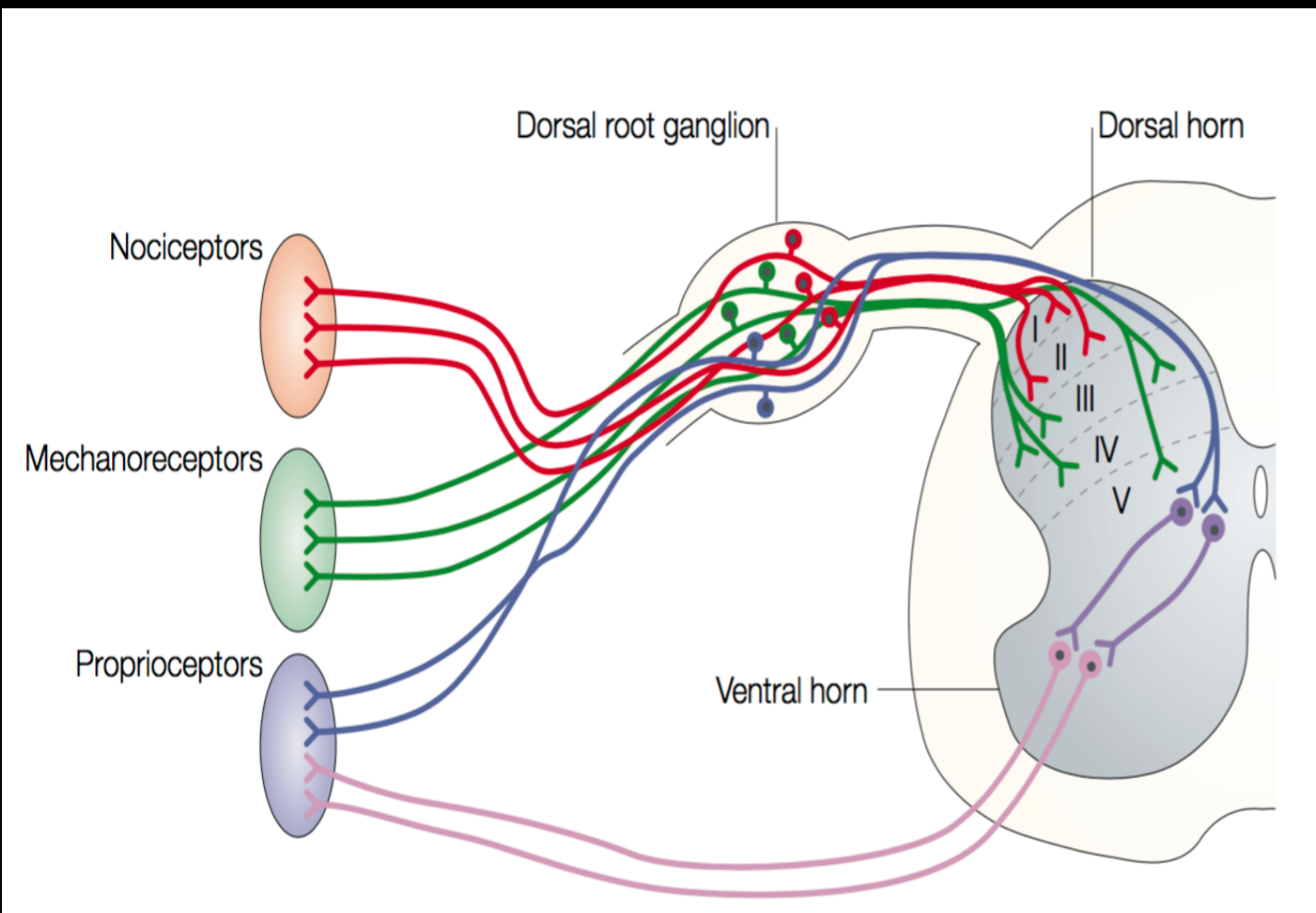
Differential mechanical coupling between cardioblasts guides heart morphogenesis

Timothy Saunders

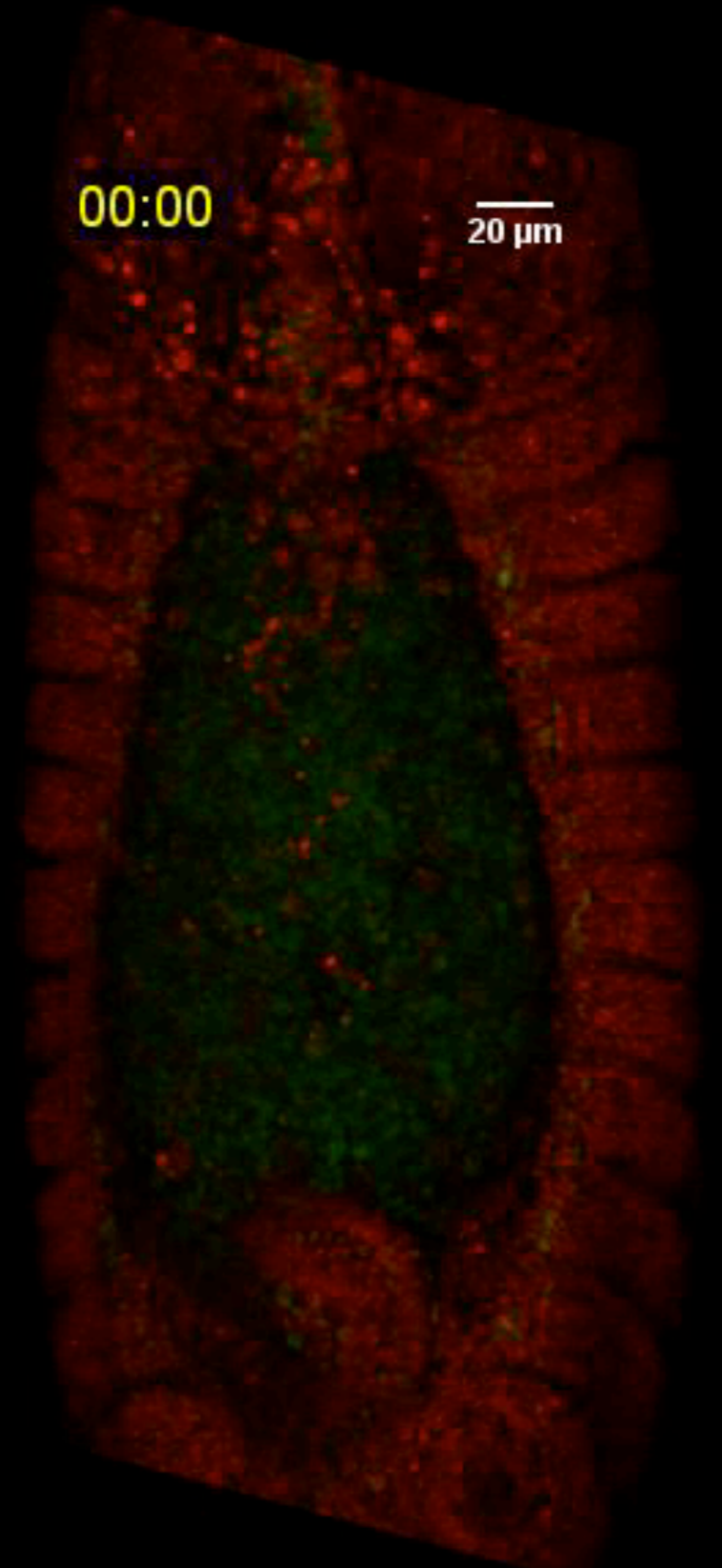
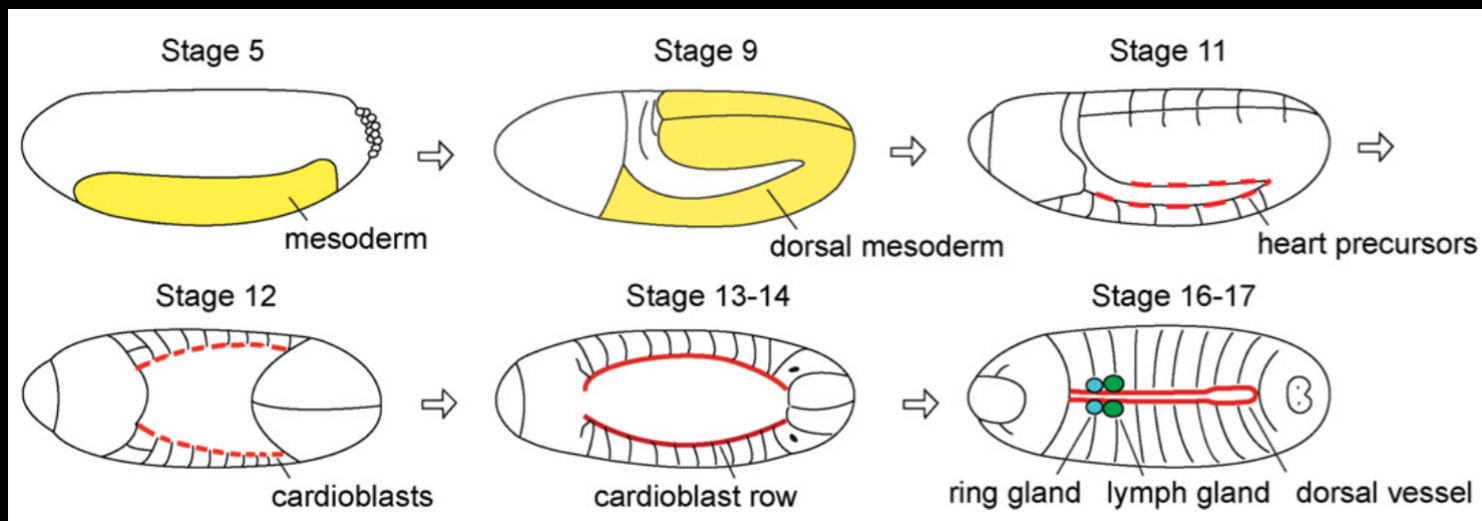
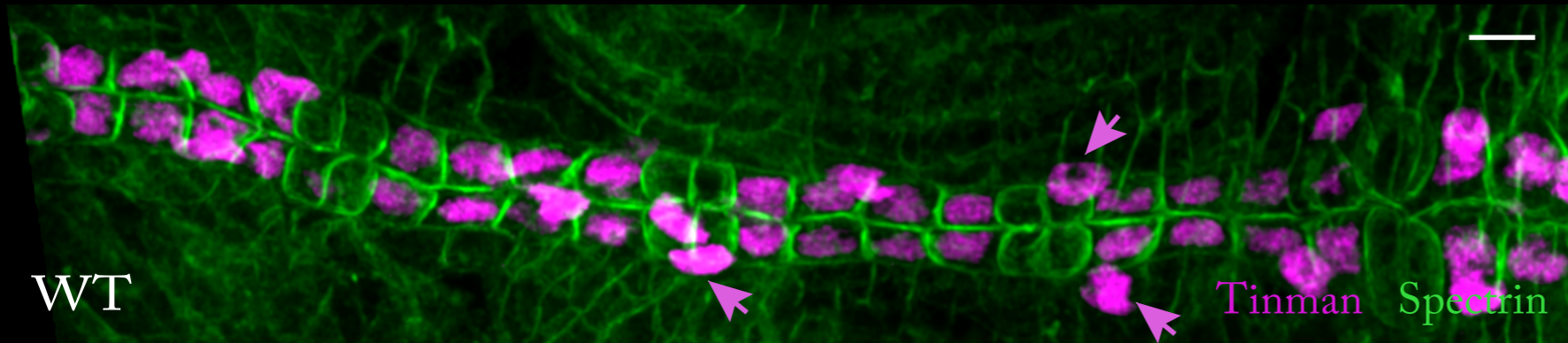
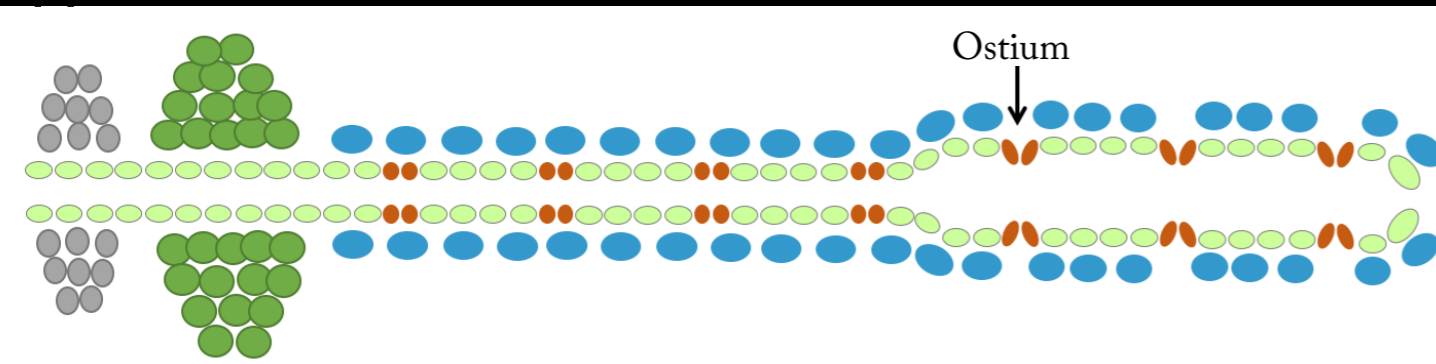


Cell matching is essential in development

Caspary & Anderson, Nature Reviews Neuroscience 2003

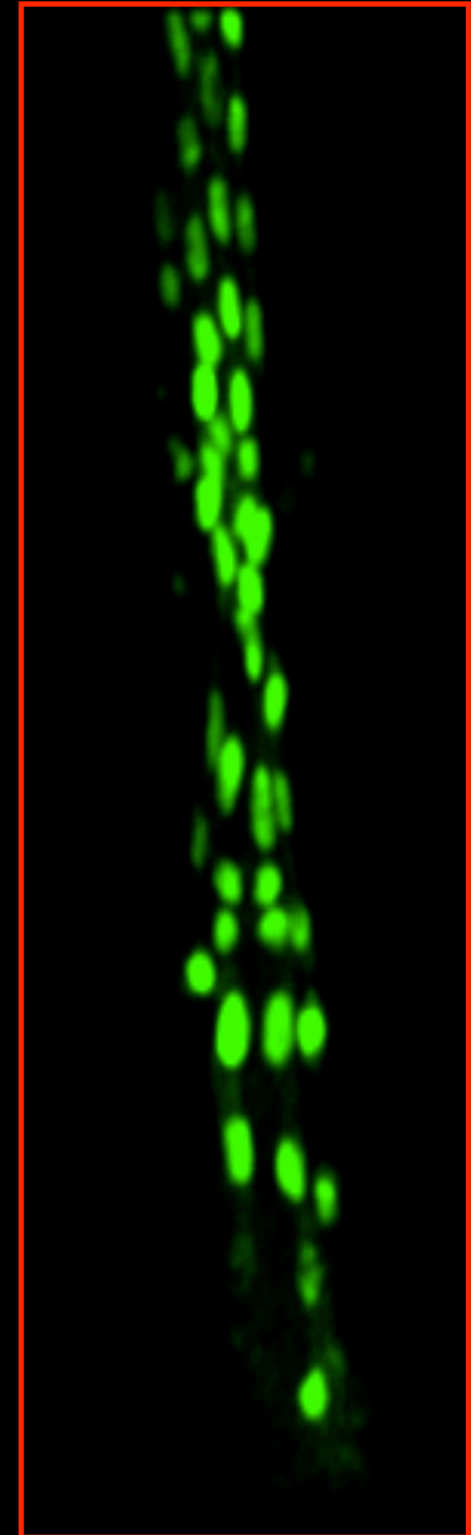
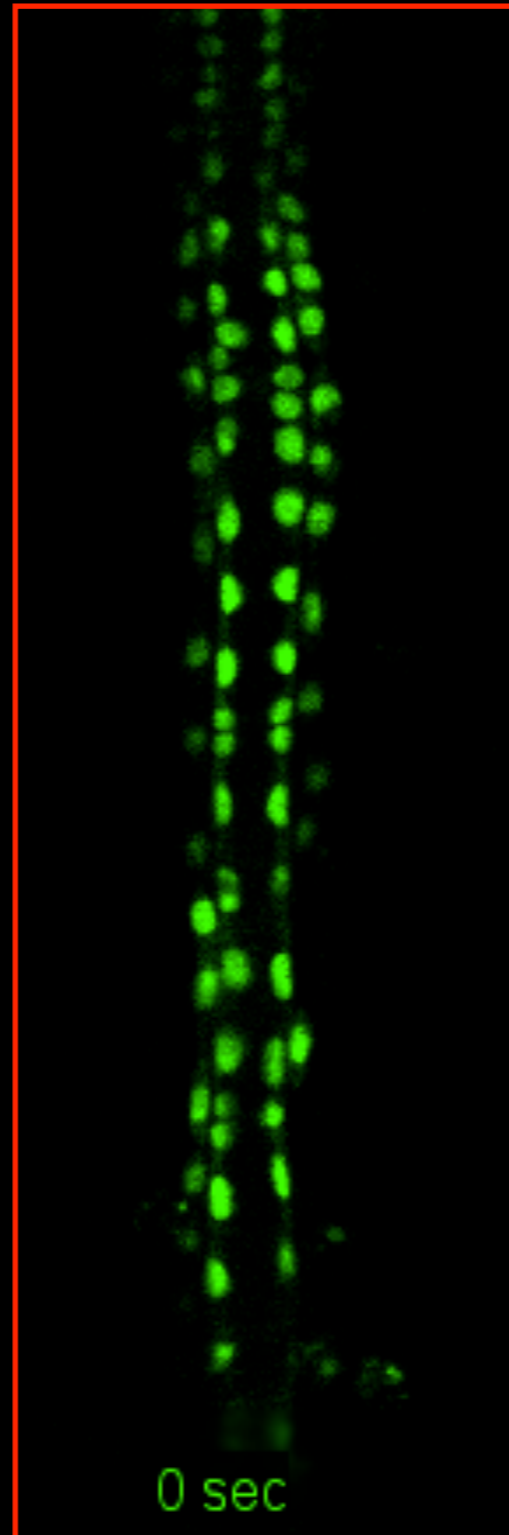
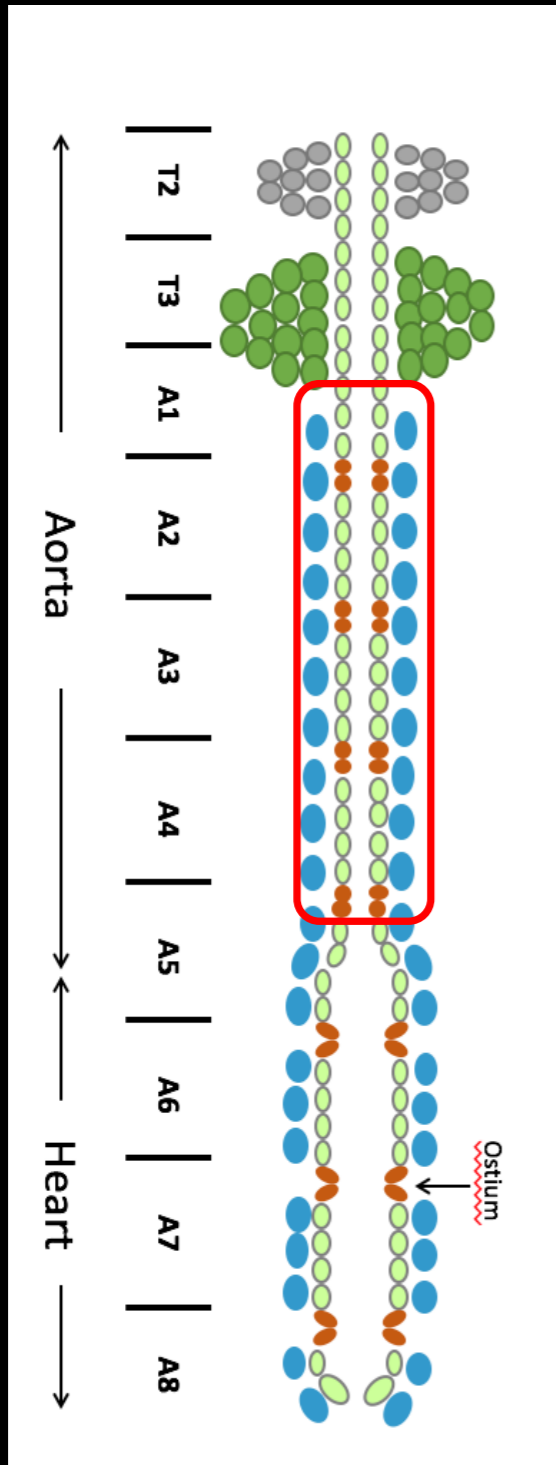


Drosophila cardiogenesis as simple matching system



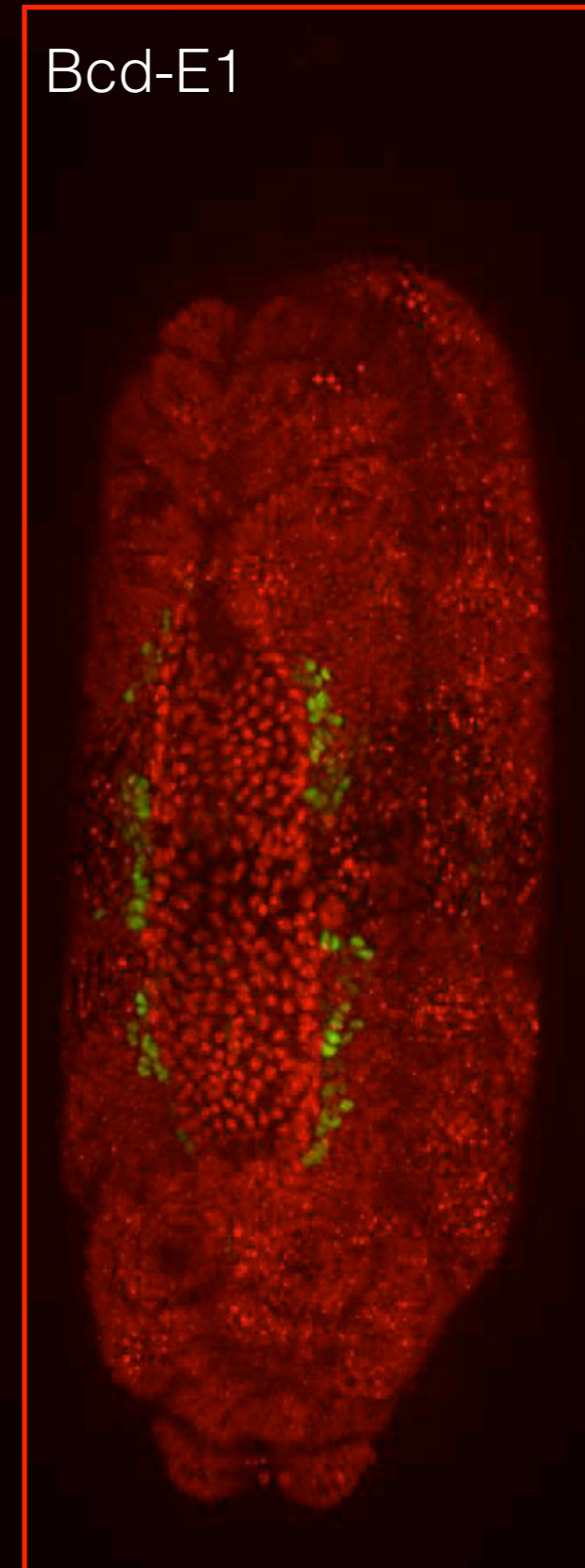
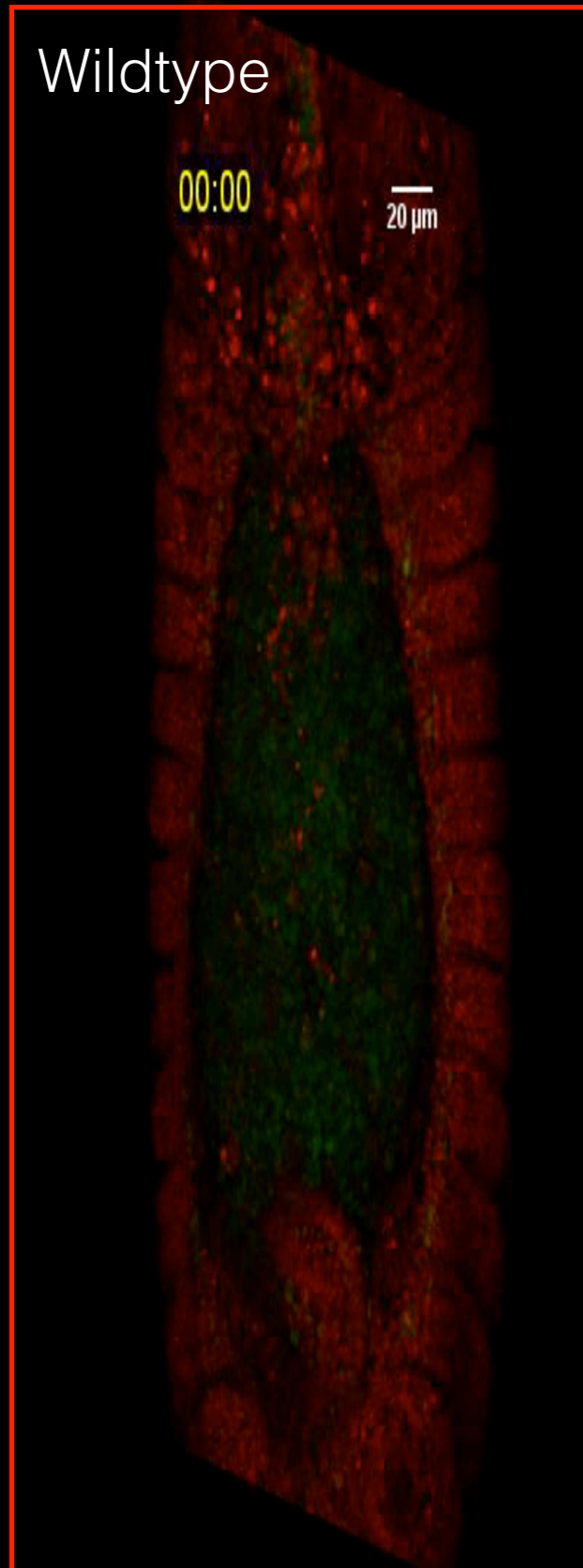
Hand>GFP, H2b-mCherry

Mismatched hearts have beating defects

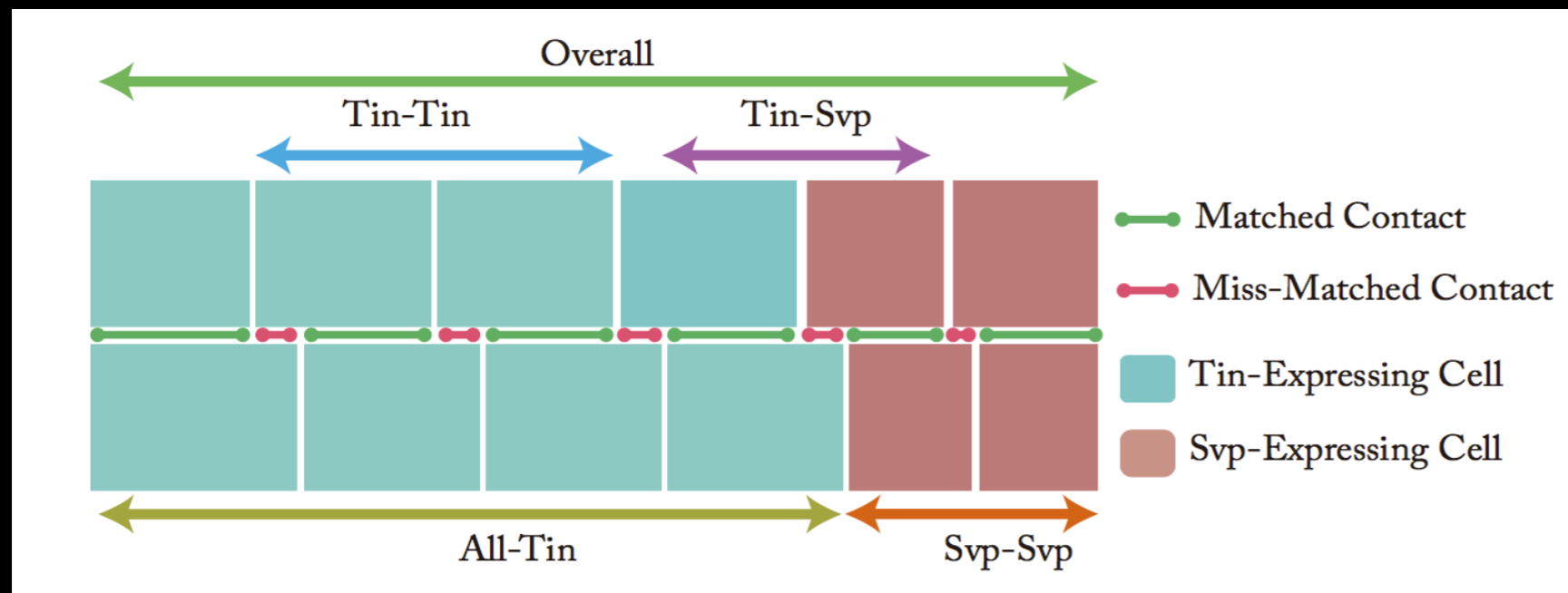
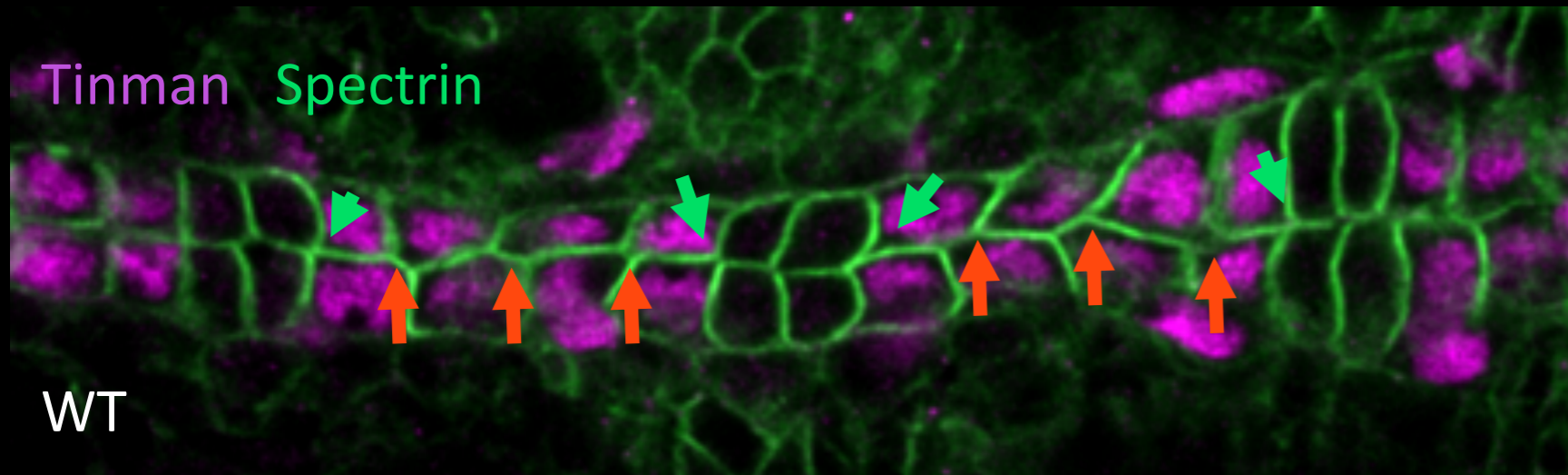


Hand-GFP

Cell matching is an active process

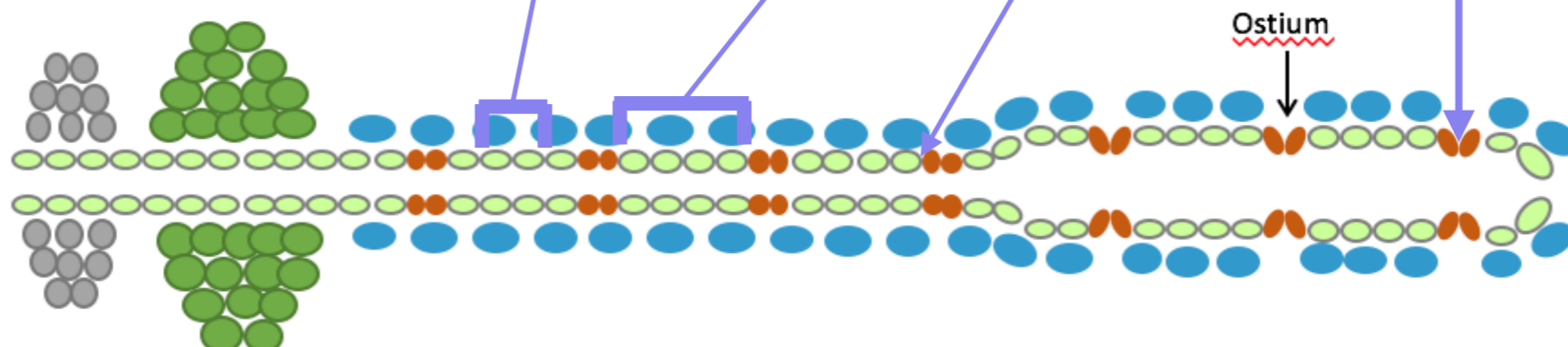
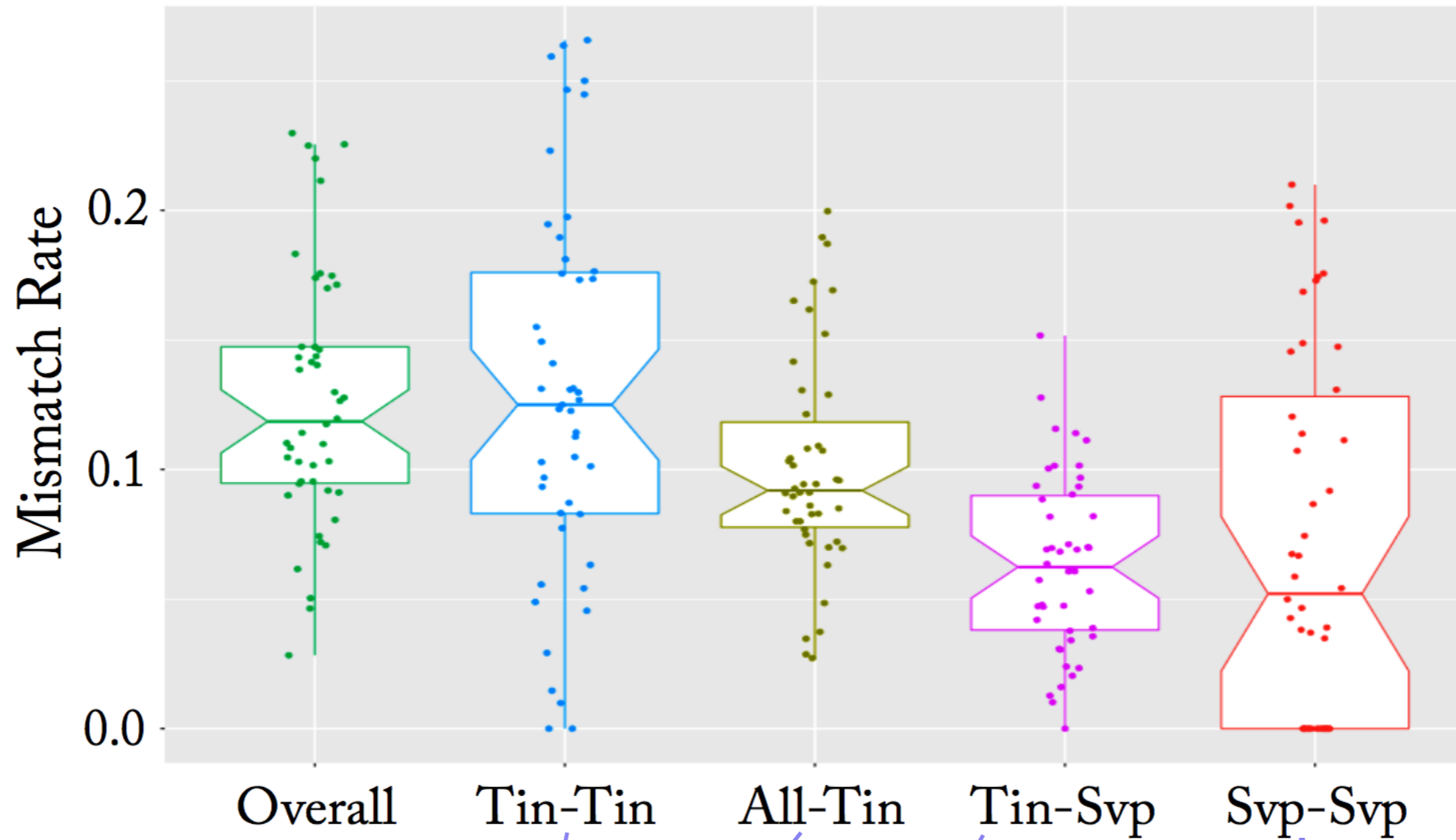


Quantifying cell matching

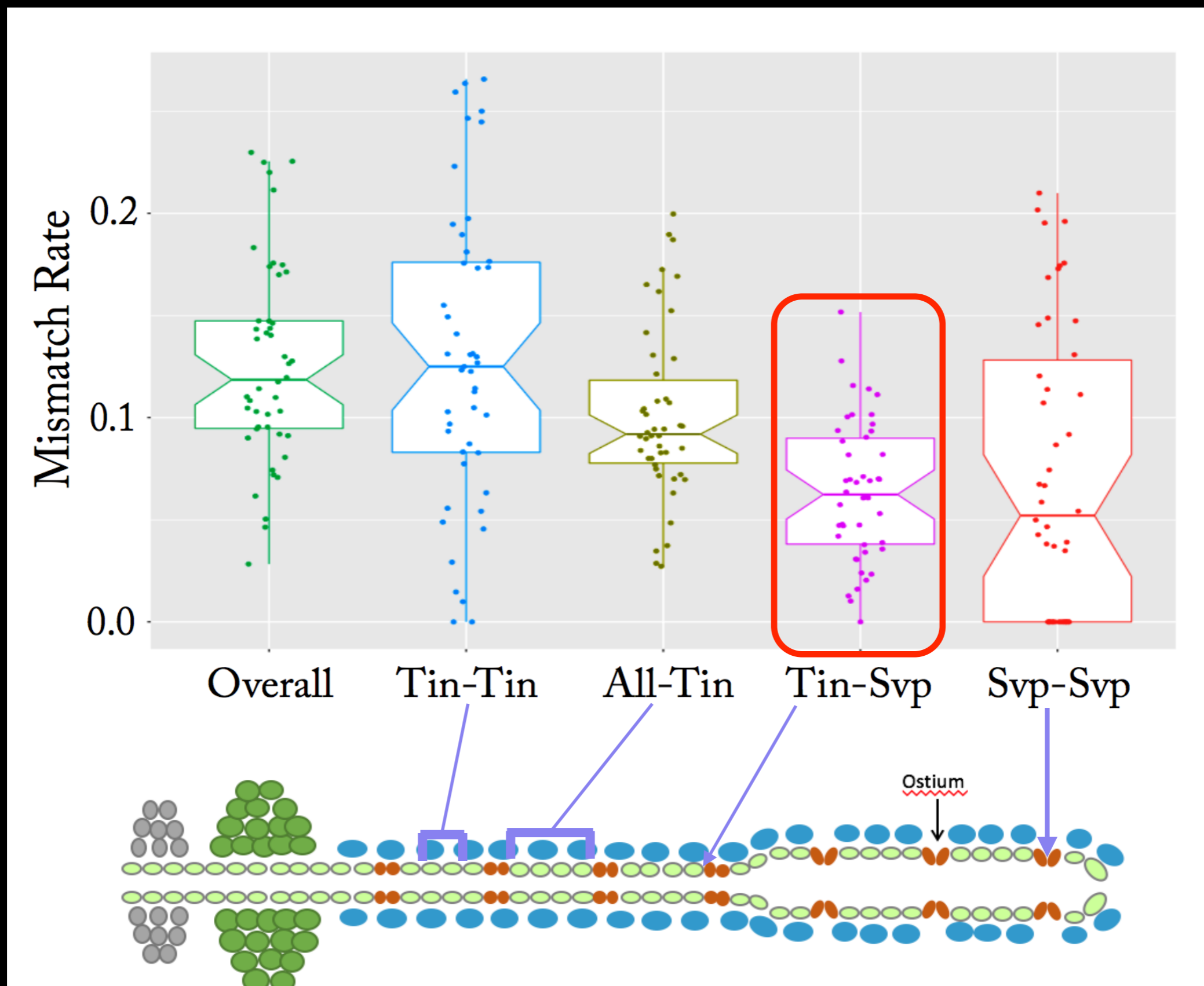


$$\frac{\text{Mis-Matched Contact Length}}{\text{Total Contact Length}} = \text{Mismatch}$$

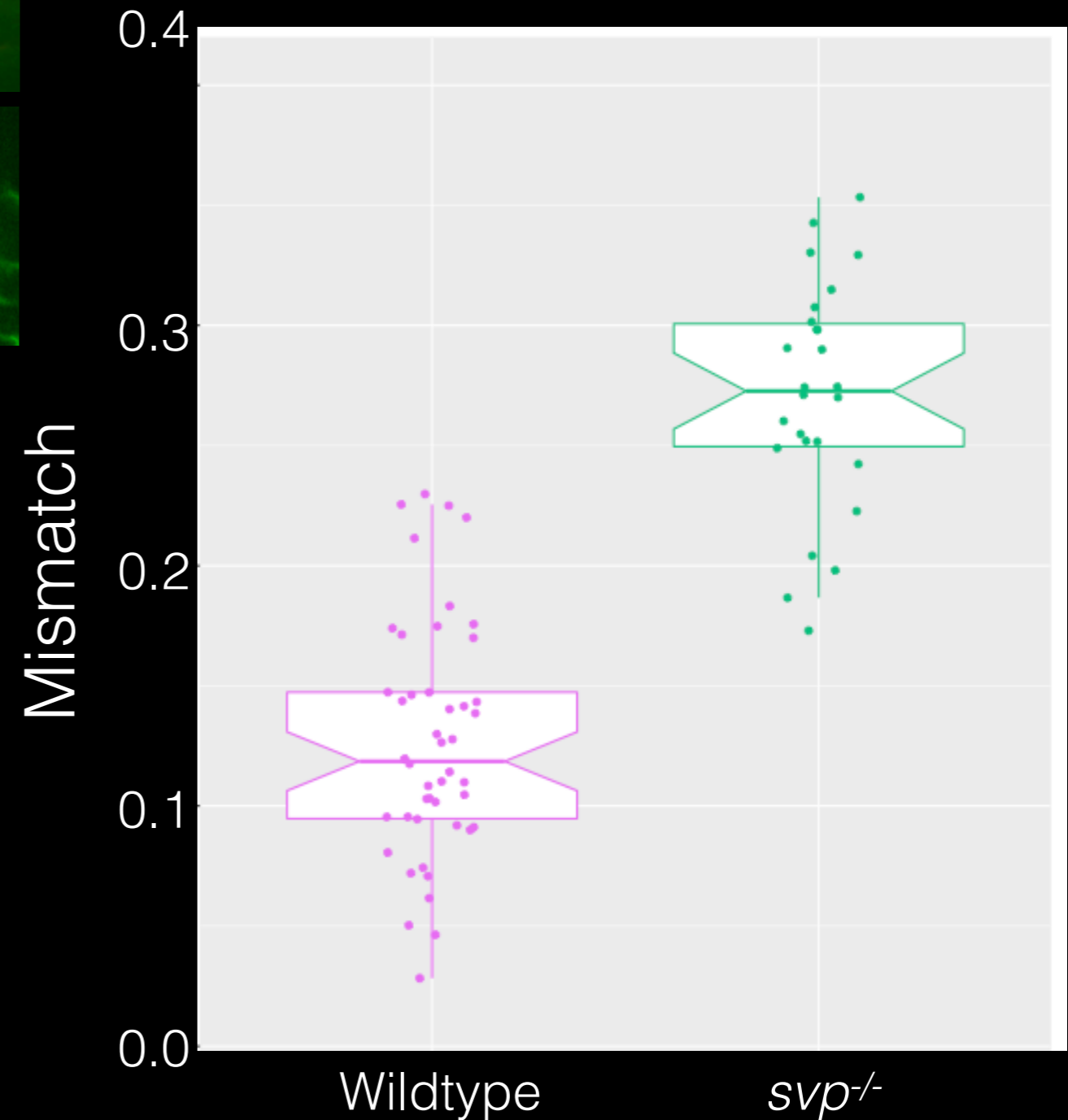
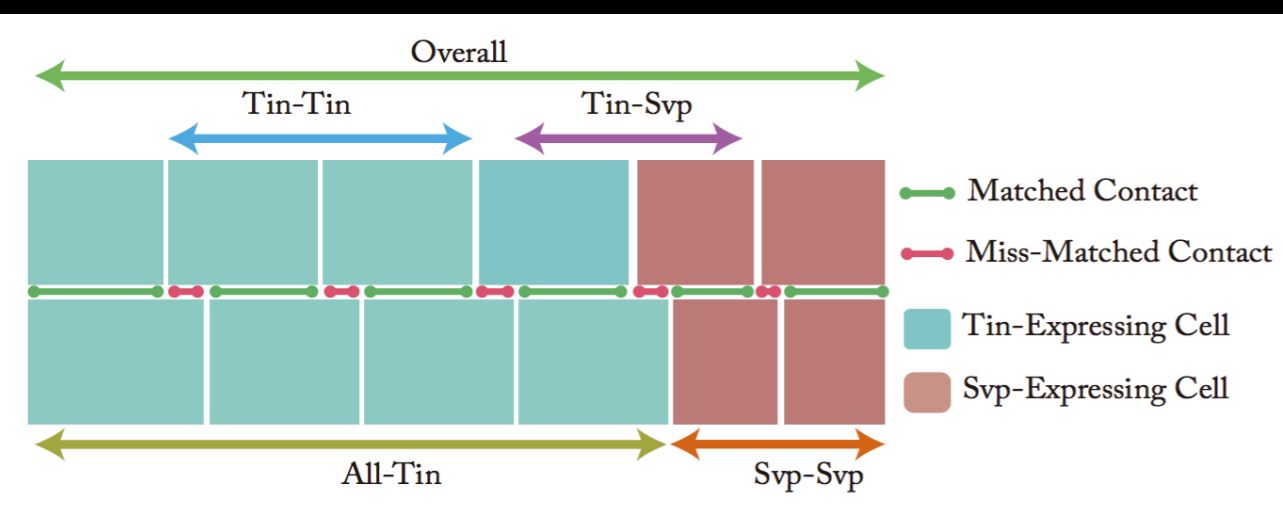
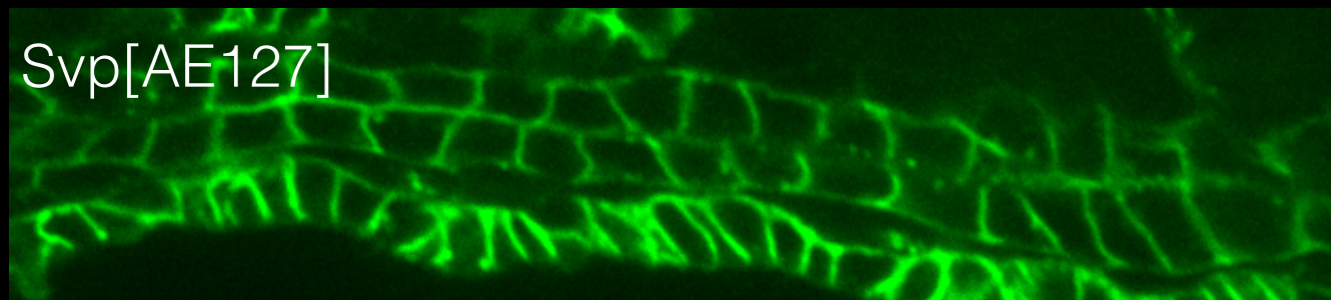
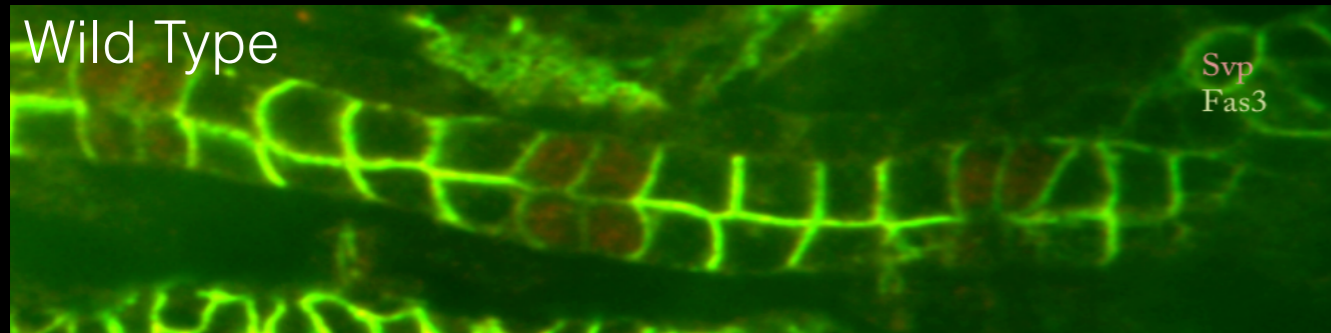
Quantifying cell matching



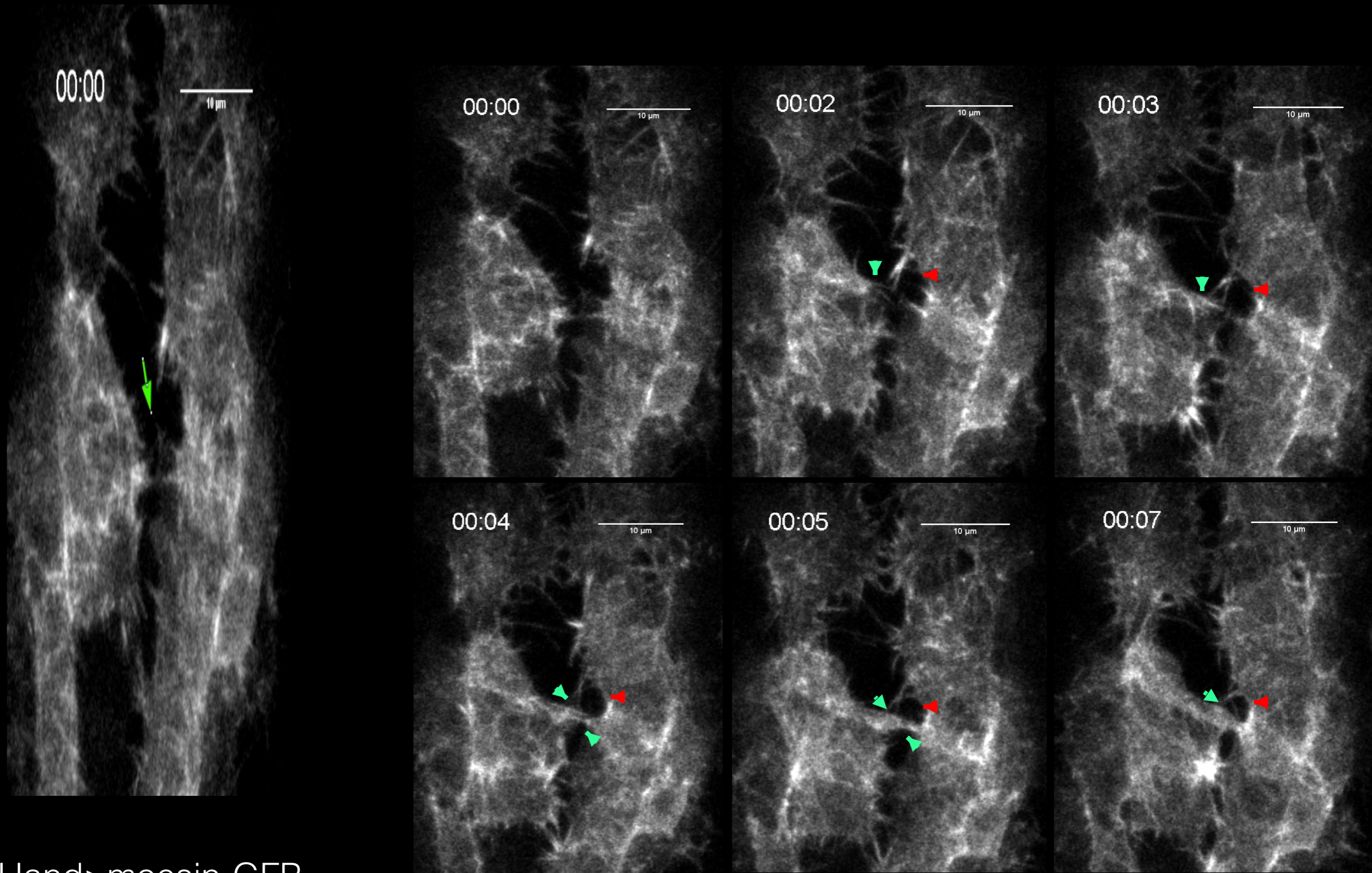
Cell matching robust at boundary of different cell types



Cell matching imprecise without different cell types



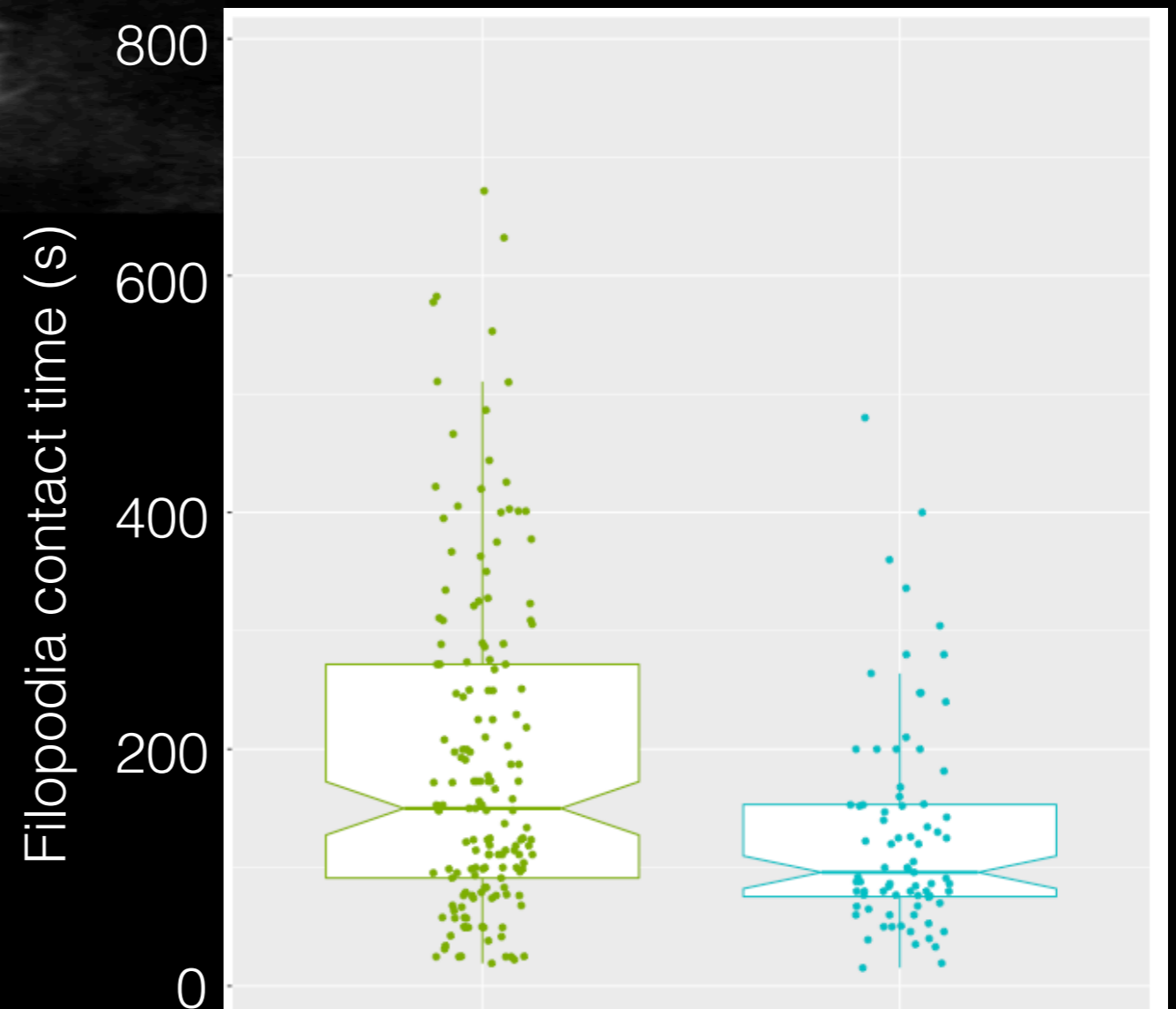
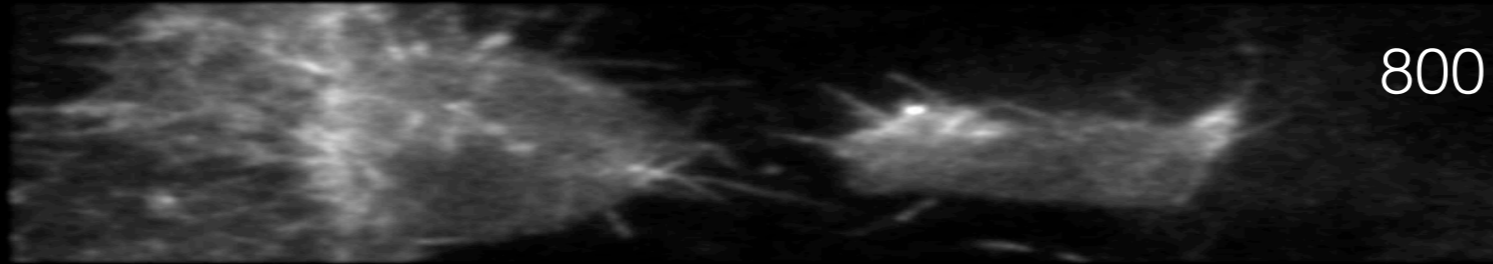
Filopodia are selectively binding to distinct cell types



Hand>moesin-GFP

Svp heart cells have weaker filopodia binding

Svp>moesin-GFP



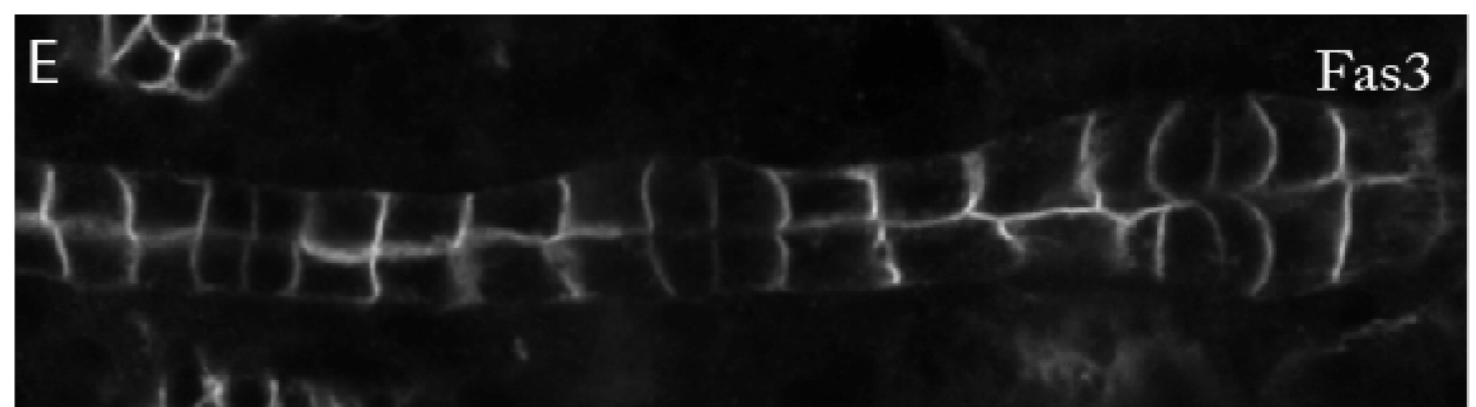
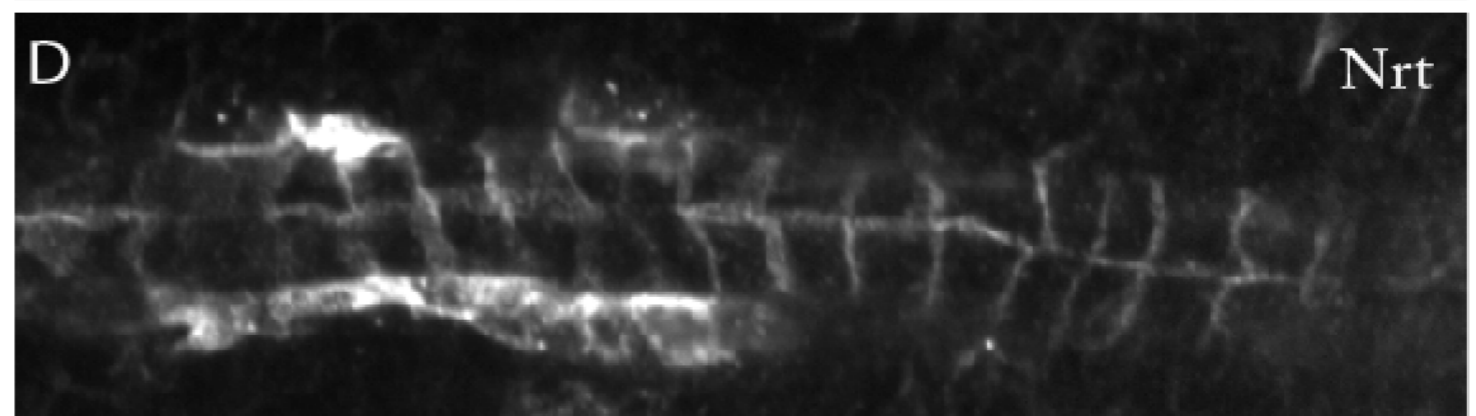
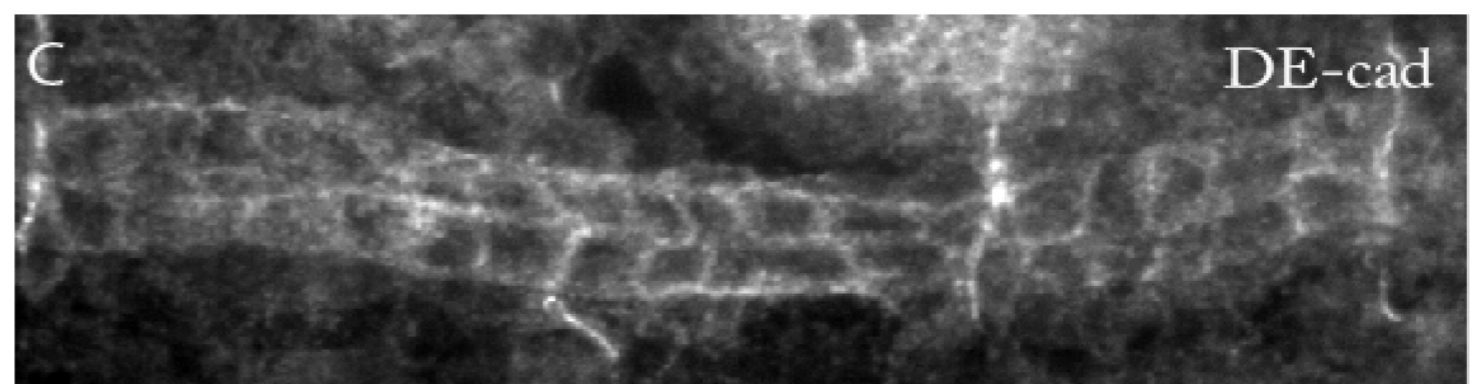
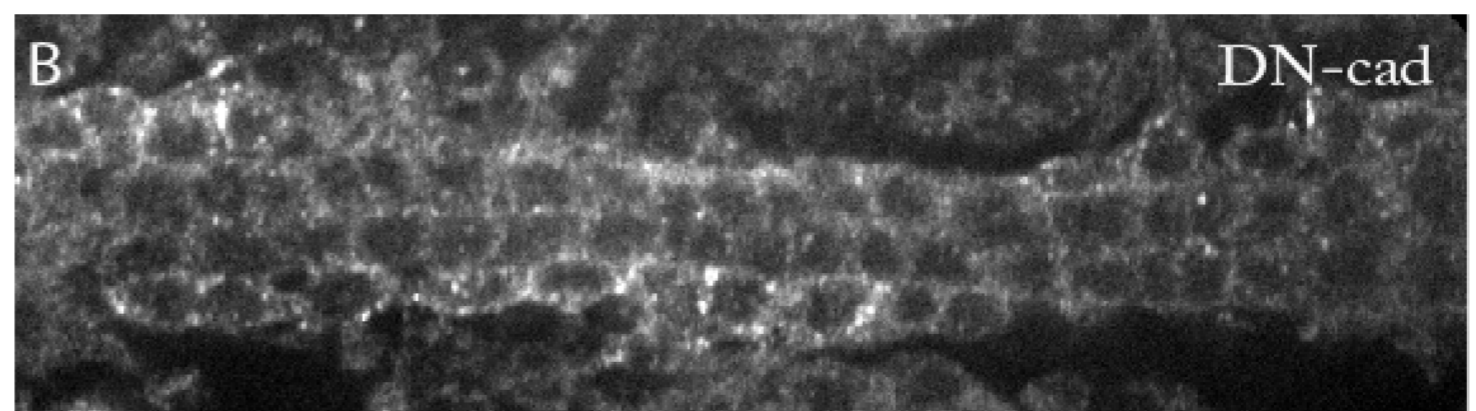
So what is the underlying mechanism?

All cardioblasts

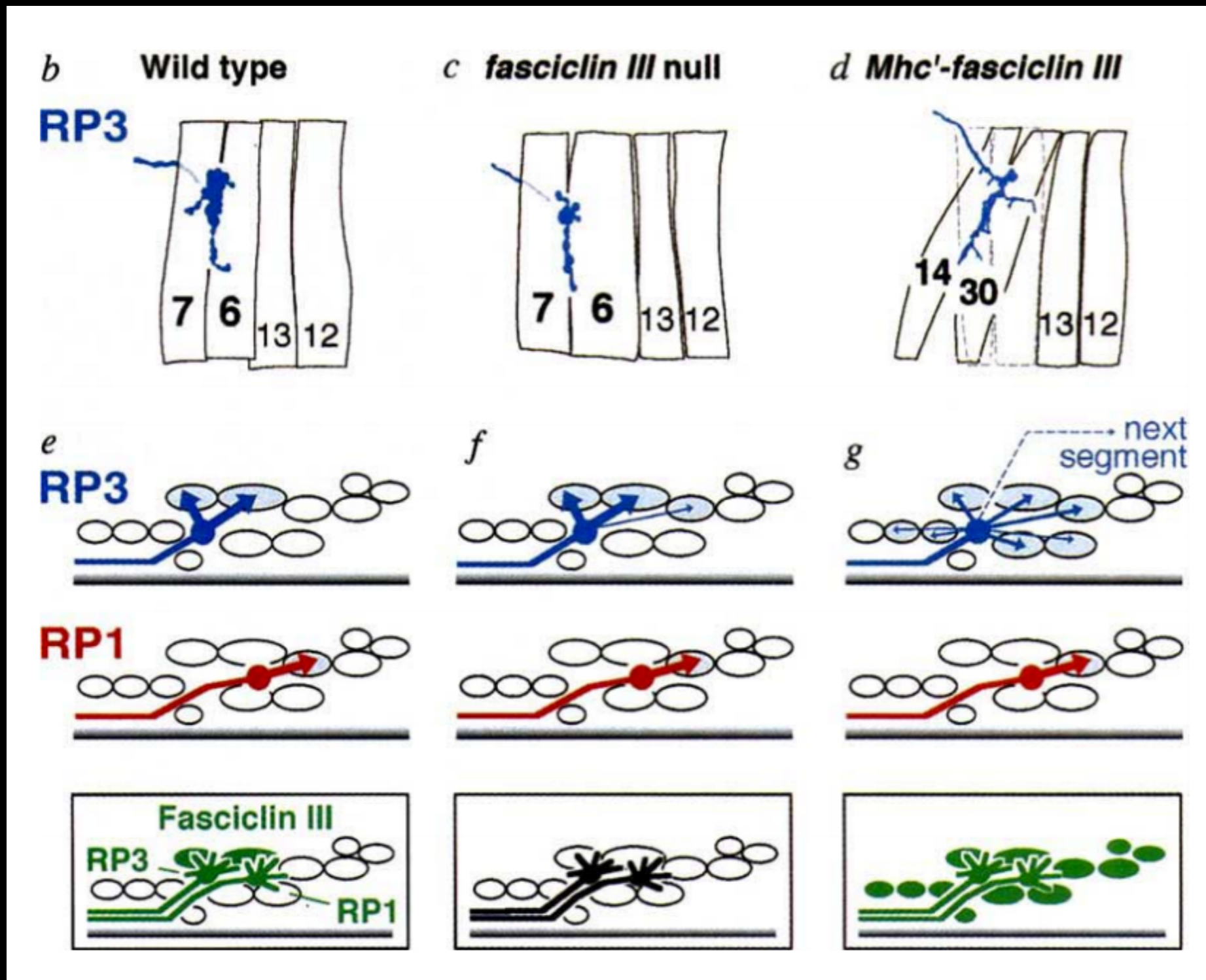
Svp-positive cardioblasts

Screen focusing on known neural cell matching molecules

Gene	Protein
shg	DE-cadherin
CadN	DN-cadherin
Nrt	Neurotactin
Nrg	Neuroglian
Con	Connectin
Fas1	Fascilin I
Fas2	Fascilin II
Fas3	Fascilin III
Ten-m	Tenascin major
Ten-a	Tenascin accessory
Dscam 1	Down syndrome cell adhesion molecule 1
Pvf3	PDGF- and VEGF-related factor 3



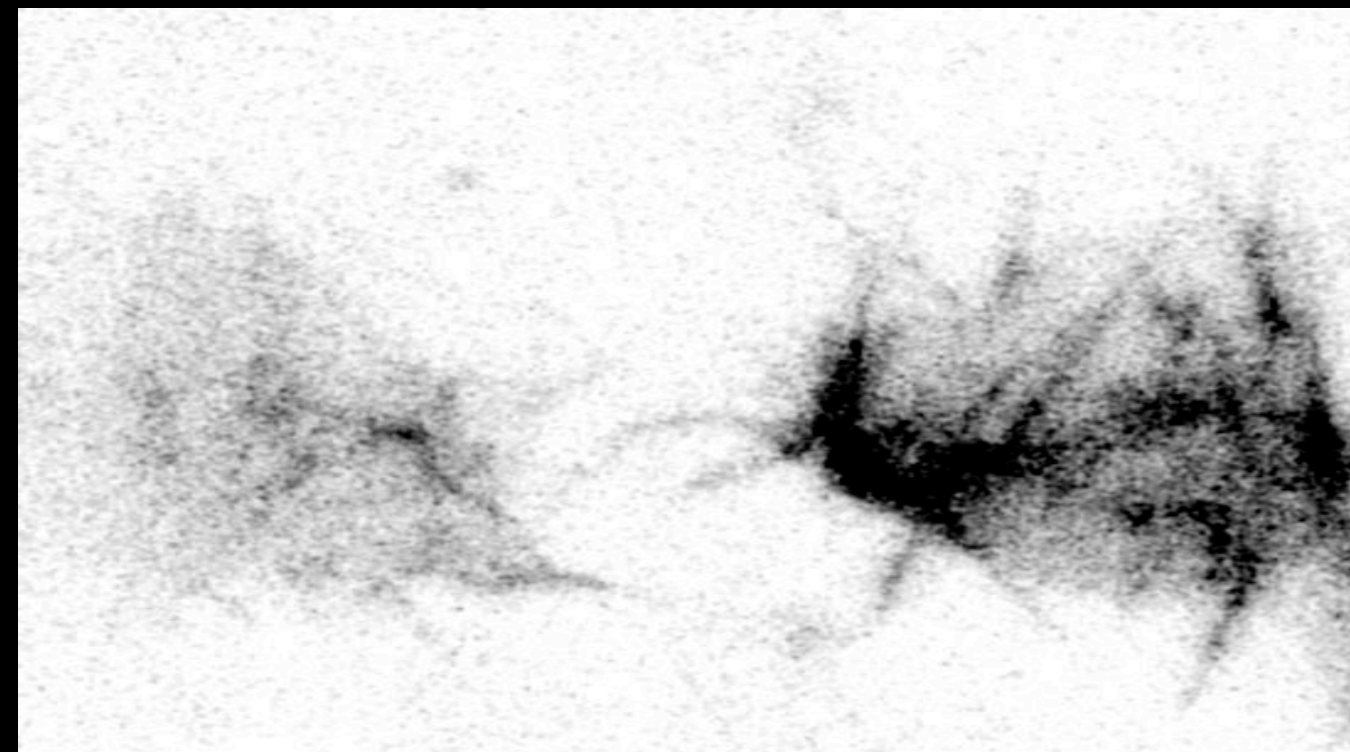
Fas3 is involved in neuronal cell matching



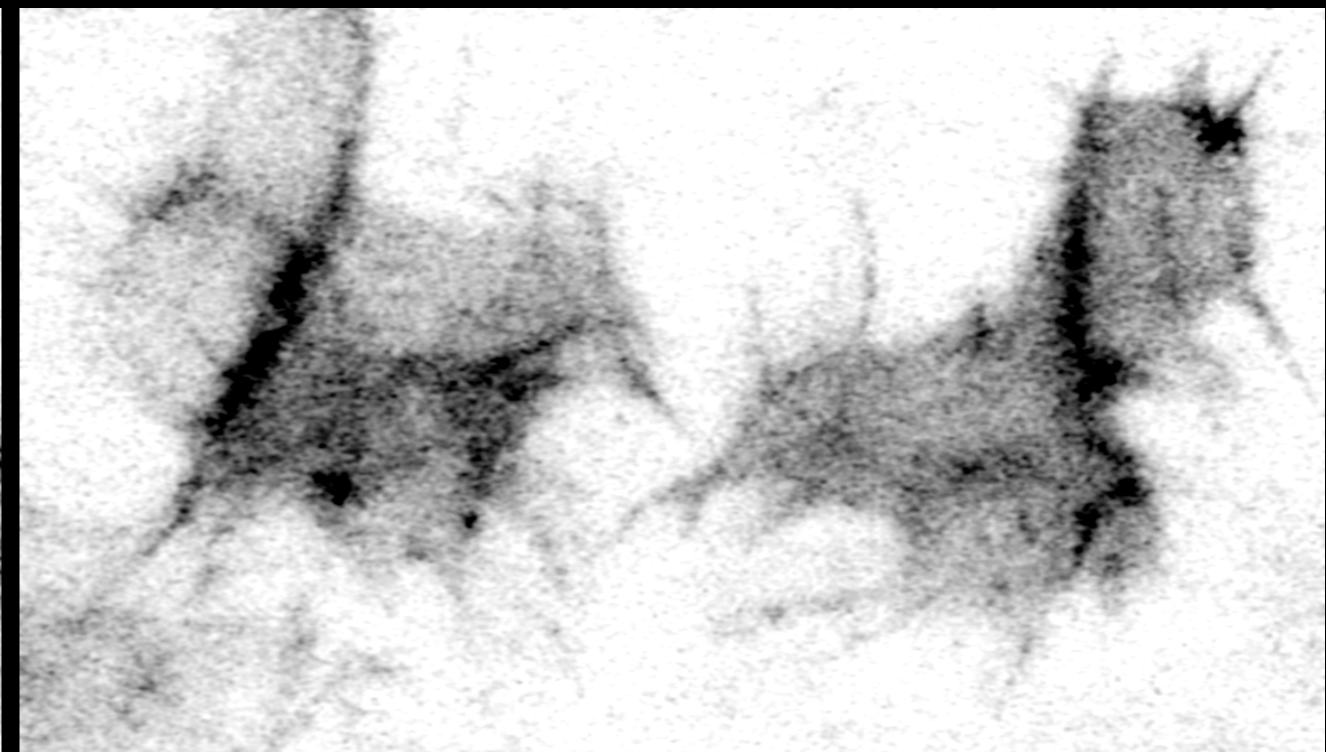
No observed phenotype in null mutant

Altering Fas3 expression effects filopodia contact time

Fas3 up-regulation

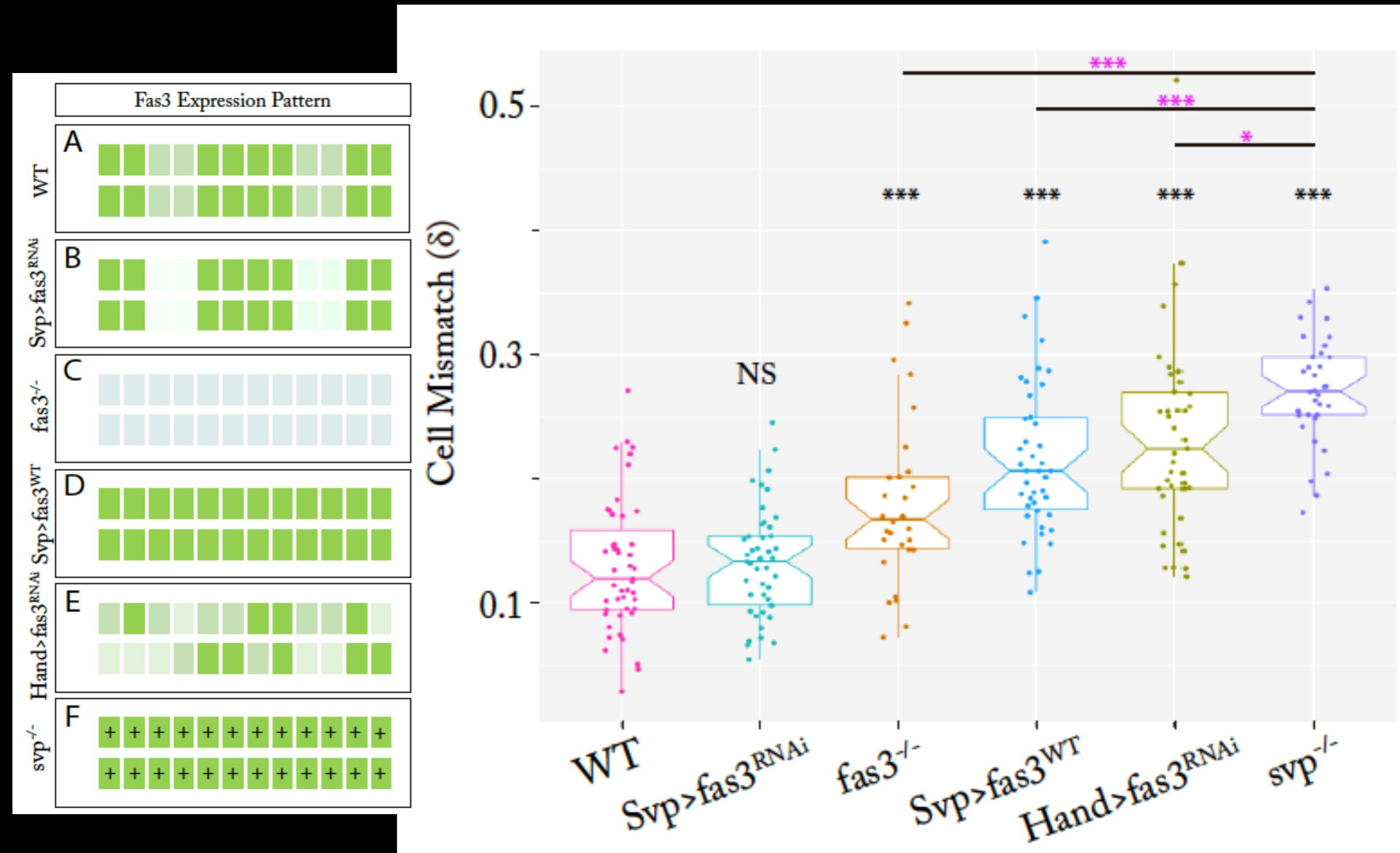


Svp>UAS-moesin-GFP

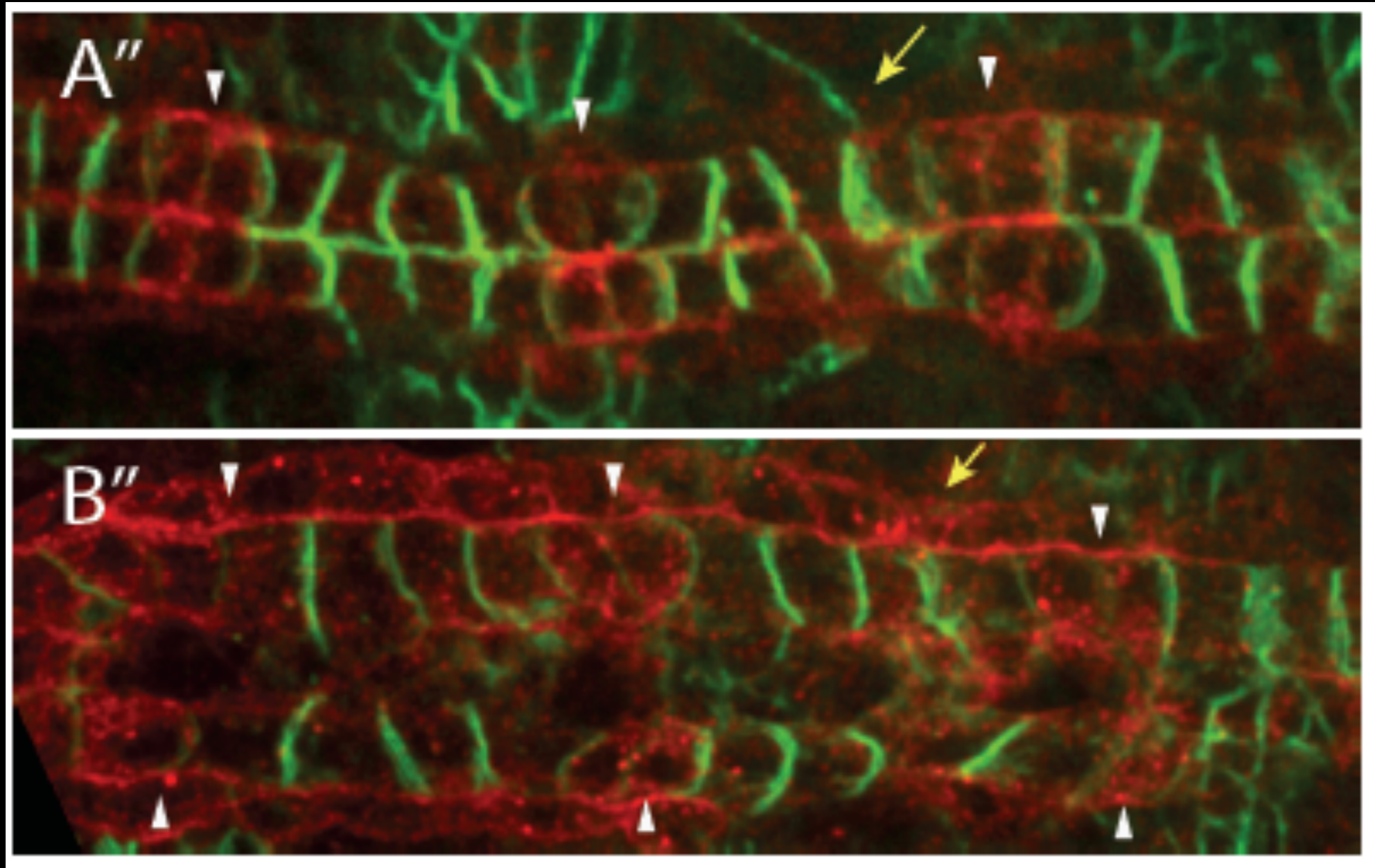


Svp> UAS-moesin-GFP
UAS-Fas3

How does the Fas3 expression pattern affect cell matching?



Ten-m is expressed in a complementary fashion to Fas3



Fas3
MiMIC-Ten-m-GFP

LETTER

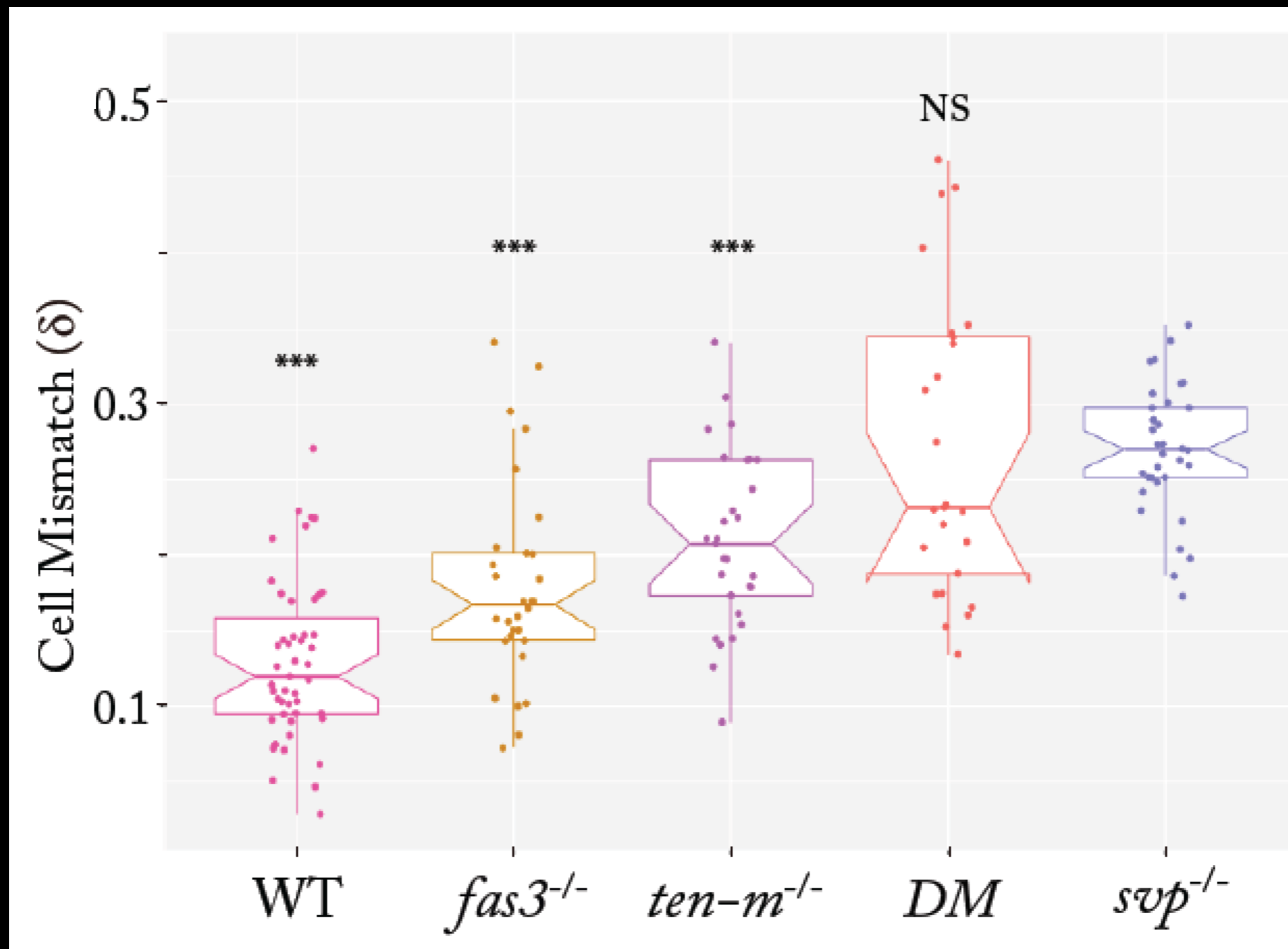
doi:10.1038/nature10923

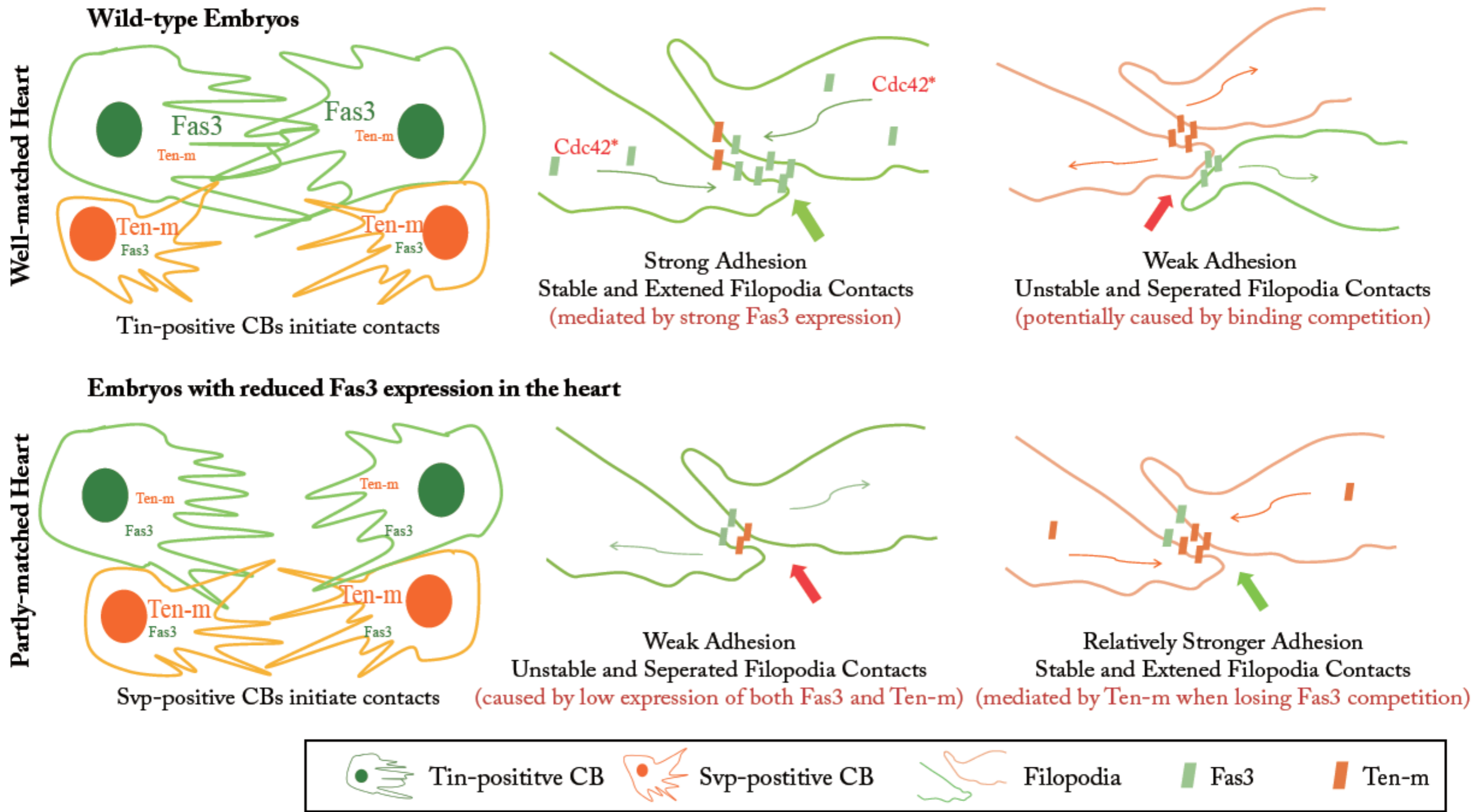
Trans-synaptic Teneurin signalling in neuromuscular synapse organization and target choice

Timothy J. Mosca^{1*}, Weizhe Hong^{2*}, Vardhan S. Dani¹, Vincenzo Favaloro¹ & Liqun Luo¹

odd oz (odz)

Fas3 / Ten-m double mutant has severe matching defects



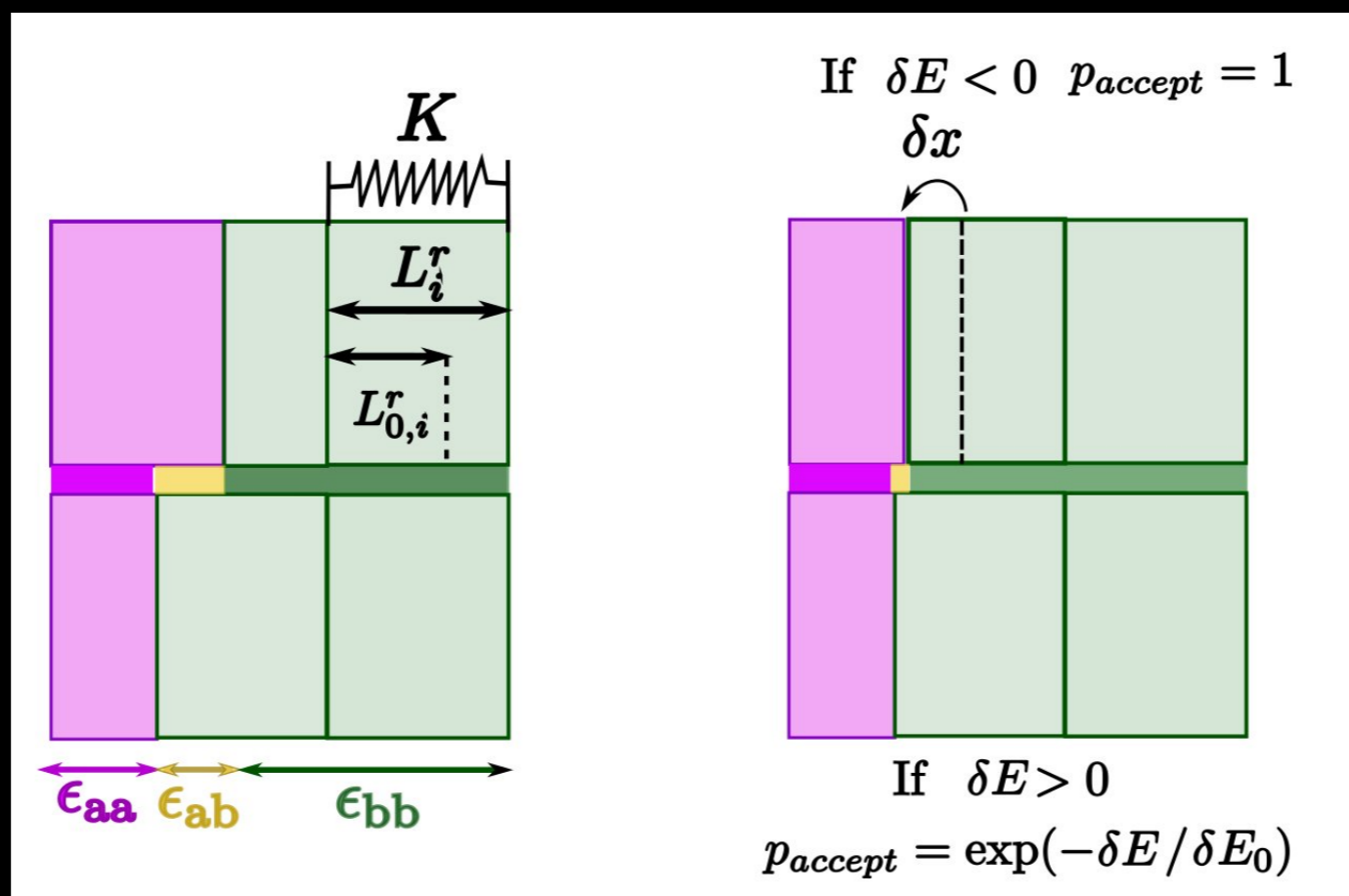


Zhang *et al.* Developmental Cell (2018)

Is differential adhesion sufficient to ensure precise cell matching?

$$E_a = K(L_{cell} - L_{0,a})^2 - (\epsilon_{aa} \cdot x + \epsilon_{ab} \cdot y)$$

$$E_b = K(L_{cell} - L_{0,b})^2 - (\epsilon_{bb} \cdot x + \epsilon_{ab} \cdot y)$$

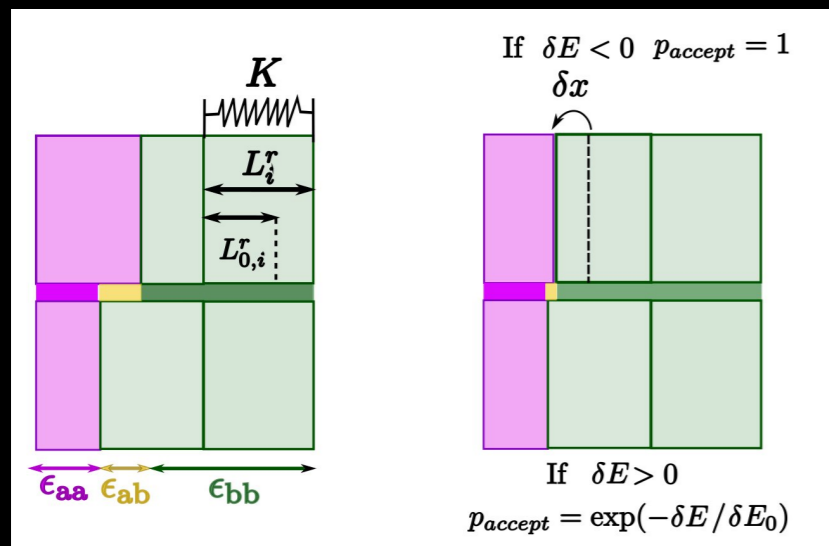


Tlili *et al.* Biorxiv 2019 (653535)

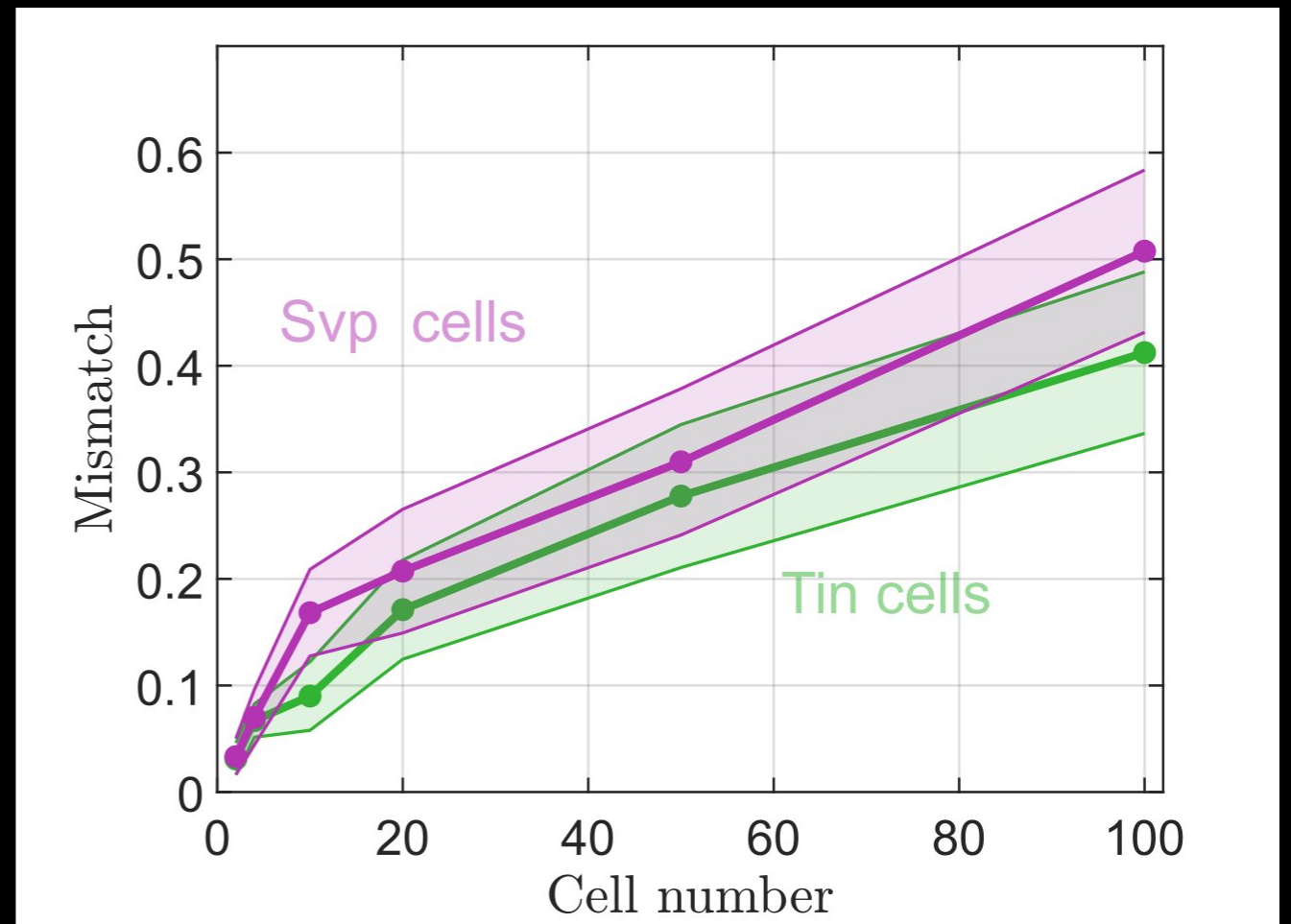
Cell matching consistent with an equilibrium energy approach

$$E_a = K(L_{cell} - L_{0,a})^2 - (\epsilon_{aa} \cdot x + \epsilon_{ab} \cdot y)$$

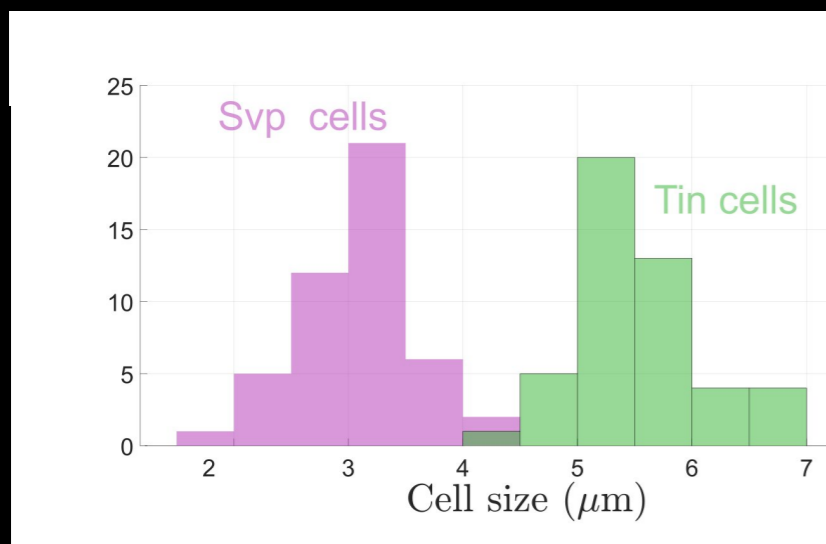
$$E_b = K(L_{cell} - L_{0,b})^2 - (\epsilon_{bb} \cdot x + \epsilon_{ab} \cdot y)$$



Equilibrate and measure mismatch



Input cell sizes

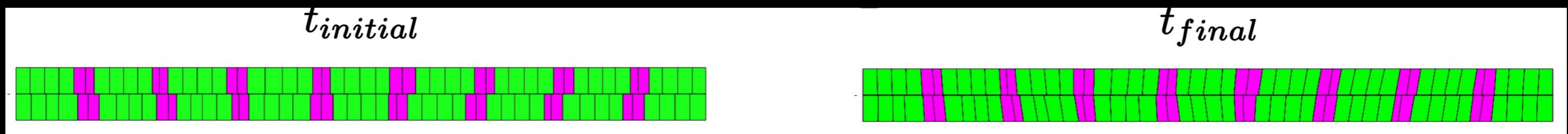


Cell matching consistent with an equilibrium energy approach

$$E_a = K(L_{cell} - L_{0,a})^2 - (\epsilon_{aa} \cdot x + \epsilon_{ab} \cdot y)$$

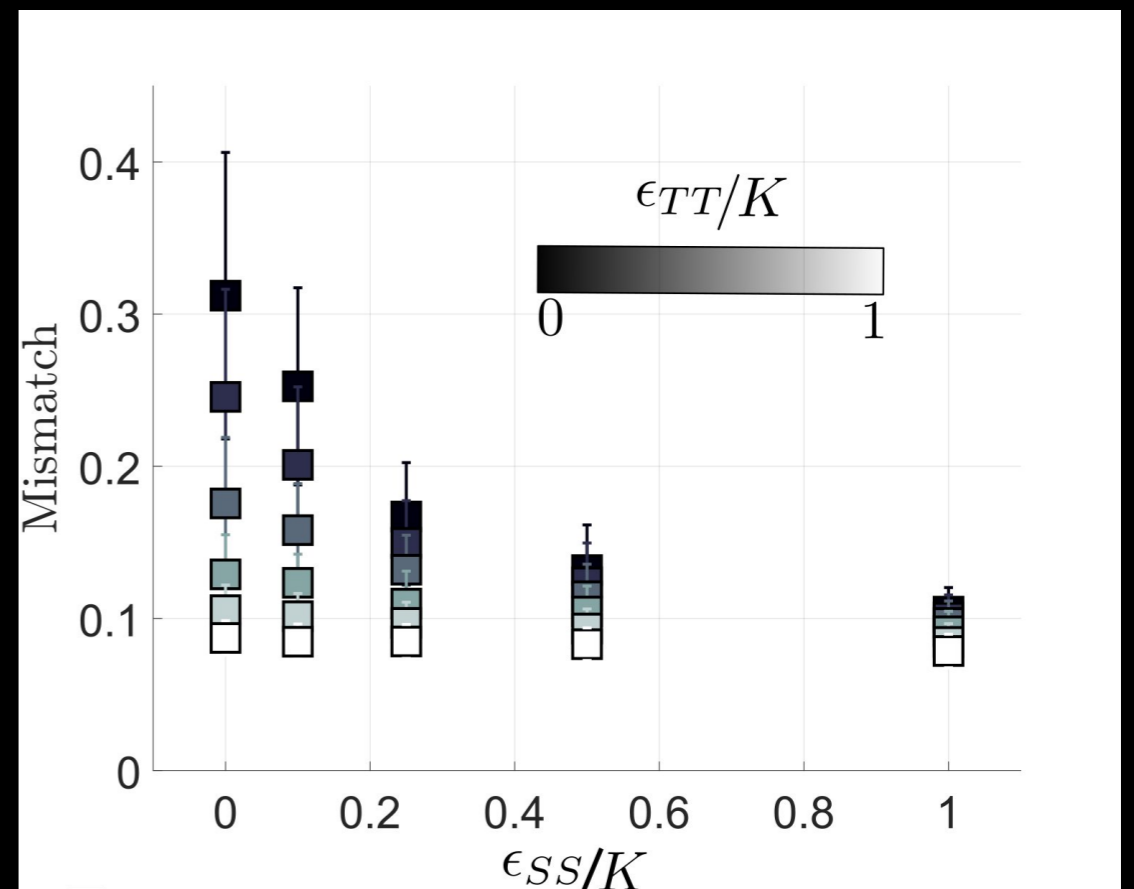
$$E_b = K(L_{cell} - L_{0,b})^2 - (\epsilon_{bb} \cdot x + \epsilon_{ab} \cdot y)$$

Potts model

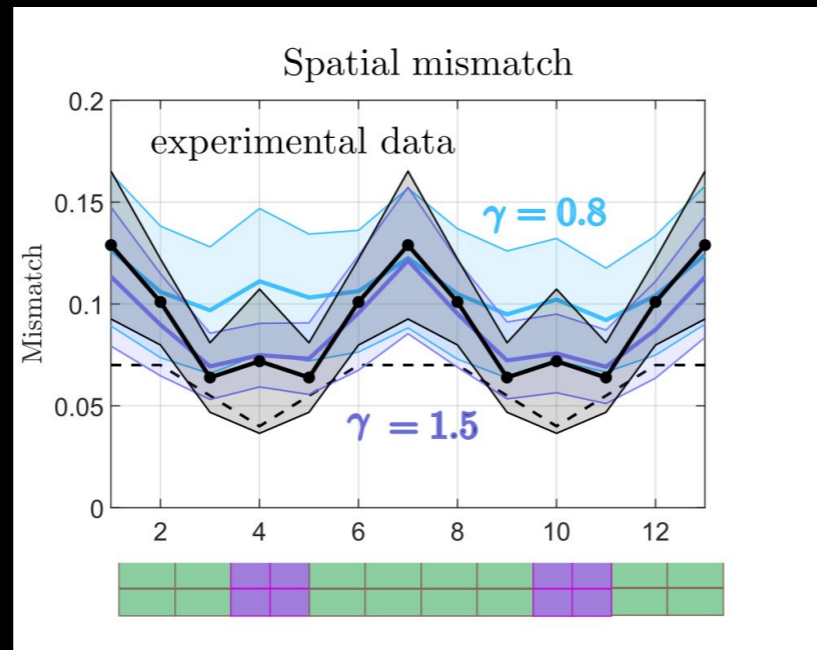


Behaviour depends on

$$\gamma = \frac{(\epsilon_{TT} - \epsilon_{ST}) + (\epsilon_{SS} - \epsilon_{ST})}{K}$$

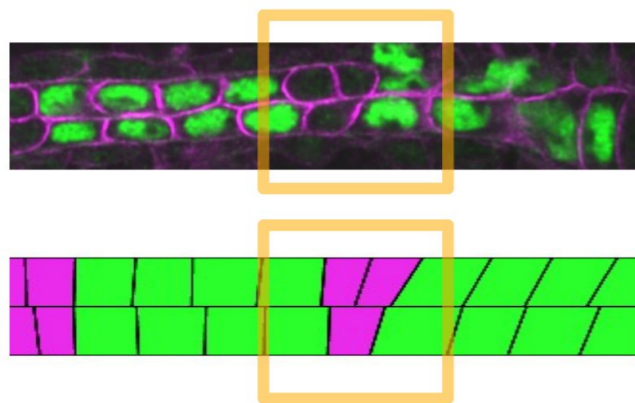


Mismatch in wild-type and deformed hearts

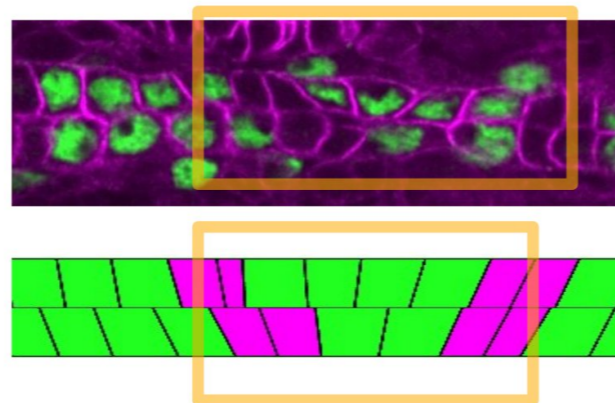


Cell alignment best at boundaries between cell types

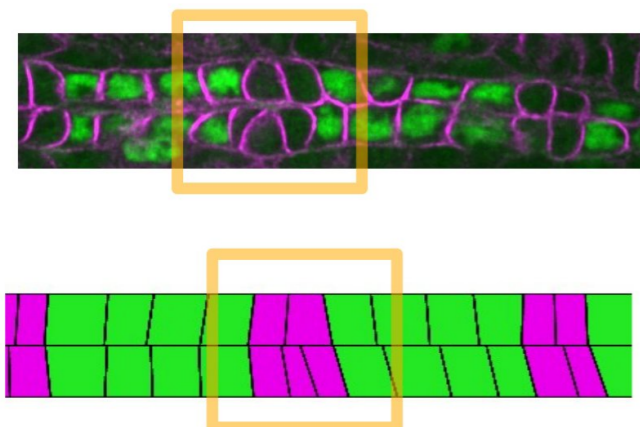
1 Svp cell



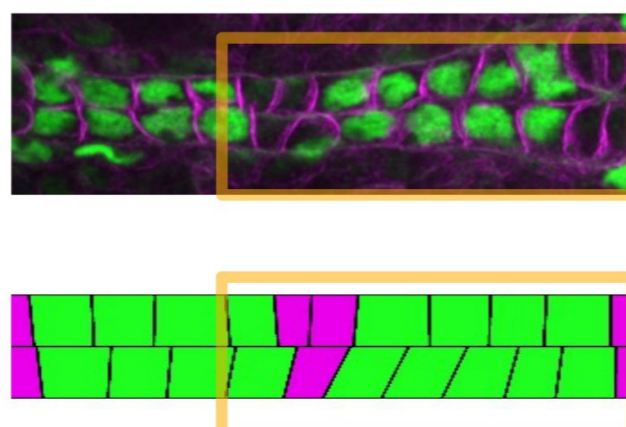
2 Tin cells



3 Svp cells

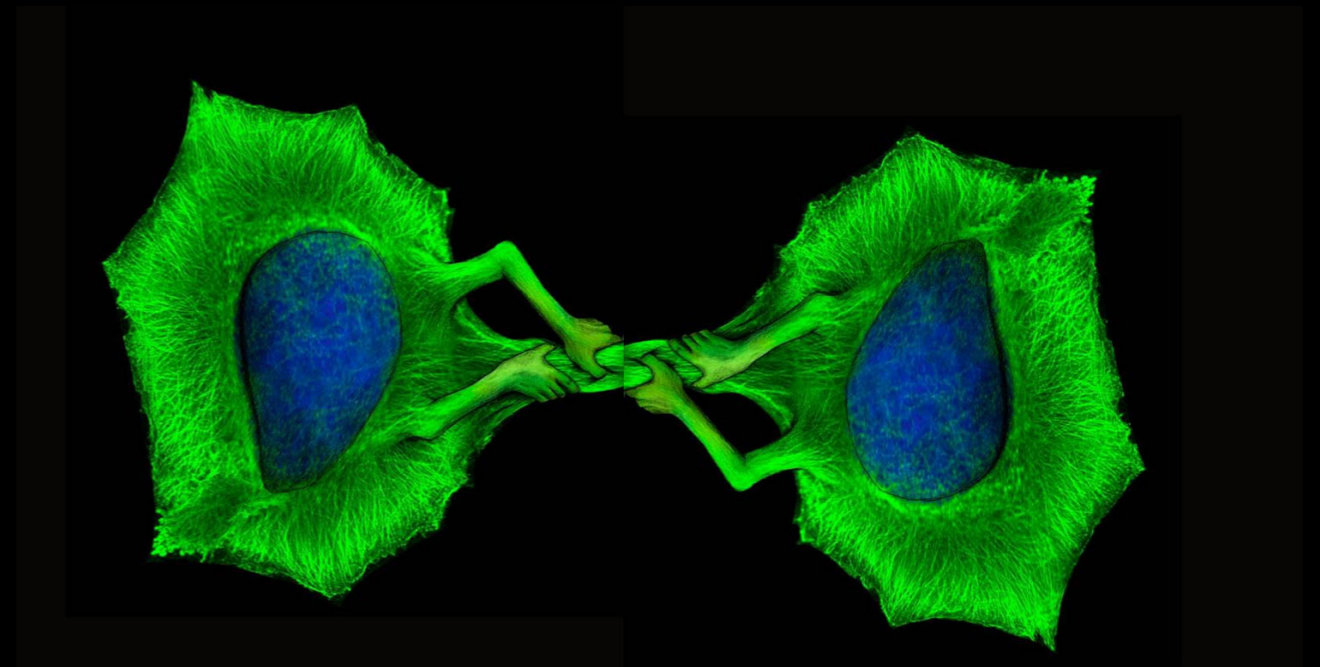
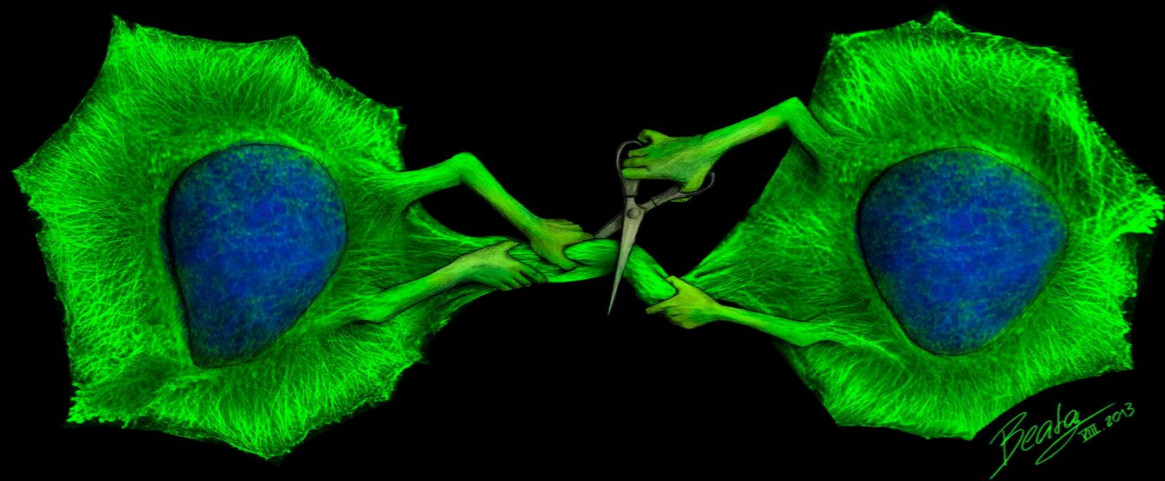


5 Tin cells 1 Svp cell

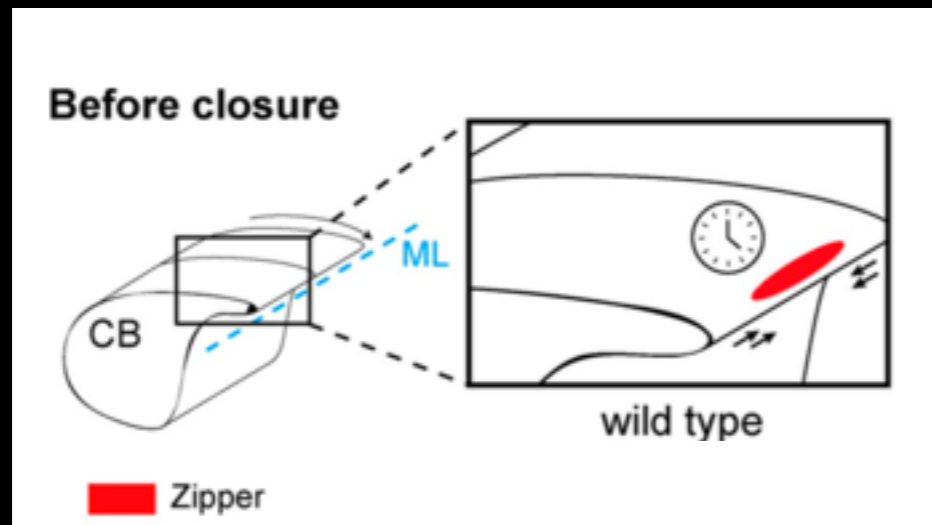


Alignment consistent with observed cases of cardioblast number variation

Coupling between waves of Myosin-II and filopodia adhesion regulates cell matching



Known waves of Myosin-II in cardioblasts during maturation

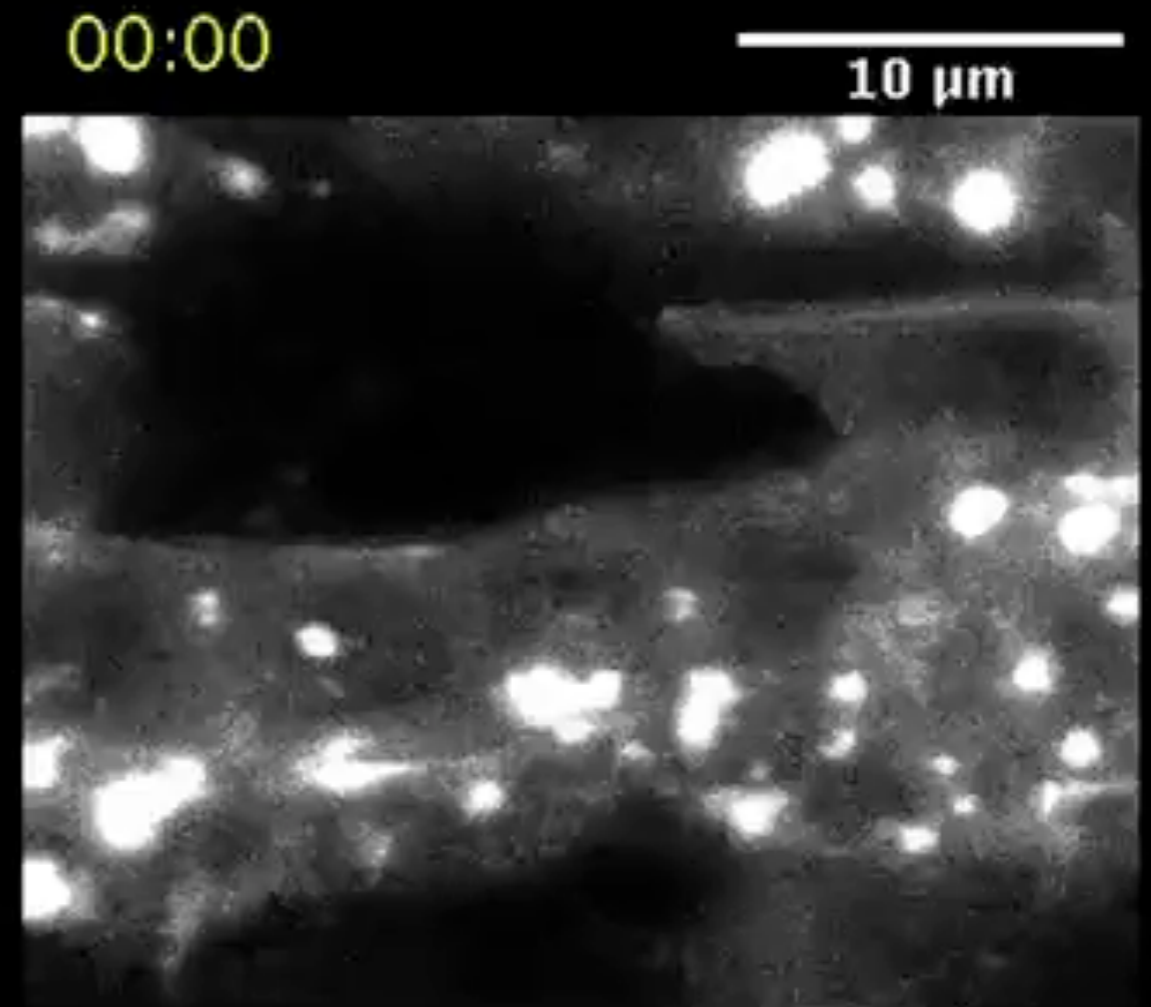


Drosophila

Zipper: Myosin II heavy chain

Squash: Myosin II light chain

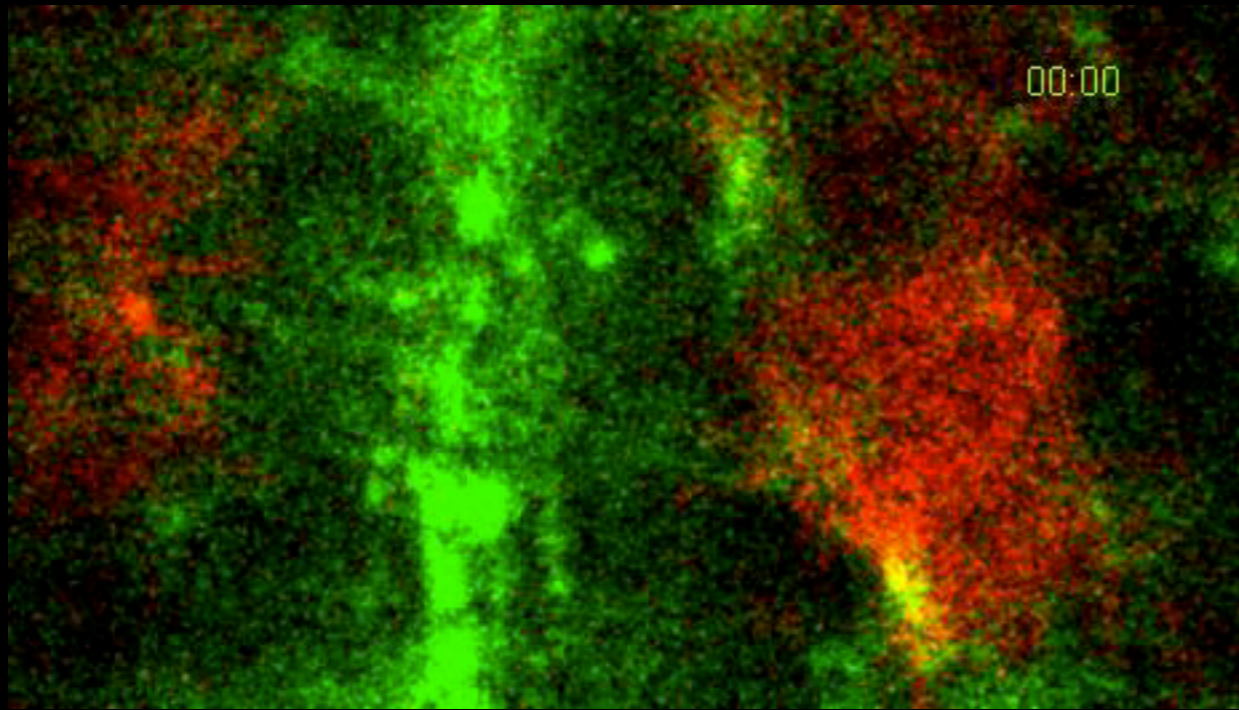
**What is the function
of this wave?**



tinCΔ4 > zipper::GFP

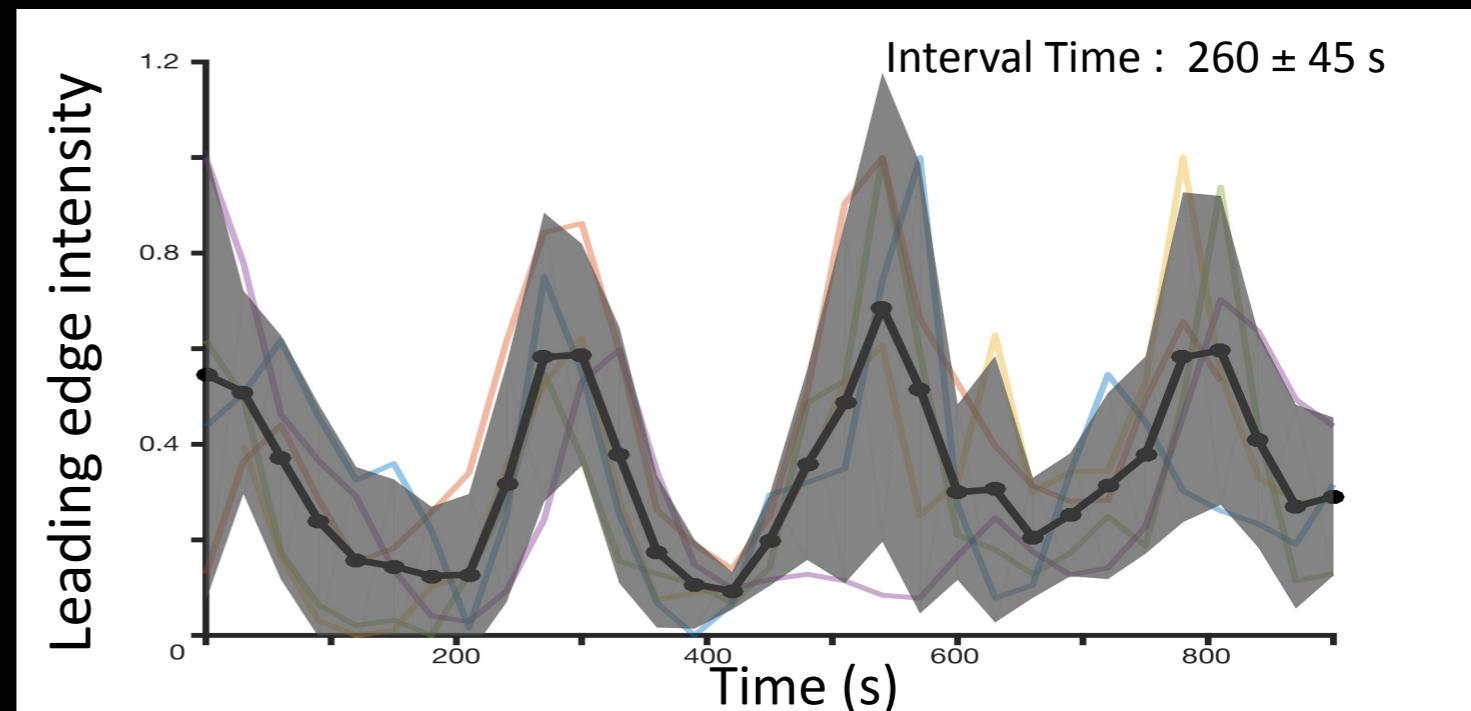
Vogler *et al.*, JCB 2014

Myosin-II waves have period ~ 4 minutes

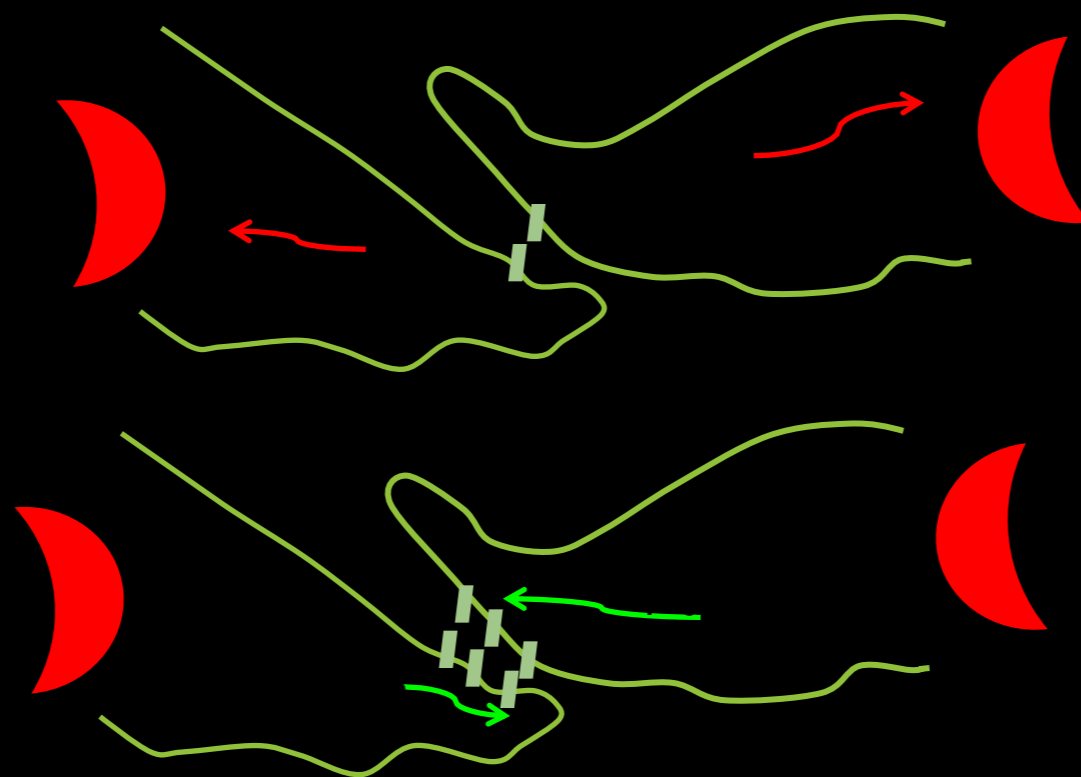
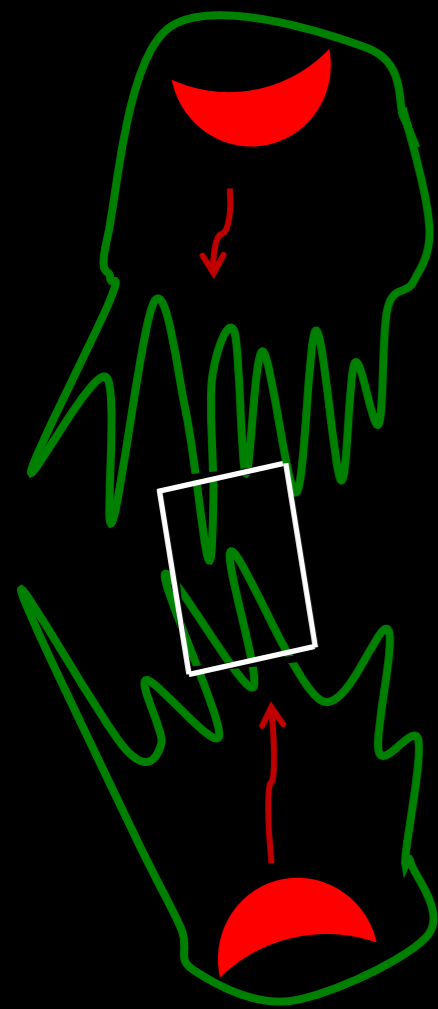


Zipper::GFP

Hand>Moesin::mCherry



Counterbalance of filopodia adhesion and Myosin II



Weak adhesion

Broken contacts
Retracted filopodia

Strong adhesion

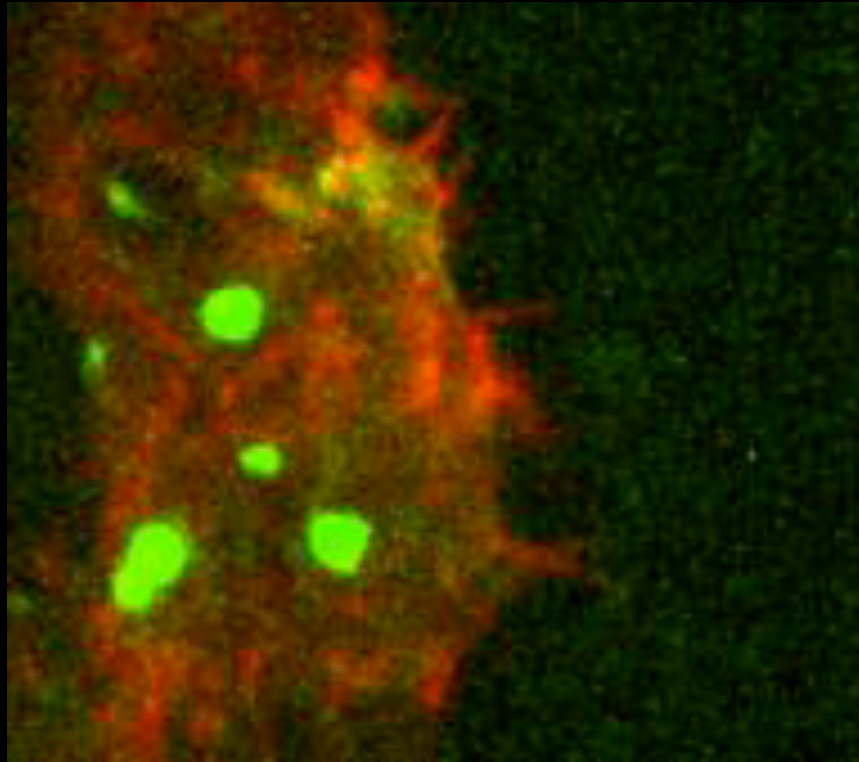
Contacts stabilise
Drives cells matching

Actin-Dependent
Stabilisation

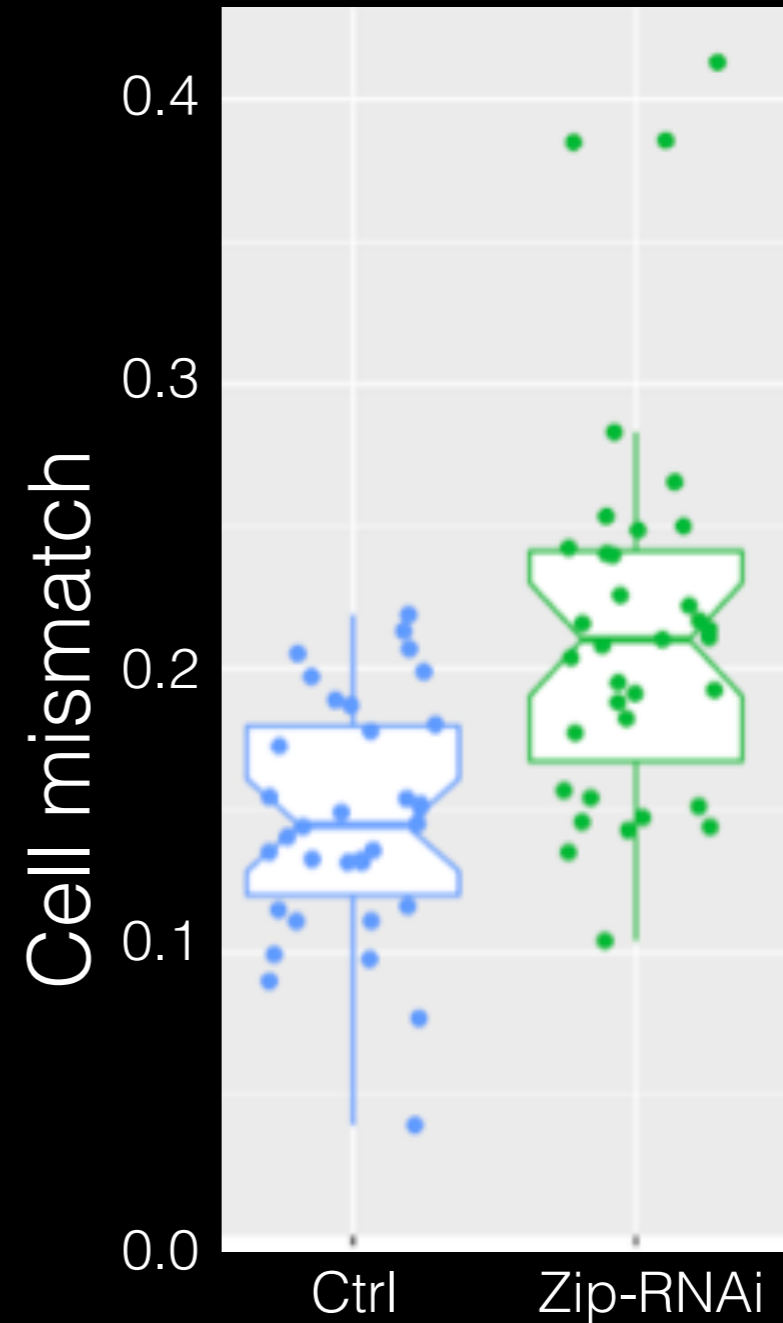
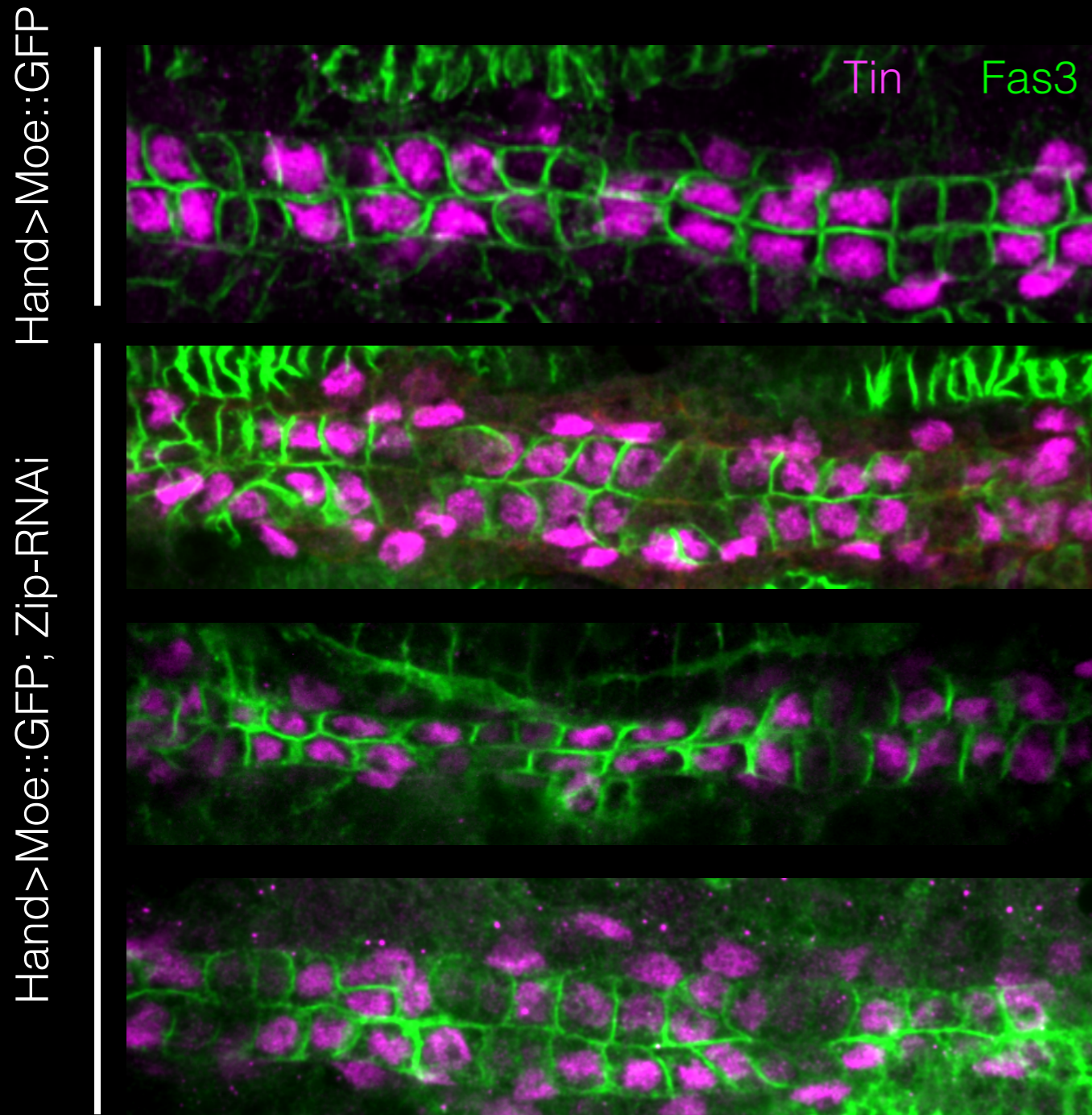


Myosin-II Dependent
Contraction

Filopodia retraction correlates with Myosin II appearance at cell leading edge

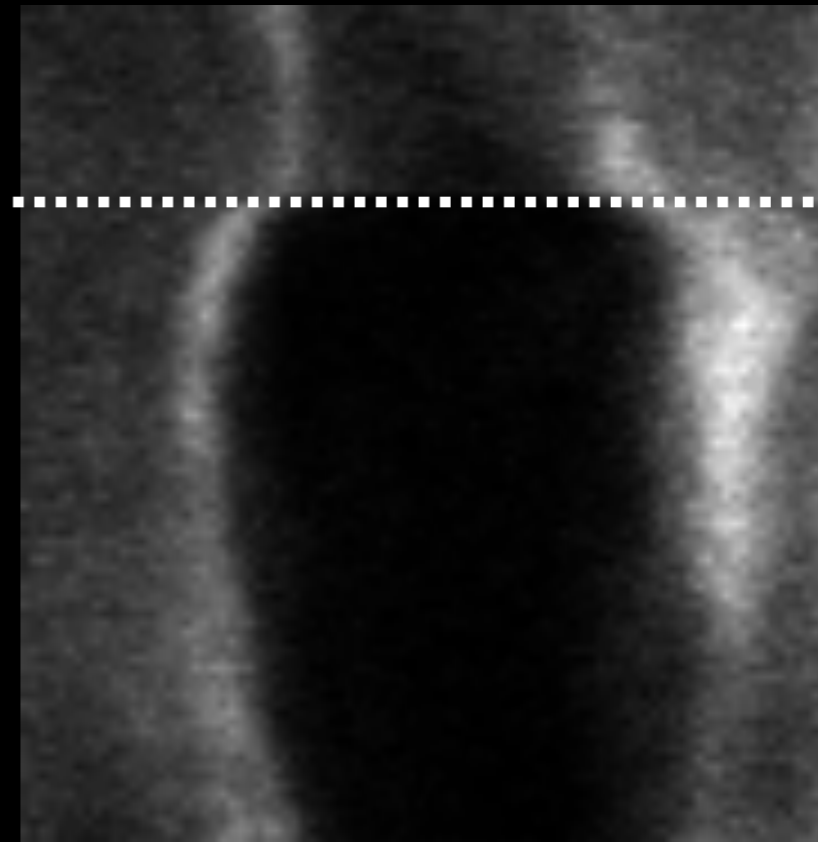
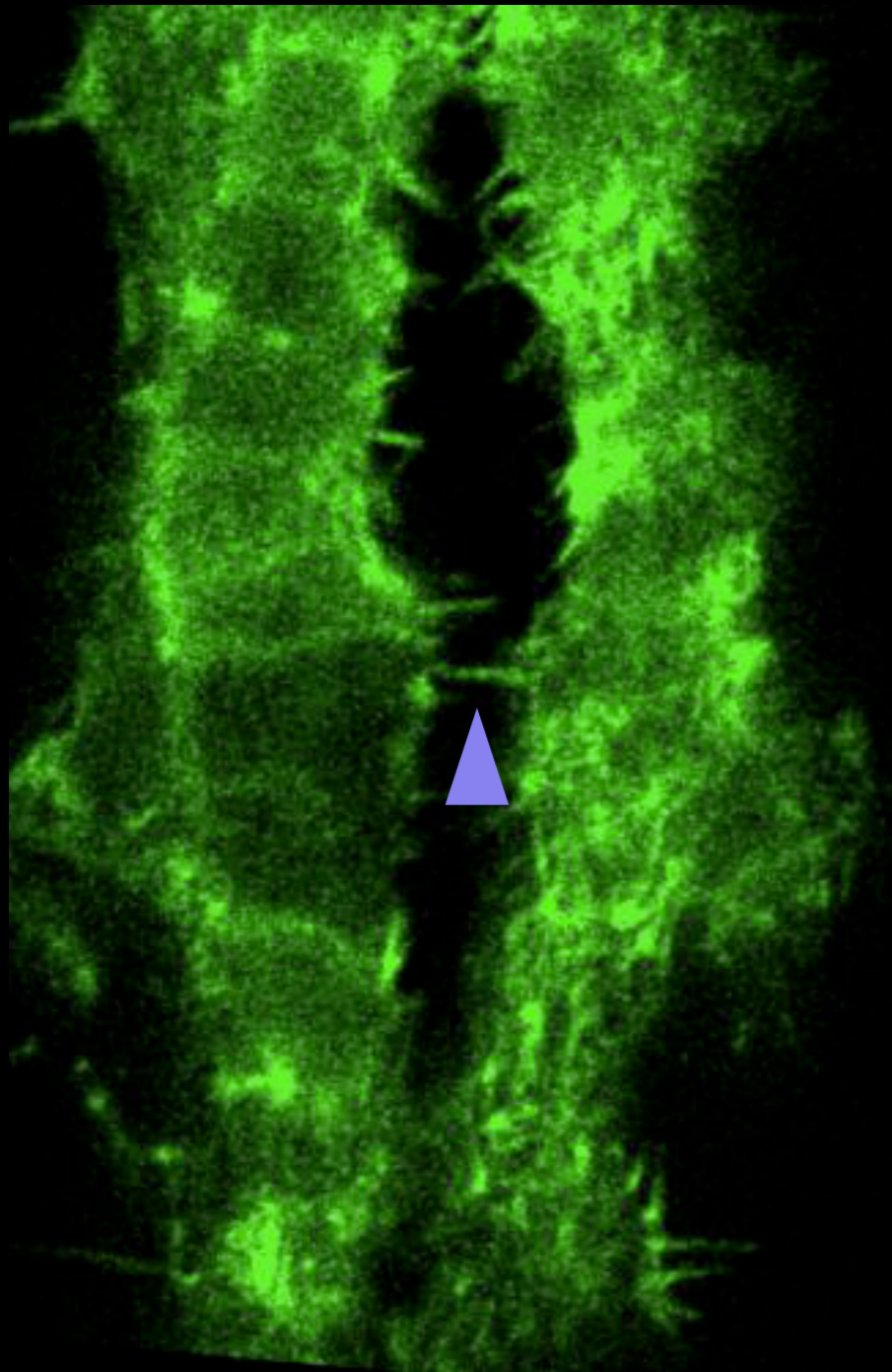


Reduction in Myosin-II results in reduced cell matching



Strong force between filopodia in wildtype hearts

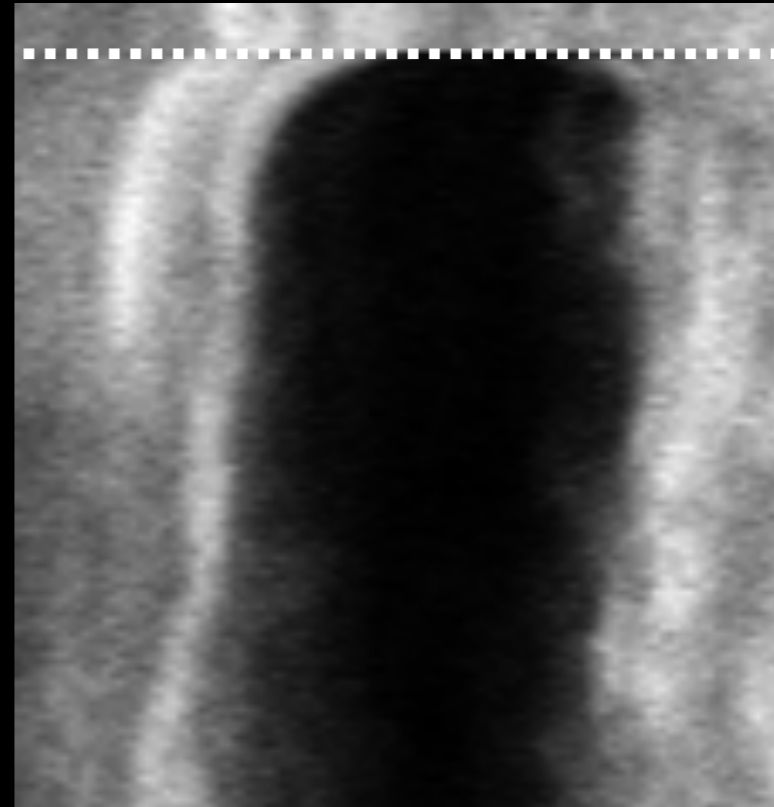
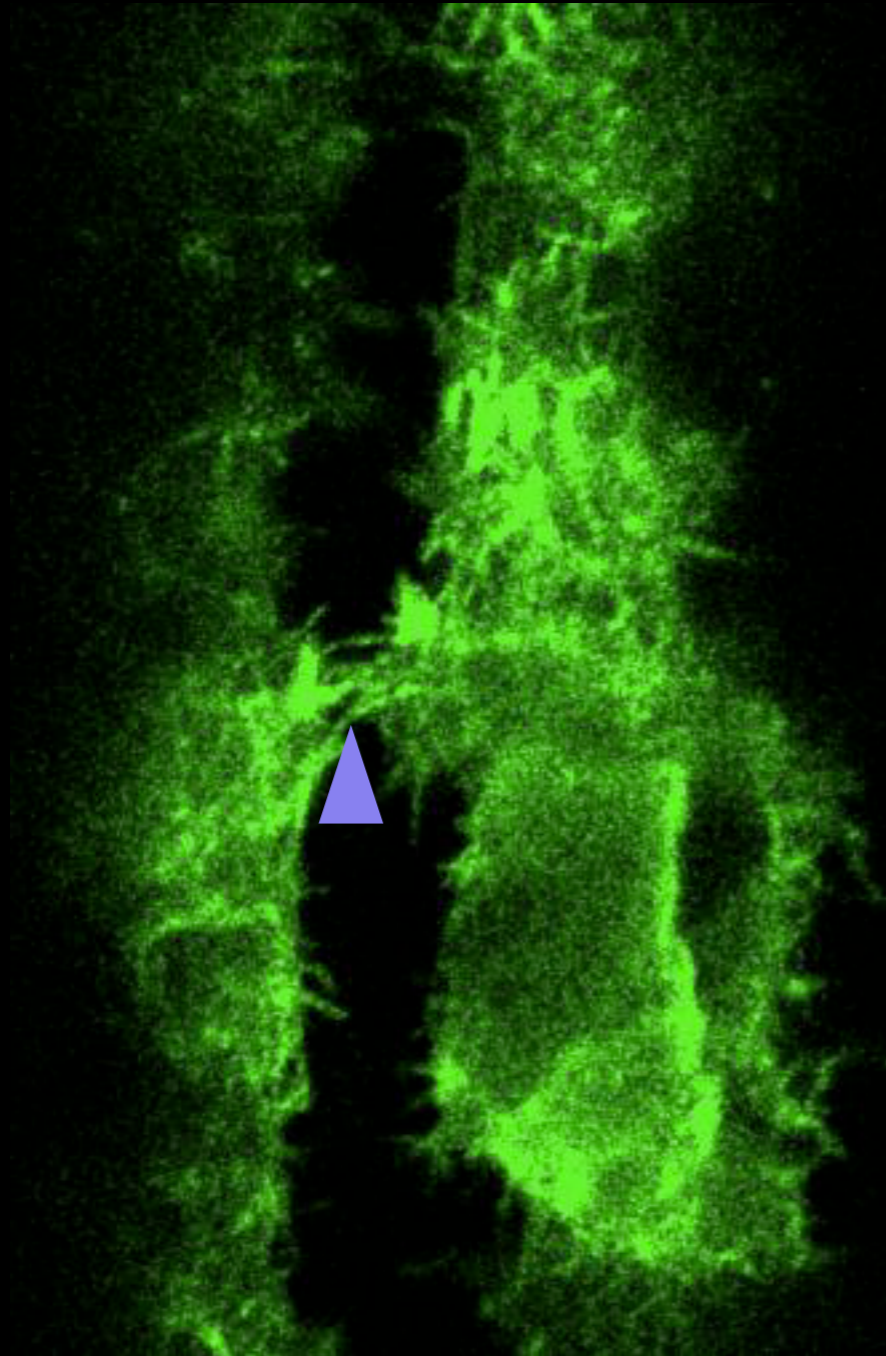
Hand>Moe::GFP



with Y. Toyama

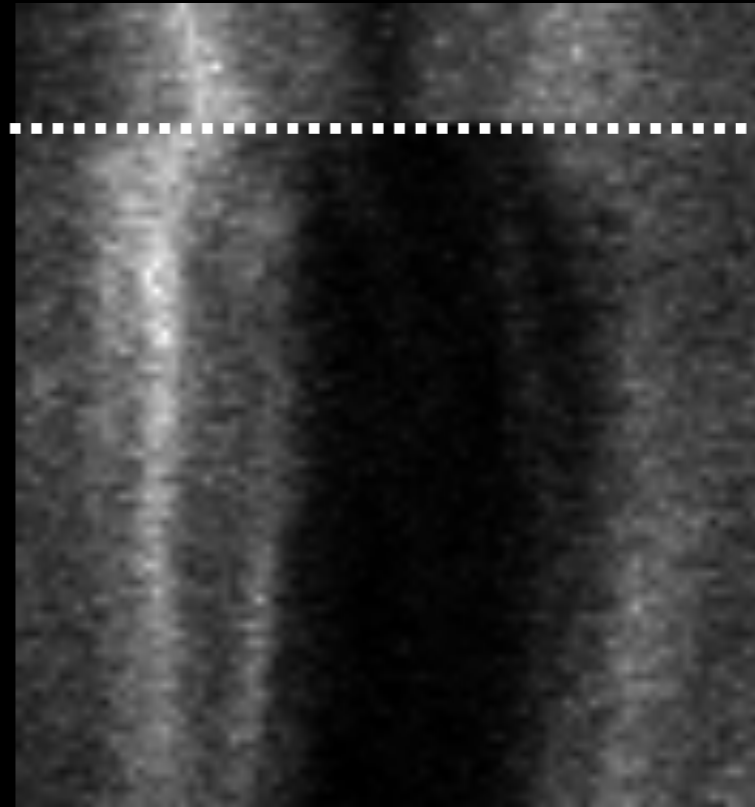
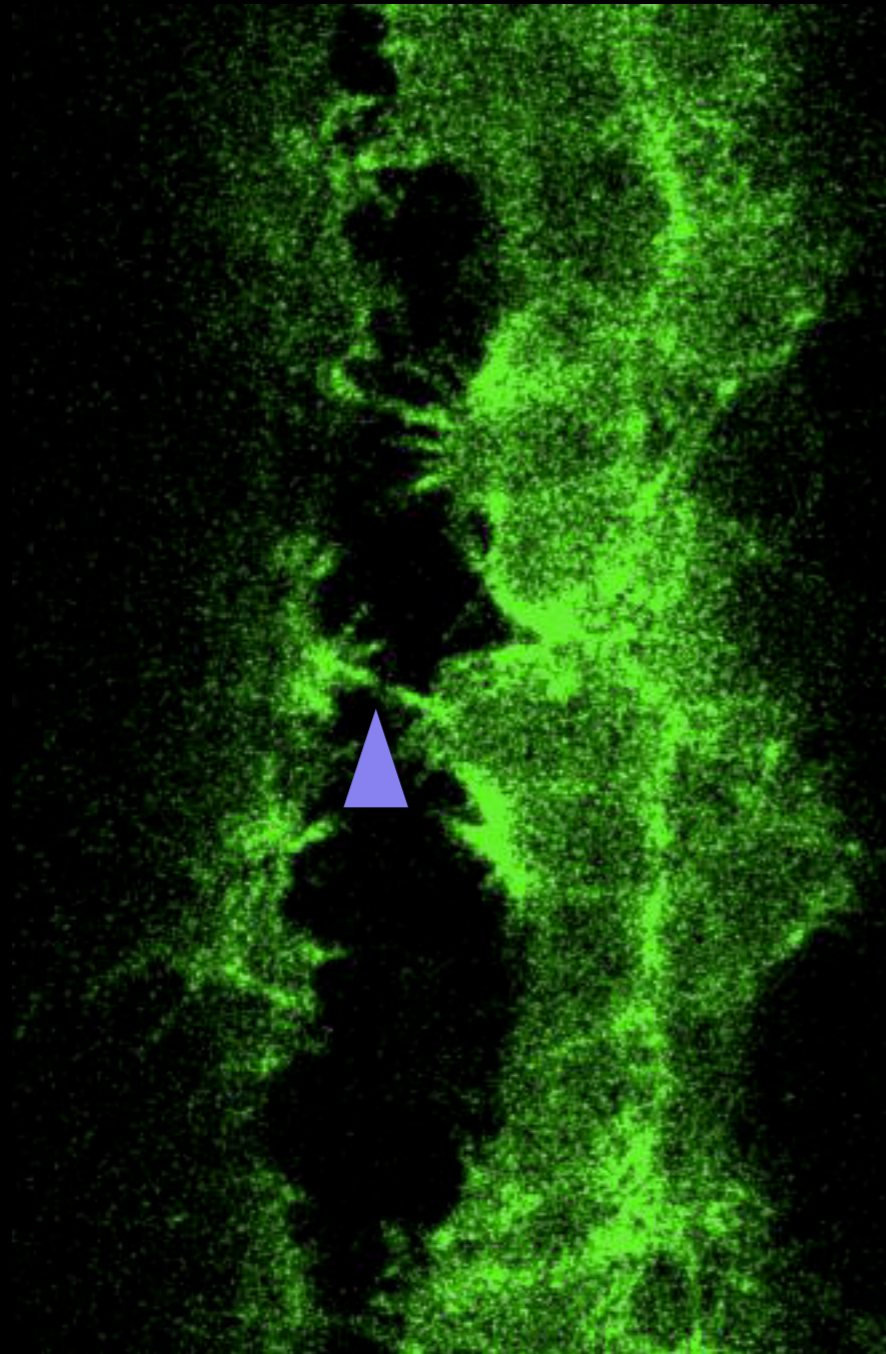
Strong force between filopodia in wildtype hearts

Hand>Moe::GFP



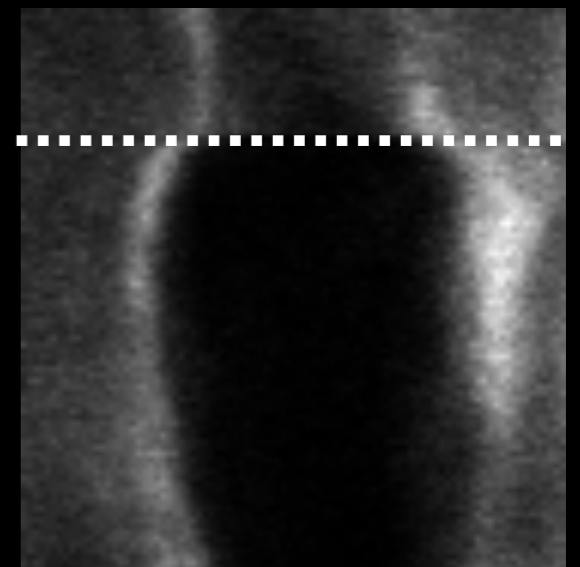
Force between filopodia reduced in Zip knockdown

Hand>Moe::GFP; Zip-RNAi



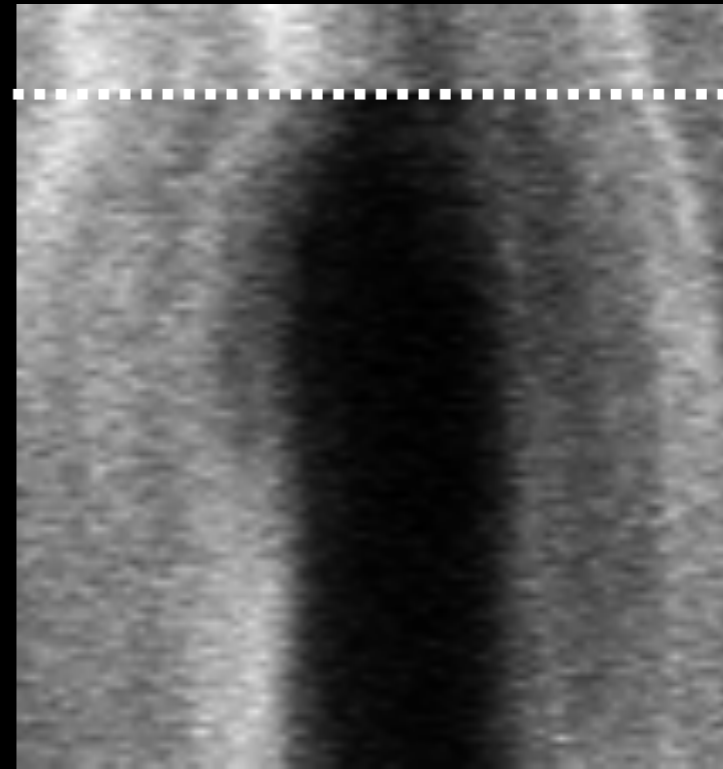
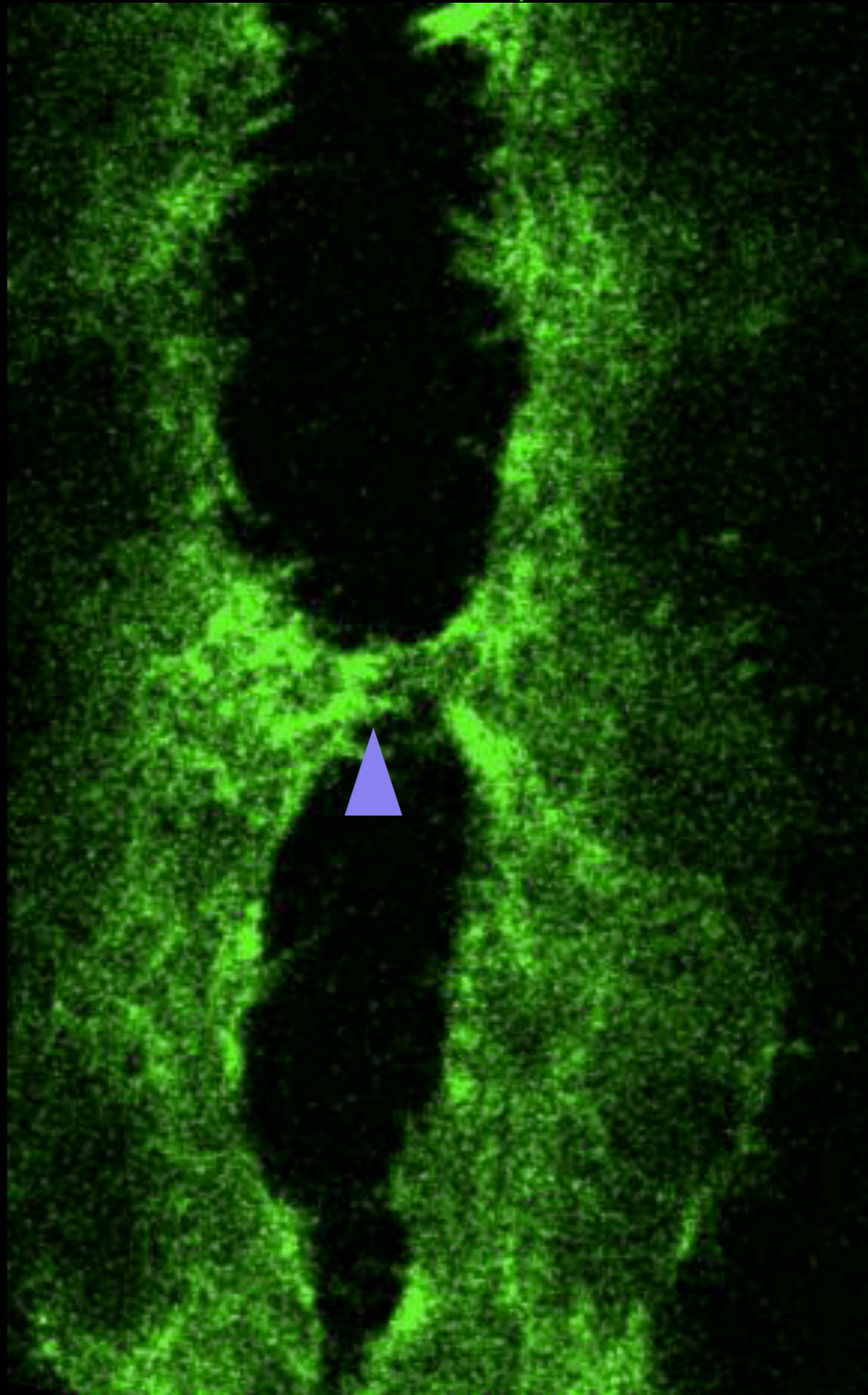
Time

wildtype



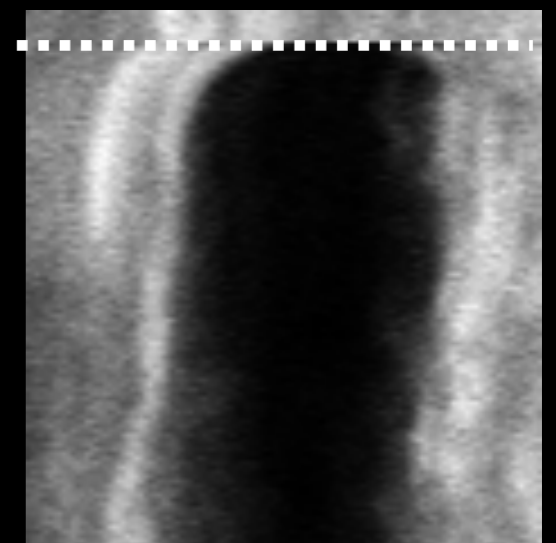
Force between filopodia reduced in Zip knockdown

Hand>Moe::GFP; Zip-RNAi

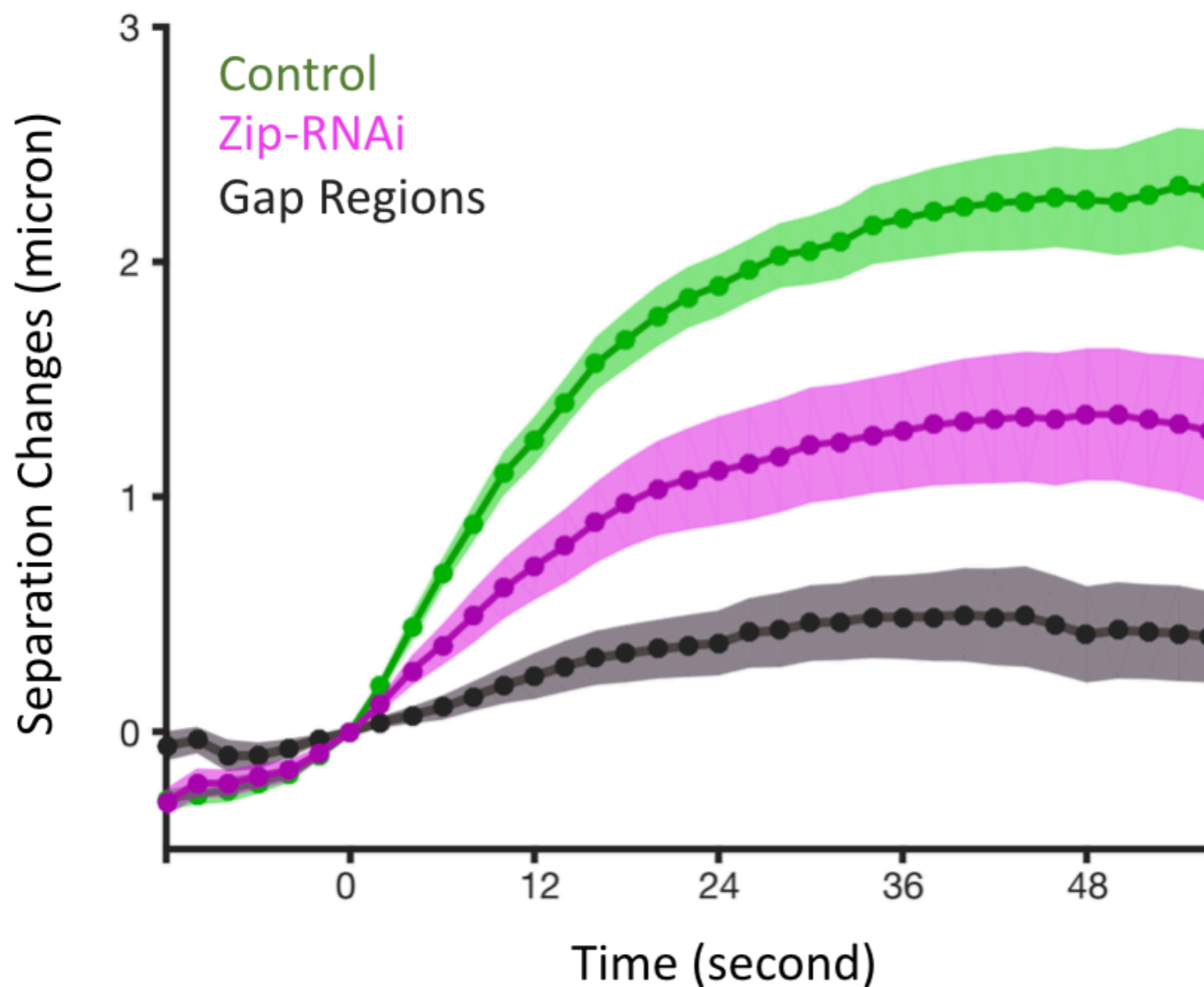


Time

wildtype



Force between filopodia reduced in Zip knockdown



Instantaneous Recoil Velocity

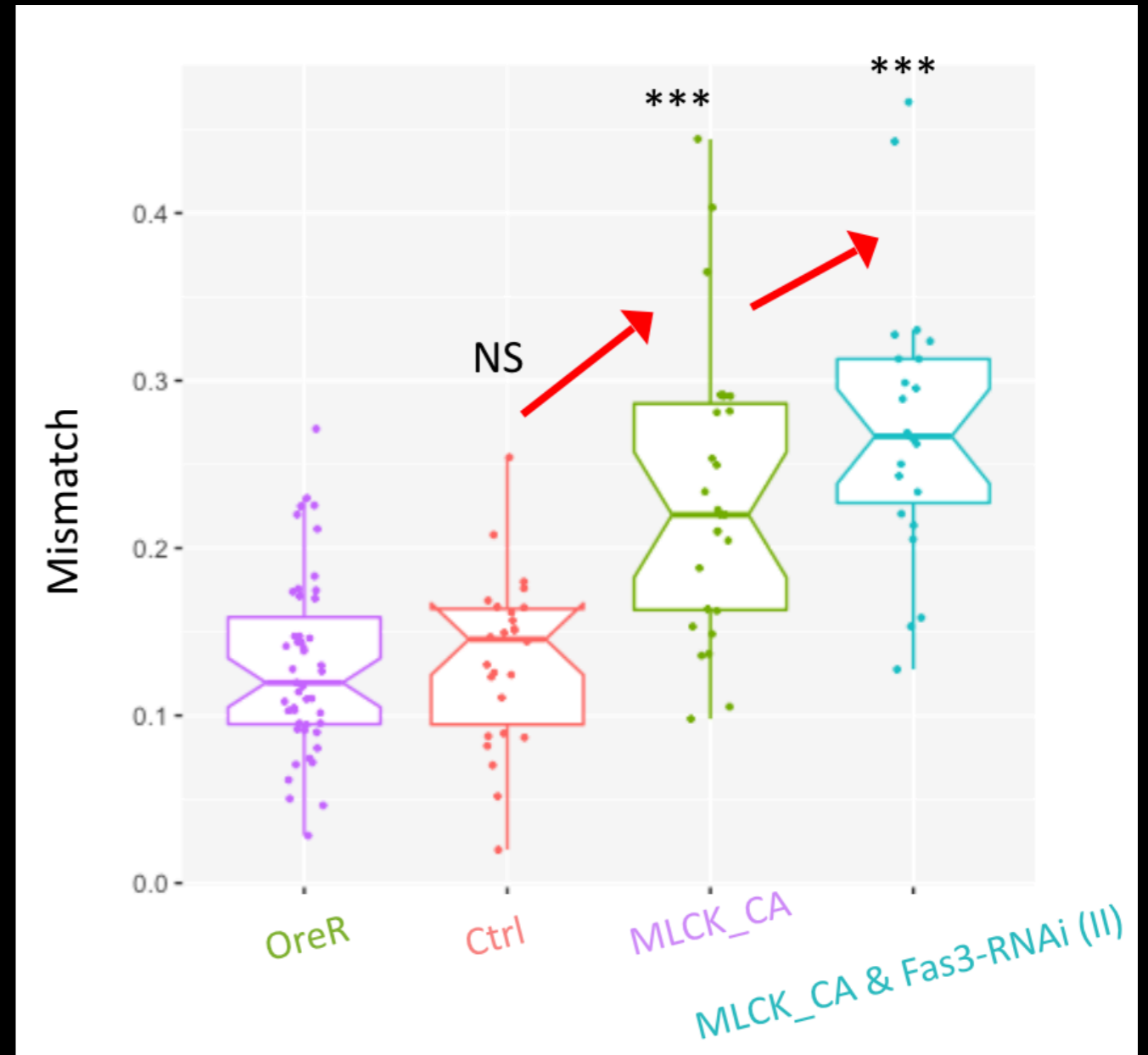
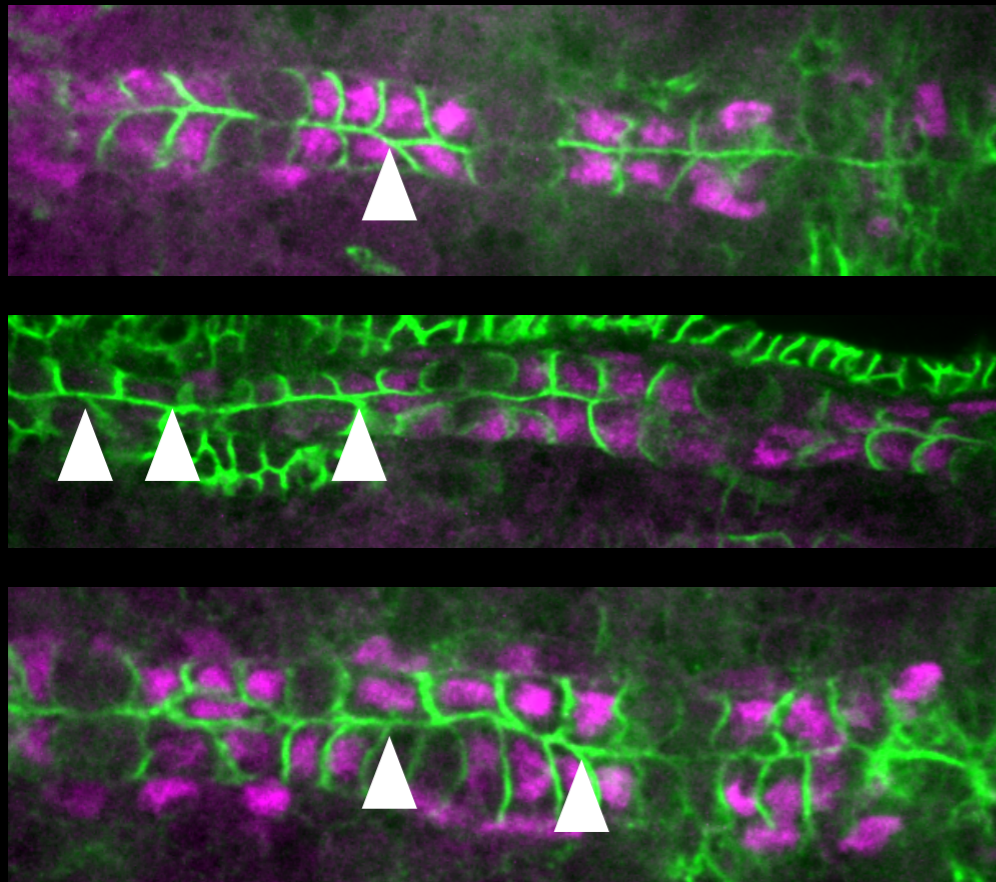
$$RV_{\text{Filopodia}} = 0.15 \pm 0.04 \mu\text{m/s}$$

$$RV_{\text{Zip-RNAi}} = 0.10 \pm 0.07 \mu\text{m/s}$$

$$RV_{\text{Gap}} = 0.02 \pm 0.03 \mu\text{m/s}$$

Myosin II over-activation also reduces cell matching

MLCK-CT



Suggests levels of Myosin are optimised to enable precise cell matching

Mechanical testing of cellular interactions

Myosin-II waves

Cdc42

Filopodia adhesions

Cdc42

Weak adhesion

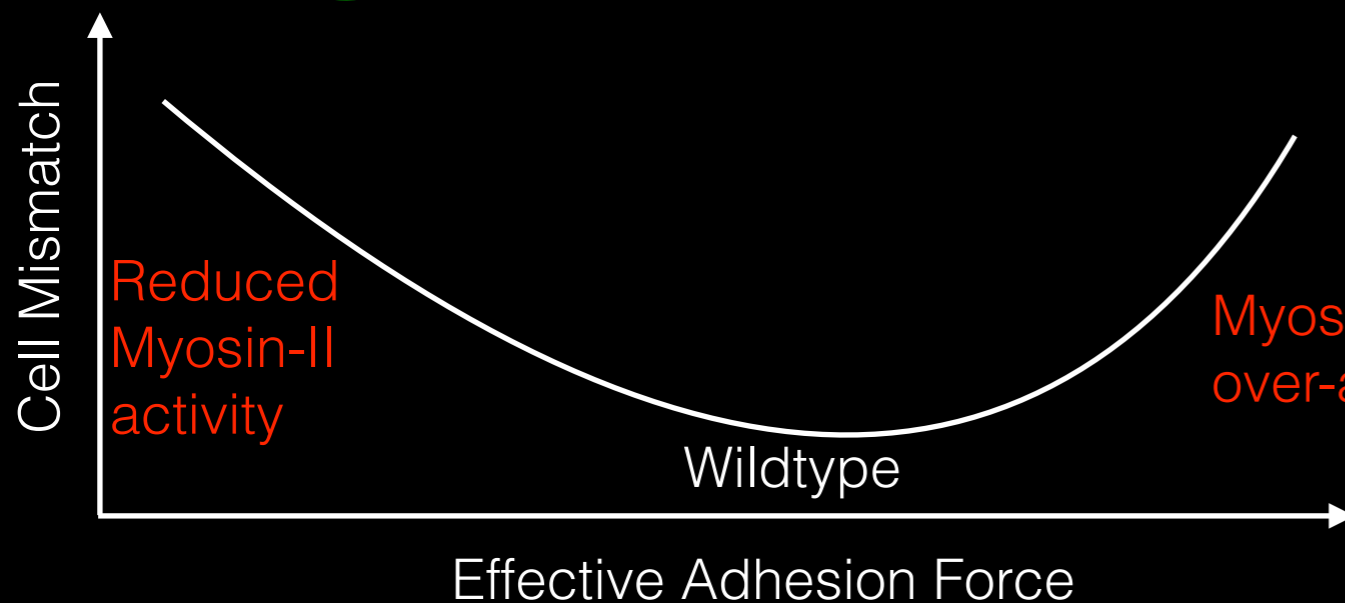
- Broken contacts
- Retracted filopodia

Strong adhesion

- *Some* contacts stabilise
- Drives precise cells matching

Extra strong adhesion

- Contacts *too* stable
- Unable to correct for mistakes in matching



Acknowledgements



Collaborators

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Jose Munoz (Spain)
Ivo Telley (Portugal)
Yusuke Toyama (MBI)

Current lab

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Tricia Loo	Prabhat Tiwari
Mario Mendieta	Vaishali Yadav
Sunandan Dhar	Veena Venugopal

Former lab members

Chris Amourda	Anand Singh
Jeronica Chong	Jianmin Yin
Anqi Huang	Sham Tlili

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