

# ***Decision making in the assembly of sensory-motor circuits***



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**Morphodynamics  
KITP, Santa Barbara  
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**Kavli Institute for Theoretical Physics,  
UC Santa Barbara**

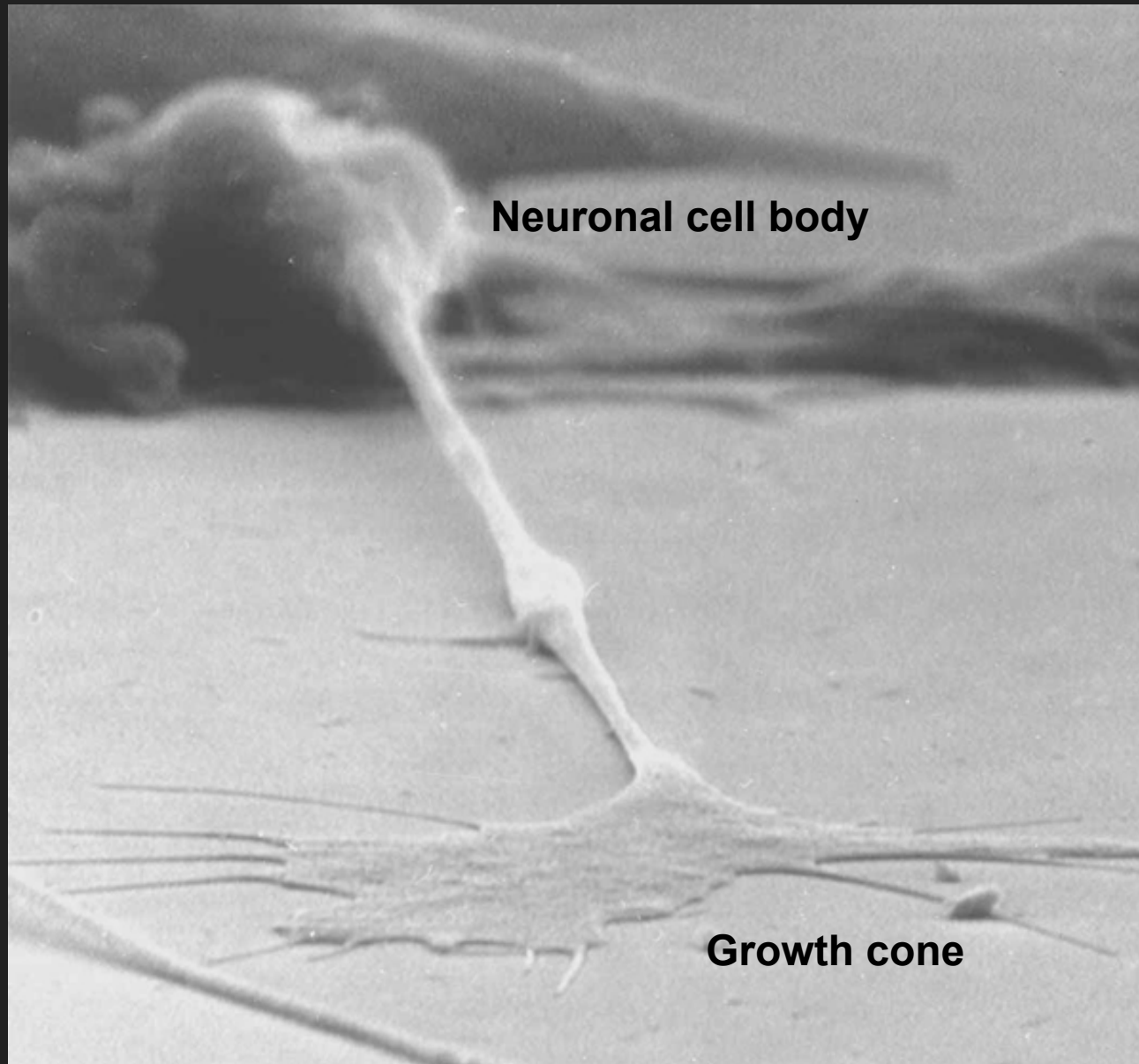


**Fred Wolf  
Sara Solla  
Gonzalo de Polavieja  
Boris Shraiman**



**Princeton  
Michael Berry**

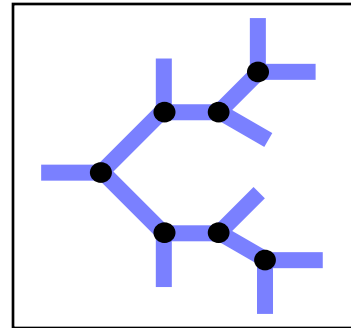
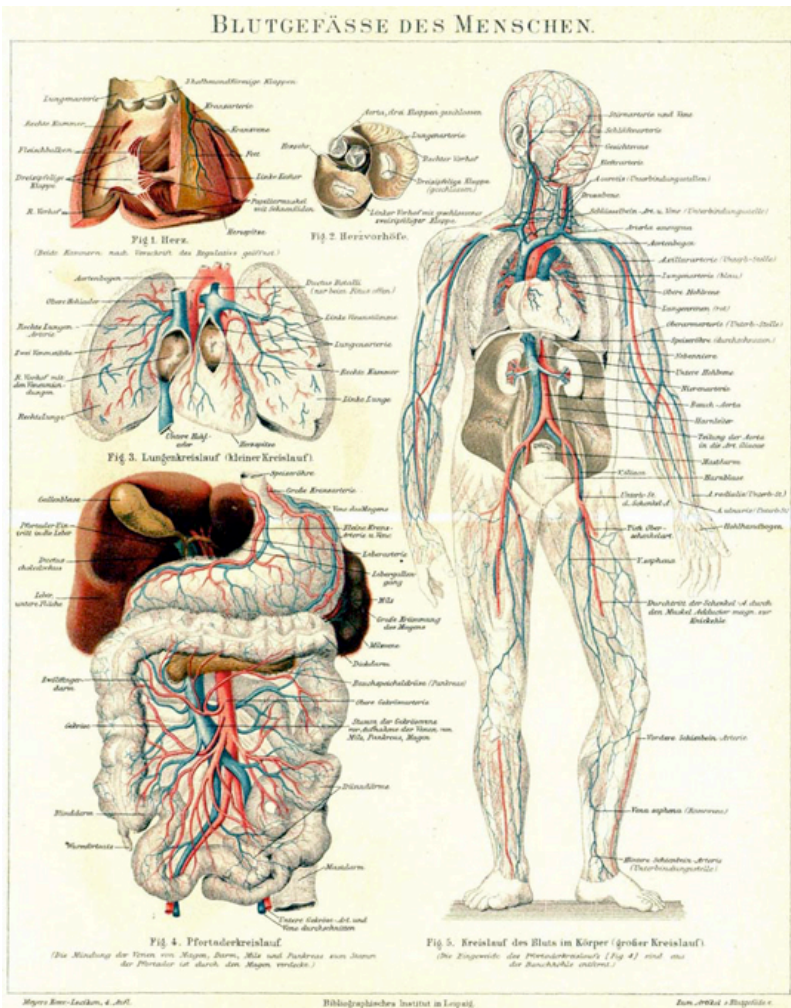
# Neurons send growth cones to faraway targets



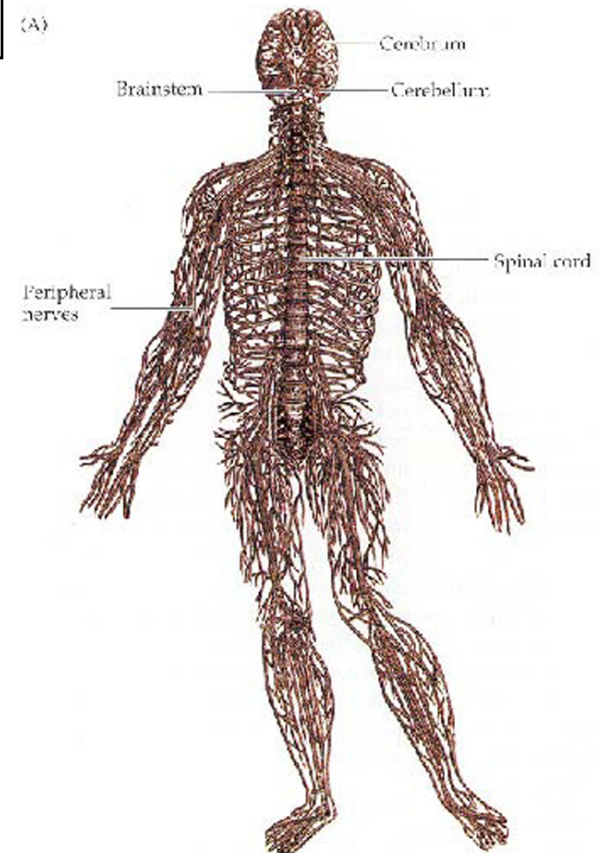


# Neural circuit construction is a *serial decision* process

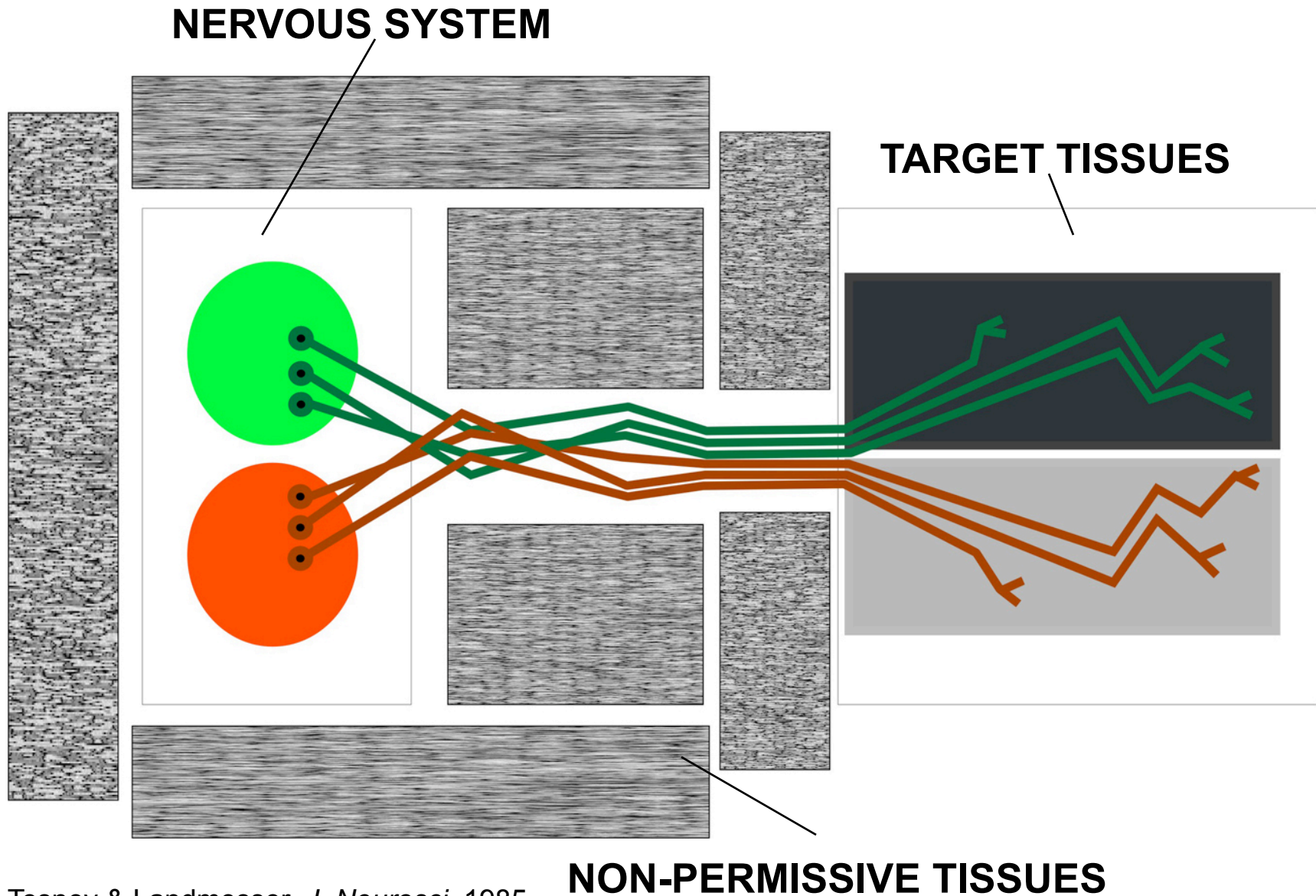
## Vascular system



## Nervous system



# Topographic mapping

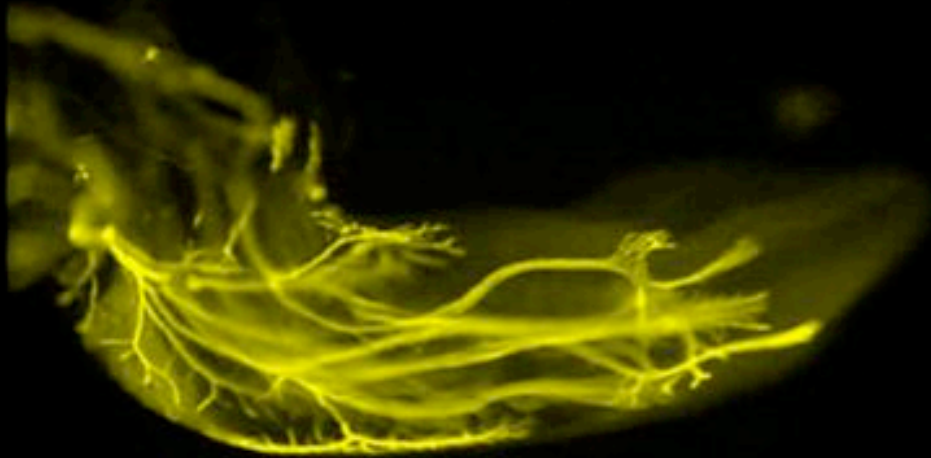
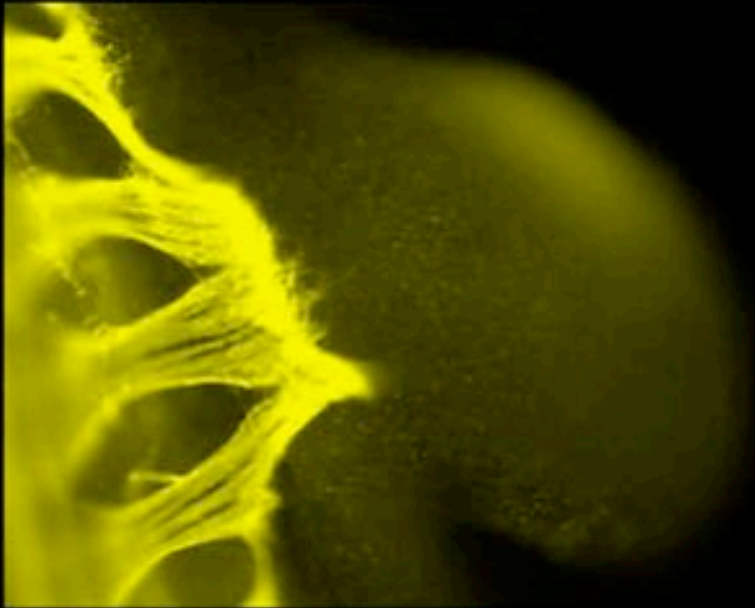


Tosney & Landmesser, *J. Neurosci.* 1985  
Jessell, *Nat. Rev. Genet.* 2000

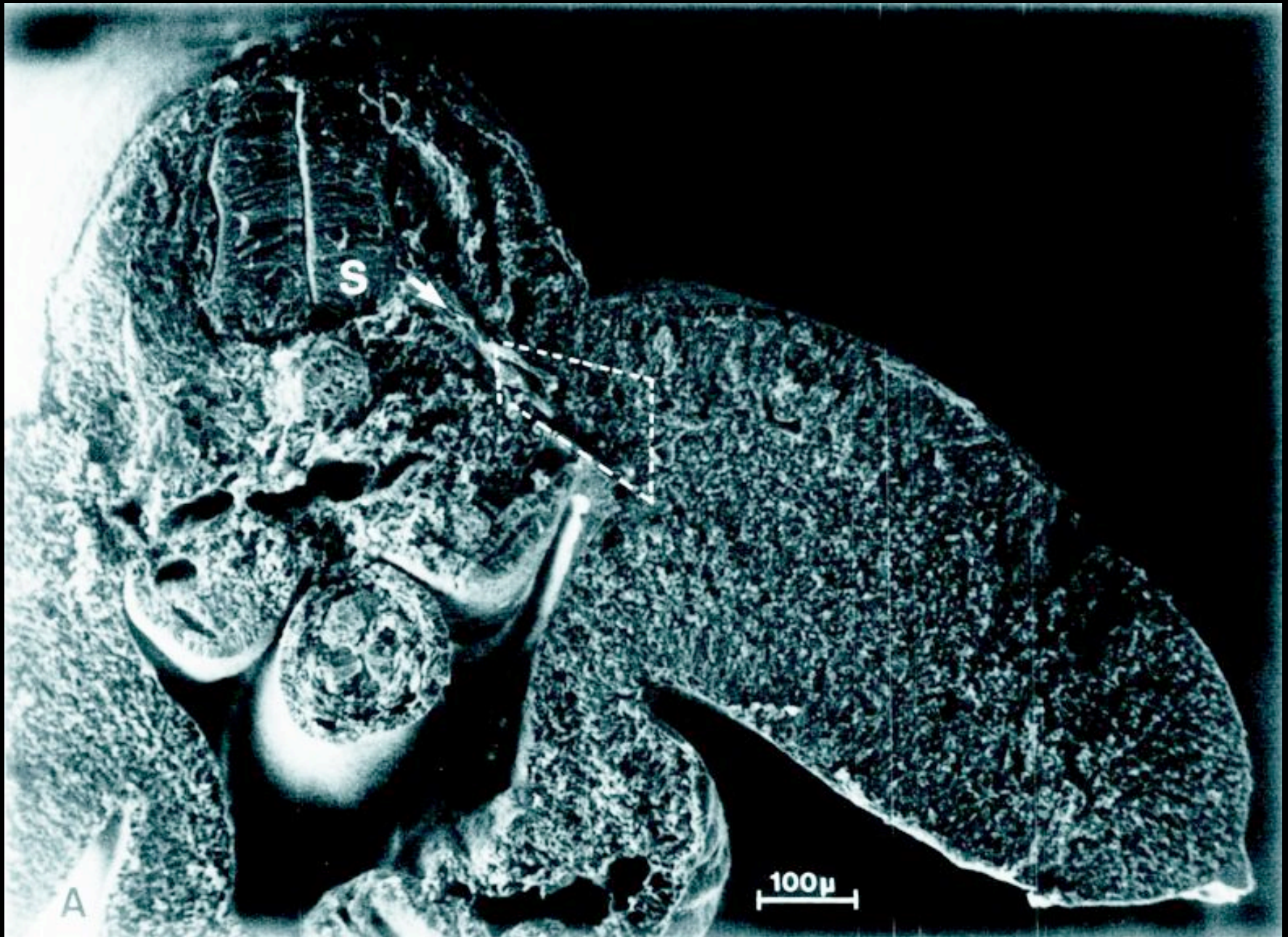
**NON-PERMISSIVE TISSUES**



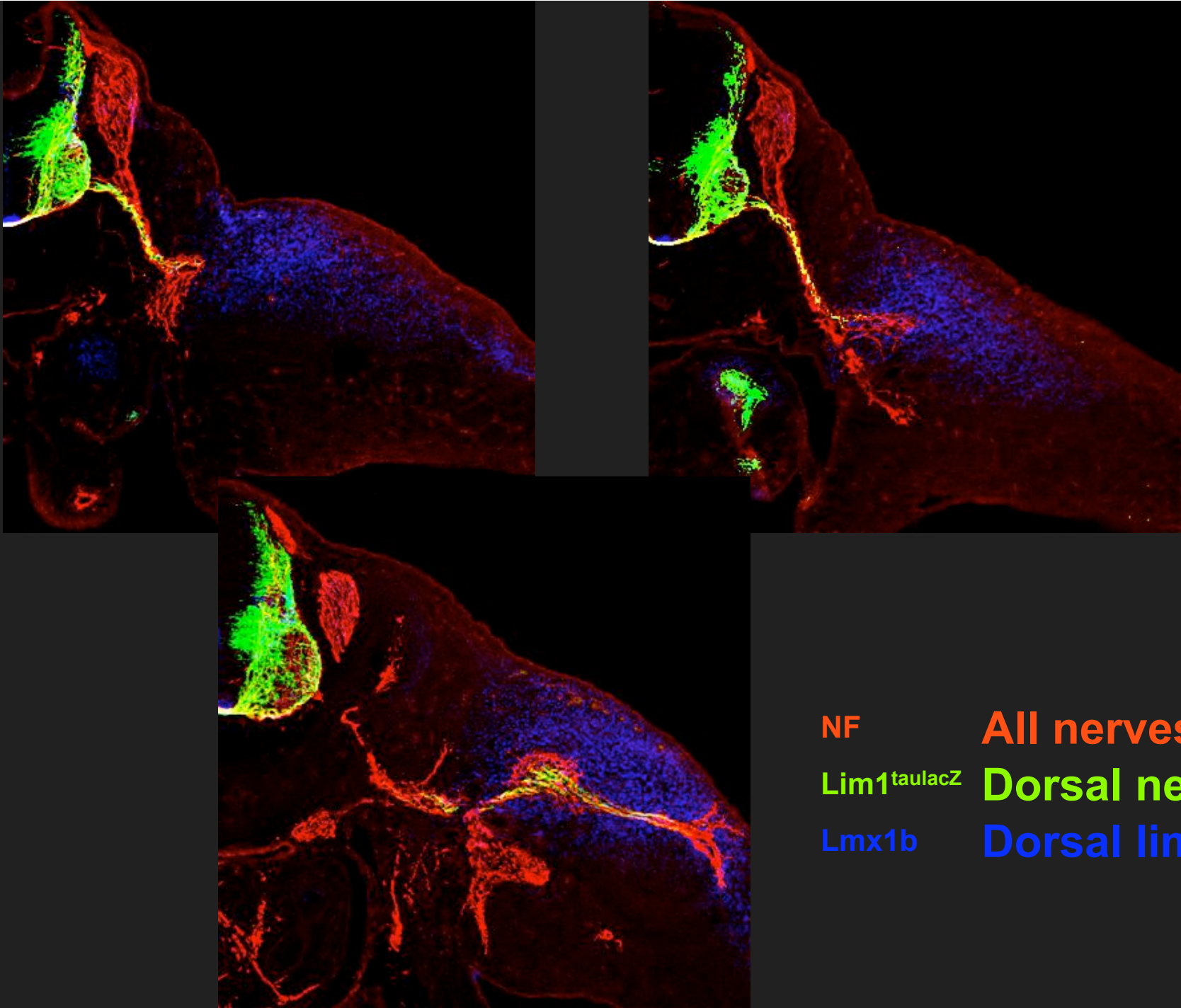








Tosney & Landmesser, *J. Neurosci.* 1985



NF

All nerves

Lim1<sup>taulacZ</sup>

Dorsal nerves

Lmx1b

Dorsal limb

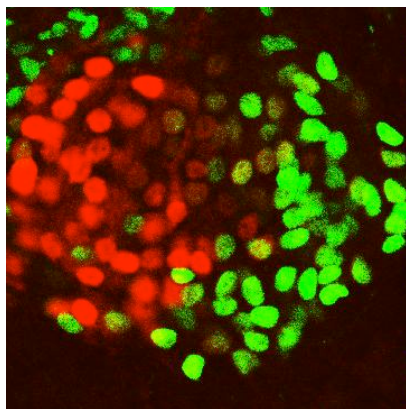


# Motor axon decision: towards *flexors* or *extensors*?

Motor column	LIM HD code
LMCI	: Lim1
LMCm	: Isl1

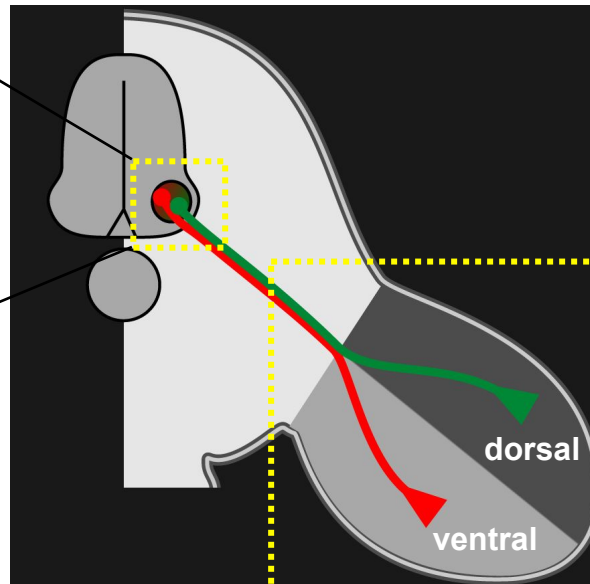
Limb mesenchyme	Lmx1b
dorsal	+
ventral	-

Isl1 Lim1



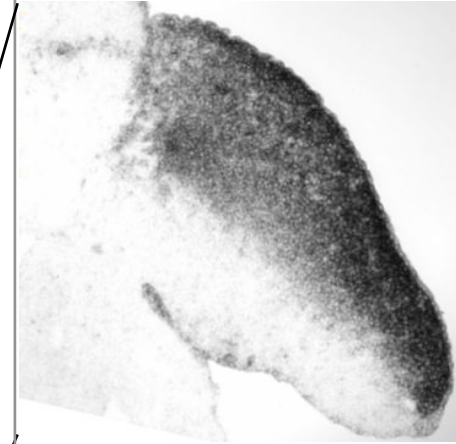
LMCm LMCi

Tsuchida et al., *Cell* 1994



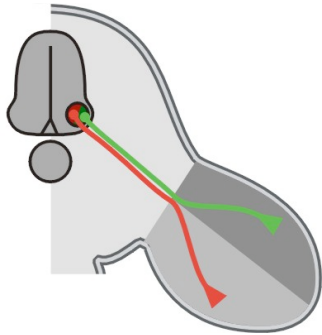
Landmesser, *J. Physiol.* 1978

Lmx1b



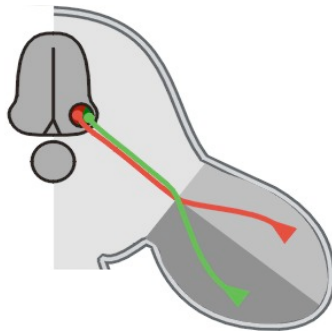
Riddle et al., *Cell* 1995

# Pathway selection is local and active



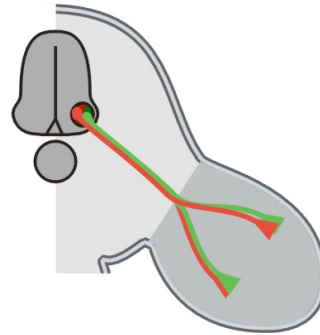
Wild type

Landmesser,  
*J. Physiol.* 1978  
Tsuchida et al.,  
*Cell* 1994



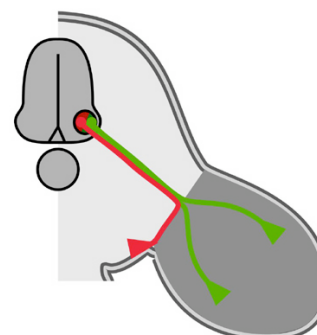
Reversal

Whitelaw &  
Hollyday,  
*J. Neurosci.* 1983



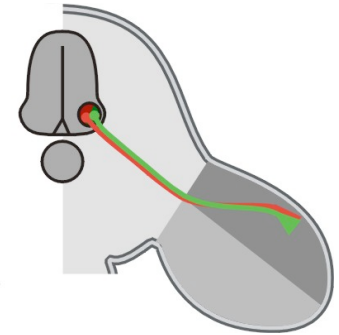
Bi-ventral  
*Lmx1b*<sup>-/-</sup>

Kania et al.,  
*Cell* 2000



Bi-dorsal  
*Bmpr1a*<sup>-/-</sup>

Luria et al.,  
*Neural Dev.* 2007



*EphA4* +/+

Eberhart et al.,  
*Dev. Biol.* 2002  
Kania & Jessell,  
*Neuron* 2003

LMC(I)	Dorsal Limb
LMC(m)	Ventral Limb

+	Ventral
+	Dorsal

+	-
+	+

+	+
+	-

+	+
+++	+

Reversed trajectories

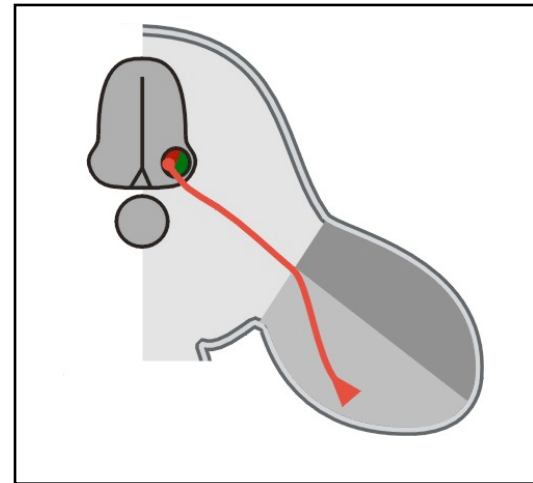
LMCm & LMC(I) randomized trajectories

LMC(I) randomized trajectories, LMCm misrouted

LMCm axons misrouted dorsally

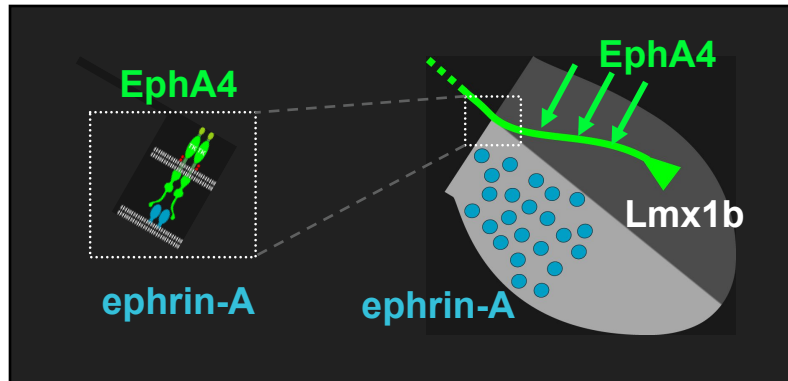


***I. MOLECULAR LOGIC - What are the effector molecules that control **LMCm** trajectories?***



***II. Quantitative models, experimental predictions and tests***

# I. What are the effector molecules that control **LMCm** trajectories?

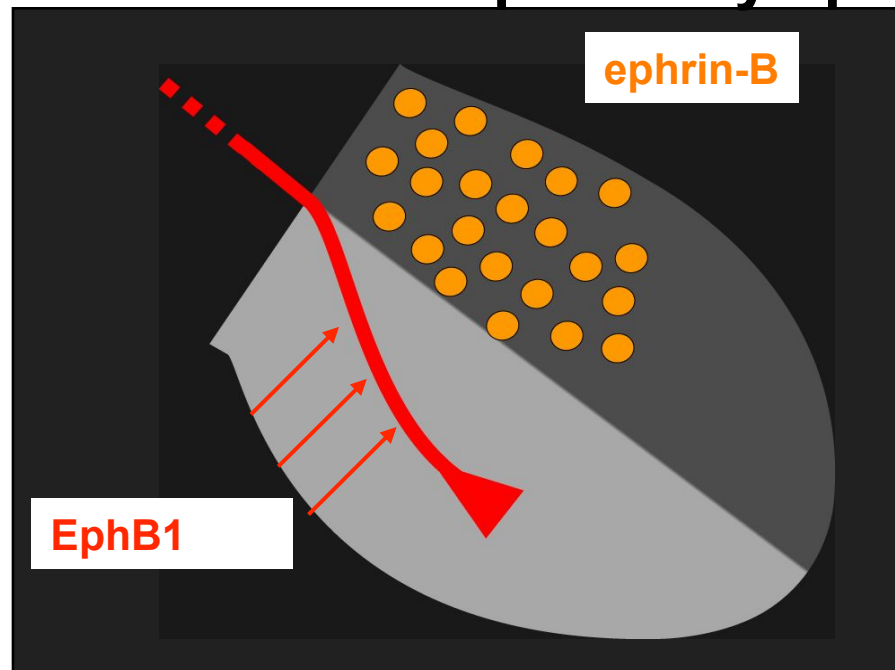


ephrin-A/EphA, GDNF/c-ret, Sema/Npn signaling influence **LMCI** trajectories

- Kania et al., 2000
- Helmbacher et al, 2000
- Eberhart et al, 2002
- Kania and Jessell, 2003
- Huber et al., 2005
- Kramer et al., 2006

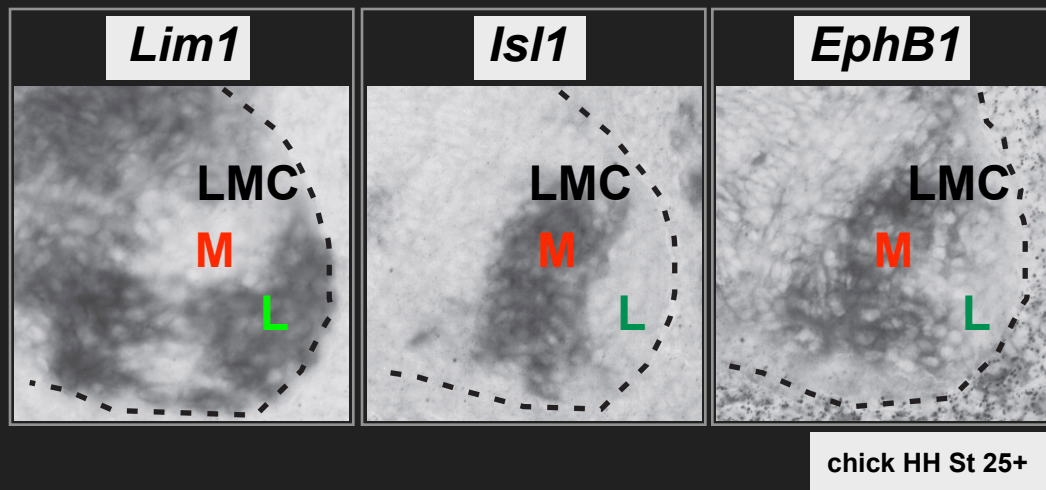
## Hypothesis

**EphB+** **LMCm** axons are repelled by ephrin-B+ dorsal limb

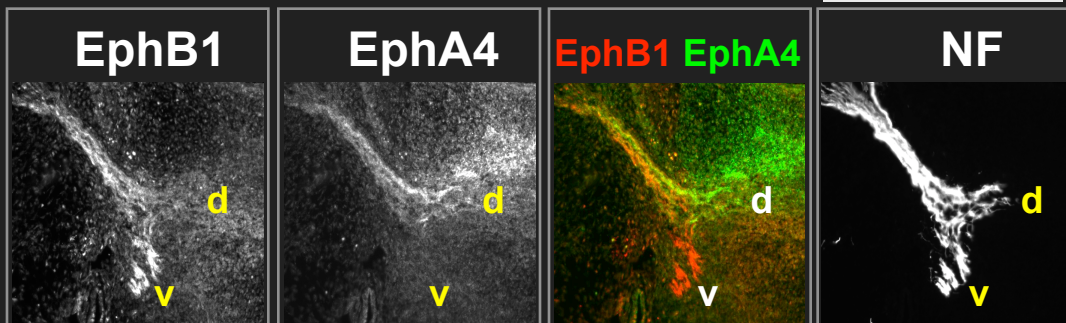
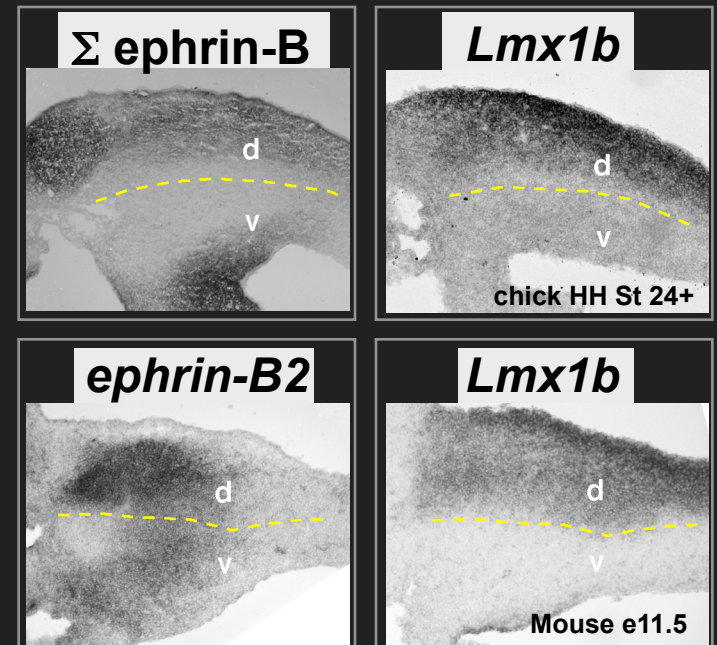


# Distribution of EphBs receptors and ephrin-Bs ligands

## RECEPTOR in NEURONS

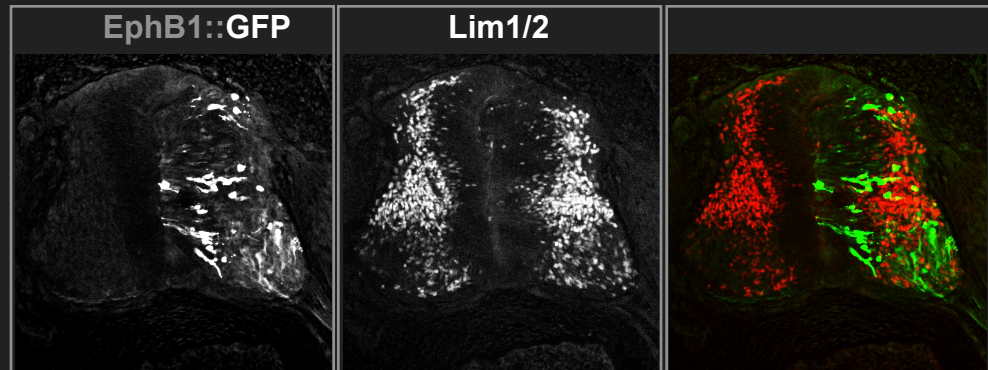
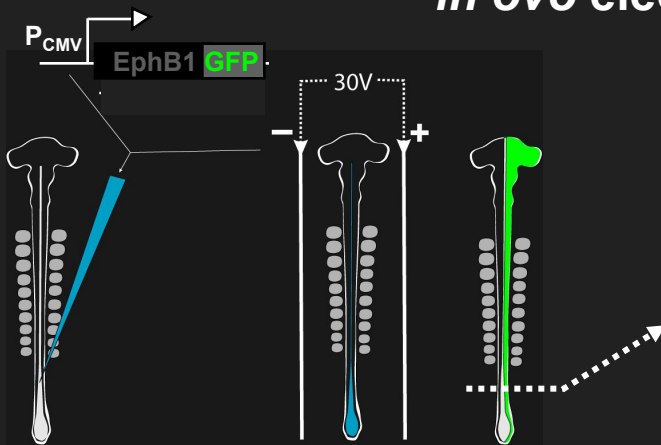


## LIGAND in NON-TARGET TISSUE



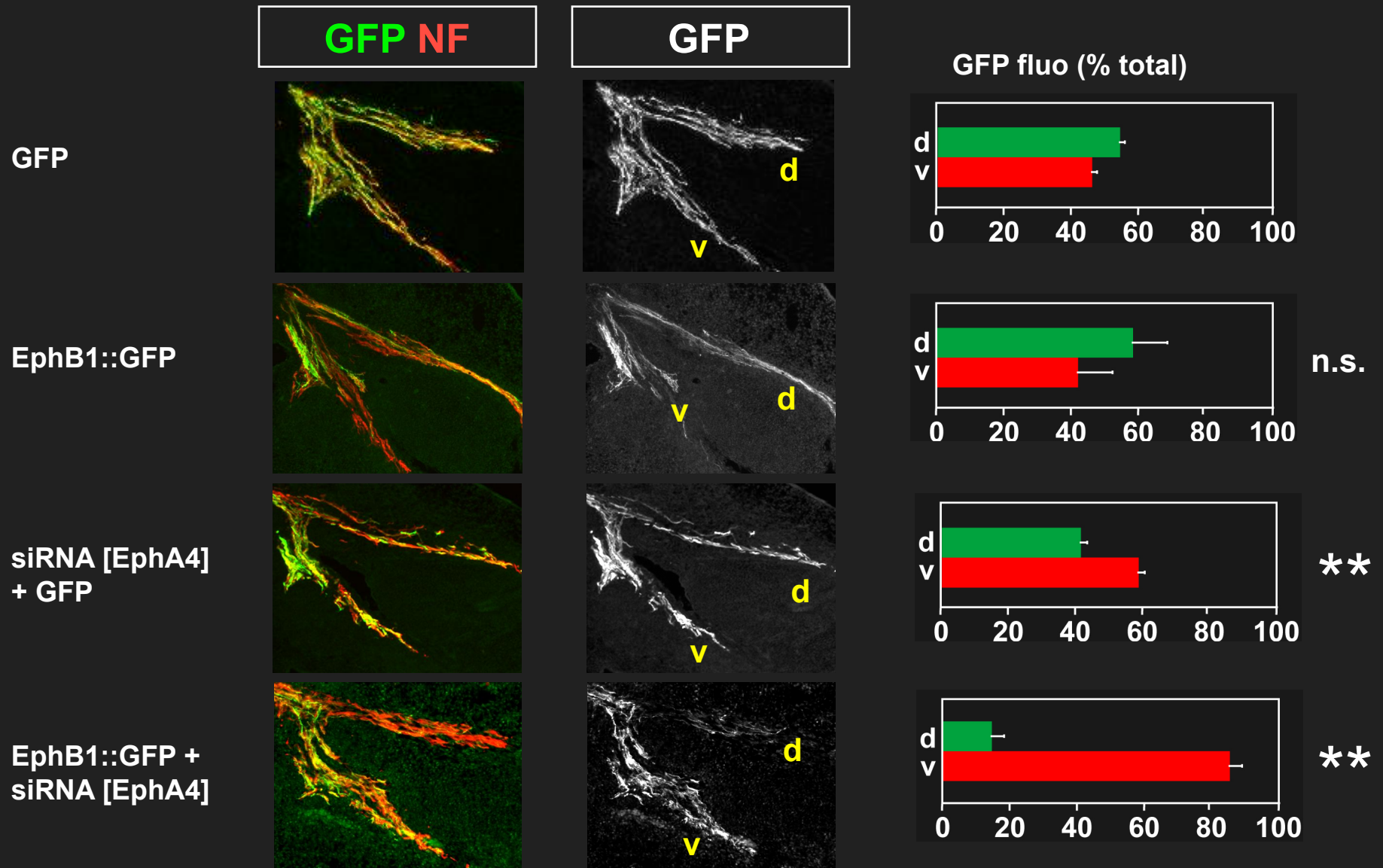
# Is EphB1 sufficient to guide LMC axons to the ventral limb?

*in ovo* electroporation of *EphB1::GFP*

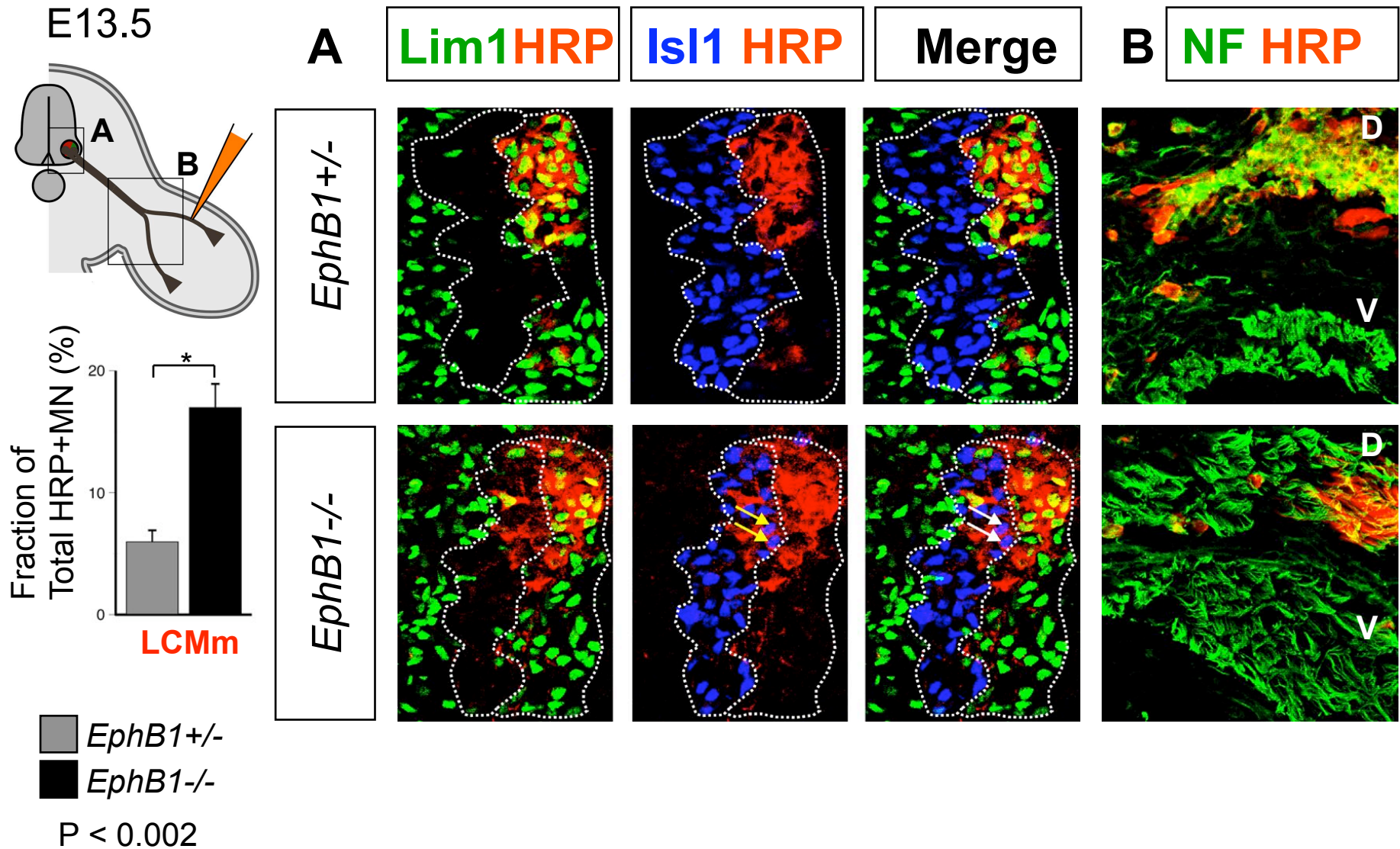




# EphB1 can *redirect* LMC axons to the ventral limb



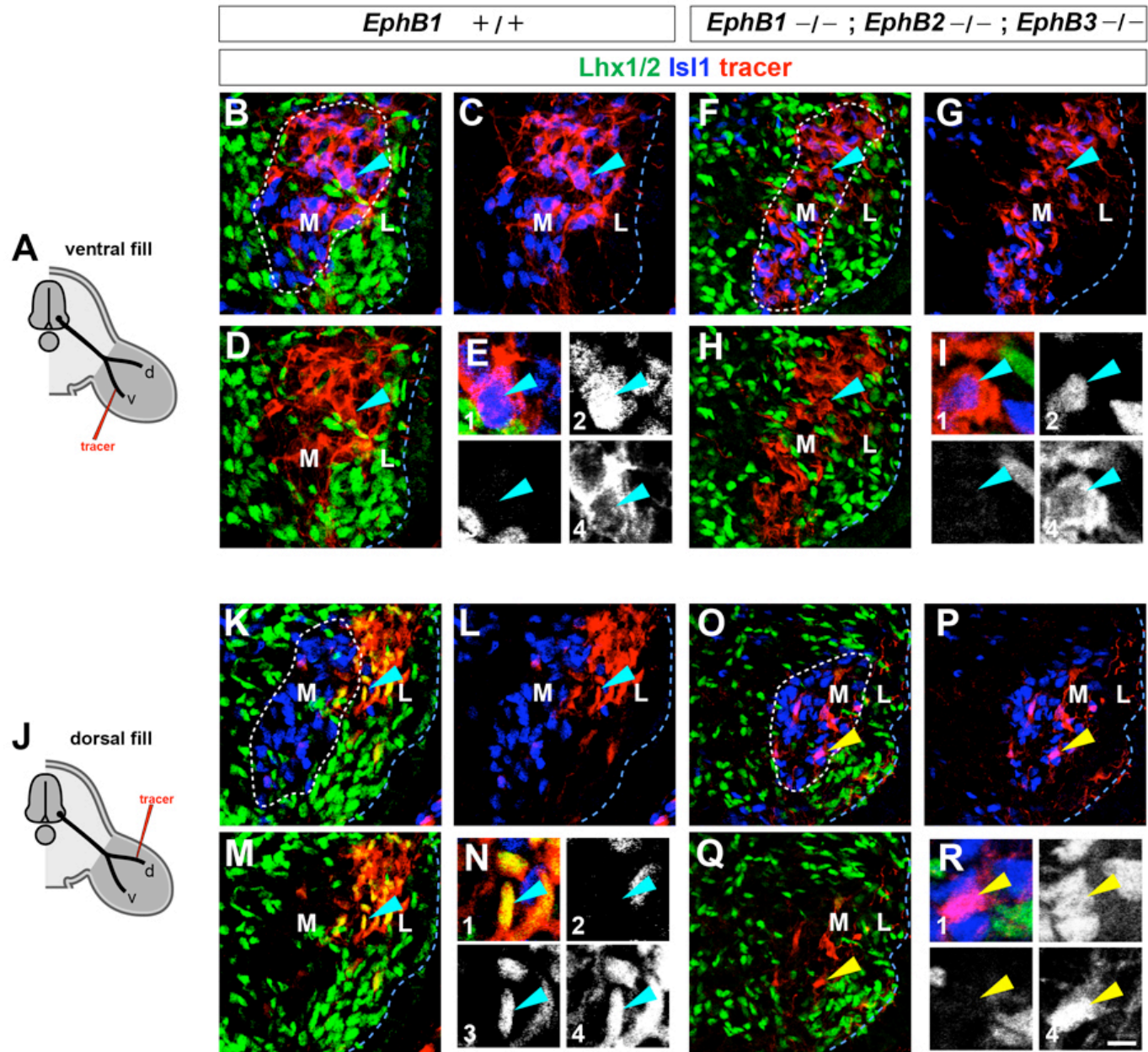
# *EphB1* is necessary for LMCm ventral targeting





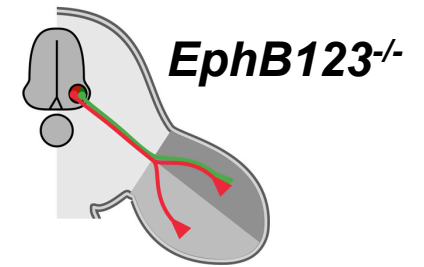
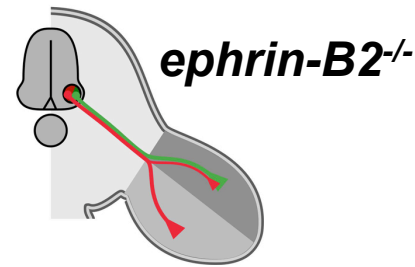
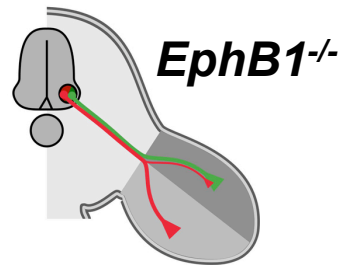
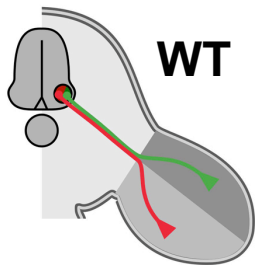


# *EphBs* are necessary for **LMCm** ventral targeting

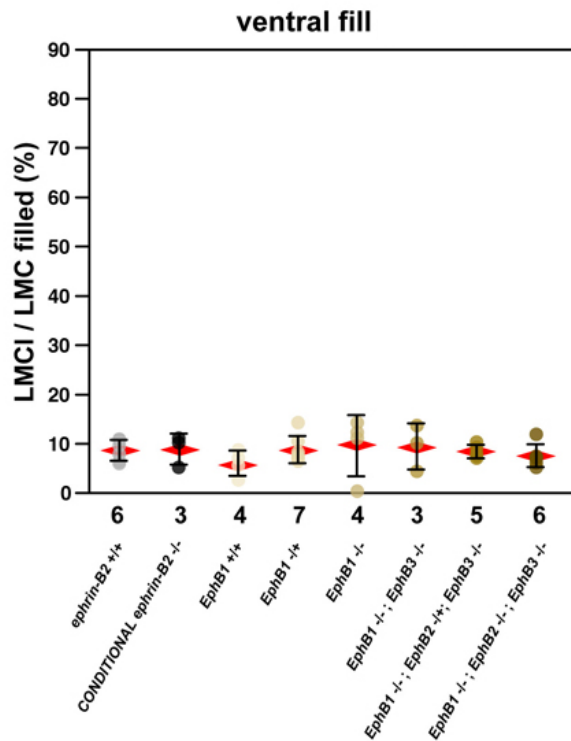




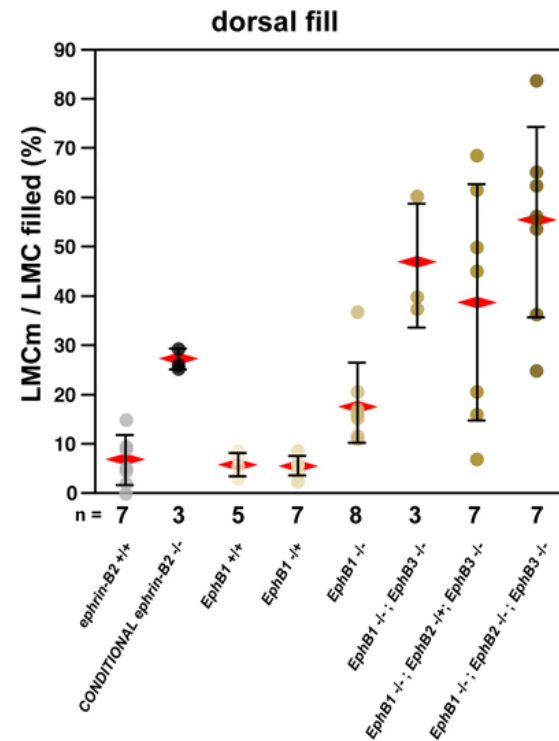
# LMCm projections are *randomized* AND *more variable* in compound mutants



## mistargeted LMCI



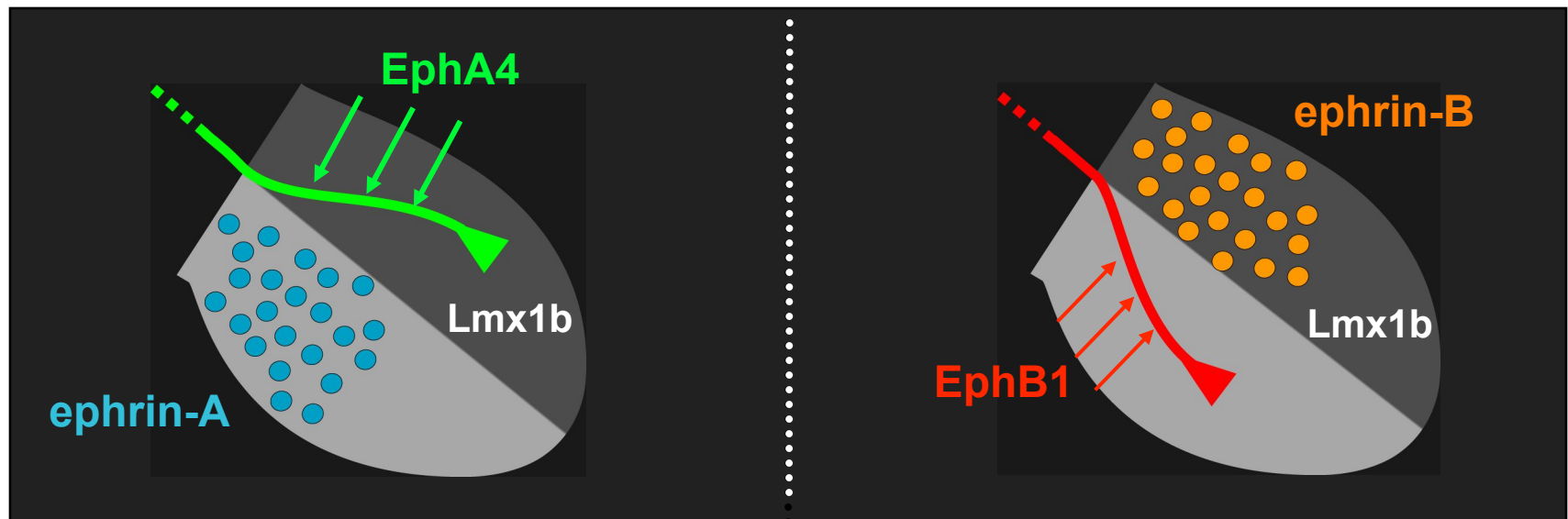
## mistargeted LMCm



# Mirror symmetry ephrin-Eph signaling controls motor axon trajectories to the limb

	Ligands	Receptors
<b>EXPRESSION</b>		
<b>Dorsal limb</b>	<b>b</b>	<b>A</b>
<b>Ventral limb</b>	<b>a</b>	<b>B</b>

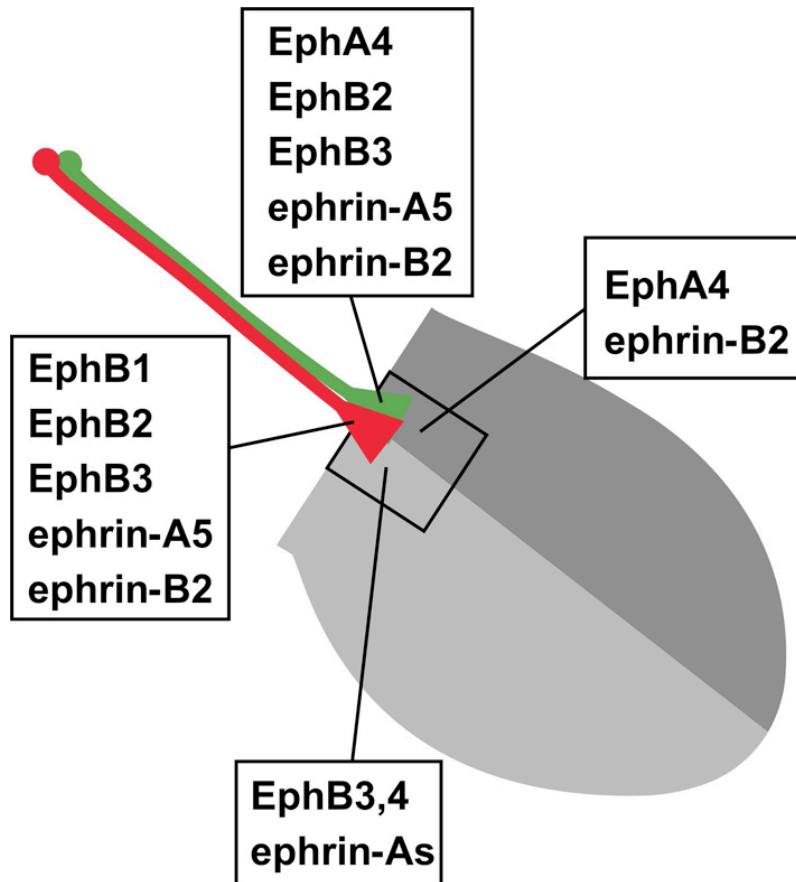
## FUNCTIONAL



LMCI → EphA4 → Dorsal limb    LMCm → EphB1 → Ventral limb

## II. Quantitative models, experimental predictions and tests

### Axons integrate multiple cues at limb entry



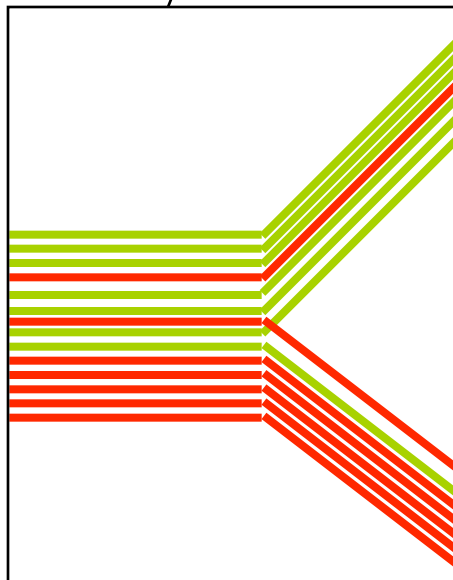
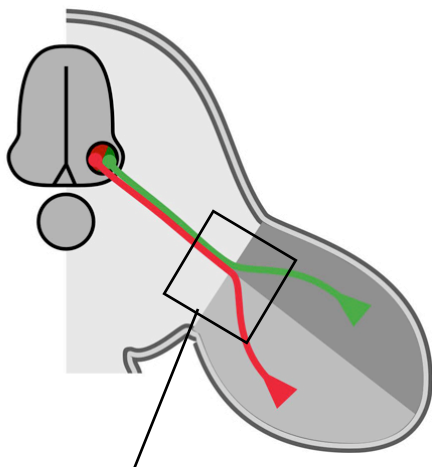
Axon	Interaction	Limb	Symbol
EphB1	>—<	ephrin-B2	$B_1b_2$
EphB2	>—<	ephrin-B2	$B_2b_2$
EphB3	>—<	ephrin-B2	$B_3b_2$
EphB2	>—<	ephrin-As	$B_2a$
EphA4	>—<	ephrin-As	$A_4a$
EphA4	>—<	ephrin-B2	$A_4b_2$
ephrin-B2	<—>	EphB3,4	$b_2B_{34}$
ephrin-A5	<—>	EphA4	$a_5A_4$
ephrin-B2	<—>	EphA4	$b_2A_4$

$$\Sigma \text{lateral axons} = + A_4a + B_2a + a_5A_4 + b_2A_4 - A_4b_2 - B_2b_2 - B_3b_2 - b_2B_{34} > 0$$

$$\Sigma \text{medial axons} = - B_1b_2 - B_2b_2 - B_3b_2 - b_2B_{34} + B_2a + a_5A_4 + b_2A_4 < 0$$

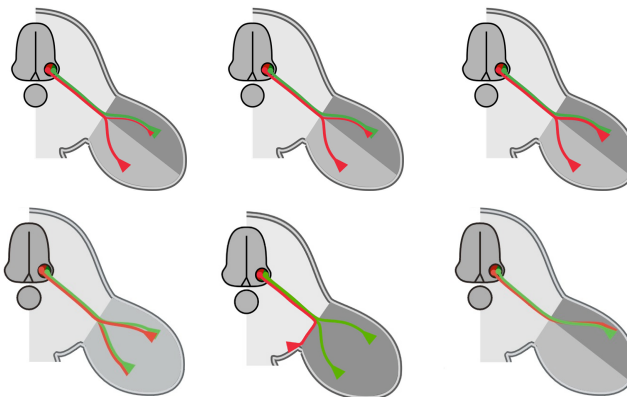
# Neural connectivity is *almost* exact, but defects are **NOT** corrected

## Wild type



~95% - 5%

## Various guidance gene mutants



d'Avella & Bizzi,  
*PNAS* 2005

## Shared and specific muscle synergies in natural motor behaviors

Andrea d'Avella<sup>1\*</sup> and Emilio Bizzi<sup>1,3</sup>

<sup>1</sup>Department of Neuromotor Physiology, Istituto di Ricovero e Cura a Carattere Scientifico Fondazione Santa Lucia, 00179 Rome, Italy; <sup>2</sup>Department of Brain and Cognitive Sciences and McGovern Institute for Brain Research, Massachusetts Institute of Technology, Cambridge, MA 02139; and <sup>3</sup>European Brain Research Institute, 00143 Rome, Italy

***Lmx1b*<sup>-/-</sup>**

Kania et al., *Cell* 2000

***Bmpr1a*<sup>flox/-</sup>**

Luria et al.,  
*Neural Dev.* 2007

***EphA4*<sup>-/-</sup>**

Helmbacher et al.,  
*Development* 2000

***Sema3F*<sup>-/-</sup> , *Npn2*<sup>-/-</sup>**

Huber et al.,  
*Neuron* 2005

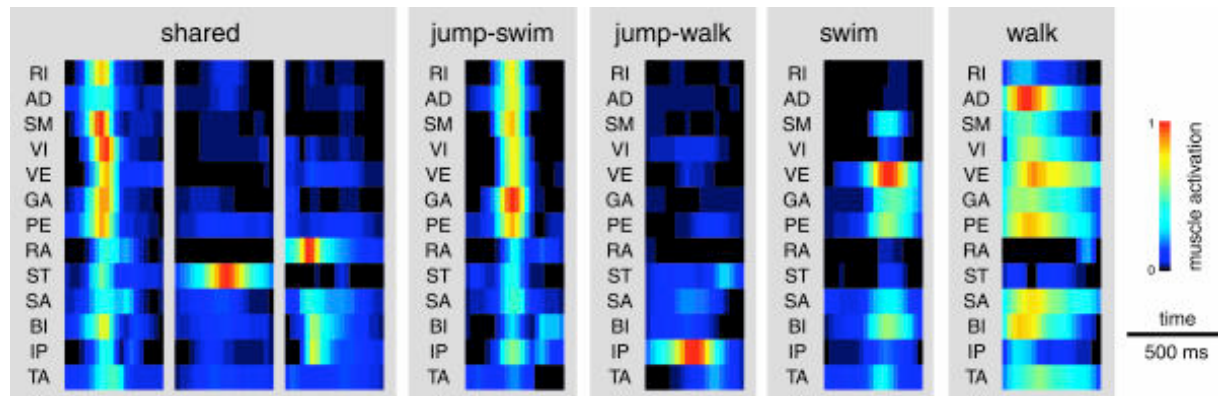
***EphB1*<sup>-/-</sup>  
in optic chiasma**

Williams et al.,  
*Neuron* 2003

***EphB1,3,13,123*<sup>-/-</sup>**

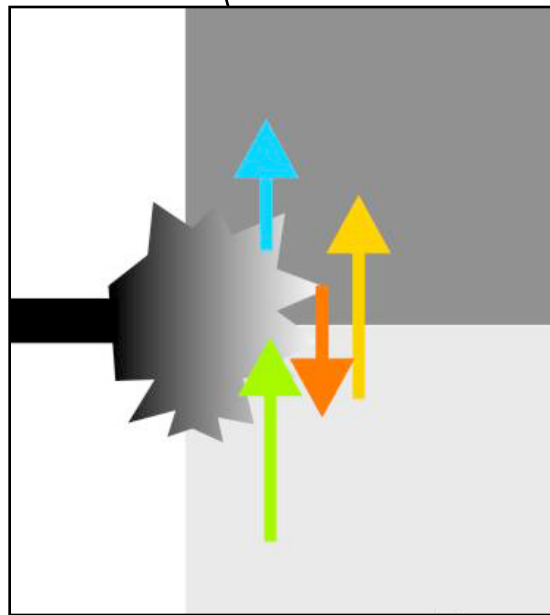
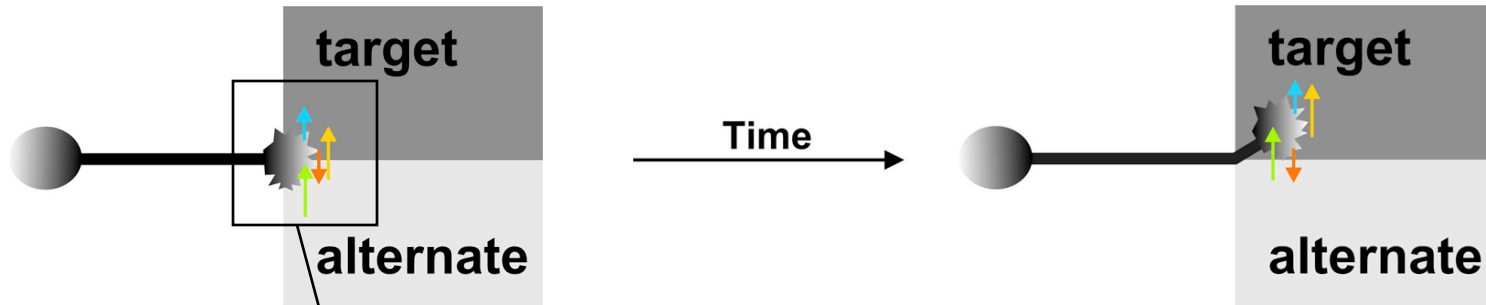
***ephrin-B2*<sup>-/-</sup>**

Luria et al.,  
*Neuron* 2008





# Axons integrate multiple cues at choice points



**A** repels axons from non-target tissue

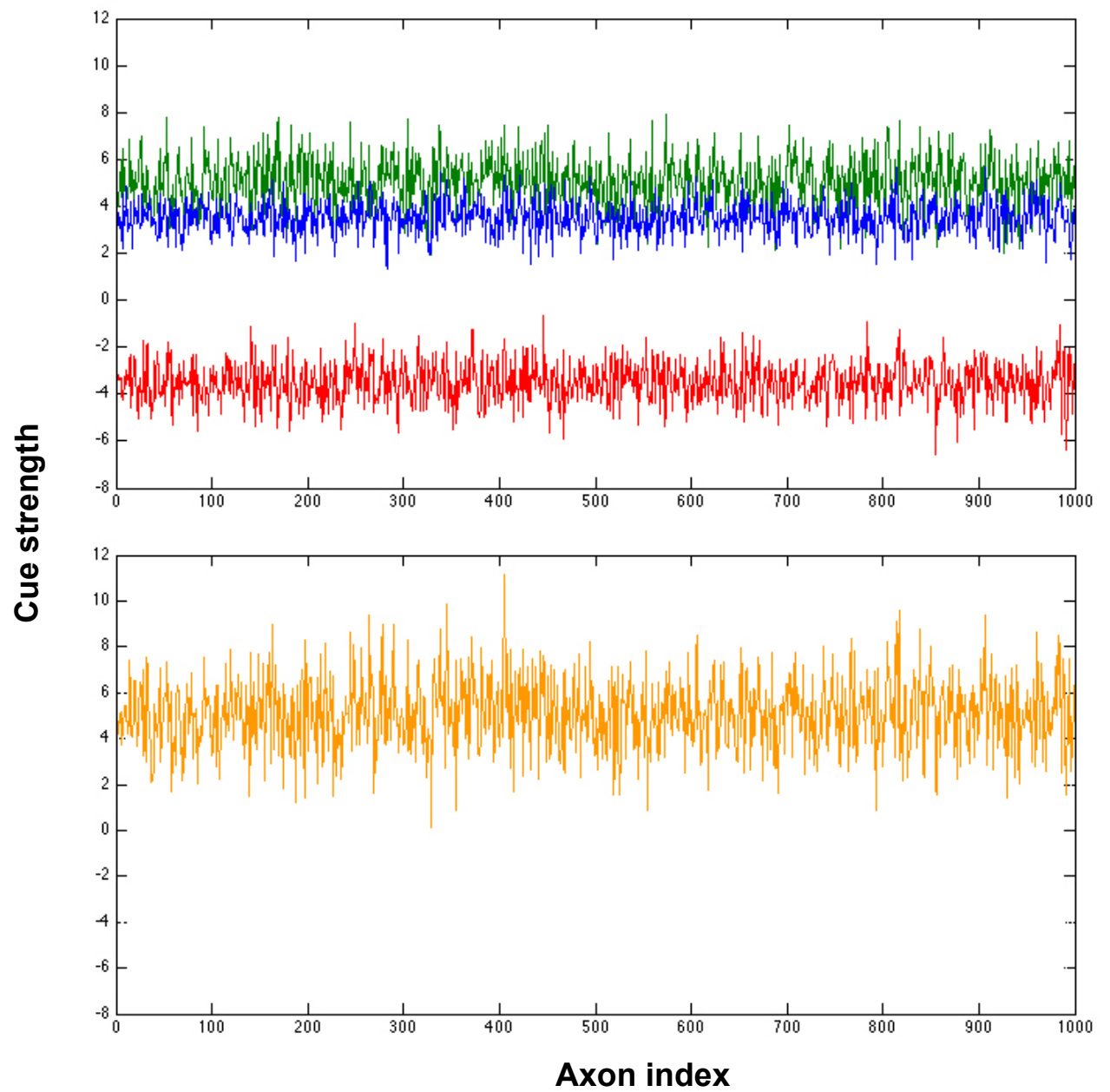
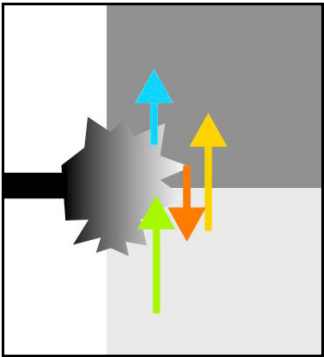
**B** attracts axons to target tissue

**C** repels axons from target tissue

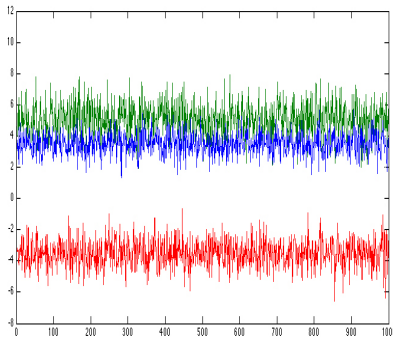
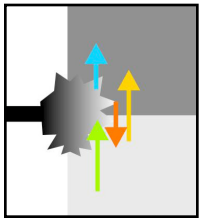
$$\Sigma = A + B + C$$

# Individual cues are noisy, the summed cue even more so

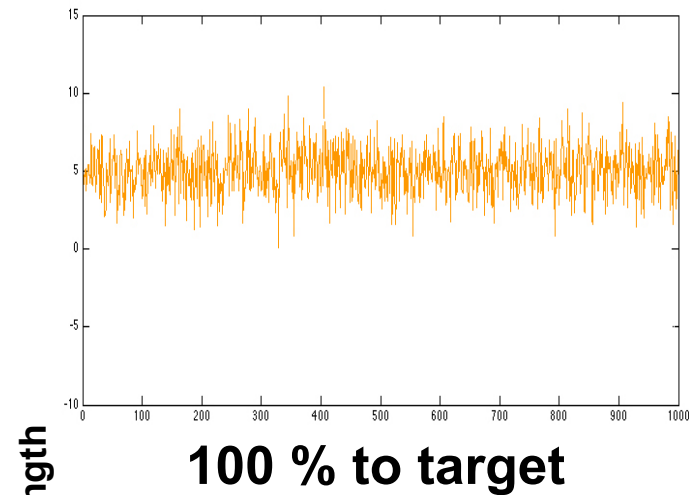
$$\Sigma = A + B + C$$



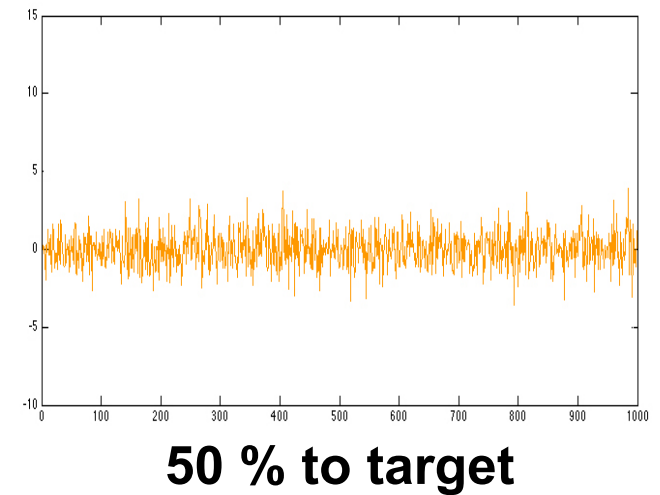
# Cue noise can explain guidance defects of mutants



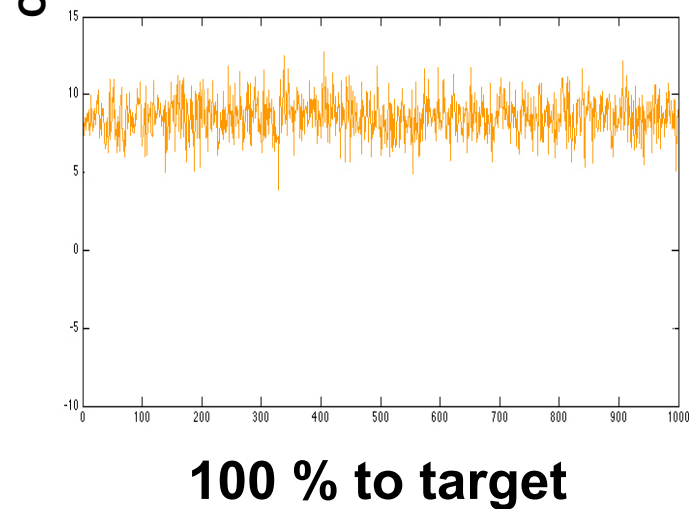
$$\Sigma = A + B + C$$



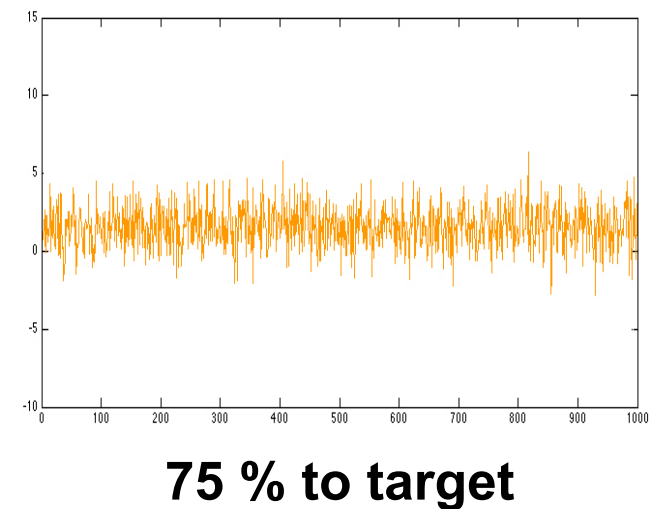
$$\Sigma = B + C$$



$$\Sigma = A + B$$

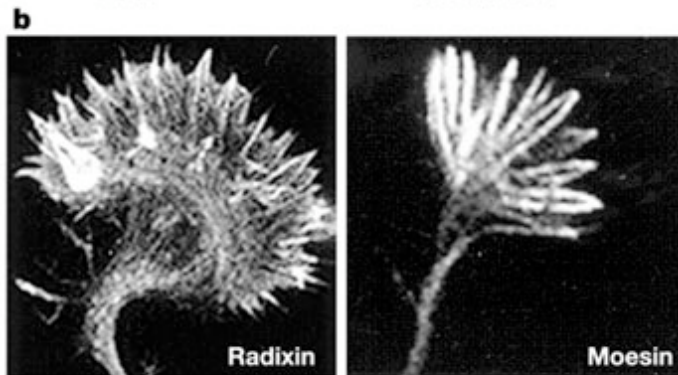
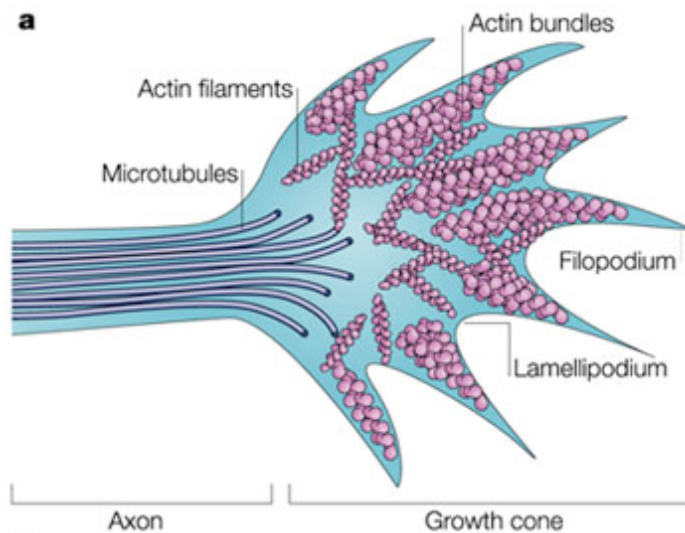


$$\Sigma = A + C$$

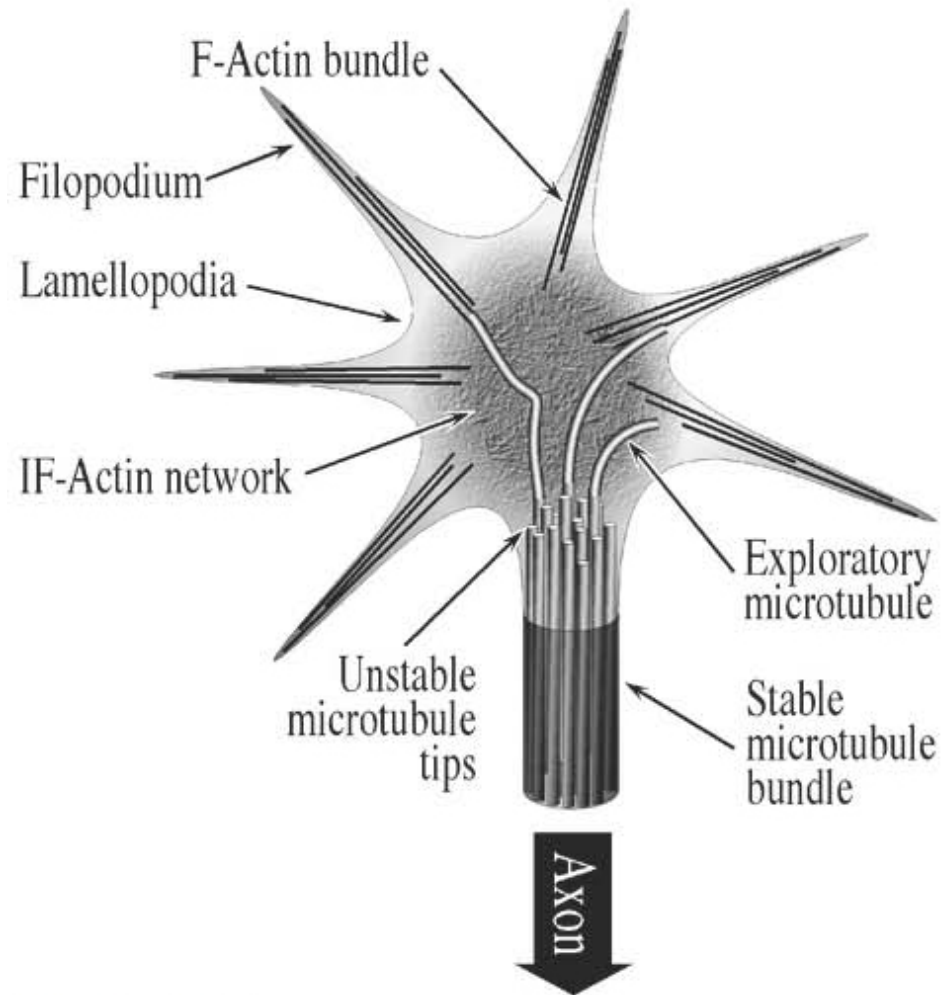


Axon index

# Growth cone machinery is a dynamic structure susceptible to stochastic events



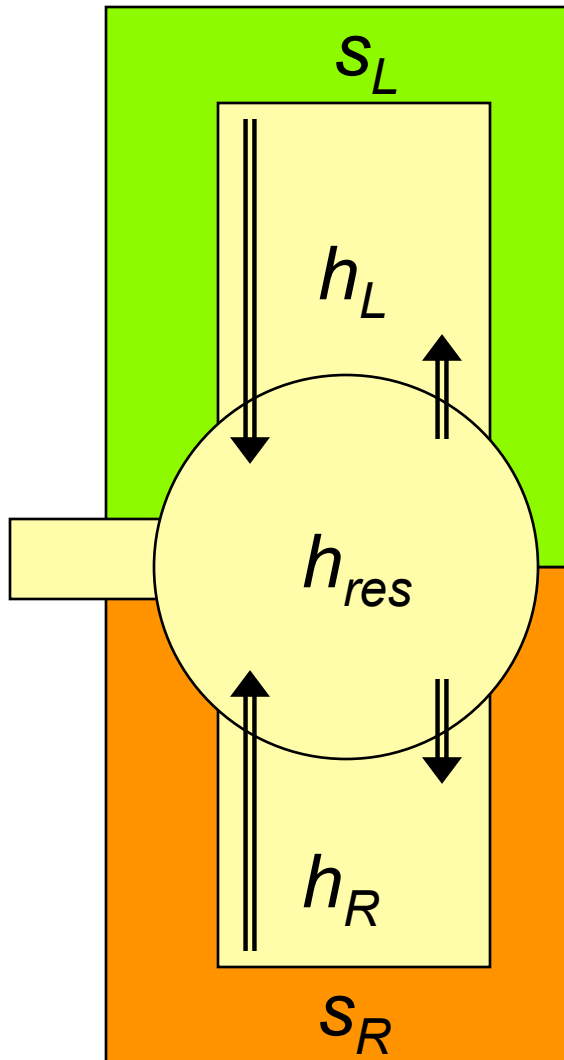
Vijaya Ramesh  
*Nature Reviews Neuroscience* 2004



Maskery and Shinbrot  
*Annual Review in Biomedical Engineering* 2005



# Dynamical model of cue integration



Summation: 
$$s_{L,R} = \sum_i c_i^{L,R} + \eta_i$$

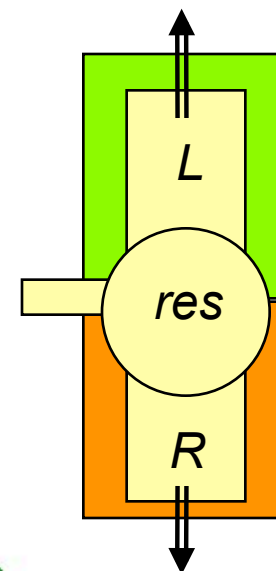
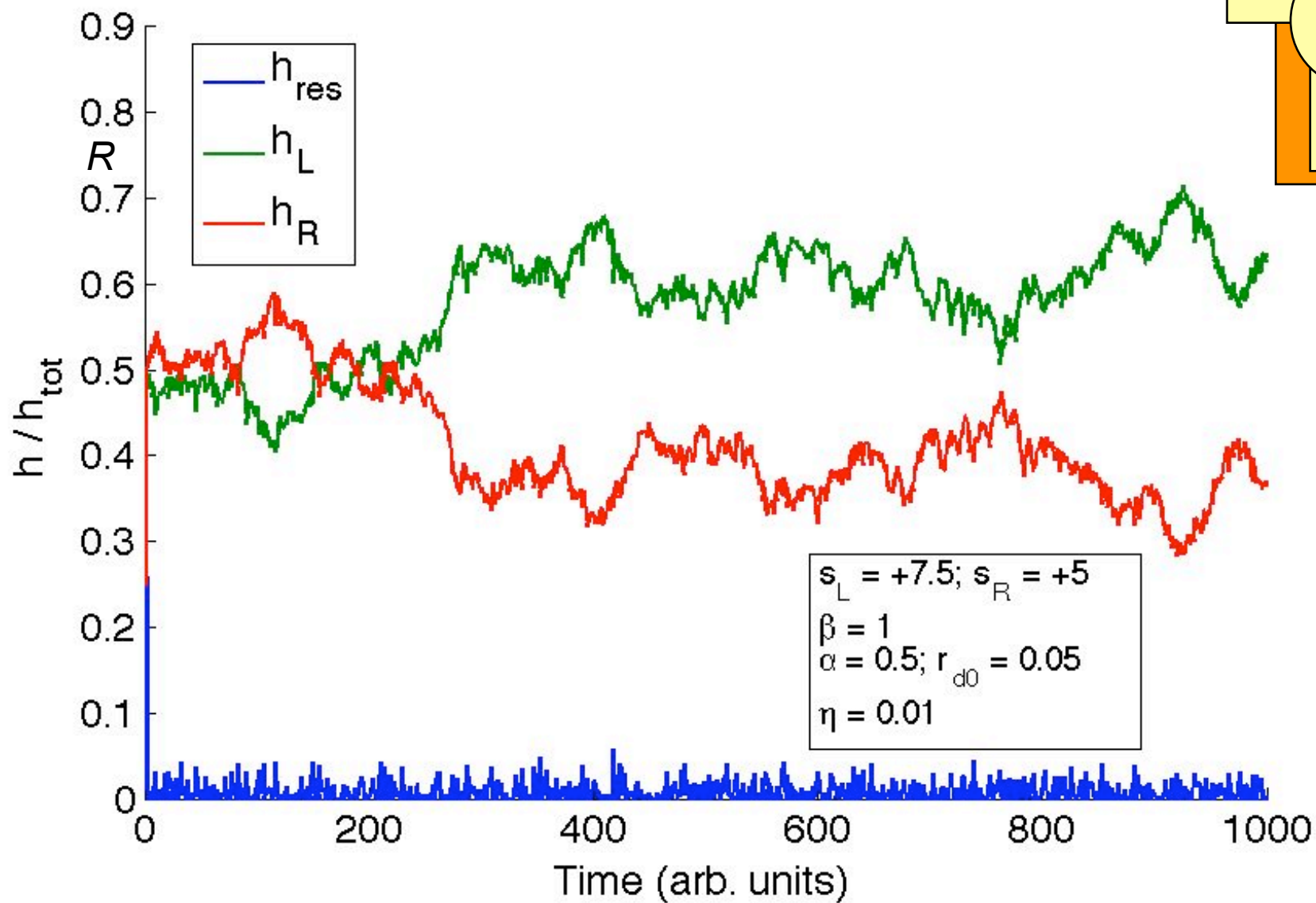
Dynamics: 
$$\frac{dh_{L,R}}{dt} = \alpha h_{res} - r_d(s_{L,R}) + \eta$$

$$\frac{dh_{res}}{dt} = -\frac{dh_L}{dt} - \frac{dh_R}{dt}$$

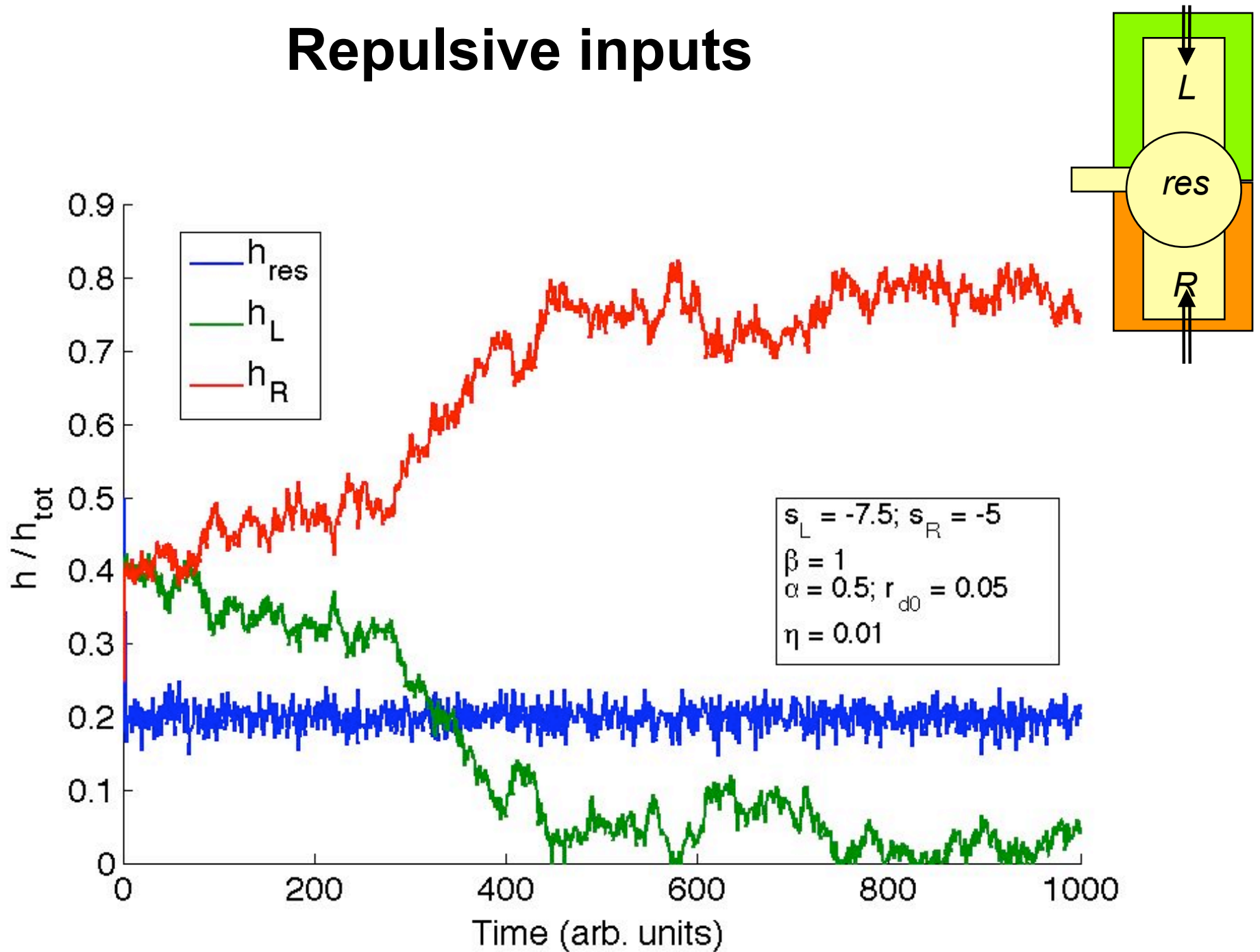
Degradation rate: 
$$r_d(s) = 2r_0 \left( \frac{1}{1 + \exp(\beta s)} \right)$$

Conservation: 
$$h_{tot} = h_L + h_R + h_{res} + h_{noise}$$

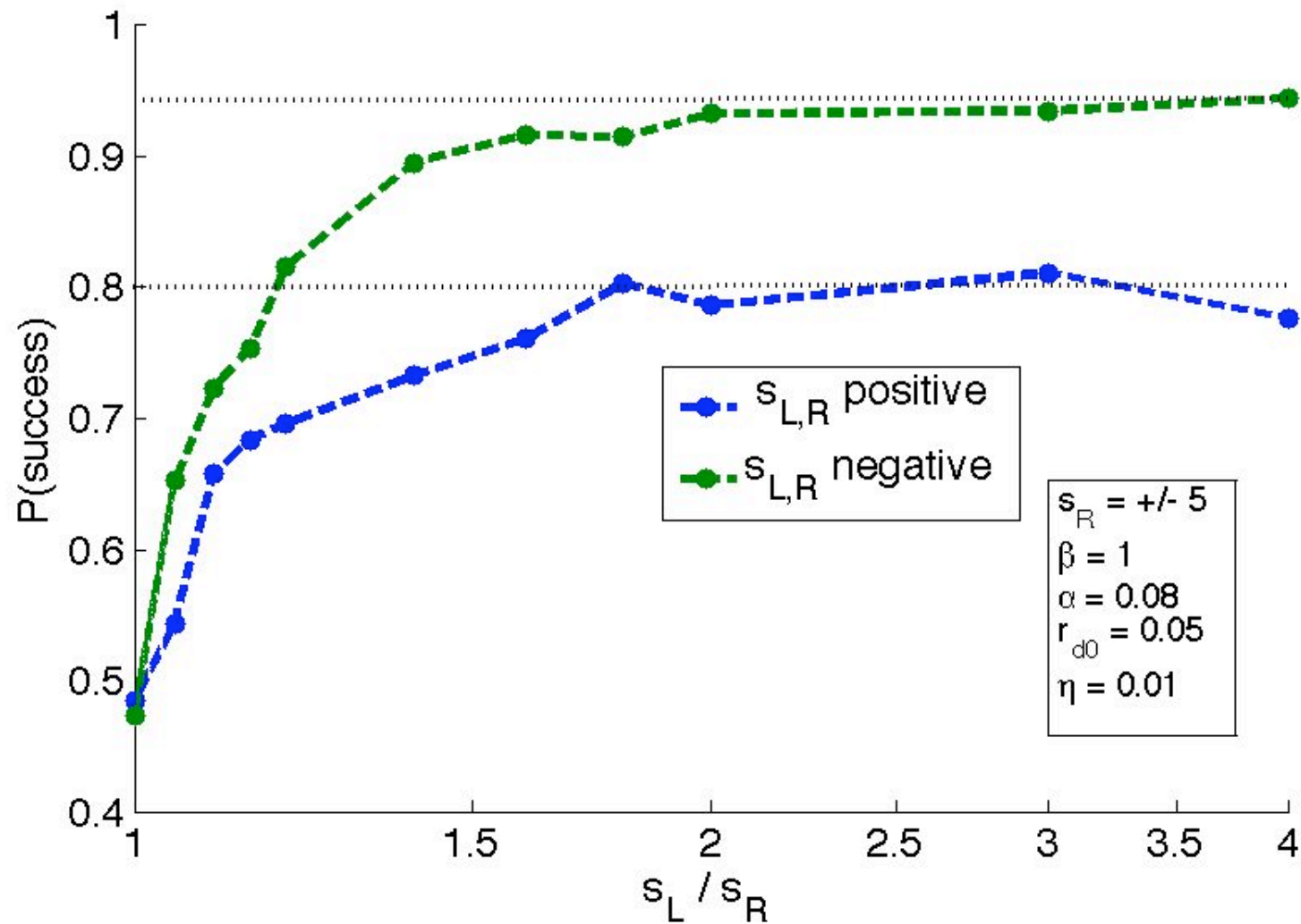
# Attractive inputs



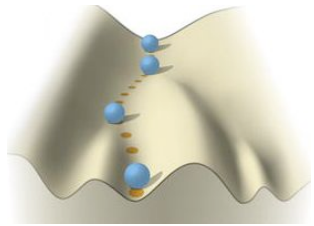
# Repulsive inputs



# Stochastic choice behavior & positive/negative asymmetry

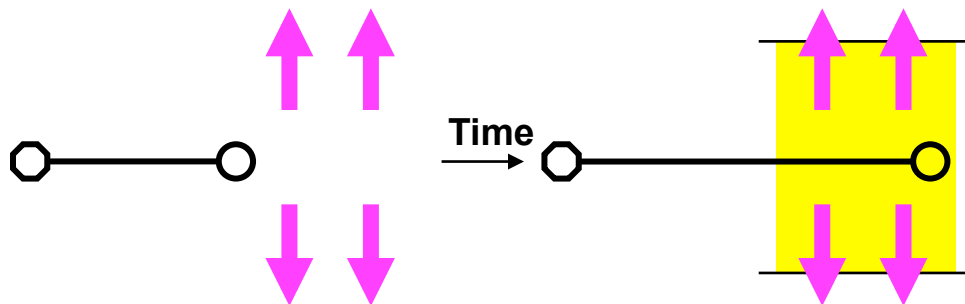




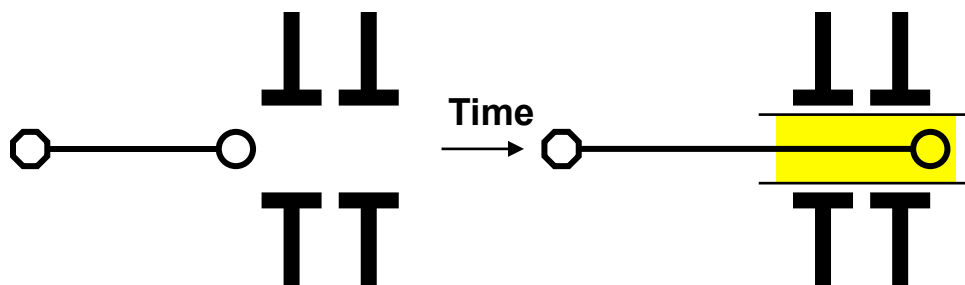


# Competing cues and system stability

**ATTRACTIVE cues**  
--> *unstable* trajectories



**REPULSIVE cues**  
--> *stable* trajectories



# Summed cue strength is limited by sensor noise and amplification

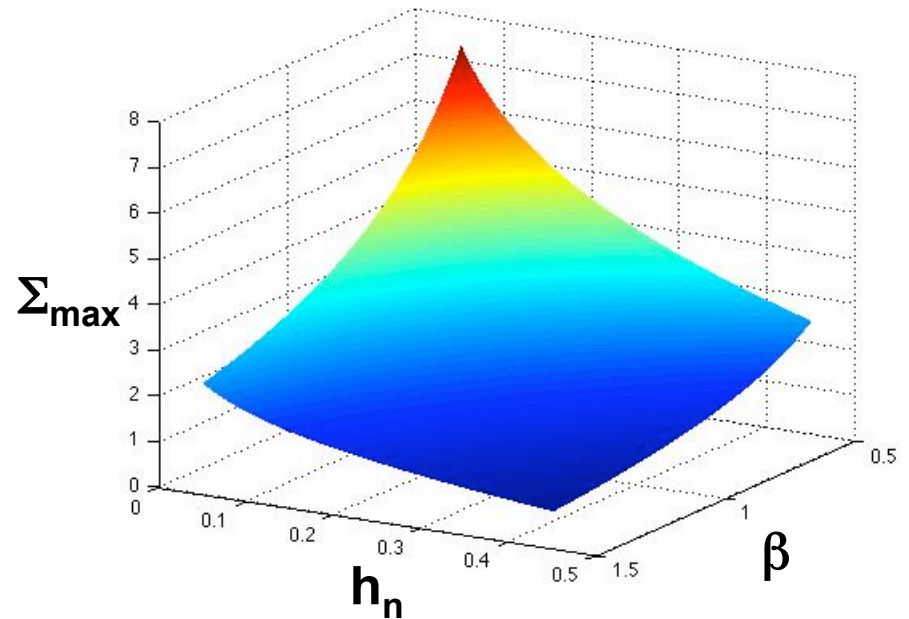
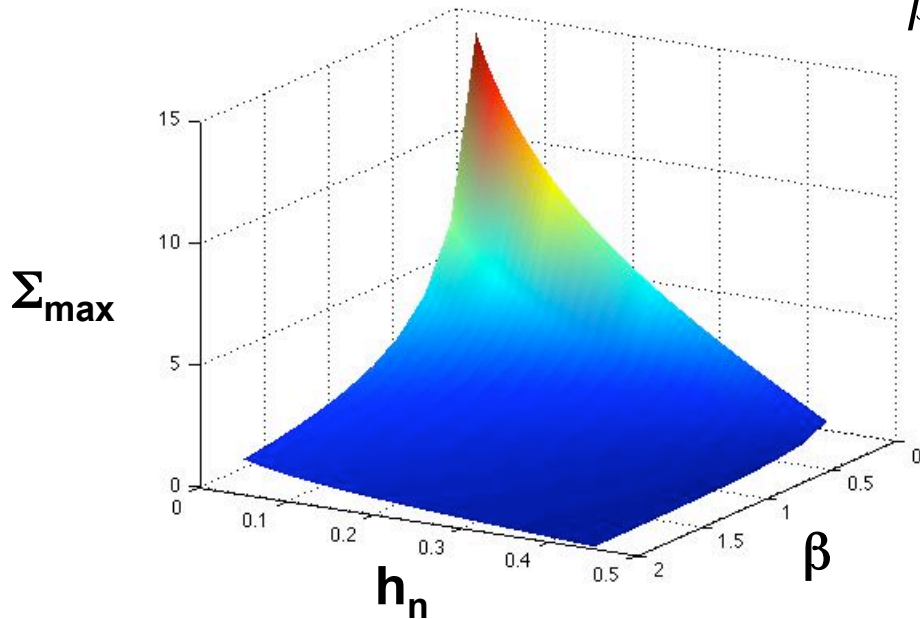
$$h = h_{engaged} + h_{available} + h_{noise}$$

$$h_{engaged} = h \left( \frac{1}{1 + \frac{1}{e^{\beta \Sigma}}} \right)$$

$$h_{engaged} \leq h - h_{noise}$$

$$\Sigma_{max} = \frac{\log\left(\frac{h - h_{noise}}{h_{noise}}\right)}{\beta}$$

$h$  = sensing cytoskeletal material  
 $\Sigma$  = summed cue  
 $\beta$  = signal transduction strength



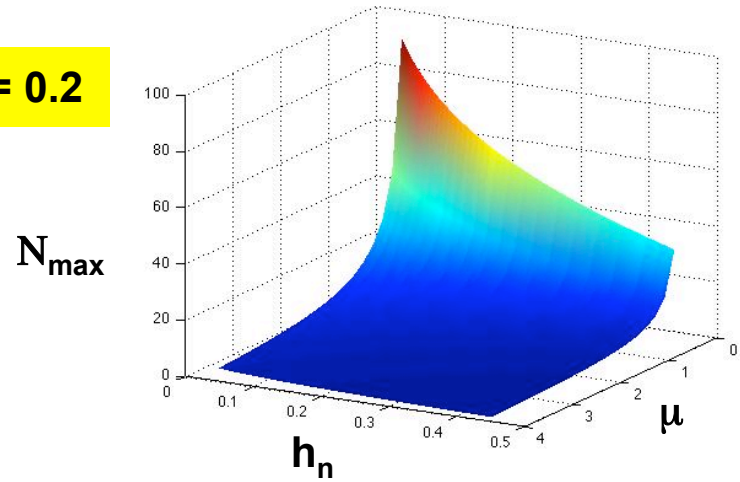
# Cue number depends on sensor noise and mean cue strength

$$\Sigma_{\max} = N_{\max} \times \mu$$

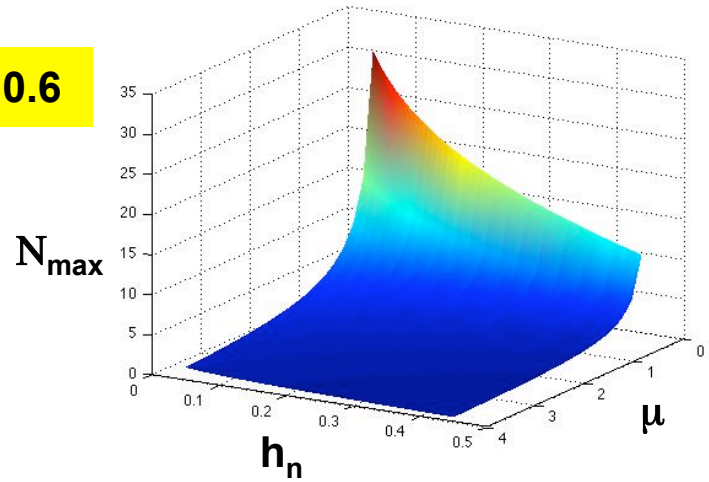
$$N_{\max} = \frac{\log\left(\frac{h - h_{\text{noise}}}{h_{\text{noise}}}\right)}{\beta \times \mu}$$

$h$  = sensing cytoskeletal material  
 $\Sigma$  = summed cue  
 $\beta$  = signal transduction strength  
 $\mu$  = mean cue strength  
 $N$  = number of cues

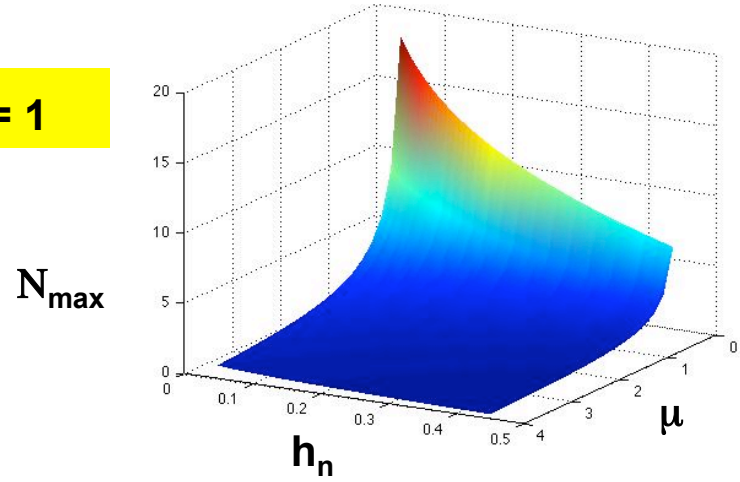
$\beta = 0.2$



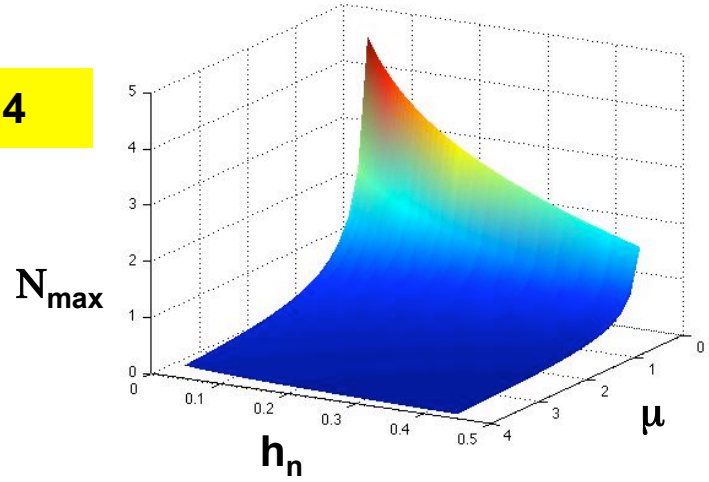
$\beta = 0.6$



$\beta = 1$



$\beta = 4$





# **Too-much-information hypothesis**

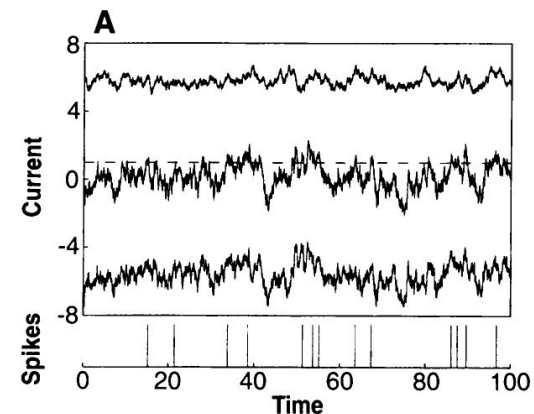
Noise and finite sensing capacity *limit* information

## Construction of neural circuits

Luria et al., *Neuron* 2008

## Neuronal firing rates

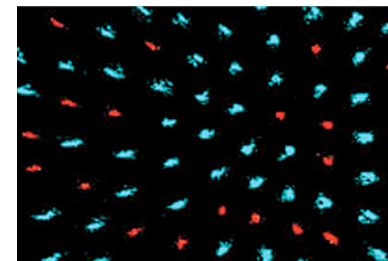
Van Vreeswijk & Sompolinsky, *Science* 1996



## Drosophila photoreceptor fields

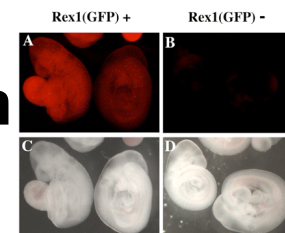
70% - 30% unequal partitioning

Wernet et al., *Nature* 2006



## Embryonic stem cells transient differentiation

Toyooka et al., *Development* 2008



# Finance - the anti-portfolio effect

Vlad et al., *PNAS* 2007

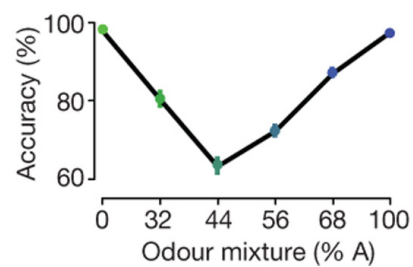
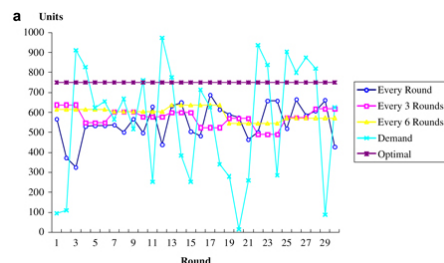
# Consumer choices

Lurie and Swaminathan,  
*Organizational Behavior and  
Human Decision Processes* 2009

# Olfactory behavior choices

Kepecs et al., *Nature* 2008

# Other behavioral decisions?...



# SUMMARY AND FUTURE WORK

## 1. Control of axon trajectories A/B ephrin-Eph *mirror symmetry*



## 2. *EphBs* suppress phenotypic variability

General variability suppressors - *Hsp90*, Rutherford and Lindquist, *Nature* 1998, 2003

Specific variability suppressors - *Eph* genes

*EphA4*, Helmbacher et al., *Development* 2000

*EphBs*, Luria et al., *Neuron* 2008

## 3. Theoretical modeling

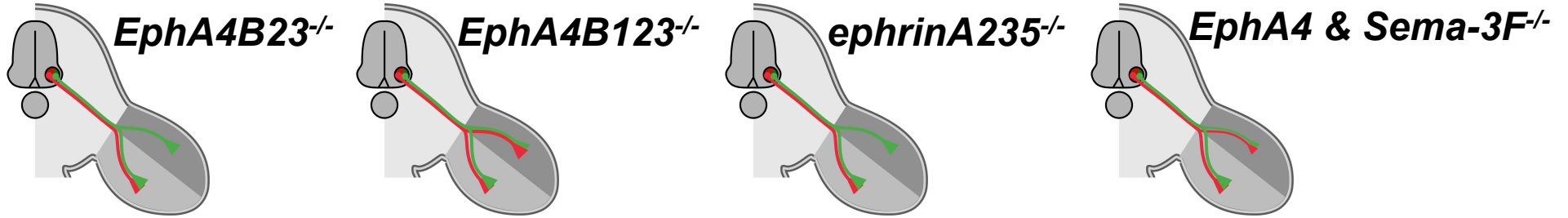
suggests how *genetic variability* is translated into *phenotypic variability*

suggests noise limits the amount of information controlling binary decisions

generates **experimental predictions**

# Genetic experiments

mutants



soft perturbations: RNAi

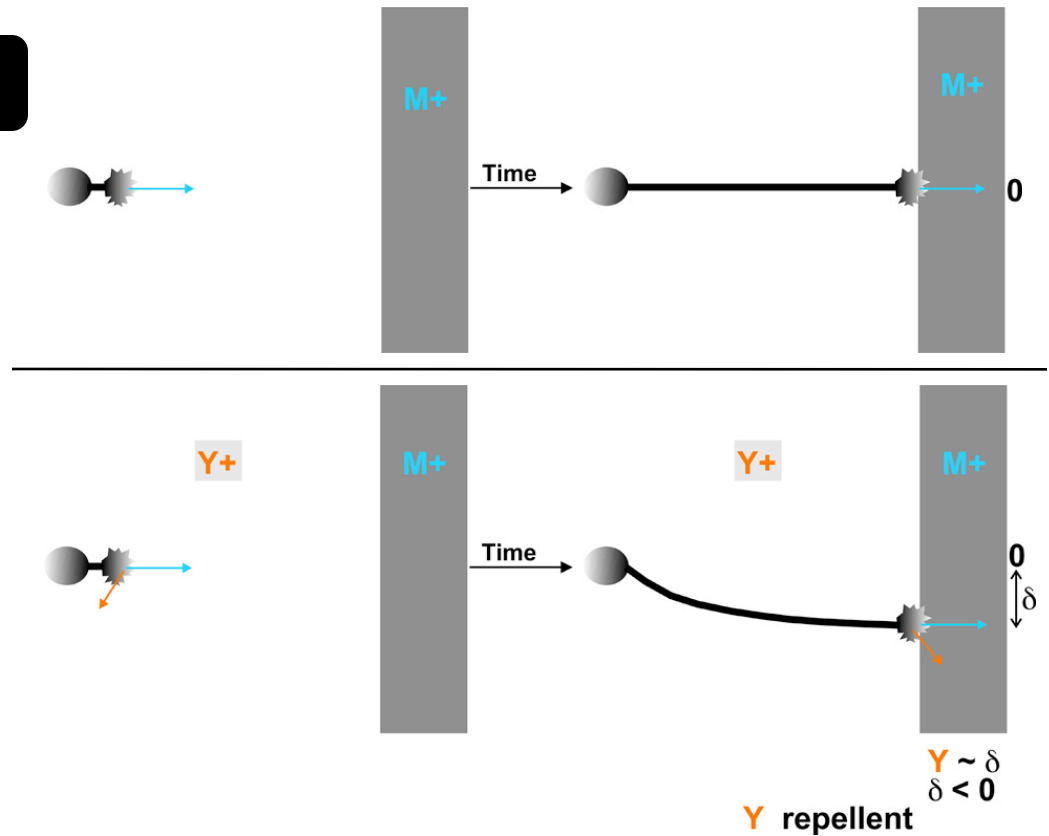
# Biophysical experiments

Diffusible

Surface-bound

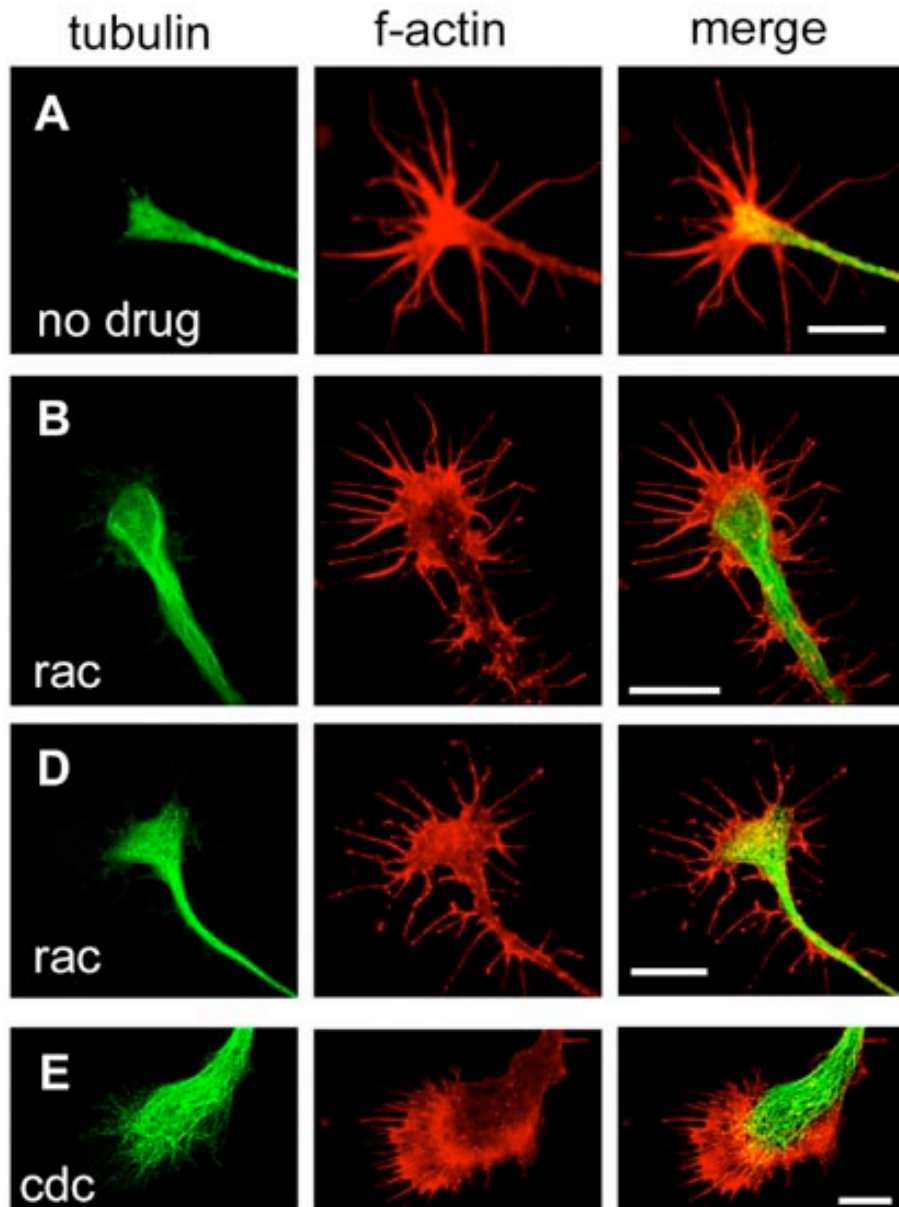
Mu-ming Poo  
Christine Holt  
Geoff Goodhill

Friedrich Bonhoeffer  
Ludwig von Philipsborn





# Growth cones reorient in response to electrical fields



(A-E) *Xenopus* spinal neuron growth cones in an electric field for 5h, with the cathode at left.

(B,D) A peptide that blocks rac effector binding stimulates filopodia relative to lamellipodia.

(E) A peptide that blocks cdc42 effector binding stimulates lamellipodia relative to filopodia.

Modified from Fig 5  
Rajnicek et al. 2006.  
*J Cell Science*  
**119**: 1736-45.

## Physiology Molecular mechanisms

Keith Robinson  
Claudio Stern  
Mu-ming Poo

**Colin McCaig**

# Cue strength: quantification using benchmarks

