

**Sarah L. Keller**

**Department of Chemistry  
University of Washington, Seattle**

***Observations of lipid membranes in (roughly) 2D  
influenced by bulk water in 3D***

***Project 1: Dynamic critical phenomena***

***Project 2: Domain coarsening***

# Postdoctoral Fellowships in Biophysics University of Washington, Seattle

*<http://depts.washington.edu/pbiopage/sackler>*

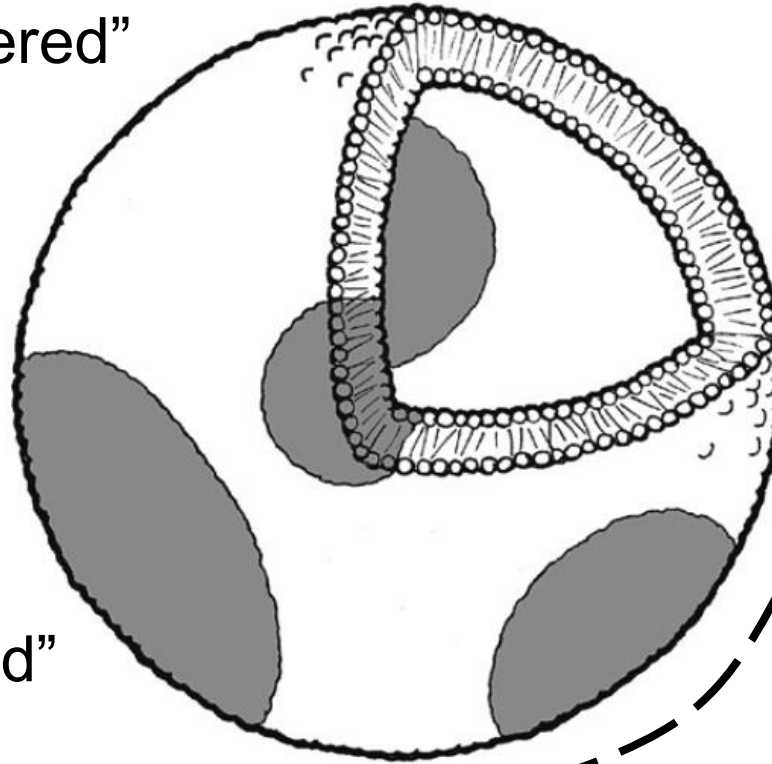
## **The Raymond and Beverly Sackler Scholars Program in Integrative Biophysics**

- Up to two years of funding (\$50,000 yearly for stipend / benefits, \$3000 yearly for professional development / conference travel)
- Encourages collaborative work by supporting fellows partnering with two principal investigators working in different fields.

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# Giant unilamellar vesicles (GUVs)

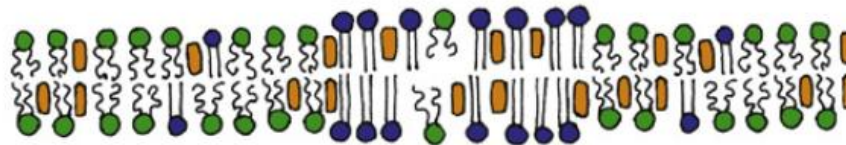
“liquid-disordered”  
phase



“liquid-ordered”



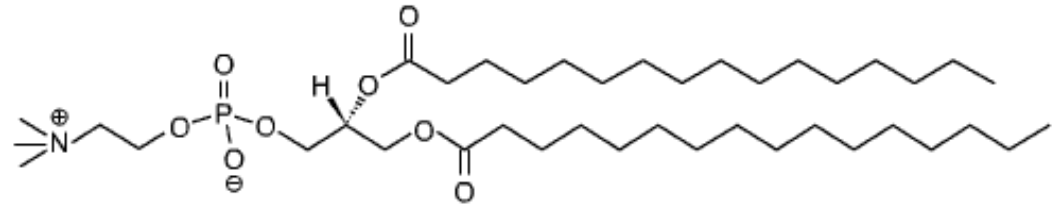
**In our expts,  
domains do not  
bulge out of the  
spherical shell**



Drawing by Aurelia Honerkamp-Smith

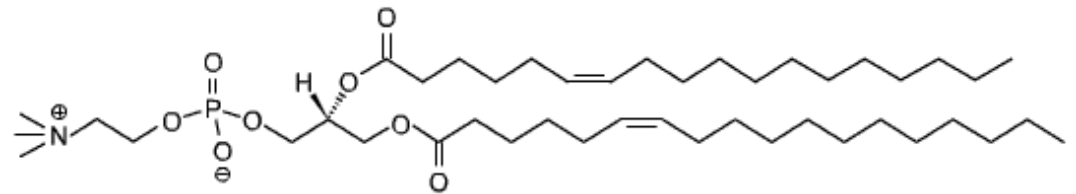
# Our vesicles (GUVs) contain 3 components (+ dye)

**High Melting Temp**  
*e.g. Saturated Lipids*  
*di(16:0)PC; DPPC*



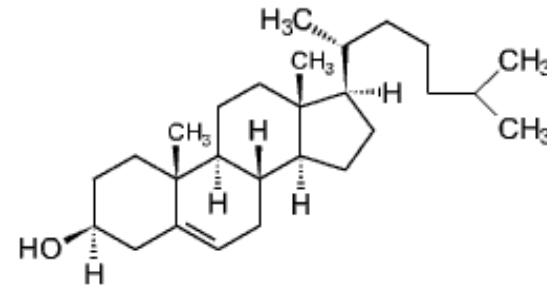
©Avanti Polar Lipids

**Low Melting Temp**  
*e.g. Unsaturated Lipids*  
*or DiphytanoylPC*



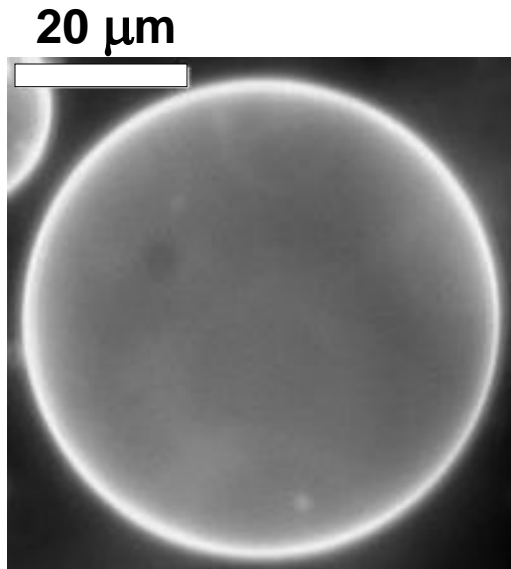
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**Cholesterol**  
*or similar sterols*

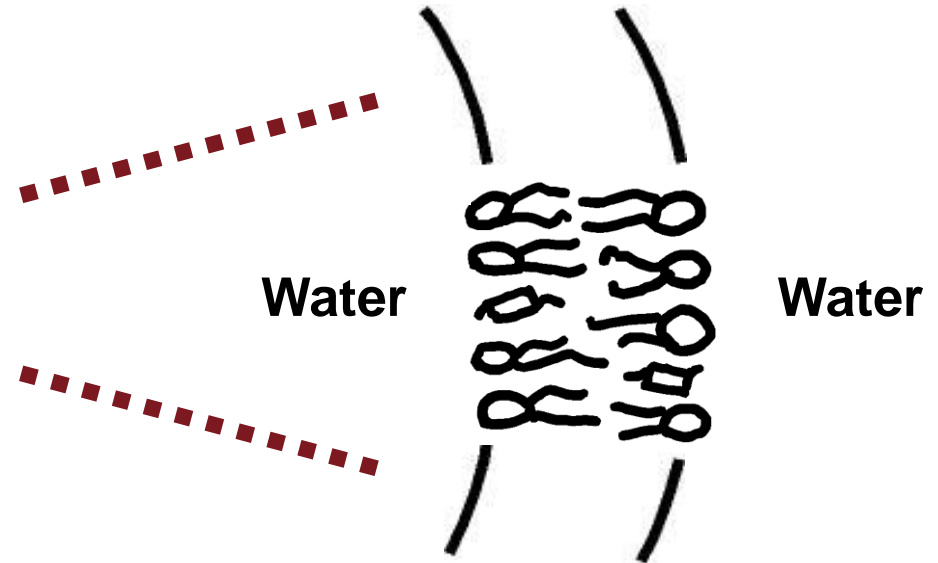


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# High temperature: uniform vesicle

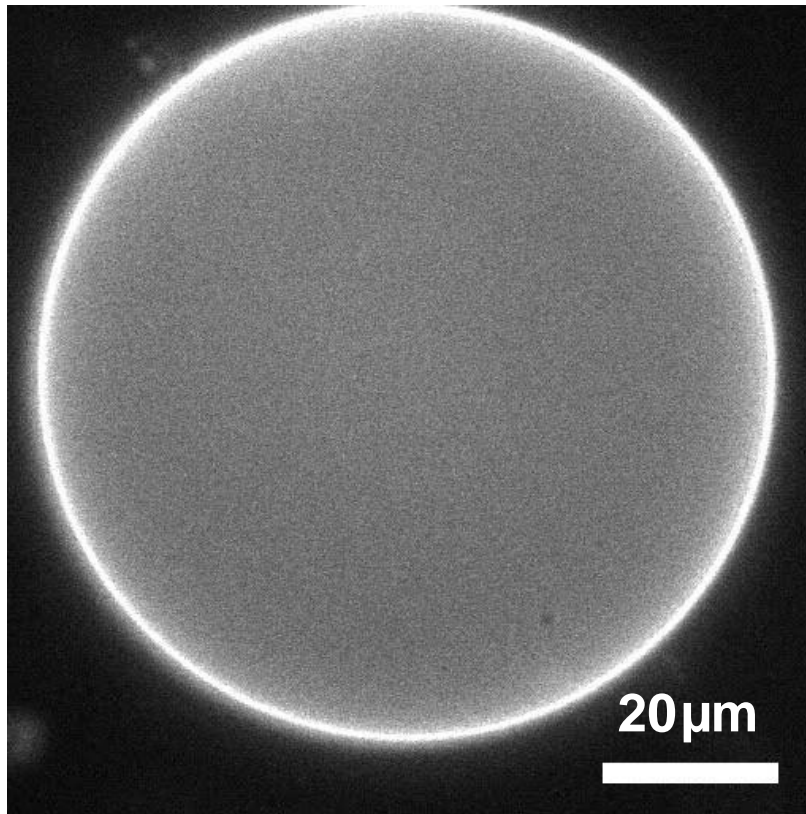


One Uniform  
Phase



Vesicles are free-floating in water,  
not attached to or sitting on any substrate.

As temperature is lowered, micron-scale domains appear and coarsen.



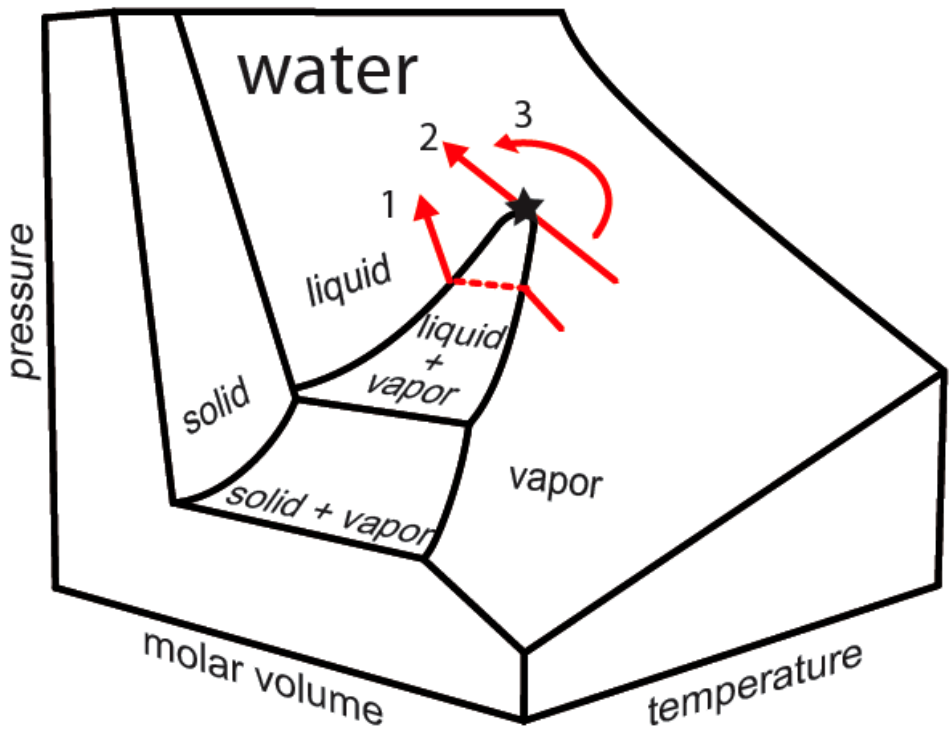
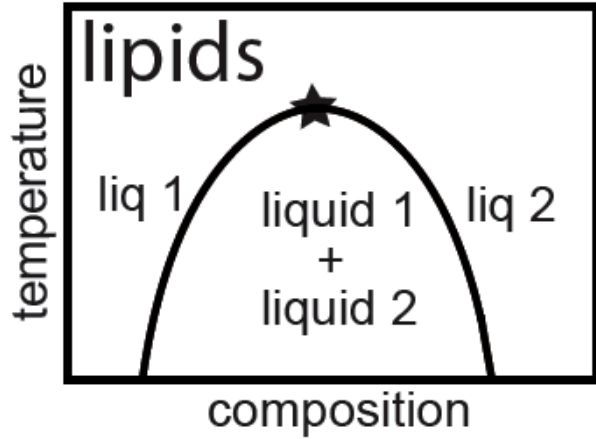
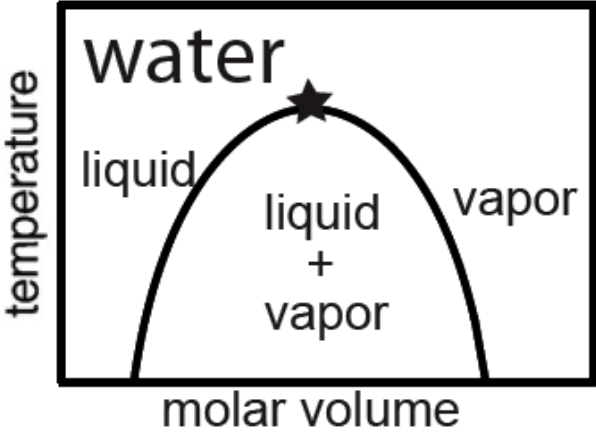
High temp:  
**Uniform** membrane

Low temp:  
**Liquid** domains /  
**liquid** background

The image is 92μm on this edge.

# ***Project 1: Dynamic critical phenomena***

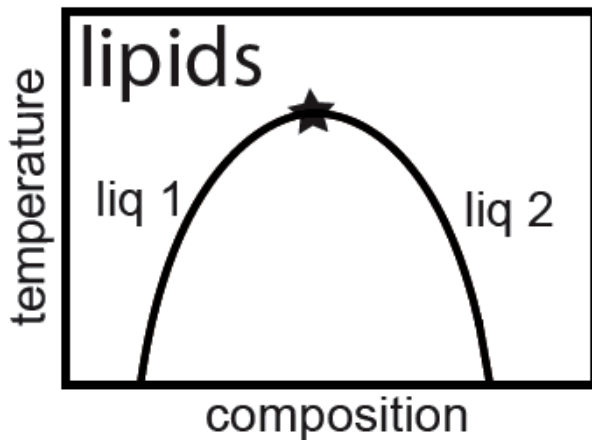
# Critical points



A.R. Honerkamp-Smith et al.  
*BBA* (2009)



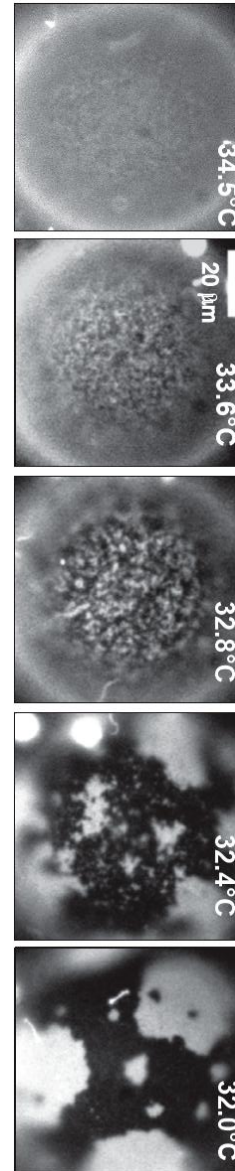
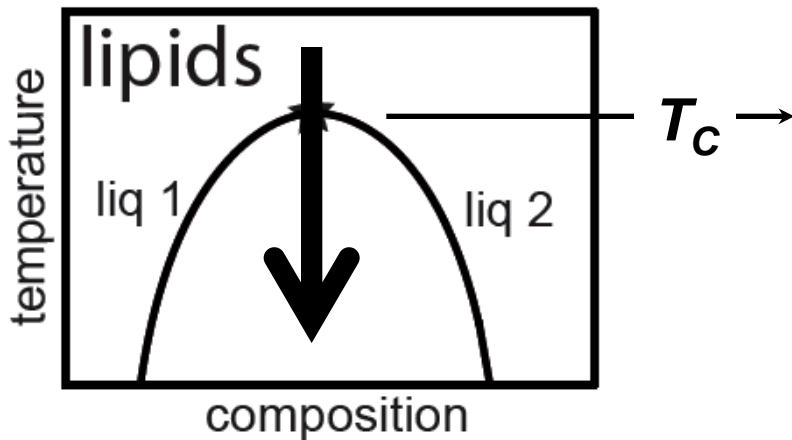
**Fluctuations  
in lipid composition  
are associated  
with critical points  
(or plait points).**



**What does this  
transition look like  
in the microscope?**

***It's beautiful.***

**Fluctuations  
in lipid composition  
are associated  
with critical points  
(or plait points).**



**Way above  $T_c$ :  
Uniform, grey**

**Above  $T_c$ :  
Tiny fluctuations**

**Which grow  
bigger...**

**And bigger...**

**Just below  $T_c$ :  
domains**

**Which become  
more circular...**

# Aurelia measured two time-independent quantities to find that membranes behave as 2D Ising models.

Aurelia's  
Data

2-D Ising

3-D Ising

$$\nu = 1.2 \pm 0.2$$

$$= 1$$

$$= 0.630$$

$\xi \sim |T-T_c|^{-\nu}$   
correlation length

**characterizes the biggest  
fluctuations we observe**

$$\beta = 0.124 \pm 0.03$$

$$= 0.125$$

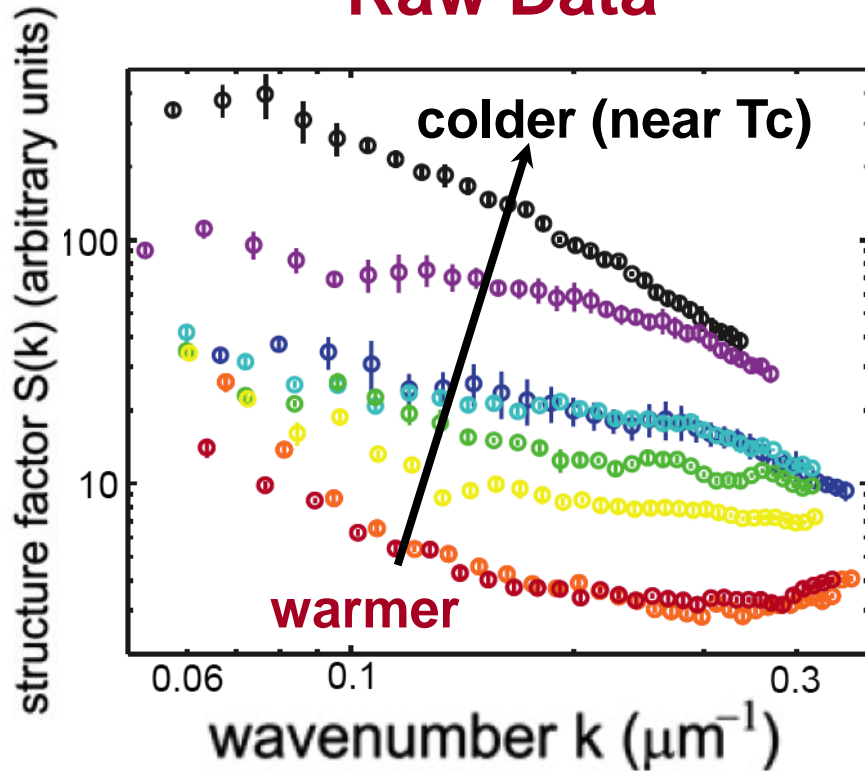
$$= 0.325$$

$\Delta m \sim |T-T_c|^\beta$   
order parameter

**characterizes the difference in  
composition of the two phases  
as  $T$  approaches  $T_c$  from below.**

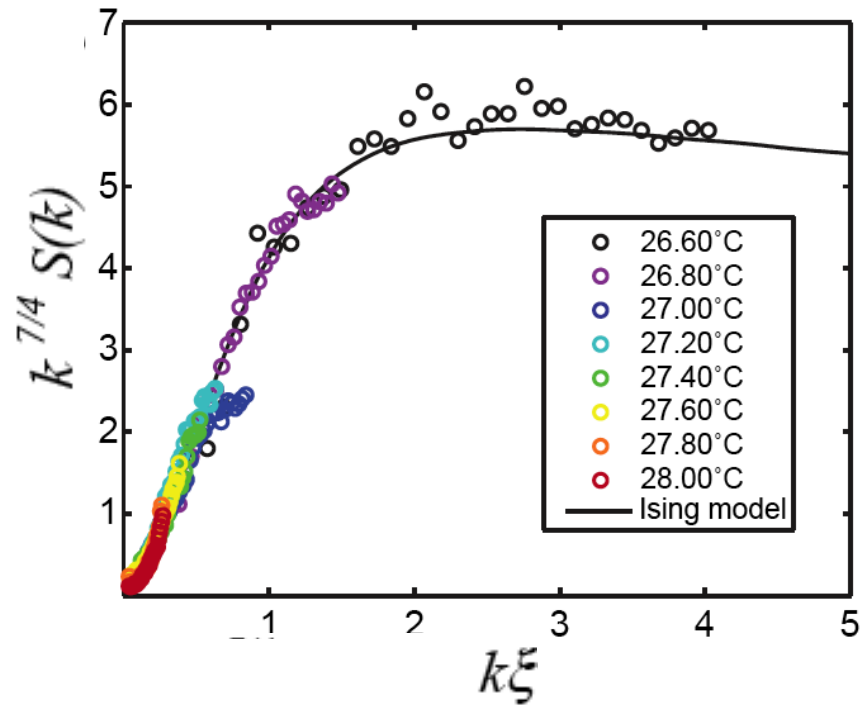
# Example from one set of experiments: Applying scaling to find critical exponents

## Raw Data



More structure near  $T_c$

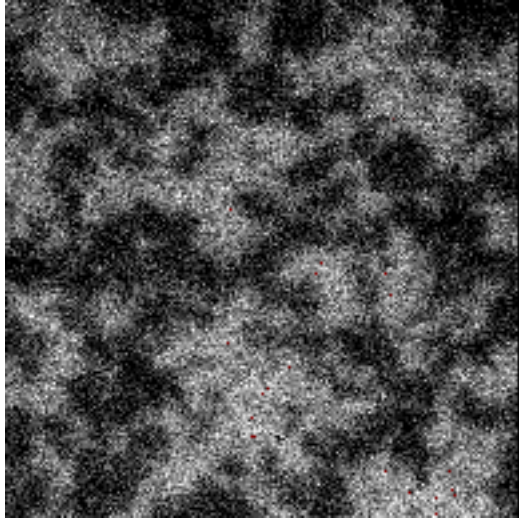
## $k^{7/4} S(k)$ vs. $k\xi$



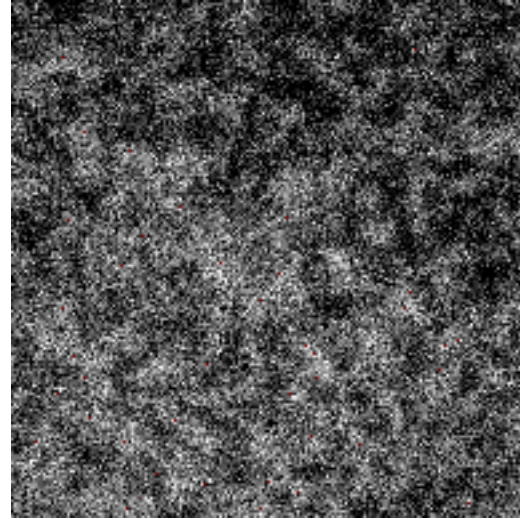
Aurelia extracted the value  
of  $\xi$  at each temp

$$\xi \sim |T - T_c|^{-\nu}$$

Now, what about any time dependent quantities?  
Above  $T_c$ , no domains persist - only fluctuations.



**Colder, 33°C  
(closer to  $T_c$ )**

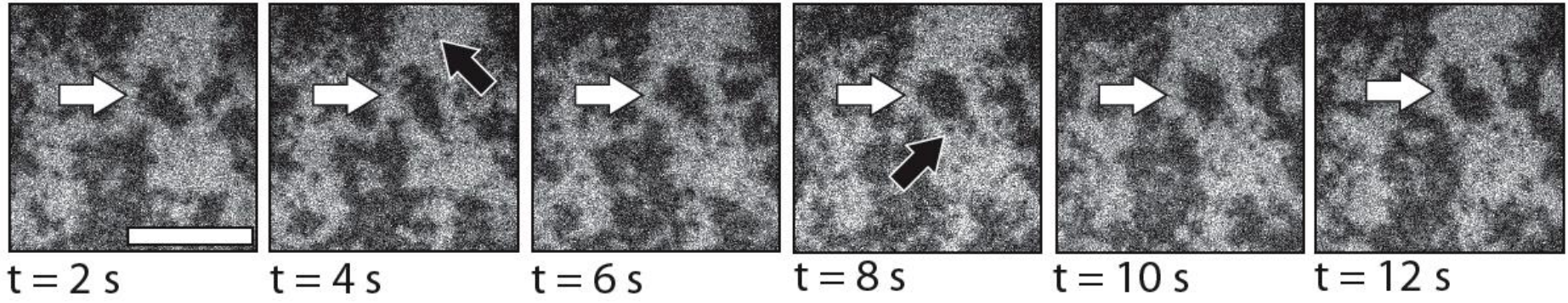


**Warmer, 34°C  
(further from  $T_c$ )**

*Approach this problem  
by using scaling and  
by thinking about  
how size relates to time*

Images  $\sim 30\mu\text{m} \times 30\mu\text{m}$

# How does size relate to time?



# How does size relate to time?

Hydrodynamic regime  
(where we usually live)

$$\Gamma = k^2 D(k, \xi)$$

rate (time<sup>-1</sup>)      wavenumber (length<sup>-1</sup>)      diffusion coefficient (μm<sup>2</sup>/s)

Critical regime  
(near a critical point)

$$\Gamma = k^z \bar{\Omega}(k\xi)$$

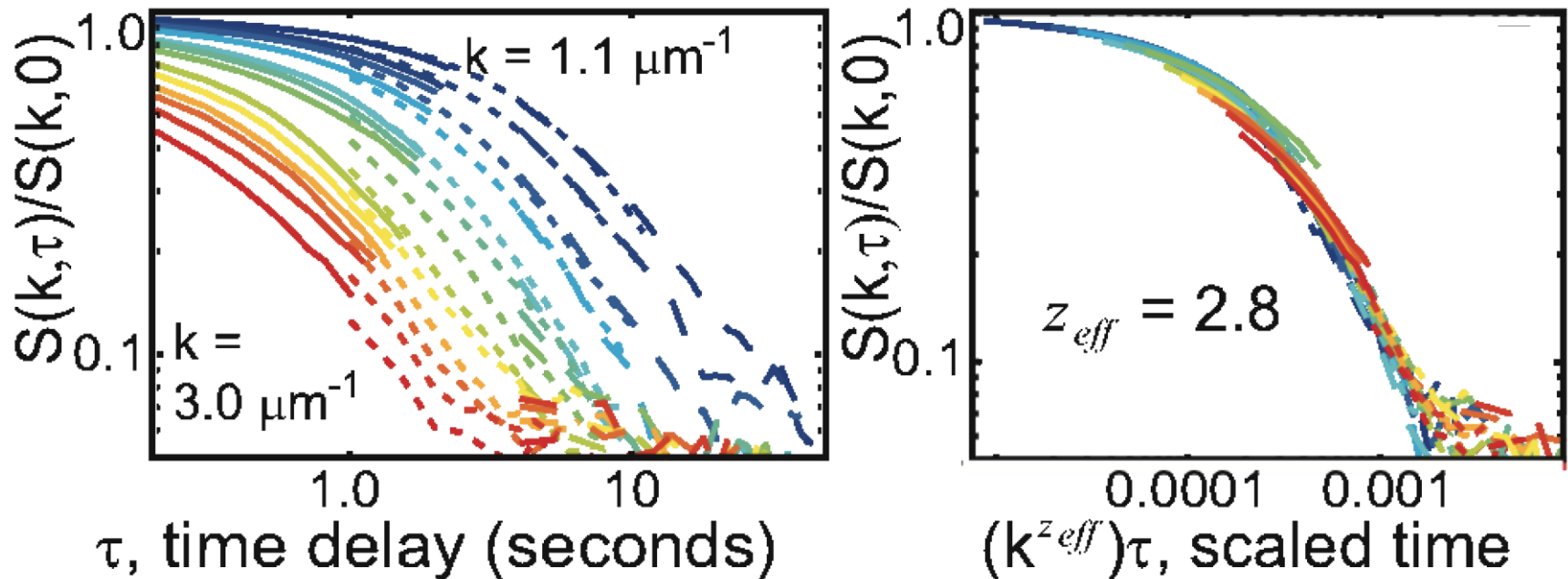
dynamic critical exponent      dynamic scaling function

## What should “z” be?

Model	Order parameter conserved?	Fluctuations dissipate via ...	Value of z
2-D Model A (magnets)	No.		N/A
2-D Model B	Yes.	2D lipid diffusion	$z = 3.75$
2-D Model H (binary fluid)	Yes.	Collective 2D hydrodynamics	$z = 2$
Model HC (hydrodynamic coupling) (Inaura&Fujitani 2008; Haataja 2009)	Yes.	Same as above, but with coupling to bulk fluid ( $L_{\text{hydrodynamic}} \neq 0$ $= \eta_{2D} / \eta_{3D}$ )	$z = 3$ ( $z_{\text{eff}}$ crosses over from 2 to 3 as $T \rightarrow T_c$ )



# Experimental data at the temperature closest to $T_c$ that can be analyzed.

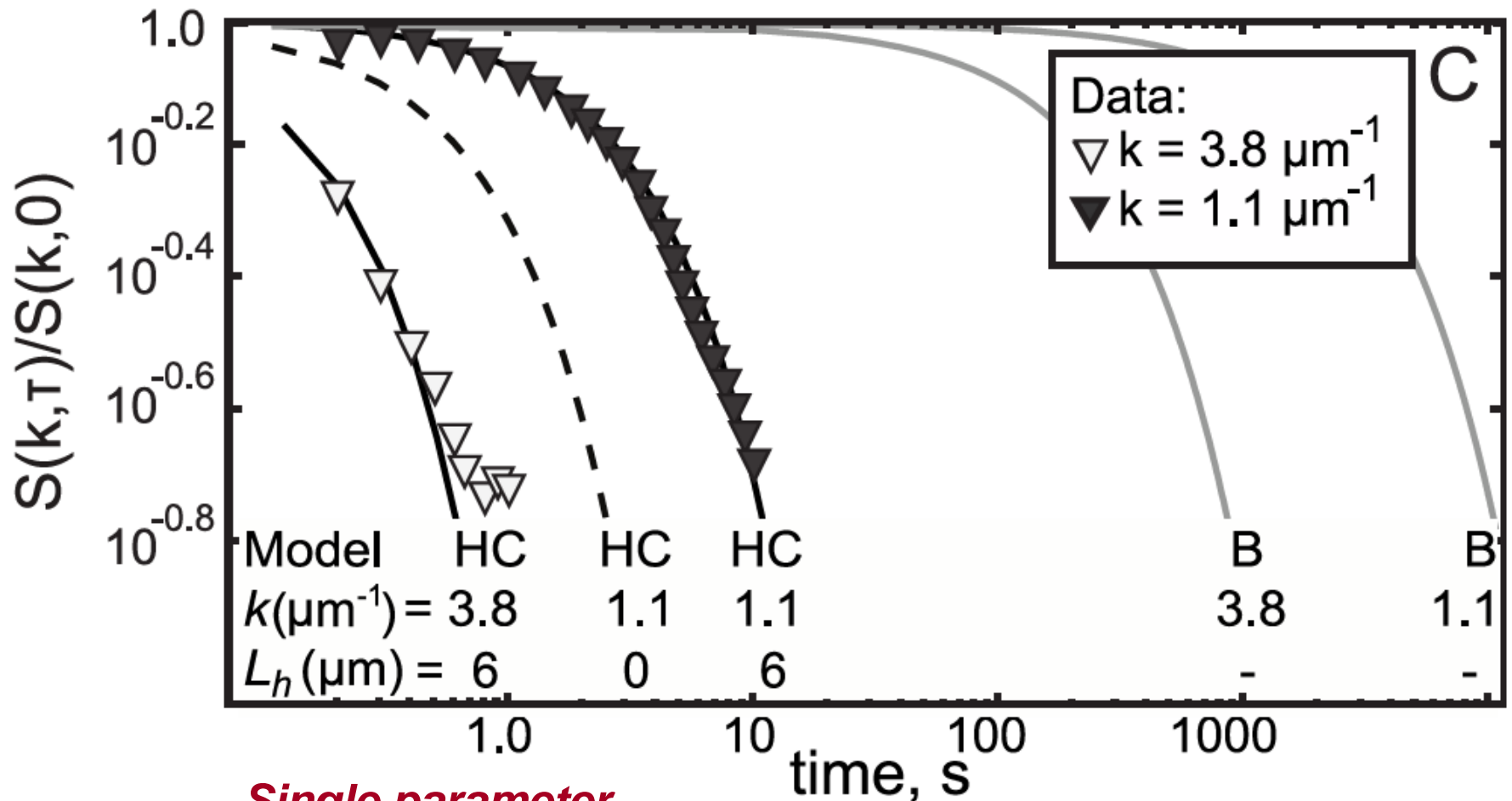


Aurelia.R. Honerkamp-Smith,  
Ben Machta, SLK  
In press *PRL* (2011)  
ArXiv:1104.2613

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# Excellent fit to structure factor predicted by “Model HC” (hydrodynamic coupling)



**Single parameter  
fit:  $L_h = 6.0 \pm 1.5 \mu\text{m}$**

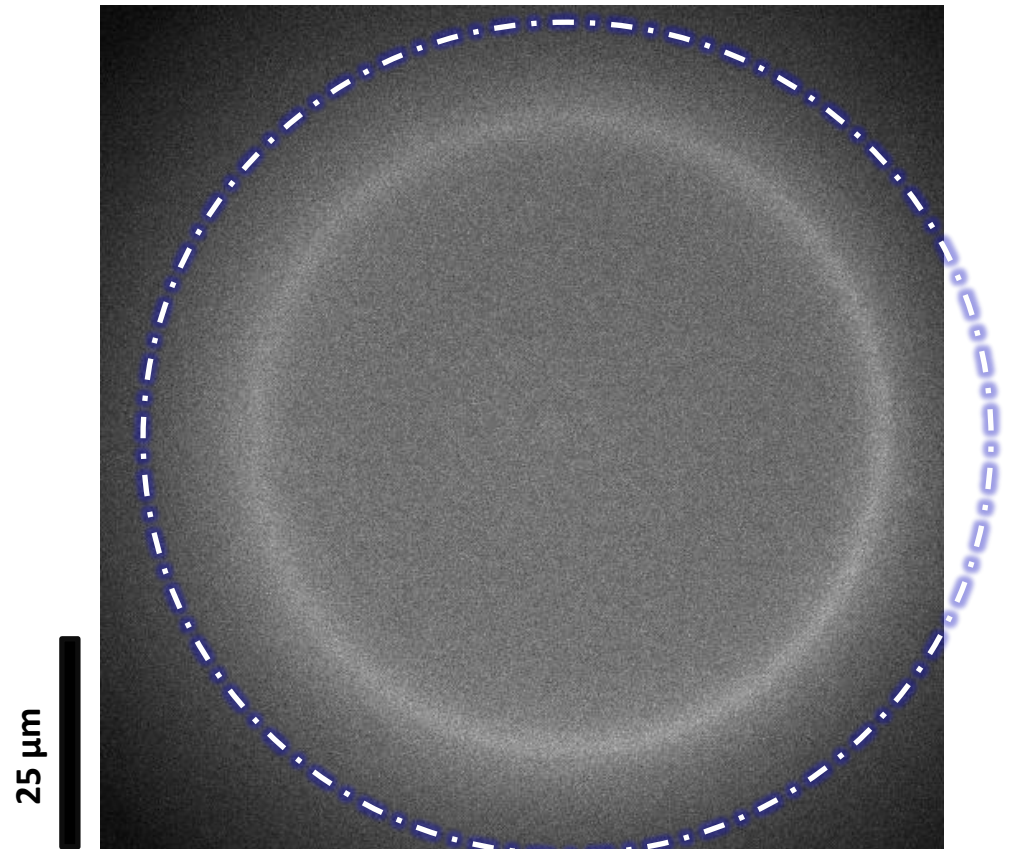
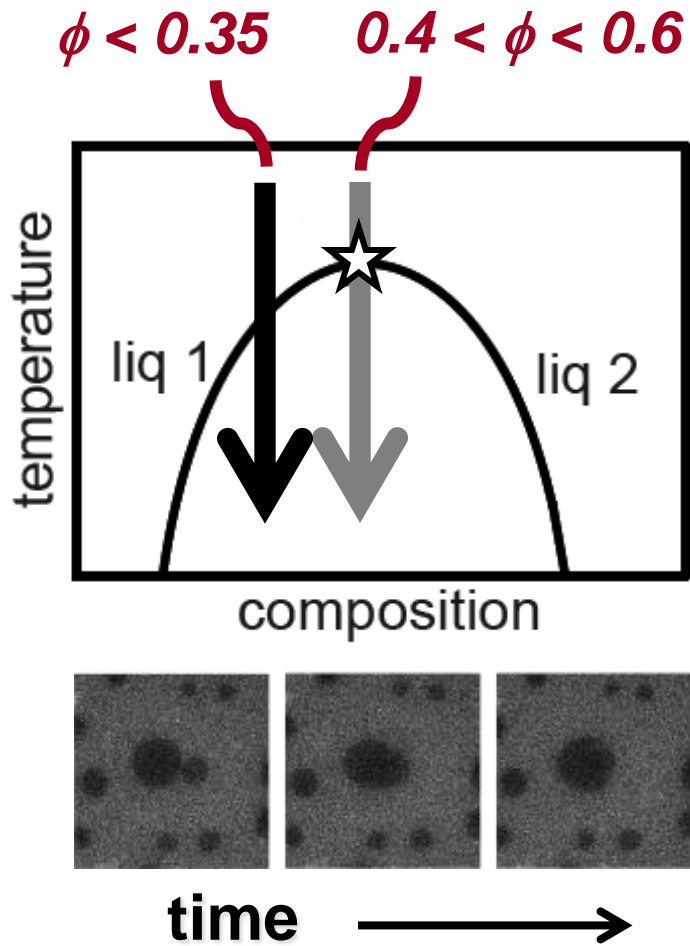
Aurelia.R. Honerkamp-Smith,  
Ben Machta, SLK  
In press *PRL* (2011)  
ArXiv:1104.2613

## **Conclusion from this section:**

Dynamics of membrane fluctuations are consistent with a recent theory that invokes hydrodynamic coupling between the membrane and bulk fluid such that Ising degrees of freedom are coupled to momentum modes.

## ***Project 2: Domain coarsening***

Quenches that are not through the critical point produce circular domains that merge with time.



Cynthia Stanich, Aurelia Honerkamp-Smith, et al.

# Which equations are relevant for diffusing domains?

## *Saffman-Delbrück (1975)*

small domains and/or high membrane viscosity

$$D(r) = \frac{k_B T}{4\pi\eta_{2D}} \left[ \ln \frac{\eta_{2D}}{(\eta_{3D})(r)} - \gamma + \frac{1}{2} \right]$$

$\eta_{3D}$  = 3D bulk viscosity of solvent

$\eta_{2D}$  = 2D viscosity of membrane

$\gamma = 0.5572$

*Hughes-Pailthorpe-White (1981)*      *de Koker (1996),*  
*Seki-Ramachandran-Komura (2011)*

large domains and/or low membrane viscosity

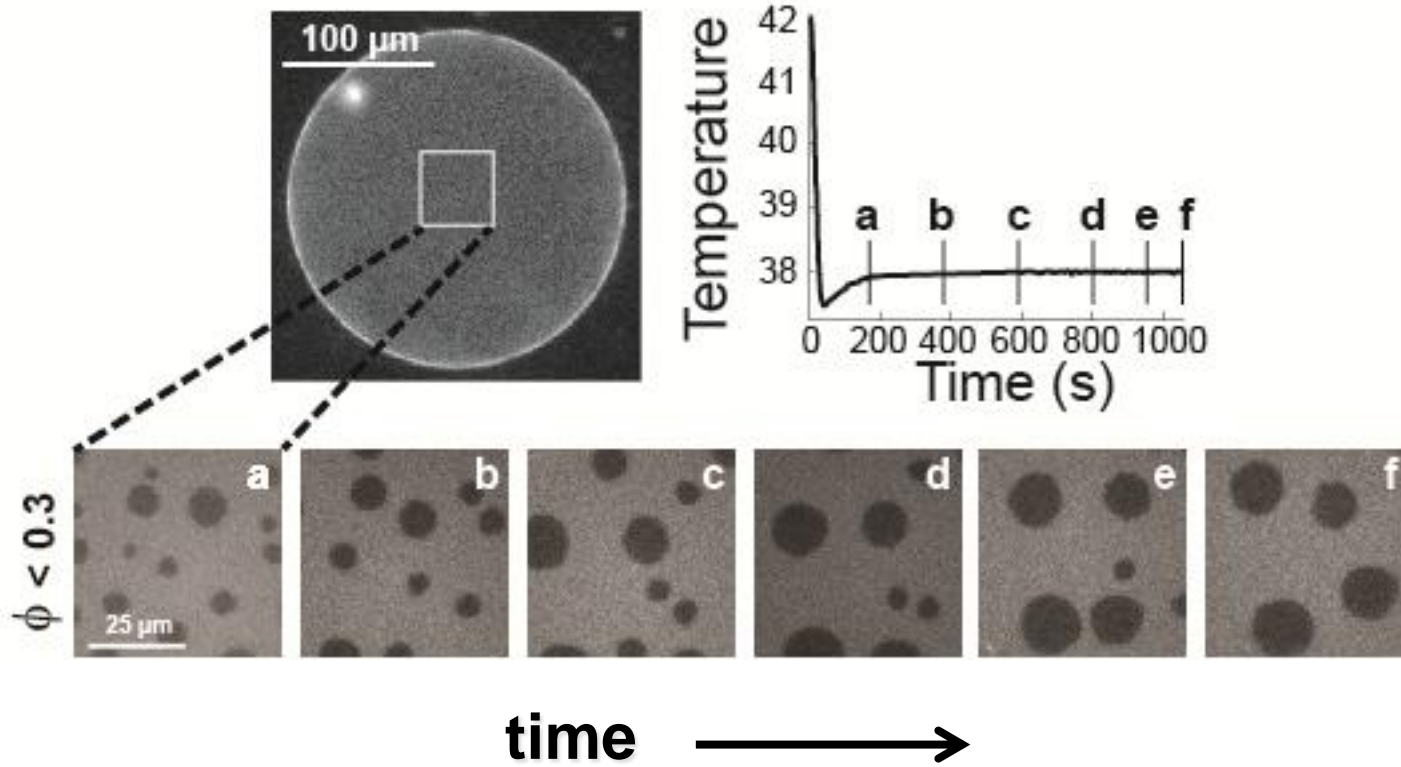
$$D(r) = \frac{k_B T}{16\eta_{3D}r} \qquad D(r) = \frac{2k_B T}{3\pi^2\eta_{3D}r}$$

A log-log plot of  $D$  vs.  $r$  should be a straight line.

*In between?*

Numerical solution by HPW, nice approximation by Petrov & Schwille

# Conceptually simple: Track domains of different size vs. time



Cynthia Stanich, Aurelia Honerkamp-Smith, et al.

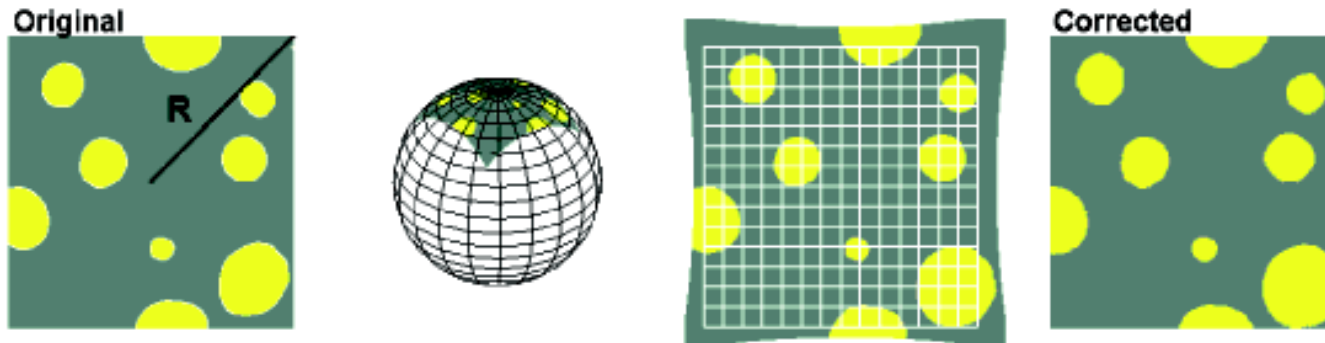


## Experimental headaches:

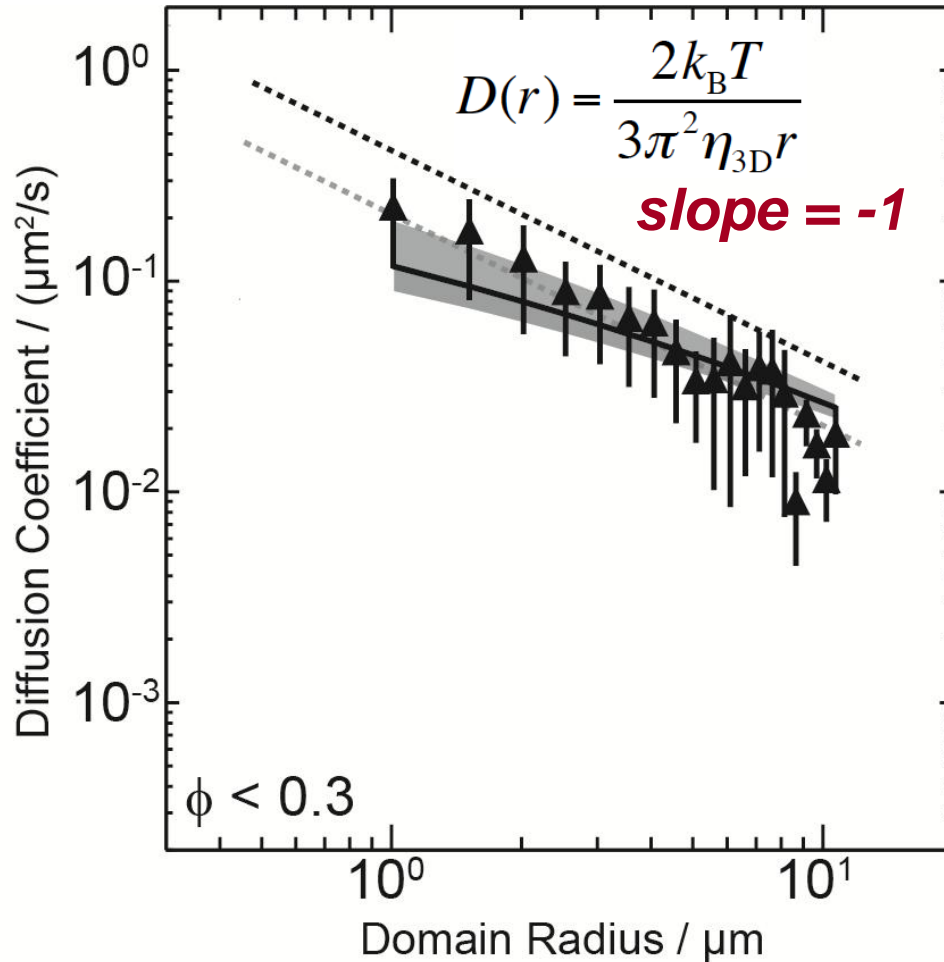
- 1) Isothermal.
- 2) Vesicles are huge (80-250 $\mu\text{m}$ ) to maximize flat area.
- 3) Vesicles are free-floating.
- 4) Rotating vesicles are deleted (17 retained out of >100).
- 5) Domains do not bulge out of the spherical surface.
- 6) Deletion of merging domains.
- 7) Parameter choice to avoid under-sampling and errors due to domains diffusing in and out of view.

$$\text{Normalized length } R = \frac{\text{Area of minority phase in view}}{\text{Perimeter of all domains in view}}$$

- 8) Projection from 2D picture to 3D surface.



# When domains are circular, $D \sim 1/\text{radius}$



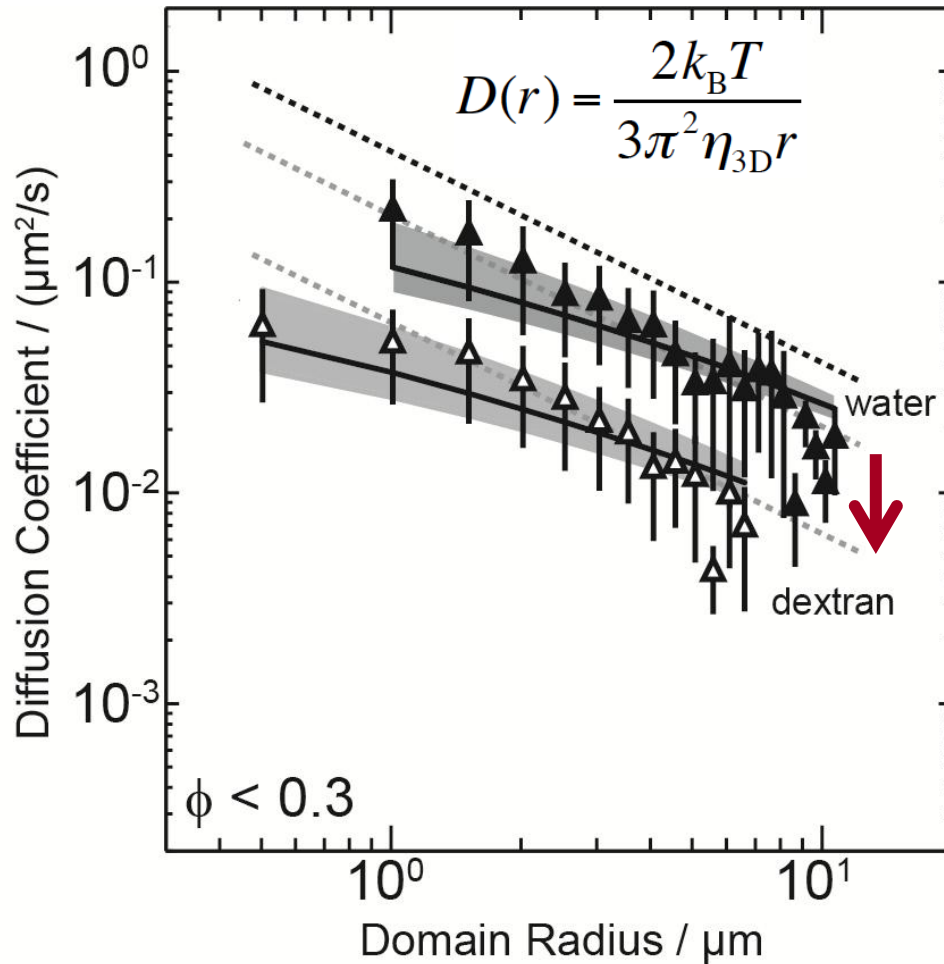
- Qualitatively,  $D \sim 1/r$

- Quantitatively, must fit through approximation.

Membrane viscosity found by the fit consistent with previous measurements.

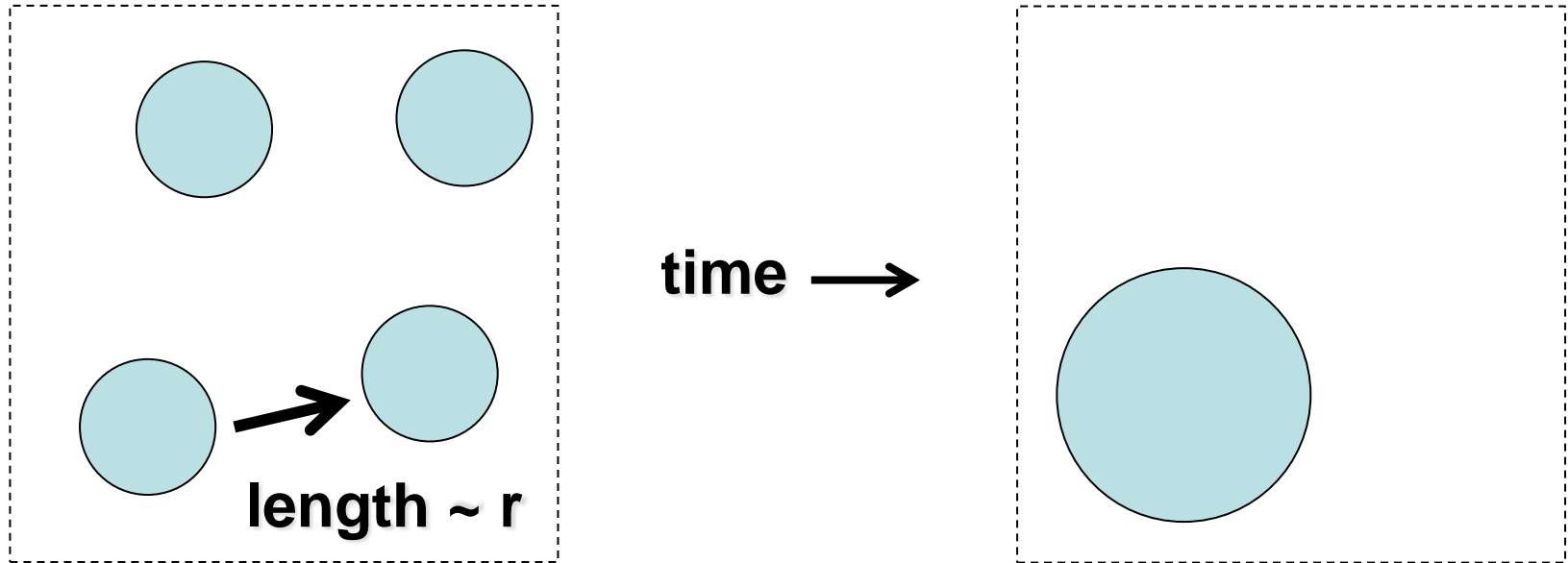
$\eta_{2D}$ (Pa s m)	Reference
$(3.3 \pm 1.1) \times 10^{-9}$	Stanich et al. 2012
$(6.0 \pm 1.5) \times 10^{-9}$	Honerkamp-Smith et al. 2012
$(4 \pm 1) \times 10^{-9}$	Camley et al. 2010

# Prediction: Increasing bulk viscosity slows diffusion



*Divide by 3.3*

We've just learned that  $D \sim 1/\text{radius}$ .  
Let's say that domain radius grows as  $r \sim t^\alpha$ . What is  $\alpha$ ?



Assume domains grow entirely by collision/coalescence.

Brownian motion:  $\text{length}^2 \sim Dt$

So  $\text{radius}^2 \sim Dt$ .

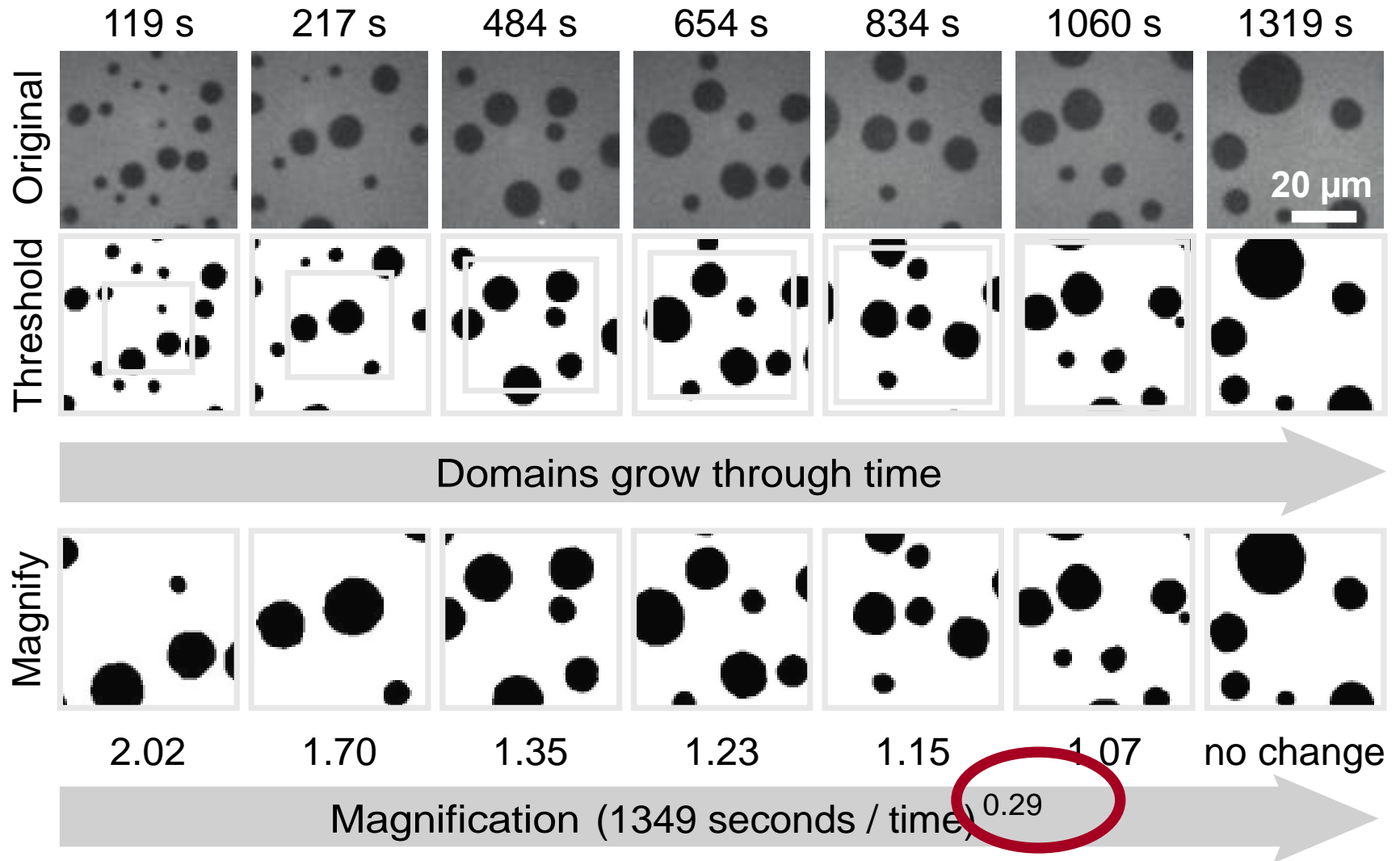
When  $D \sim 1/\text{radius}$ ,  $\alpha = 1/3$ .

## Simulations with $\phi \leq 0.3$

$\alpha$	SIMULATION	REFERENCE
0.3	Dissipative particle dynamics. $\phi = 0.3$	Taniguchi 1996
1/3	Purely dissipative dynamics. $\phi = 0.3$	Laradji et al. 2004
0.31	200 spherical caps on a vesicle. $\phi = 0.09$	Putzel (Northwestern U.)
1/3	Continuum approach with hydrodynamics	Fan et al 2010
1/3	Stochastic phase field model + hydrodynamics	Camley et al. 2010

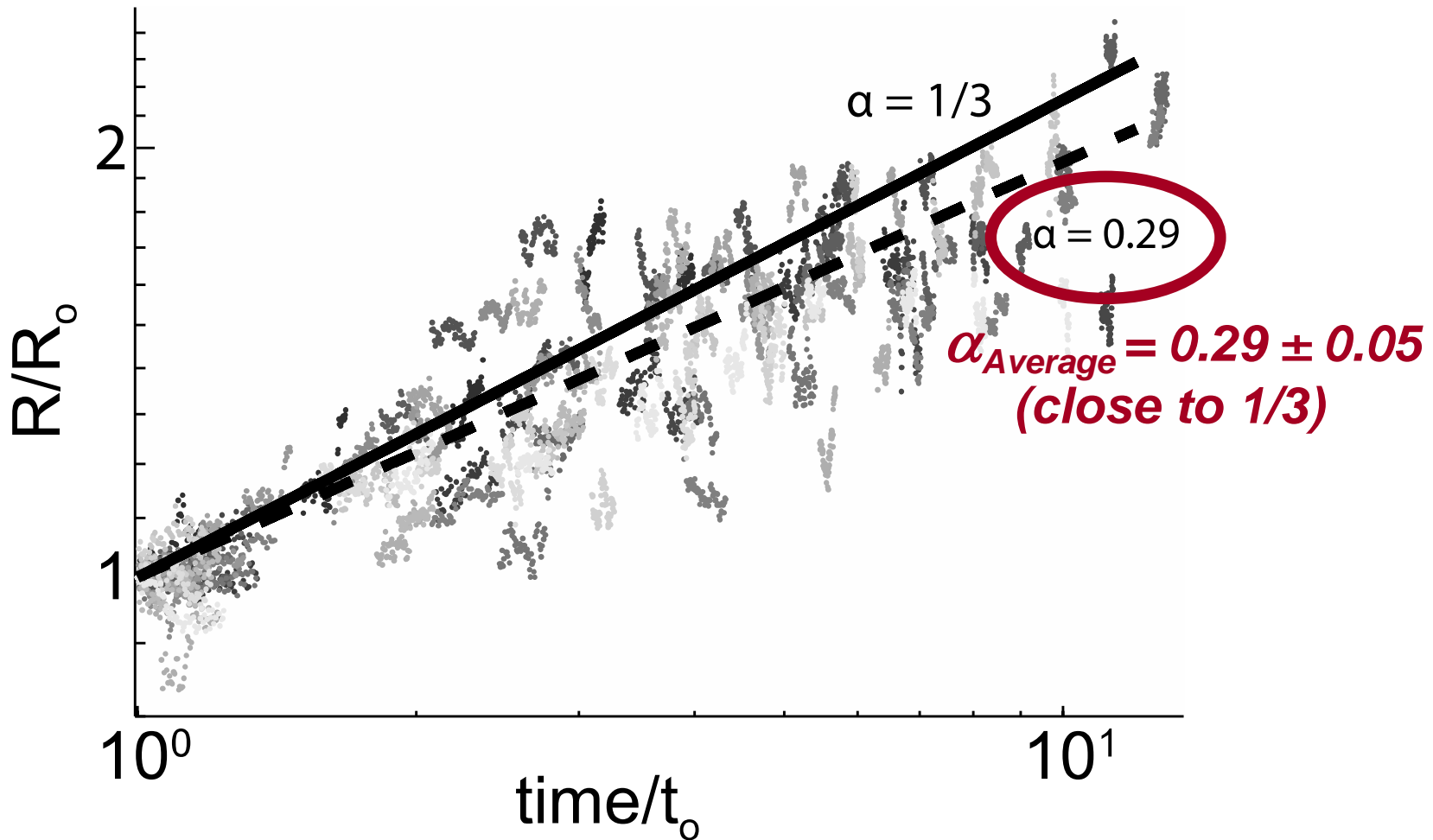
When  $D \sim 1/\text{radius}$ ,  $\alpha = 1/3$ .

# Yes, for circular domains, $\alpha \approx 1/3$ (only one experiment here)

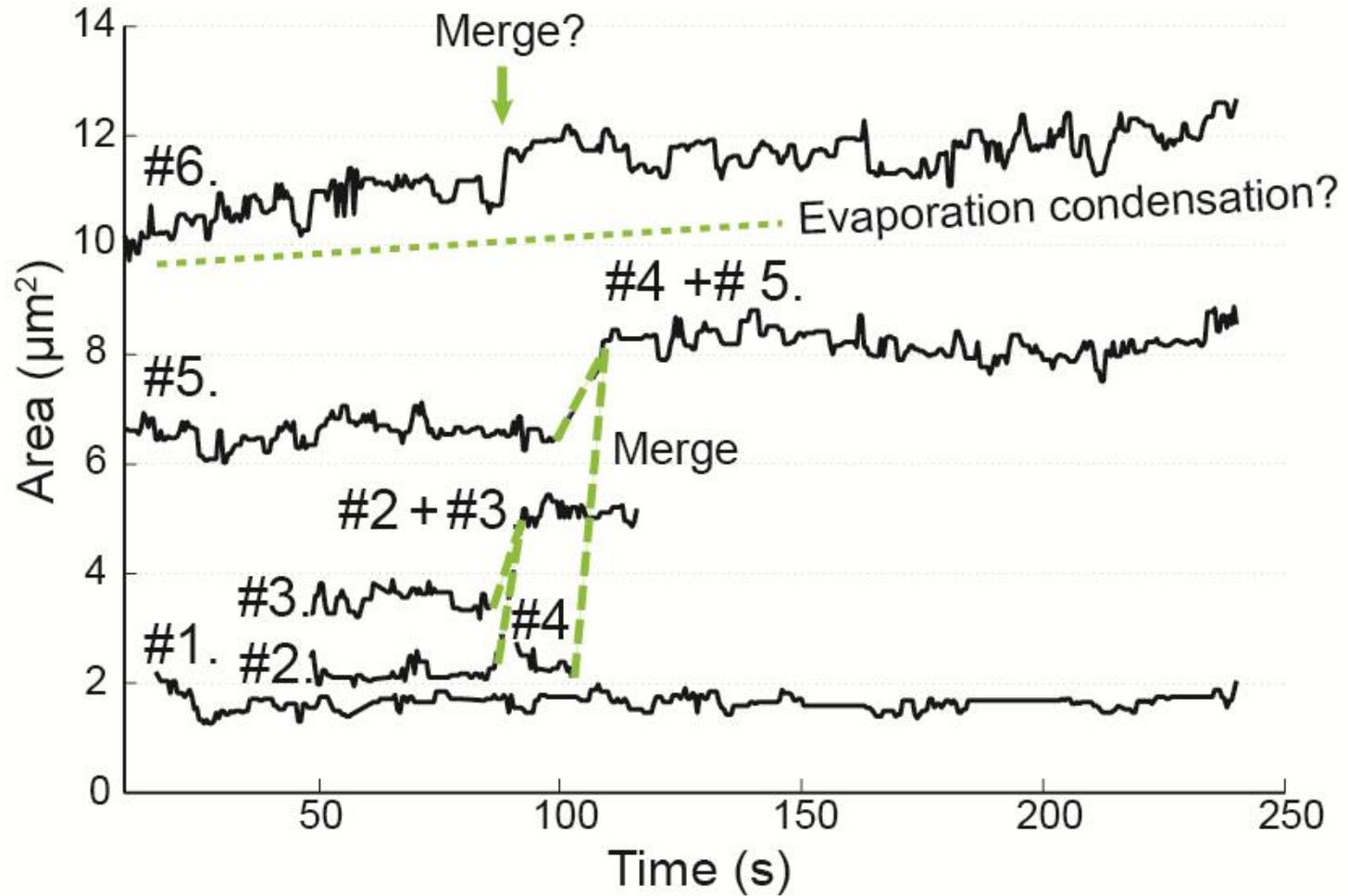


**$\alpha = 0.29$ , close to  $1/3$**

Yes, for circular domains,  $\alpha \approx 1/3$  (all 17 experiments here)



# Coarsening of domains appears to be dominated by merges (not by evaporation-condensation / Ostwald ripening)





**What happens when area fraction ( $\phi$ ) is close to  $\frac{1}{2}$ ?**

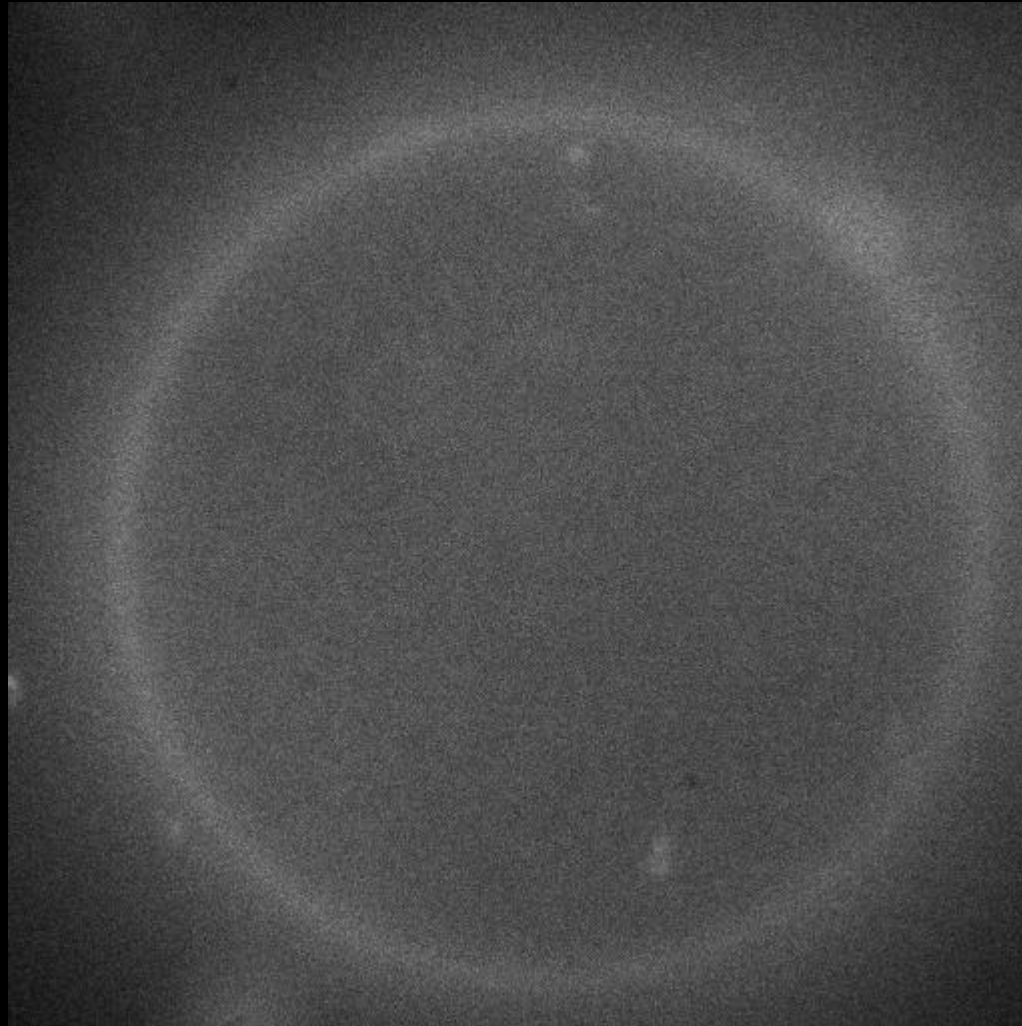
**In other words, what happens after a quench  
through a critical point?**

(Note: Isothermal critical point, or plait point)

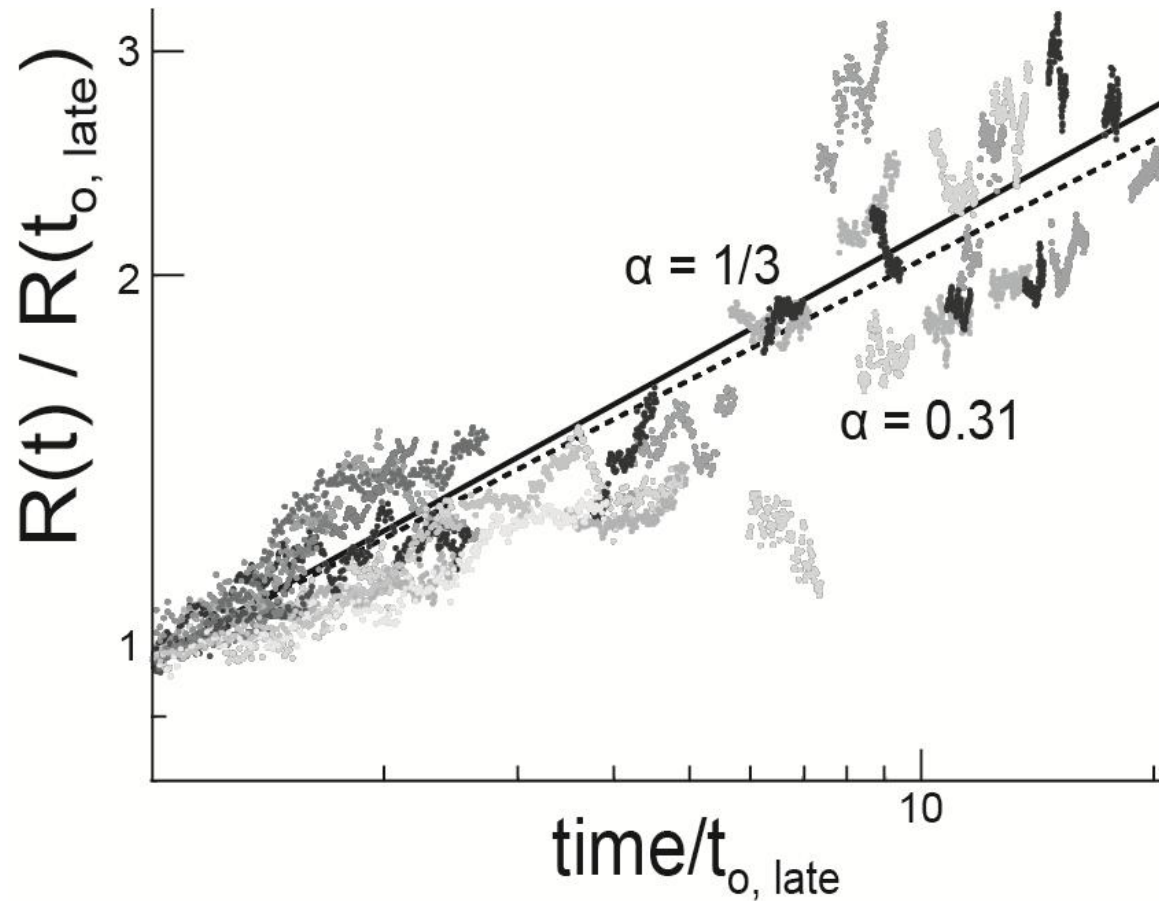
# $0.4 \leq \text{Area Fraction} \leq 0.6$

Early (temp stabilizing), Intermediate (noncircular), Late (circular)

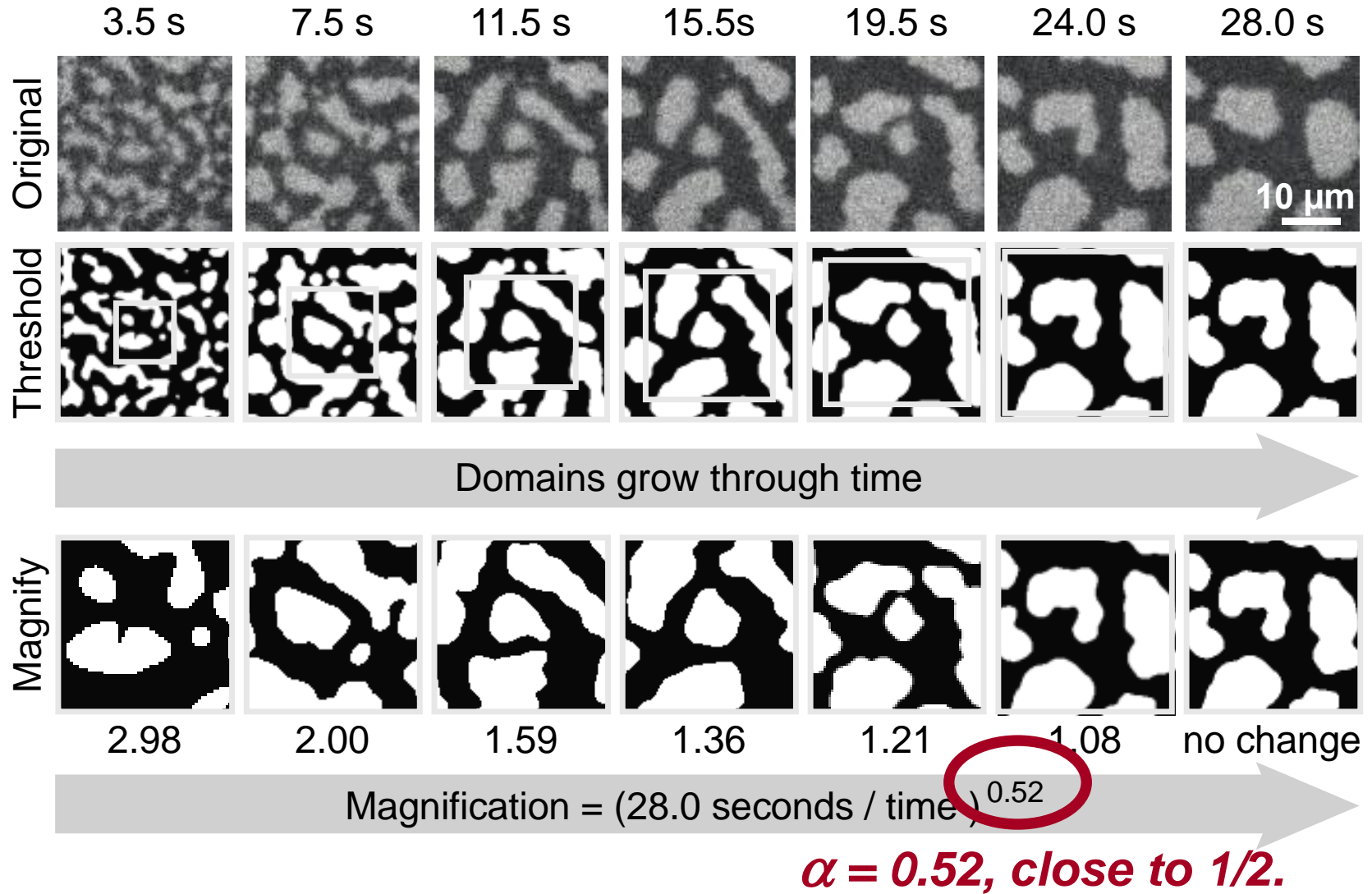
25  $\mu\text{m}$



Late times,  $0.4 \leq \phi \leq 0.6$ , 10 experiments,  $\alpha = 0.31 \pm 0.05$

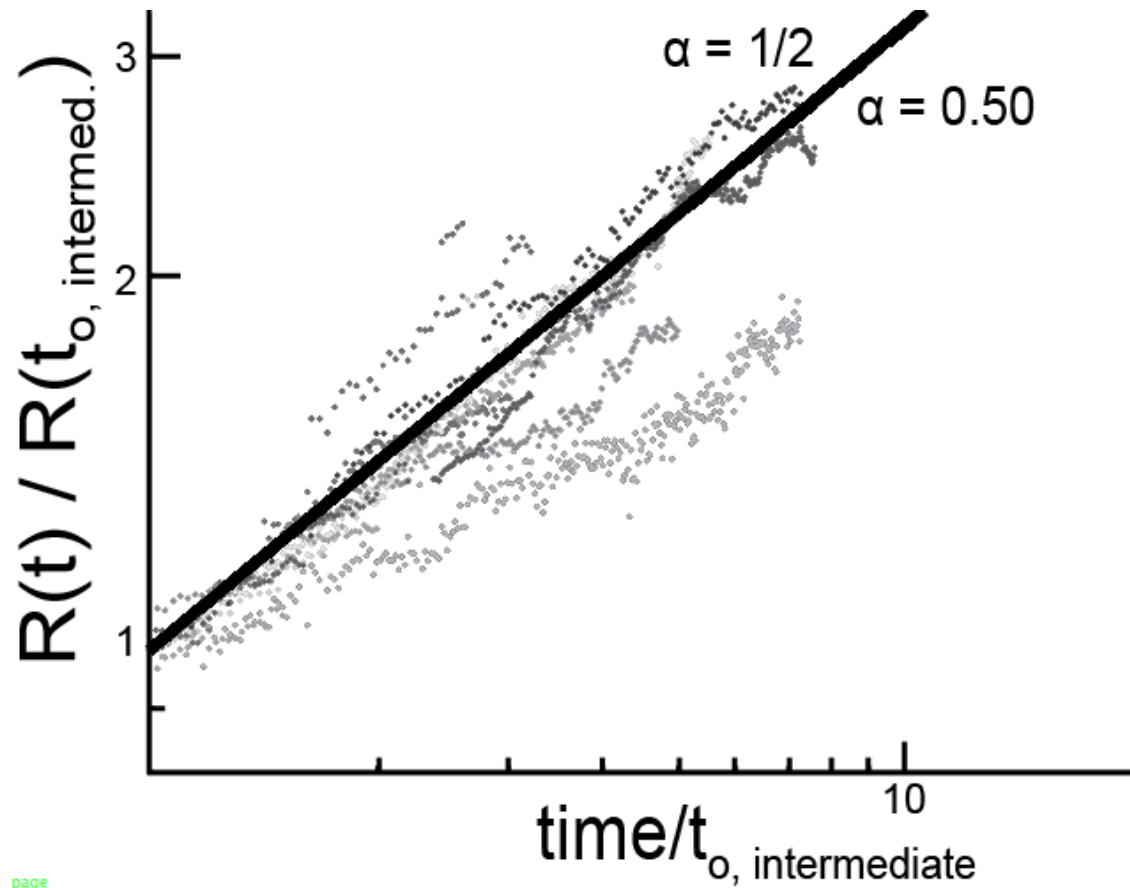


# Intermediate times, $0.4 \leq \phi \leq 0.6$ , 8 expts, $\alpha = 0.50 \pm 0.16$



*Consistent with simulations by Ramachandran et al. 2009, Laradji et al. 2005, Fan et al. 2010, and Camley et al. 2011.*

# Intermediate times, $0.4 \leq \phi \leq 0.6$ , 8 expts, $\alpha = 0.50 \pm 0.16$



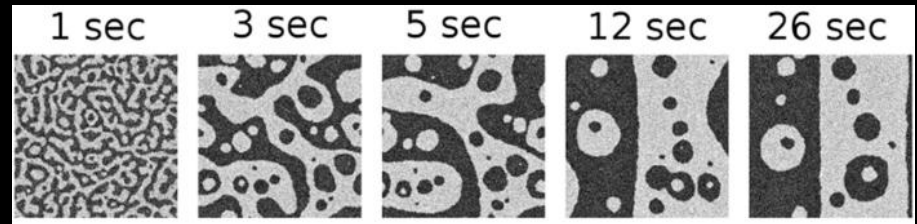
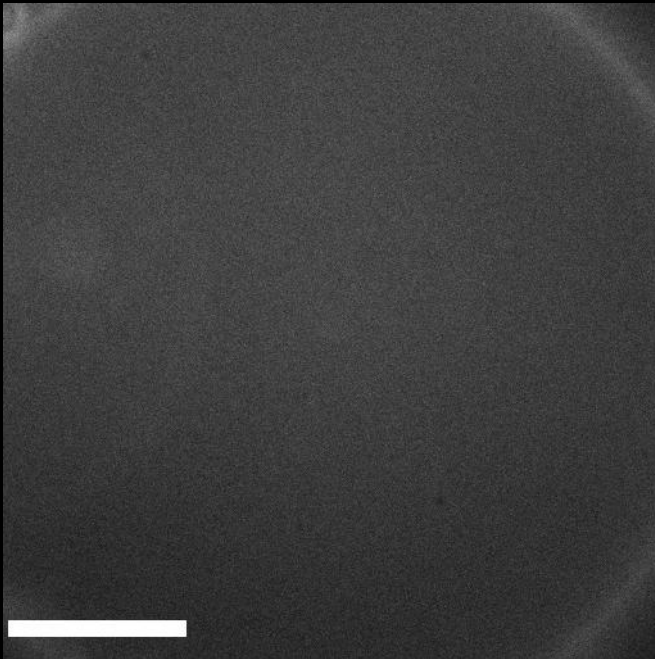
## Worth noting...

**Observing different exponents at different times means scaling is only apparent, not universal.**

This is seen in simulations by:  
Fan, Han, and Haataja (2010)  
Camley and Brown (2011)

Both groups say that if experiments see any apparent scaling in a membrane with noncircular domains near a critical point, then  $\alpha$  should be  $\sim 1/2$ .

# Example in which no scaling is observed:



Compare to simulation  
by Camley and Brown 2011

## Conclusion from this section:

- Qualitatively, micron-scale domains in membranes of our composition diffuse with  $D \sim 1/r$ .
- Quantitatively, we find membrane viscosities that agree with values found by other methods.
- Circular domains coarsen with a growth exponent of  $\alpha \approx 1/3$ .
- Noncircular domains coarsen with an apparent growth exponent of  $\alpha \approx 1/2$ .
- The hydrodynamic length scale,  $L_h$ , holds the key to understanding a range of membrane phenomena.



# Thank You

## ***Data from current group members***

*Cynthia Stanich (coarsening), Matt Blosser,  
Ben Horst, Joan Bleecker, Thomas Portet*

## ***Data/Code from group alumnae***

*Aurelia Honerkamp-Smith (U. Cambridge), Sarah Veatch (U Michigan)*

## ***Collaborations***

*Ben Machta (Cornell - this project), Pietro Cicuta (U. Cambridge),  
Michael Schick (U. Washington), Marcel den Nijs (U. Washington),  
Ray Goldstein (U. Cambridge), Sharona Gordon (U. Washington)*

## ***Funding for this project***

*NSF MCB-0744852*

## ***Partner***

*Rob Carlson*

*If you are interested in pursuing graduate or postdoctoral research in my lab,  
please contact me and let's scheme about funding.*

# Postdoctoral Fellowships in Biophysics University of Washington, Seattle

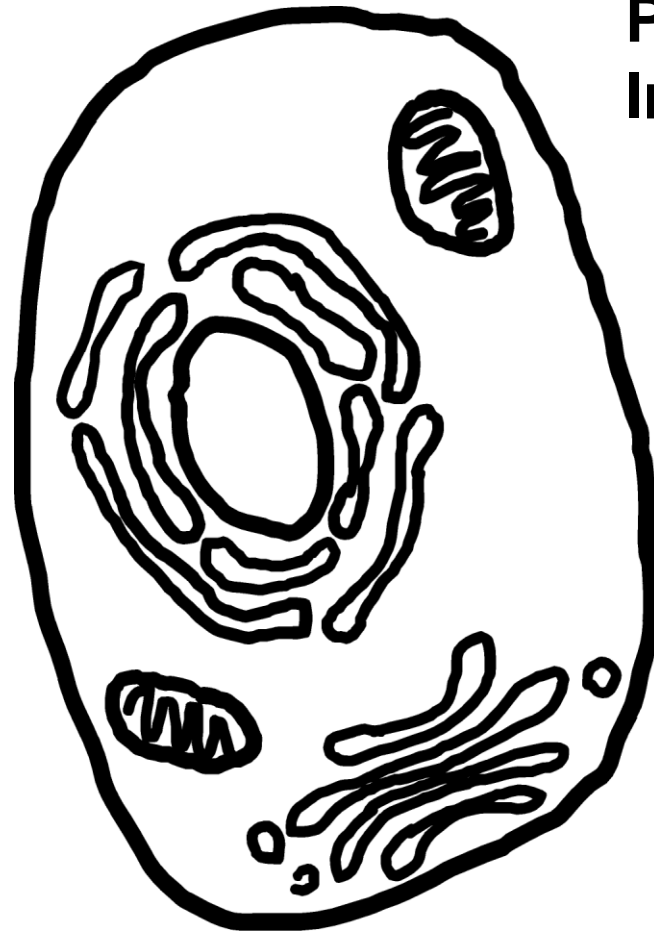
*<http://depts.washington.edu/pbiopage/sackler>*

## **The Raymond and Beverly Sackler Scholars Program in Integrative Biophysics**

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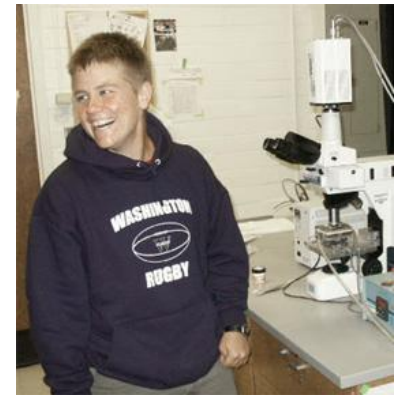
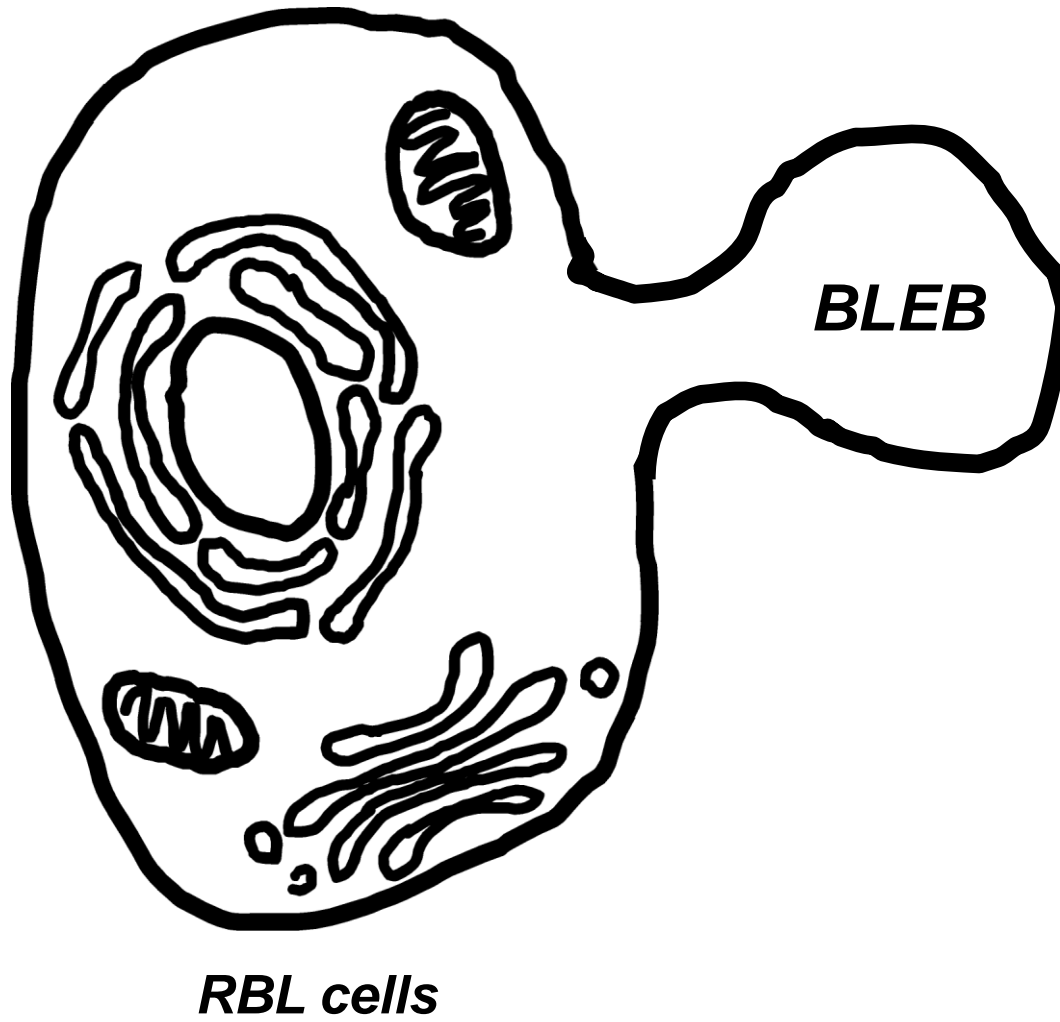
UW seeks the most outstanding early career scientists, regardless of nationality. Applications accepted on a rolling basis. Apply at *[http://depts.washington.edu/pbiopage/sackler/app\\_process.php](http://depts.washington.edu/pbiopage/sackler/app_process.php)*

# Inspiration:



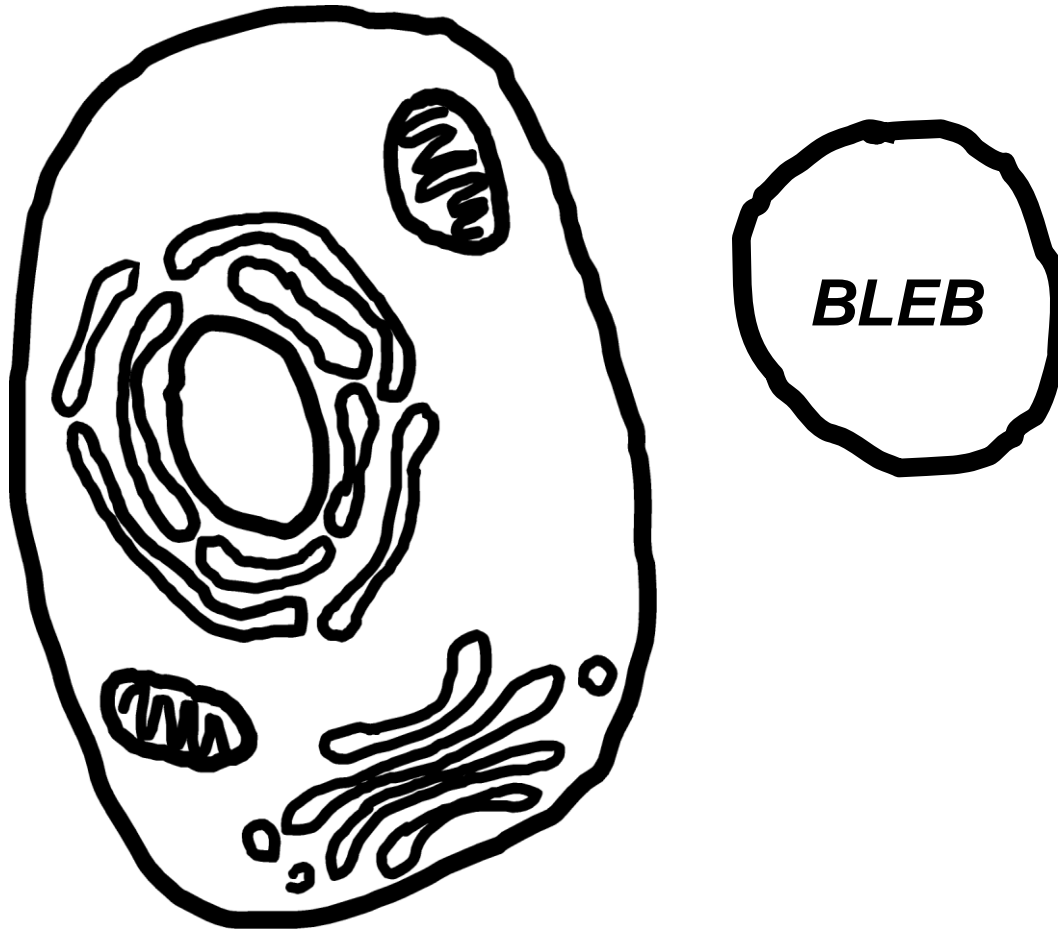
Plasma membrane  
Inhomogeneity

# Sarah Veatch's Results: In collaboration with Baird / Holowka Lab at Cornell

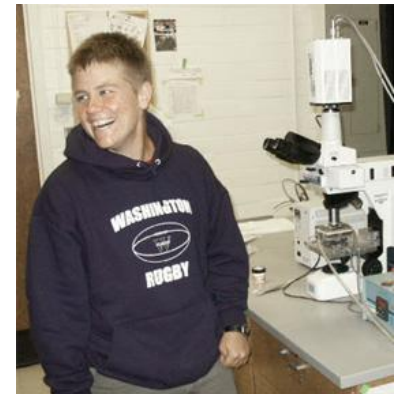


*Sarah Veatch  
Assistant Professor  
University of Michigan*

# Sarah Veatch's Results: In collaboration with Baird / Holowka Lab at Cornell



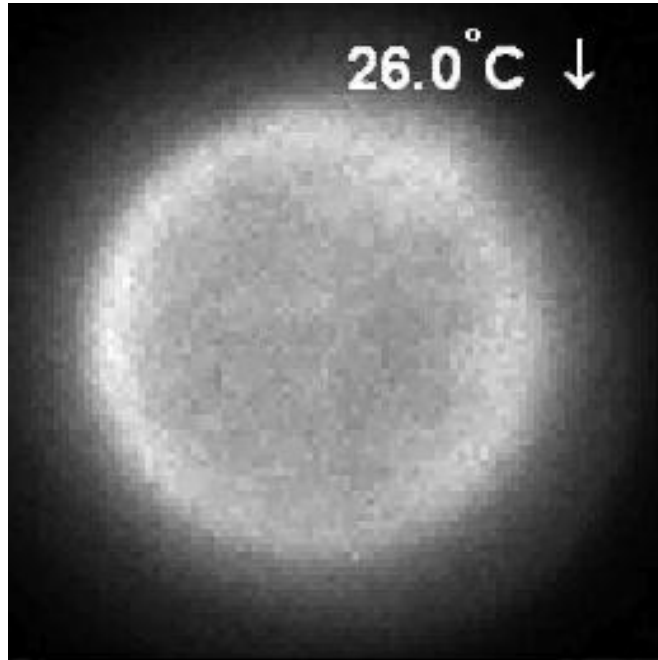
*RBL cells*



*Sarah Veatch  
Assistant Professor  
University of Michigan*

## Sarah Veatch's Results:

In collaboration with Baird / Holowka Lab at Cornell



S.L. Veatch et al.  
ACS Chem Biol (2008) 3:287

In all cases,

$$T_{Growth} > T_{Critical}$$

So, presumably, at the growth temperature, sub-micron fluctuations are indeed present in the membrane.

Pretty cool.