

Glassy behavior in biological tissues

Complexity in
Mechanics
KITP conference
UC Santa Barbara
October 22, 2014

M. Lisa Manning
Syracuse University



10 μm

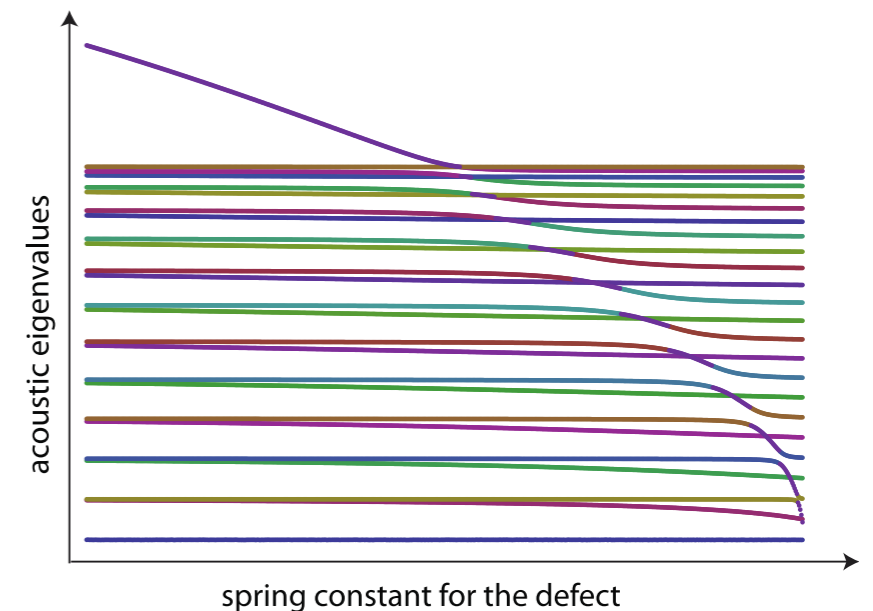
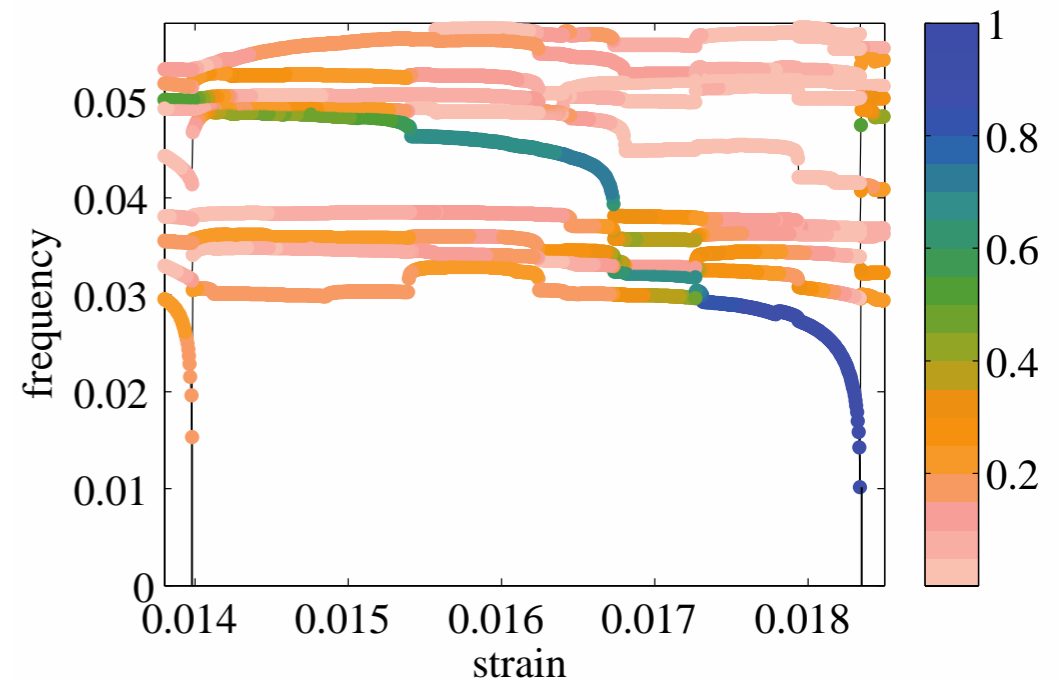
Defects and vibrational modes in disordered solids

- Eigenfunctions are not as useful away from instability
- Instead, find “flow defects” or “soft spots” or “STZs”
- A new and improved algorithm for identifying truly localized soft spots and energy barriers
- long-ranged elastic tails generate lower energy barriers
- A random matrix definition of the boson peak

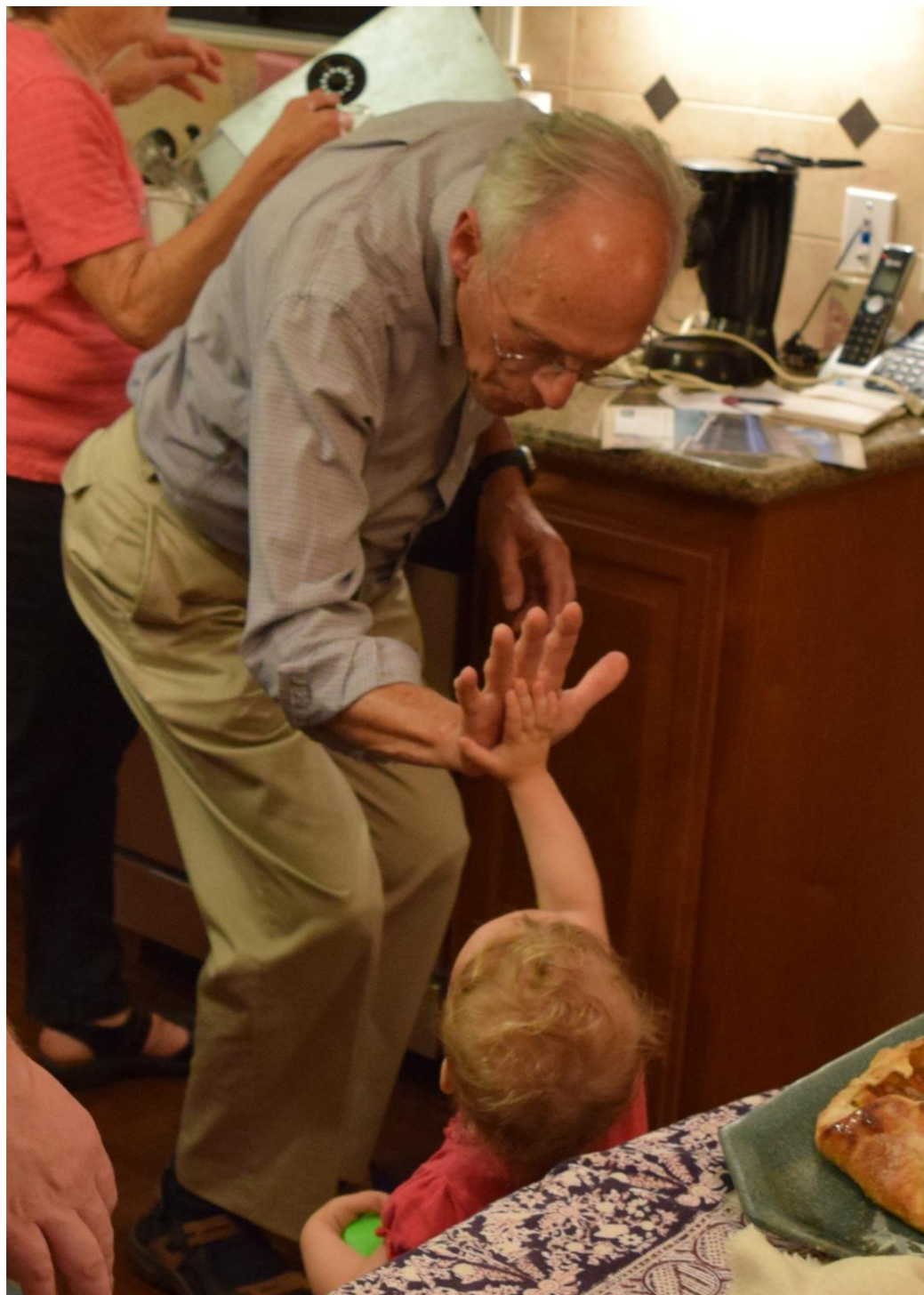
[Recorded KITP talk on this stuff](#)

<http://online.kitp.ucsb.edu> Avalanches

(Oct 10th)



Thanks, Jim!



Manning group

Dapeng “Max” Bi

Giuseppe Passucci

Sven Wijtmans

Craig Fox

Contact: mmanning@syr.edu



SU Soft Matter

Jorge Lopez

Jennifer Schwartz

Cristina Marchetti

Xingbo Yang



Amack group (SUNY Upstate
Medical)

Jeff Amack

Guangliang Wang

Agnik Dasgupta

Schoetz lab and affiliates

Eva-Maria Schoetz (UCSD)

Marcus Lanio (Princeton)

Jared Talbot (Princeton)

Ramsey Foty (Princeton)

Mal Steinberg (Princeton)

Henderson Group
(Syracuse Biomaterials
Institute)

Jay Henderson

Megan Brasch

Richard Baker

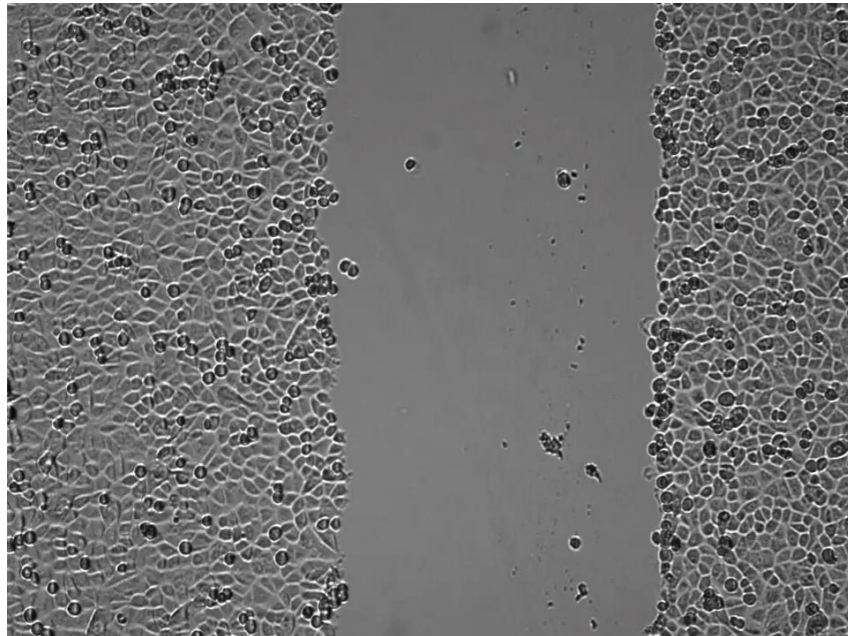
Turner Group (SUNY
Upstate)

Chris Turner

Nick Deakin

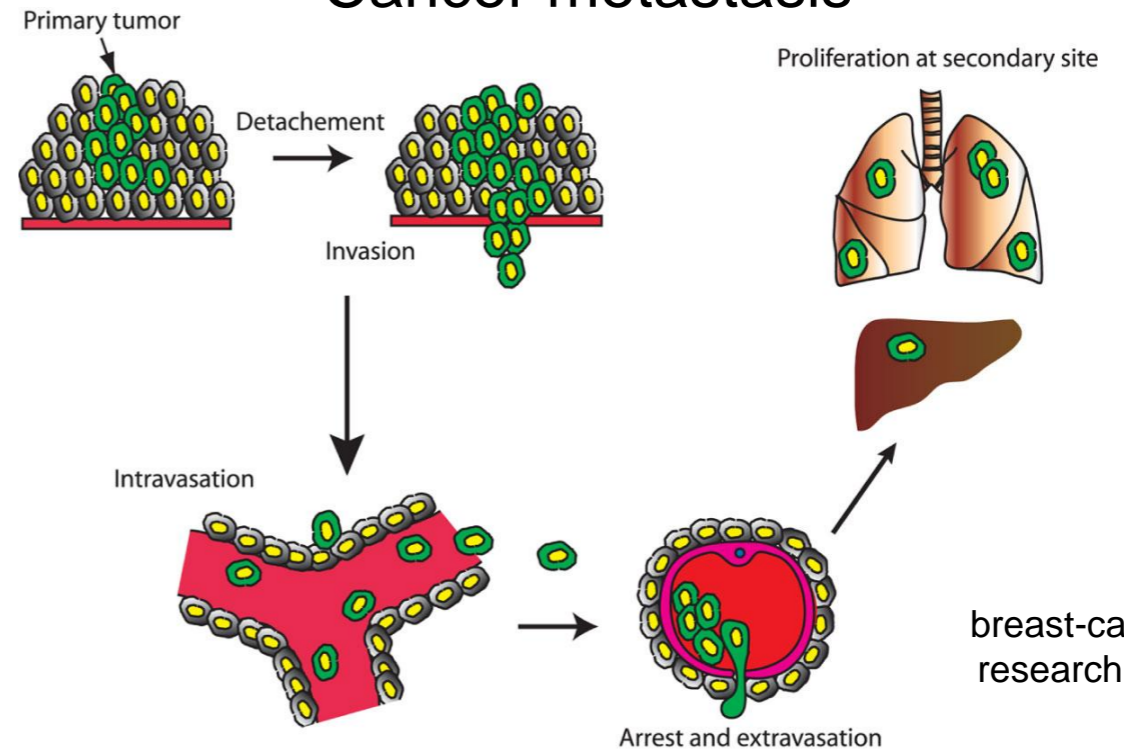
Complexity in mechanics....

Wound healing



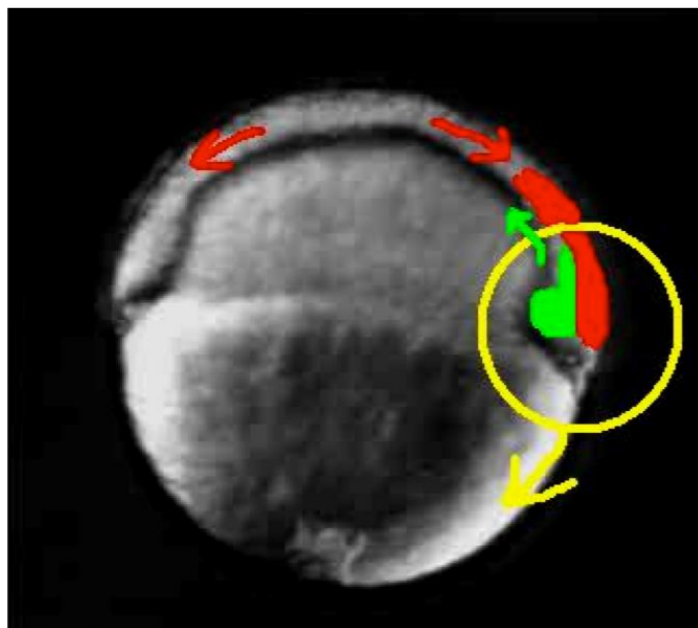
[youtube.com/watch?v=v9xq_GiRXeE](https://www.youtube.com/watch?v=v9xq_GiRXeE)

Cancer metastasis

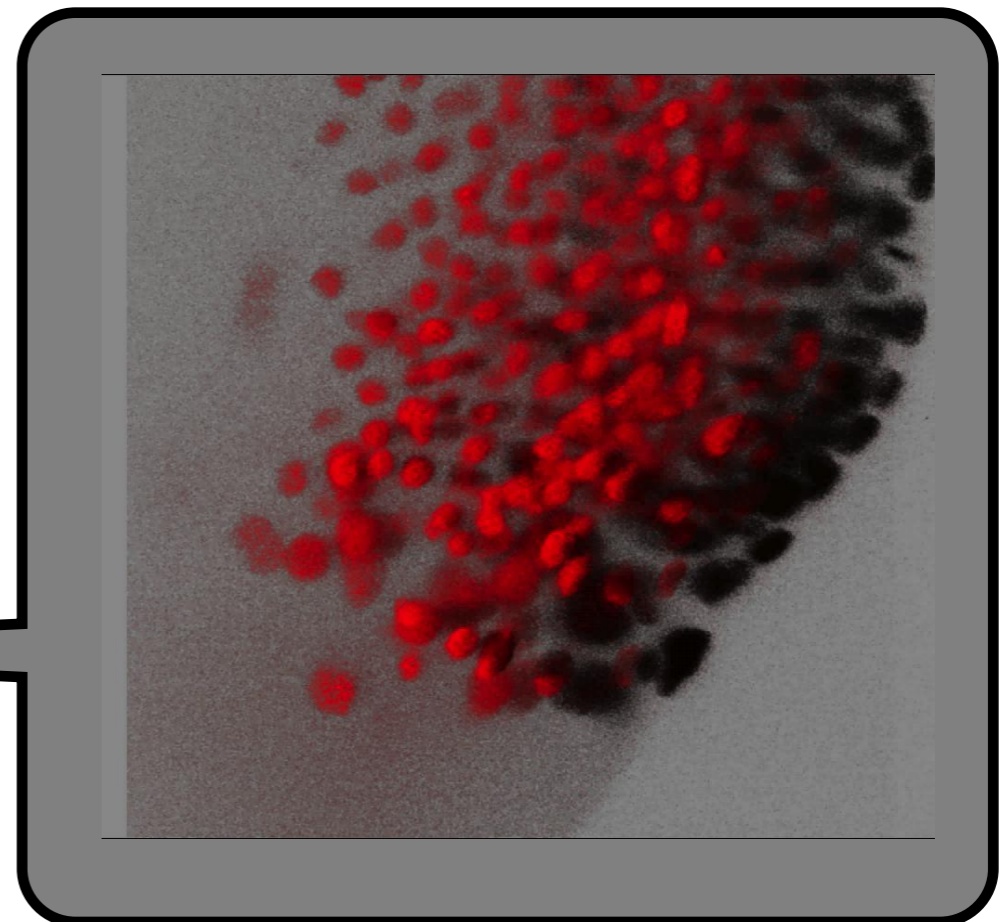


Embryonic development

Differential Adhesion Hypothesis:
Steinberg, Science 1962



Schoetz 2008

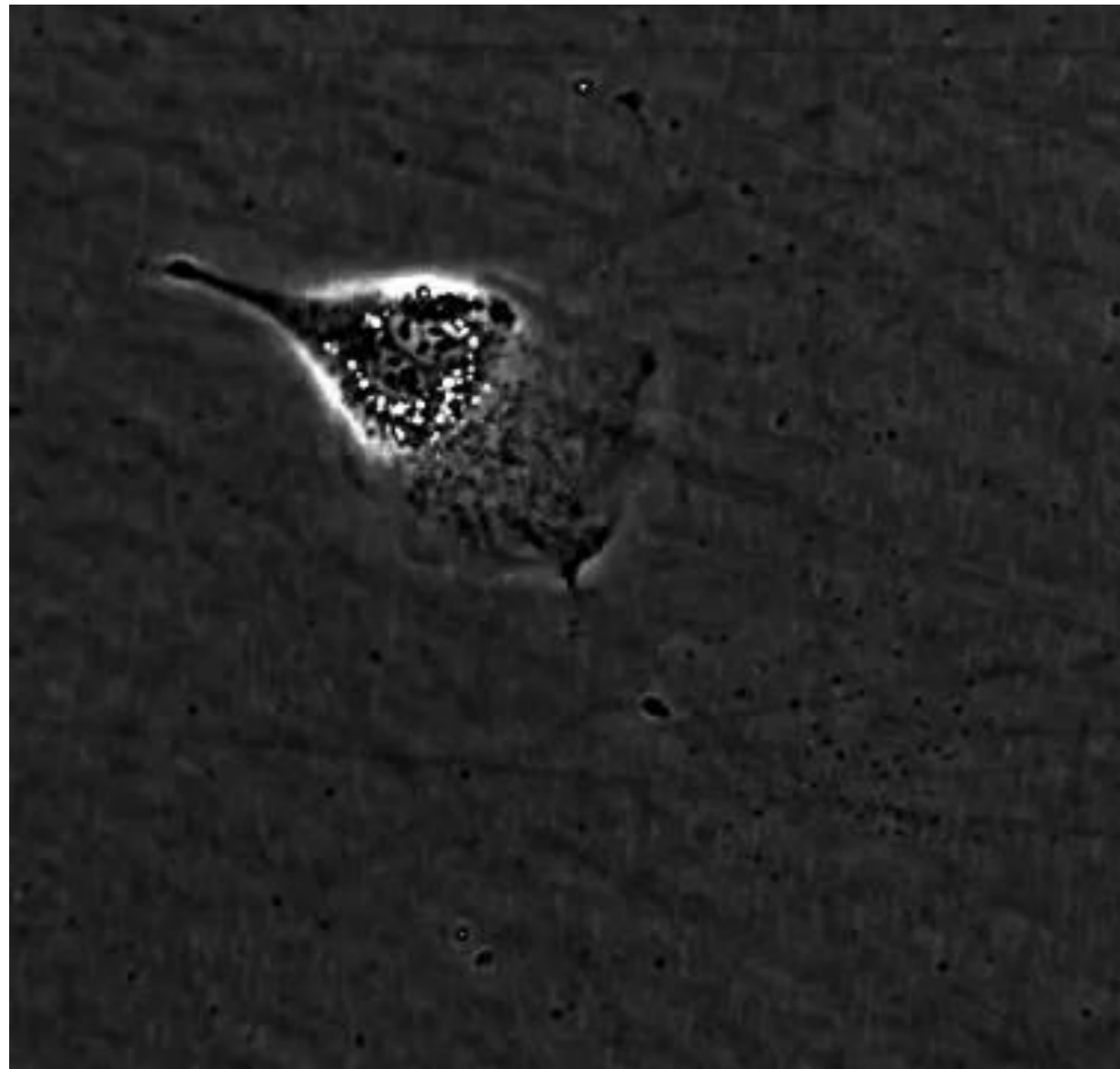


How do cells move?

In isolation

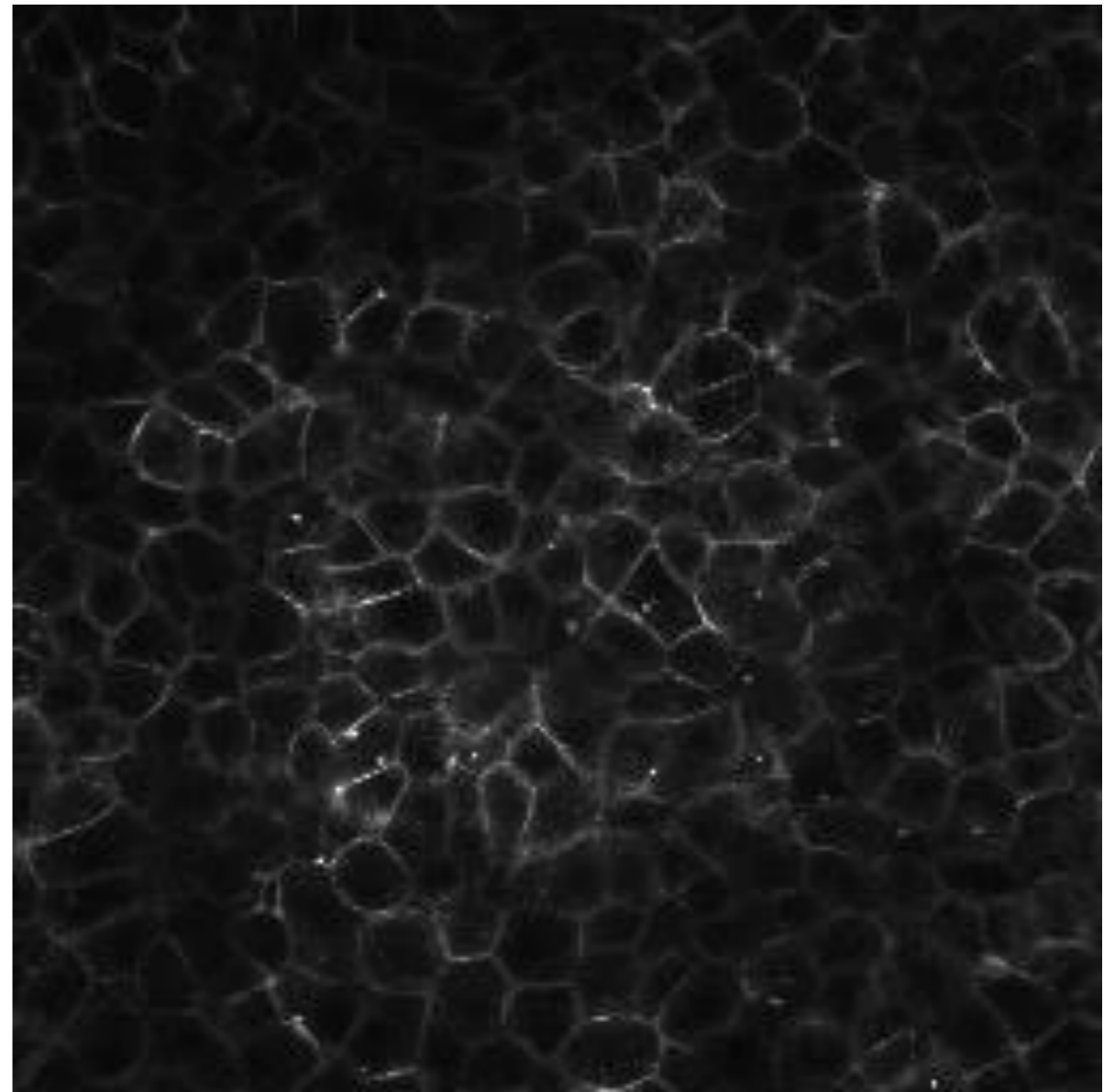
vs.

in dense tissue



<https://www.youtube.com/watch?v=FUIDCfFCTto>

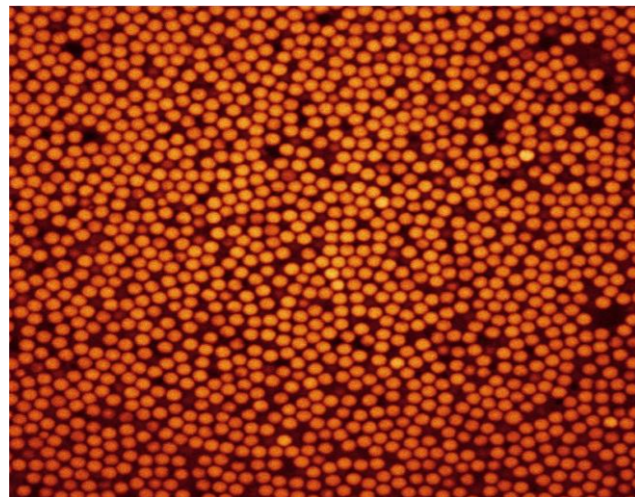
**Human bone cancer cell
on fibronectin**



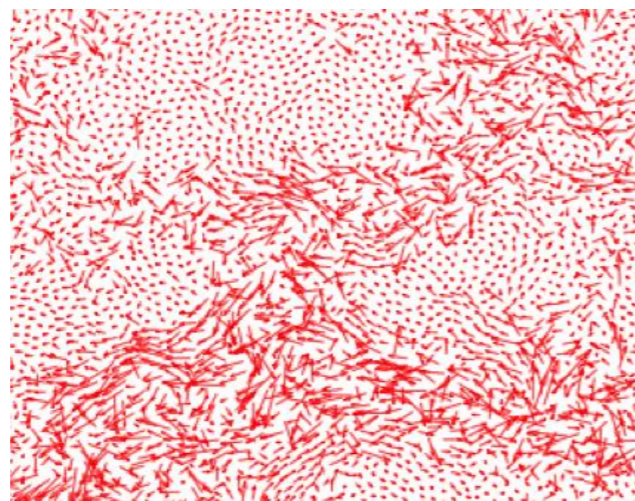
**Zebrafish embryo
Schoetz Lab, UCSD**

Dense (non-active) materials

Thermal

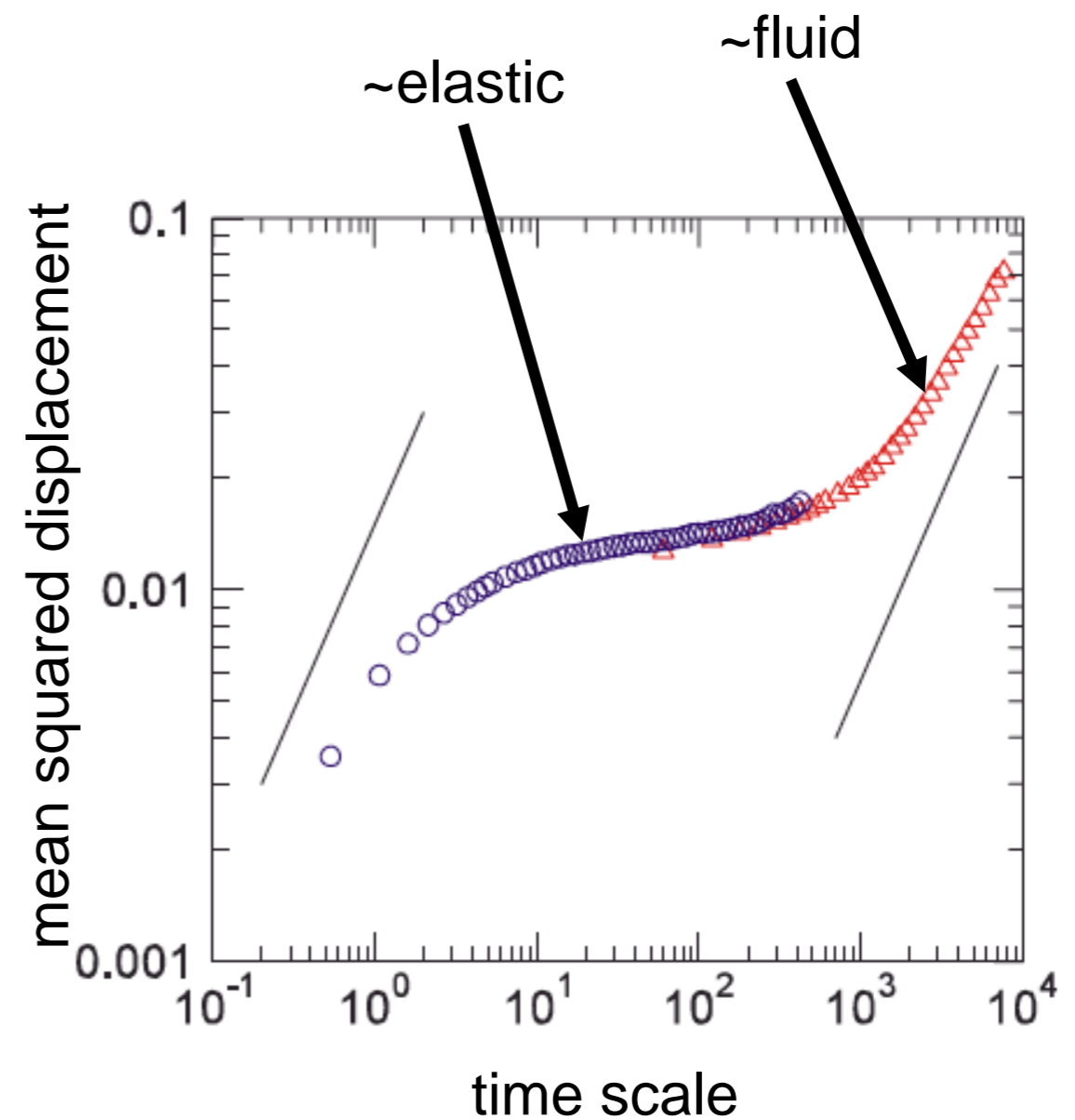


A colloidal glass.



Displacement profile in simulation of a 2-d glass former.

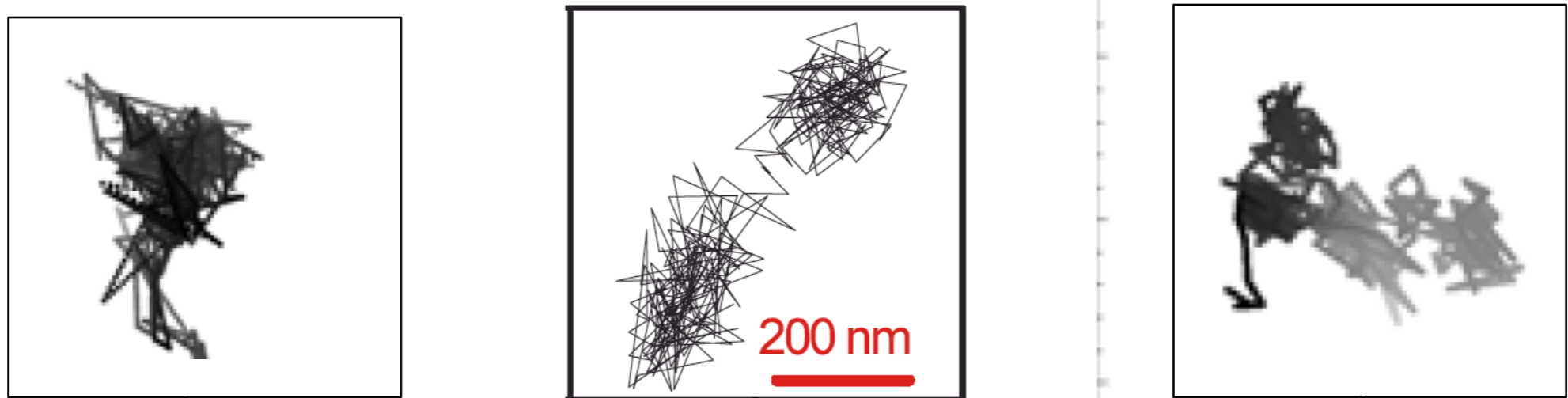
Berthier PRL 2011



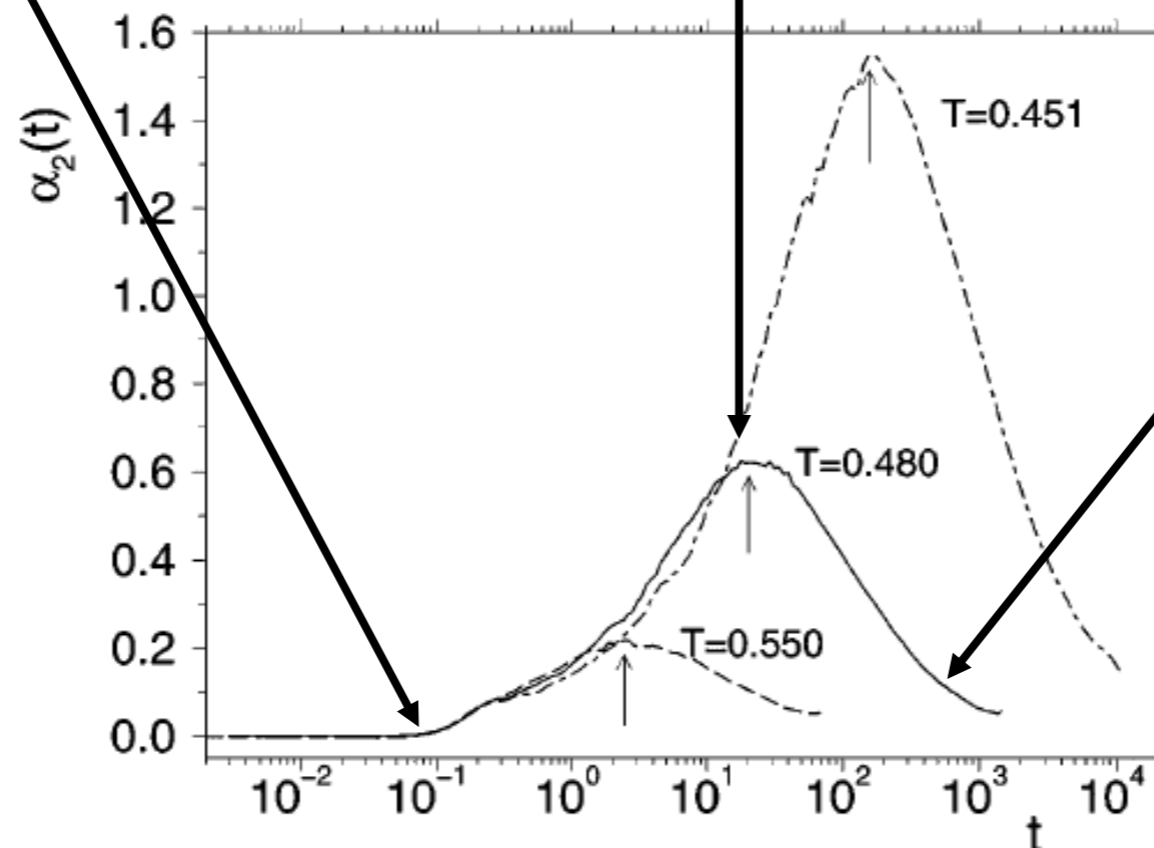
Why do mechanical properties change with timescale?

non-active materials

Weeks, Crocker, Weitz (2004)

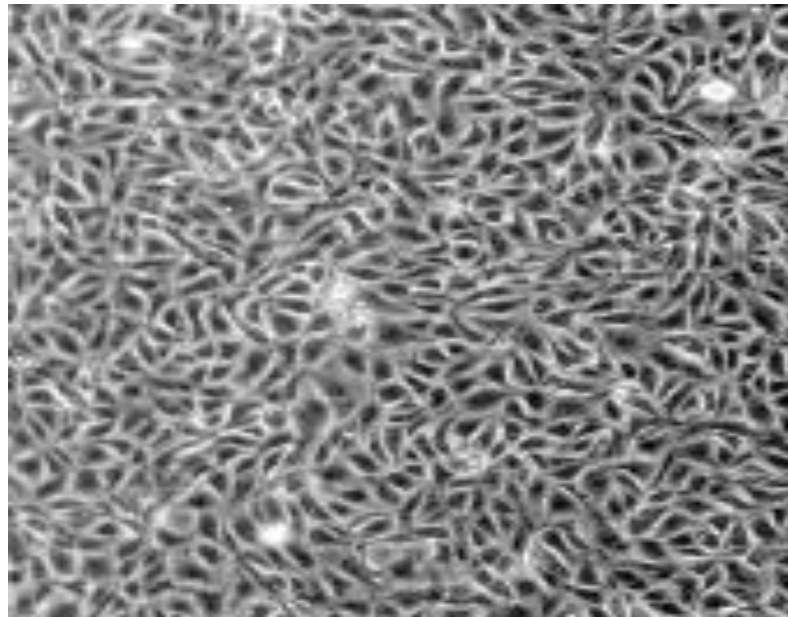


Caging behavior measured by non-gaussian parameter

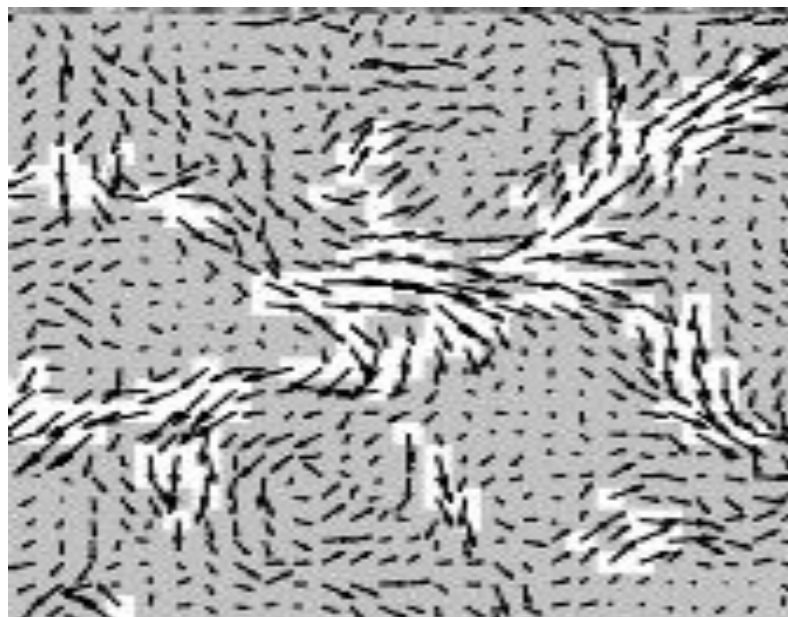


Kob et al, PRL 79 15 2827 (1997)

Dense biological tissues show the same features

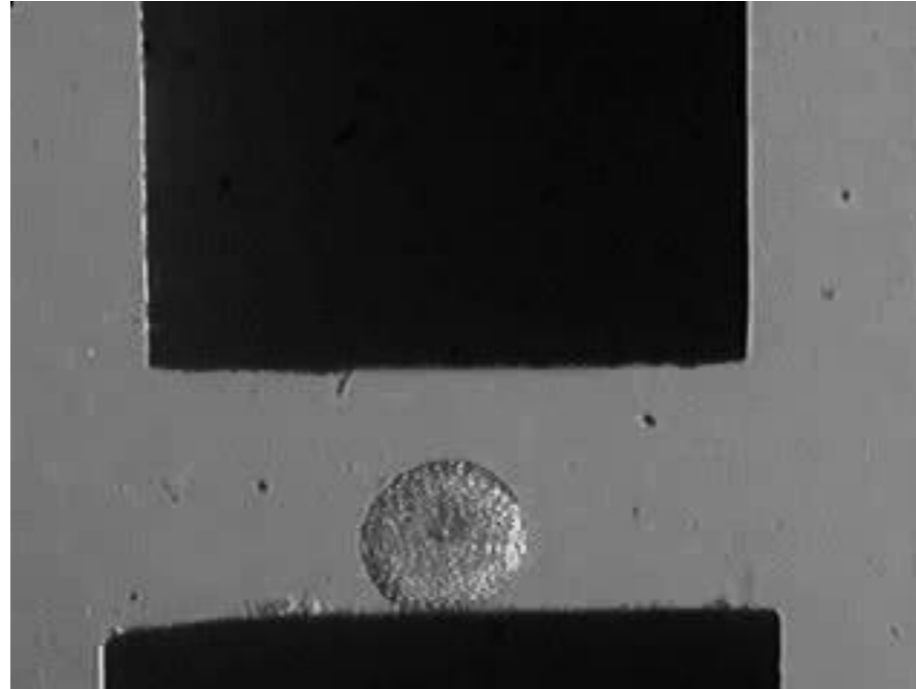


Madin-Darby canine kidney (MDCK) cells forming a 2-d confluent layer.
[Angelini et al PNAS 2010](#)

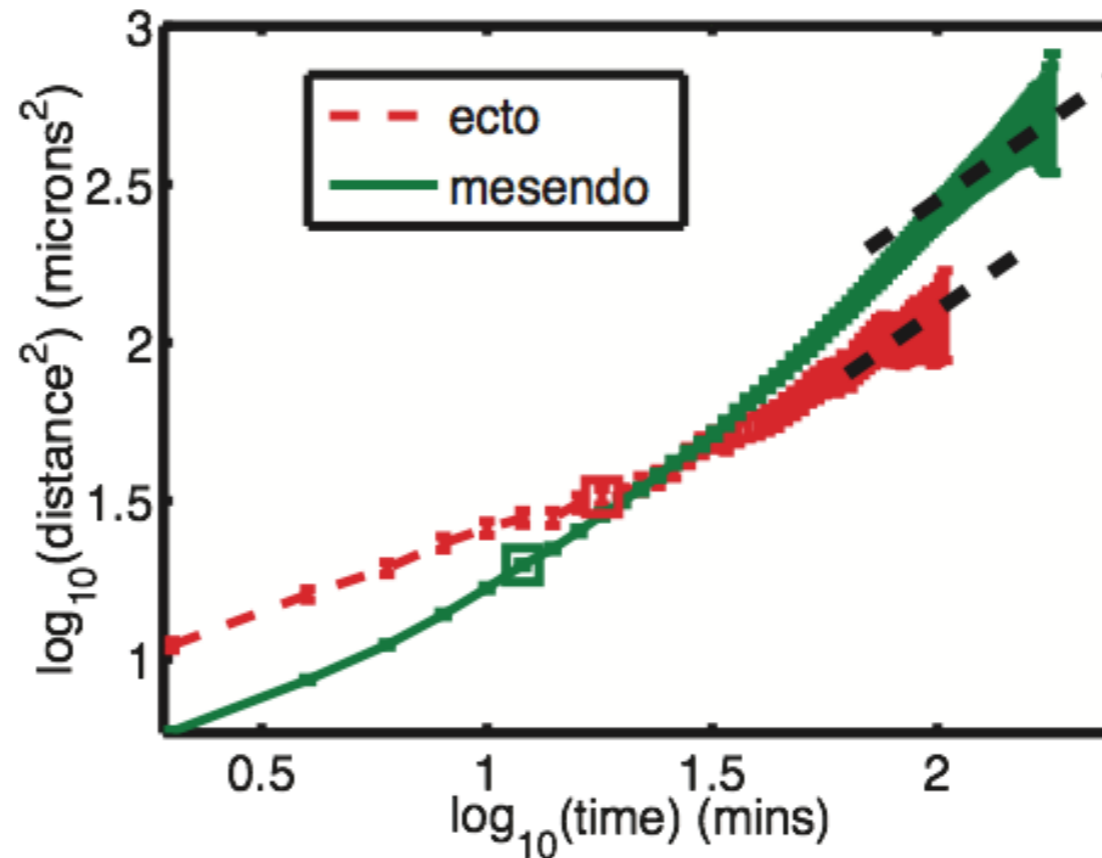
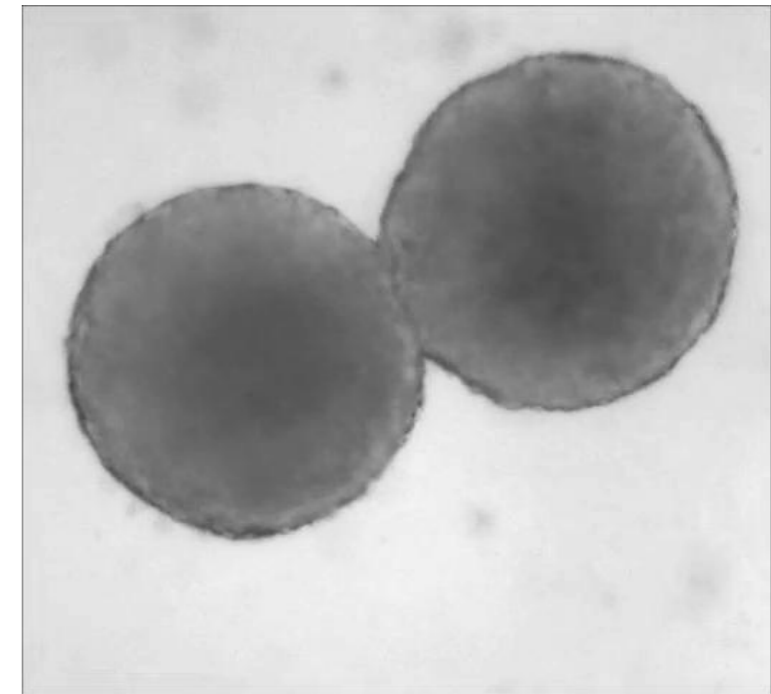


Velocity profile of cells show the spatially heterogeneous pattern in MDCK tissue.

timescale ~ seconds

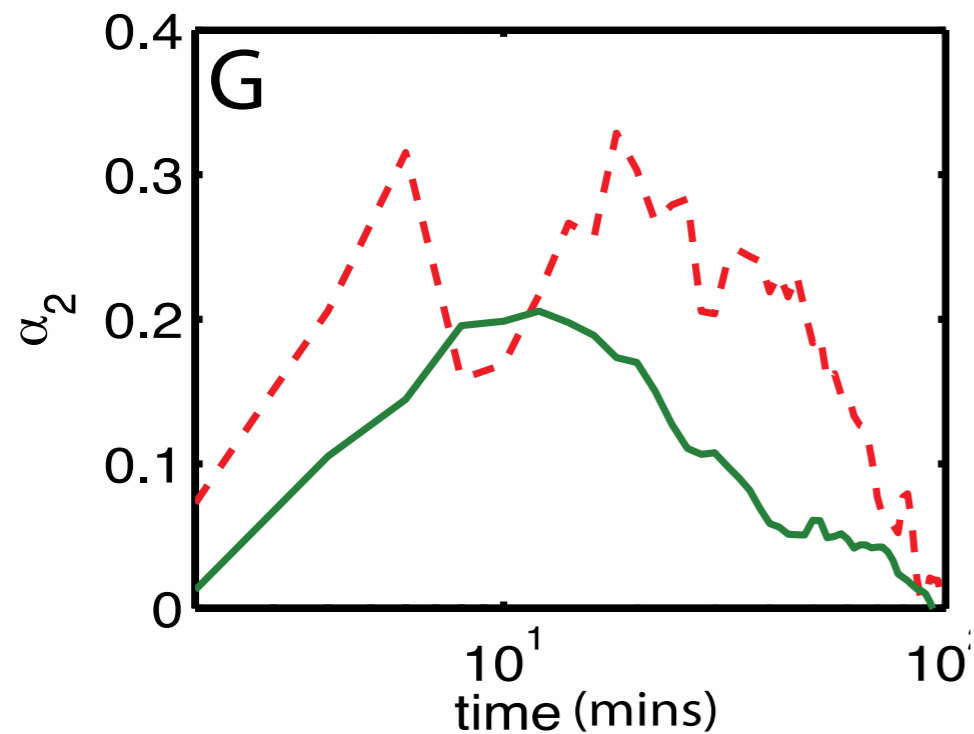
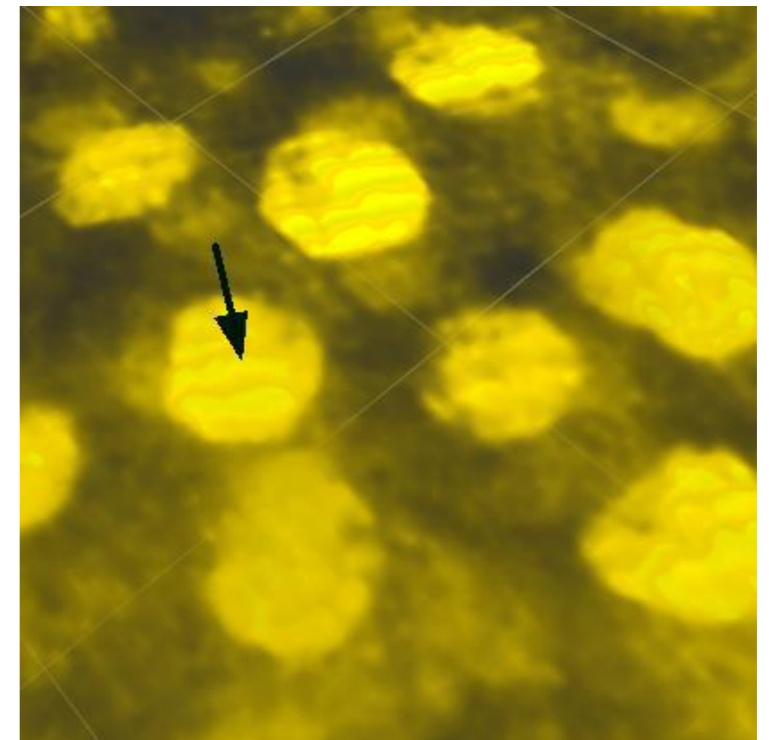
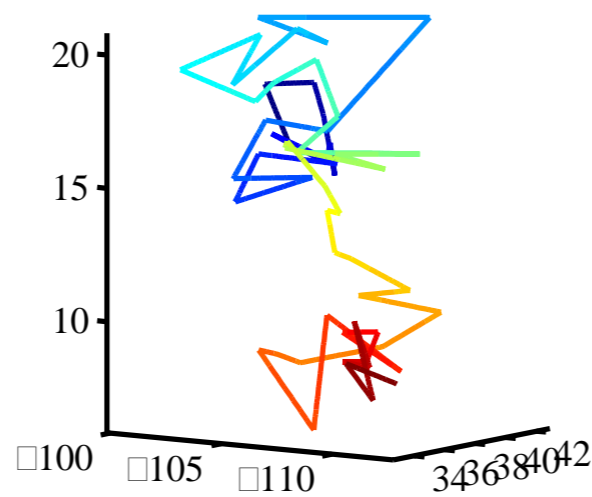
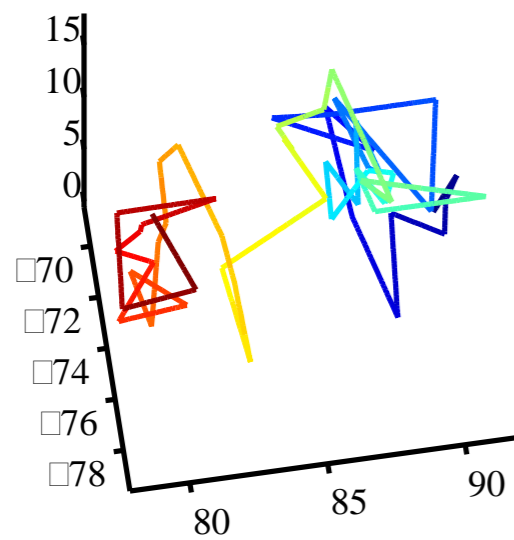


timescale ~ hours



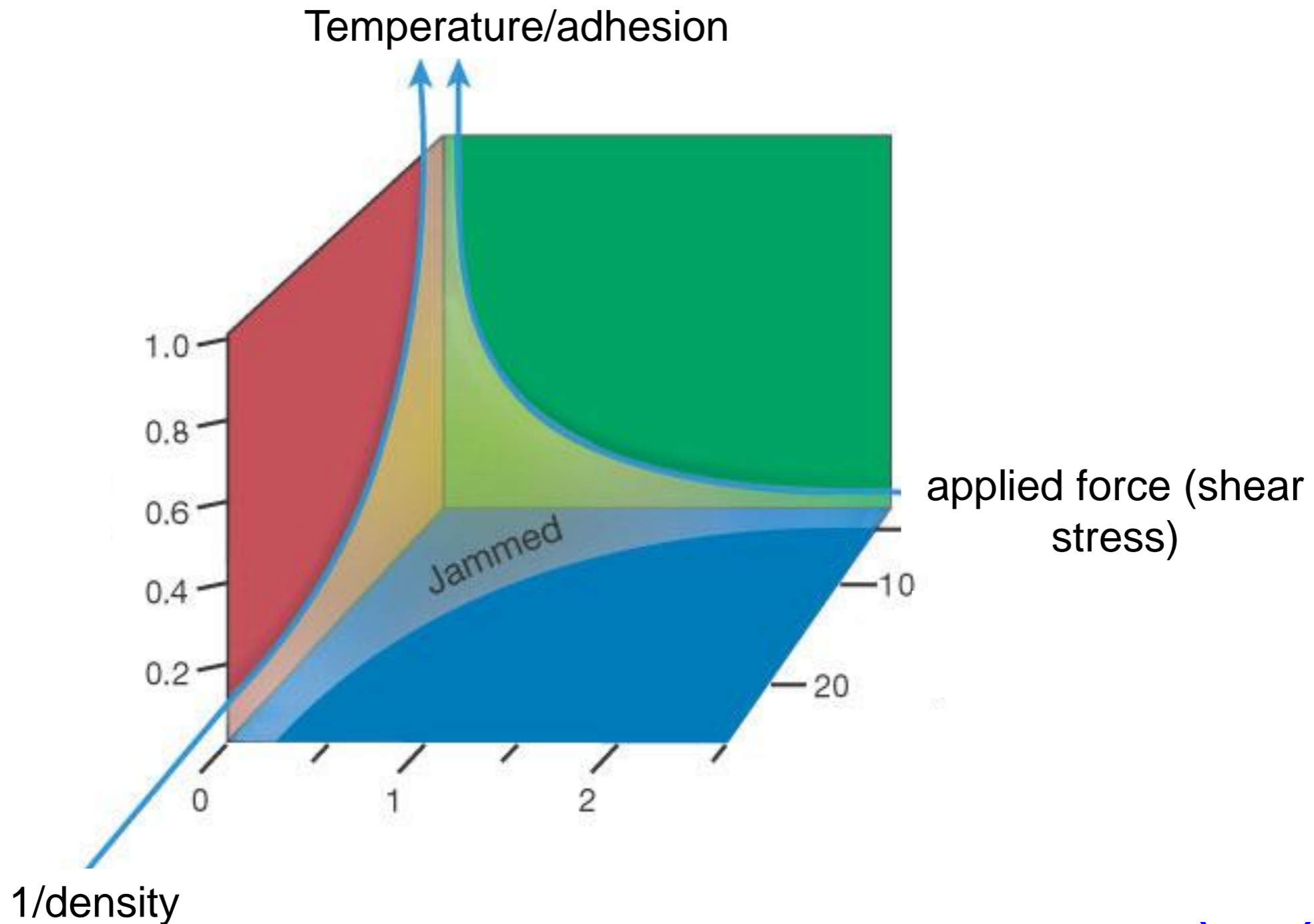
Schoetz, Lanio,
 Talbot, MLM
 J. R. Soc.
 Interface **10(89)**,
 20130726
 (2013)

Why do mechanical properties change with timescale? biological tissues



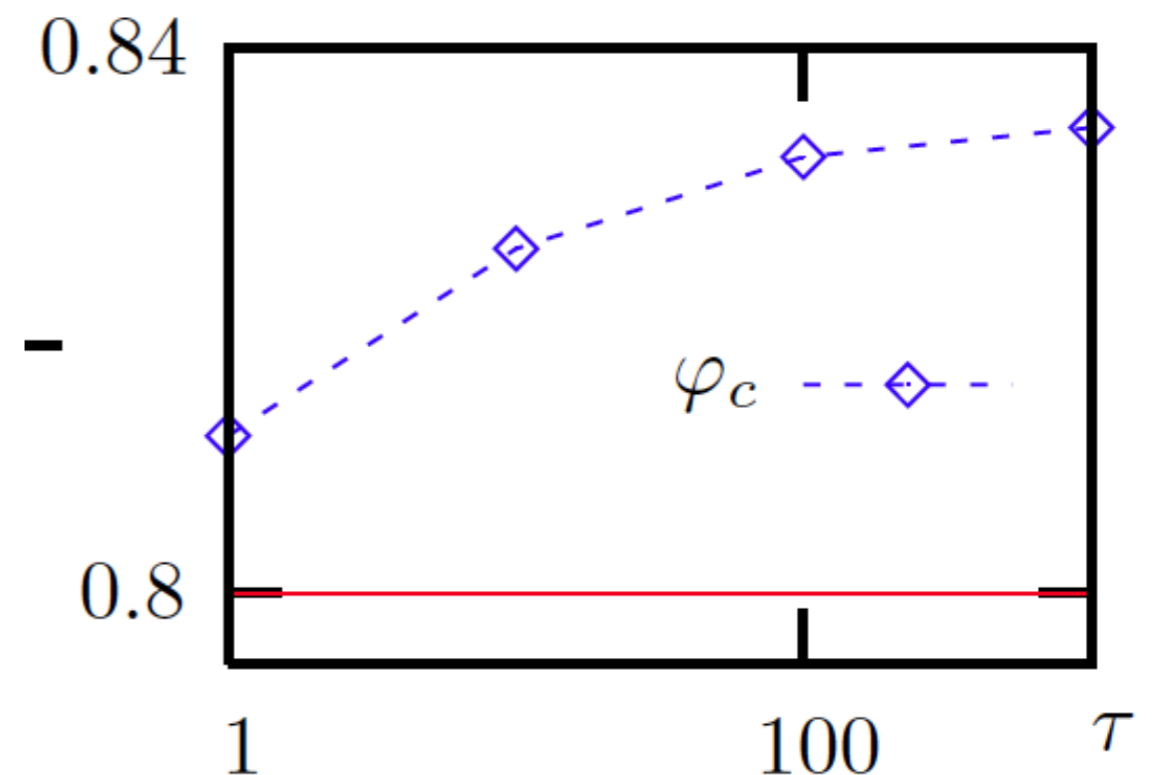
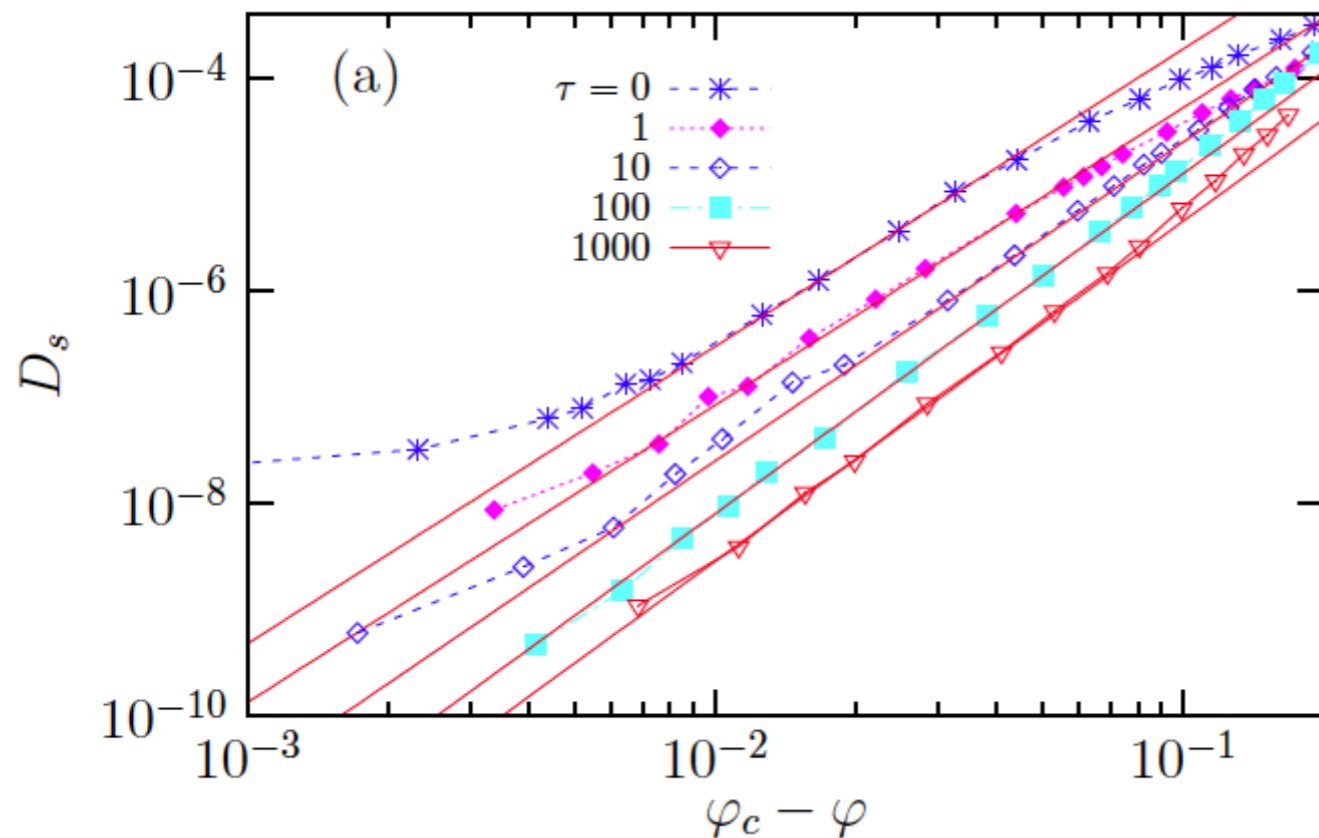
Schoetz, Lanio,
Talbot, MLM
J. R. Soc.
Interface **10(89)**,
20130726
(2013)

Jamming phase diagram for inert matter



Trappe et al, *Nature* **411**, 772-775 (2001)

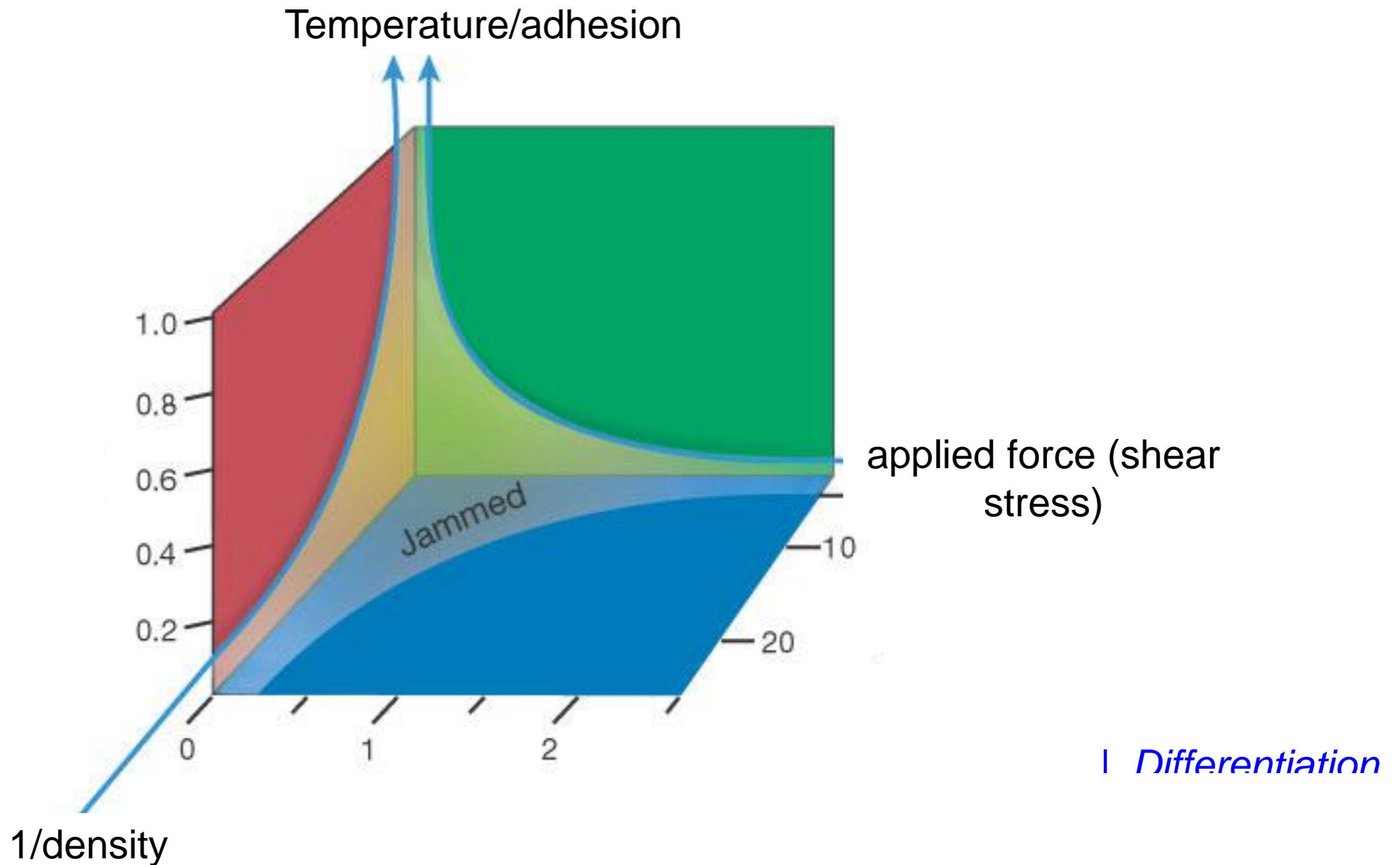
Glass transition in self-propelled particle models is identical to adhesive colloids*



Berthier, PRL 112, 220602 (2014)

*almost: The critical packing fraction changes with the persistence time of the self-propelled particles, because the activity generates an effective adhesion

Jamming phase diagram for biological tissues



Trappe et al, *Nature* **411**, 772-775 (2001)

Vector
near jamming transition

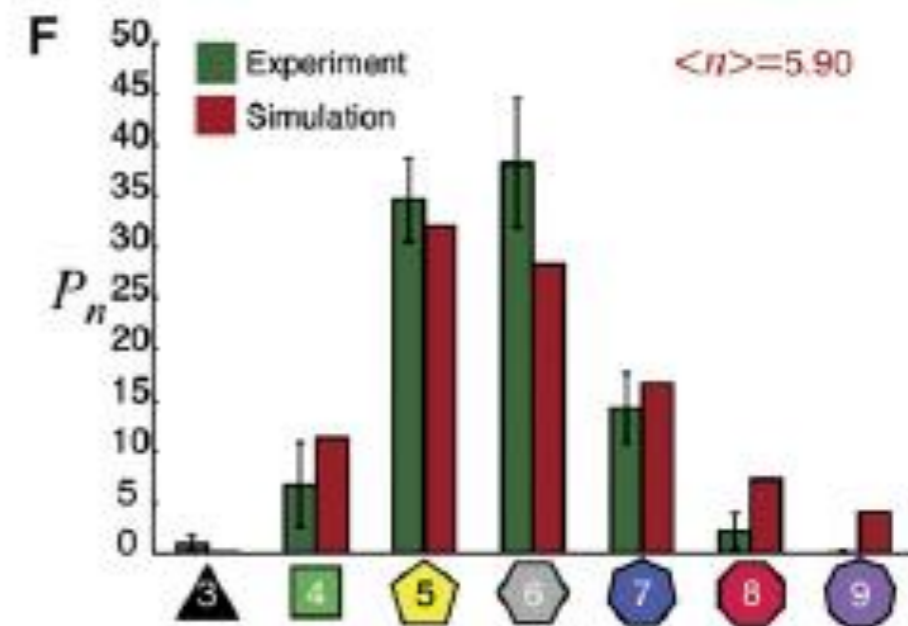
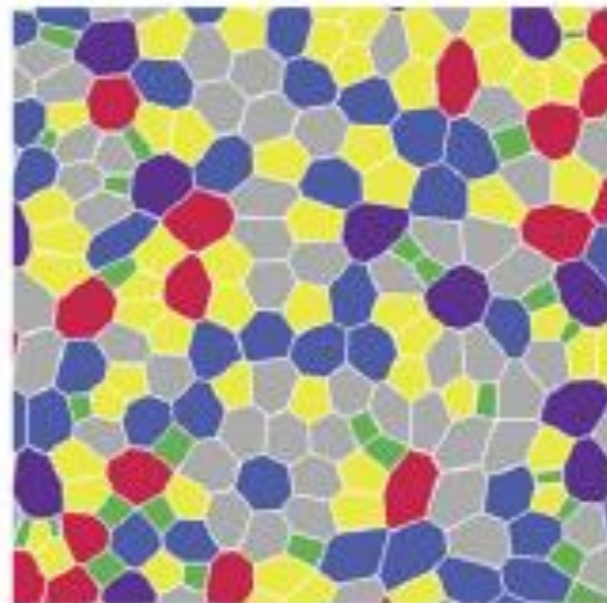
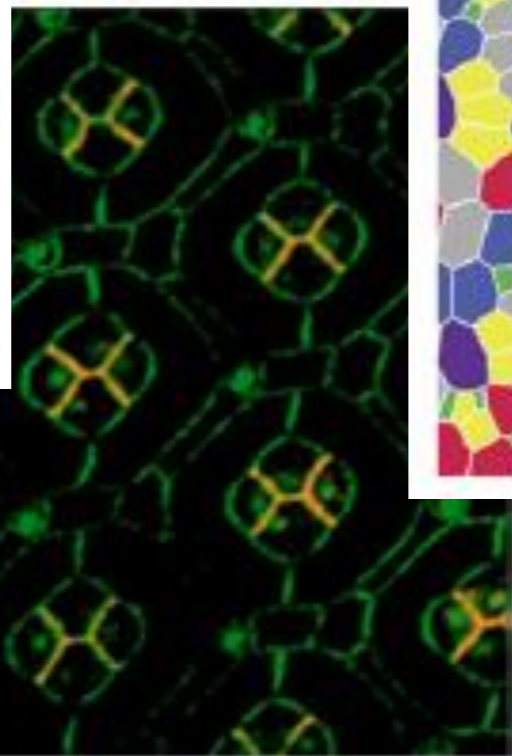
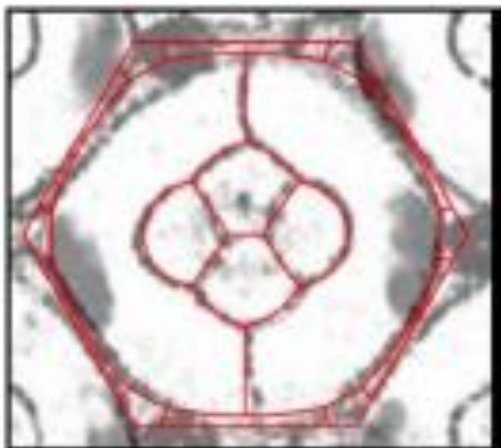
In many tissues, a density-driven transition is impossible

e.g. confluent tissues where there are no gaps between cells and the packing fraction is one

Is there some analogue to the jamming transition for confluent tissues?

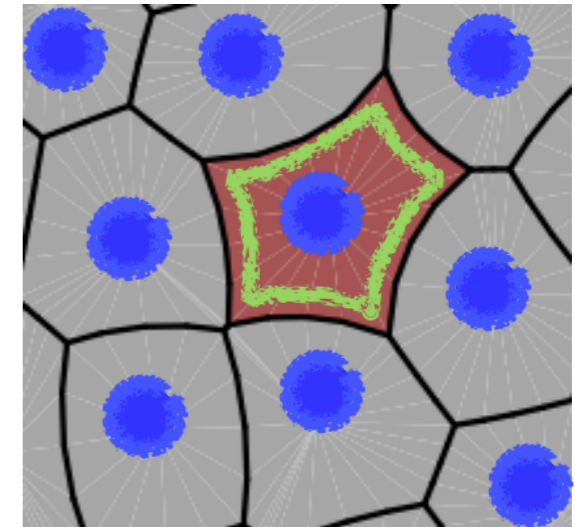
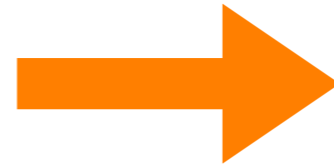
Vertex models for tissues

- Developed about 15 years ago
- Good agreement with experimentally observed cell shapes
- Explain/predict mechanically stable cell shapes and statistical properties



T. Nagai, H. Honda, *Philos. Mag. B* 81, 699 (2001)
Hufnagel et al, *PNAS* vol. 104 (10) pp. 3835 (2007)
Farhadifar et al, *Current Biology* (2007)
Jülicher et al *Phys. Rep.* (2007)
Hilgenfeldt et al, *PNAS* 105 3 907–911 (2008)
MLM et al, *PNAS* (2010)
Staple et al *EPJE* 33 (2) 117 (2010)
Chiou et al *PLOS Comp Bio* 8 (5) e1002512 (2012)

Vertex model equations



$$E_{cell} = k_A(A - A_0)^2 + k_P(P - P_0)^2$$

$$= k_A(A - A_0)^2 + k_P(P^2 - 2P_0P + P_0^2) \quad A = \text{area}, P = \text{perimeter}$$

3D
Incompressibility +
resistance to height

actomyosin contractility

Interfacial tension: adhesion and cortical tension

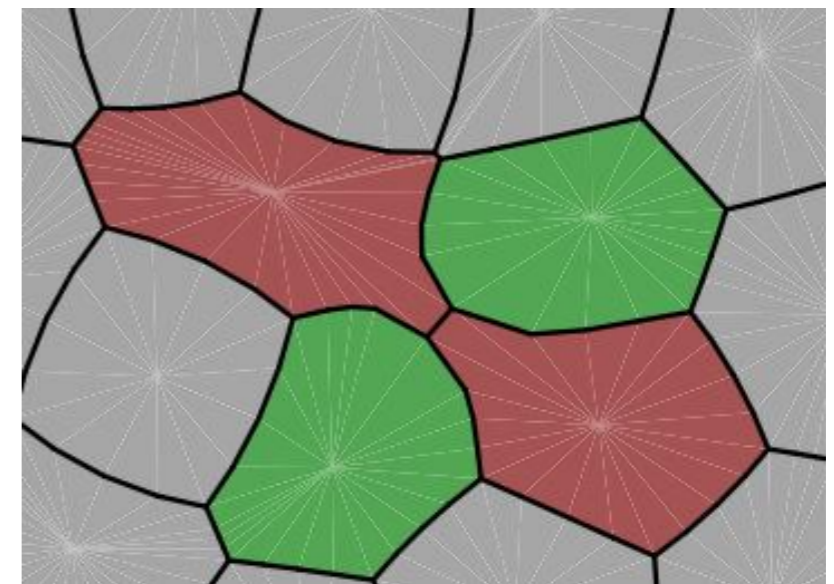
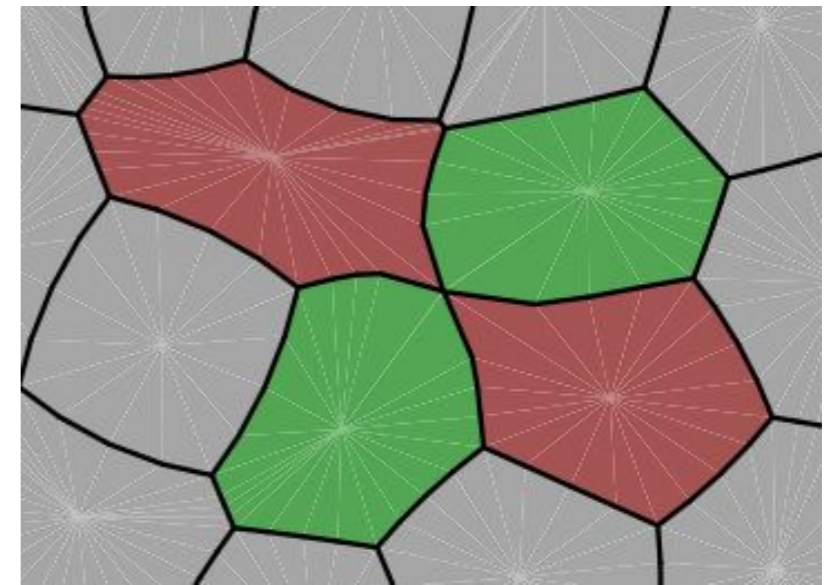
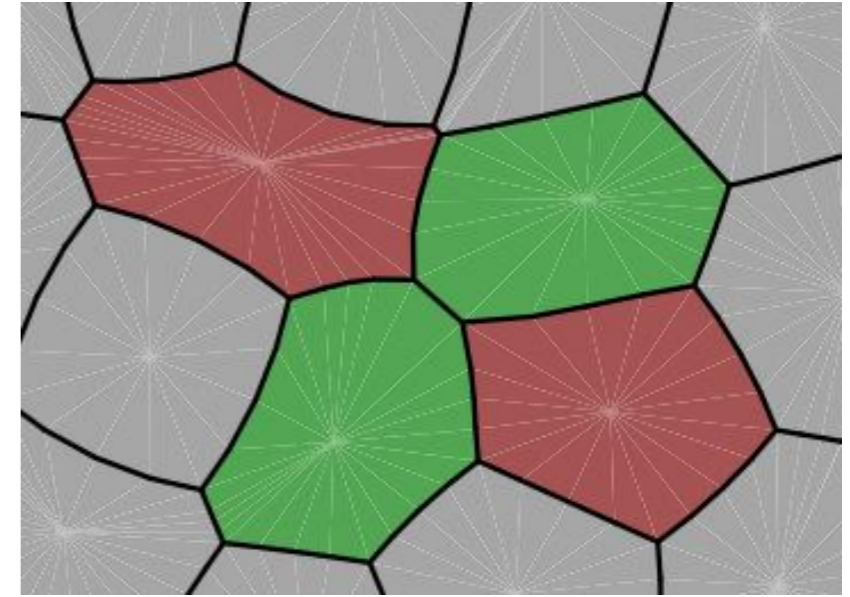
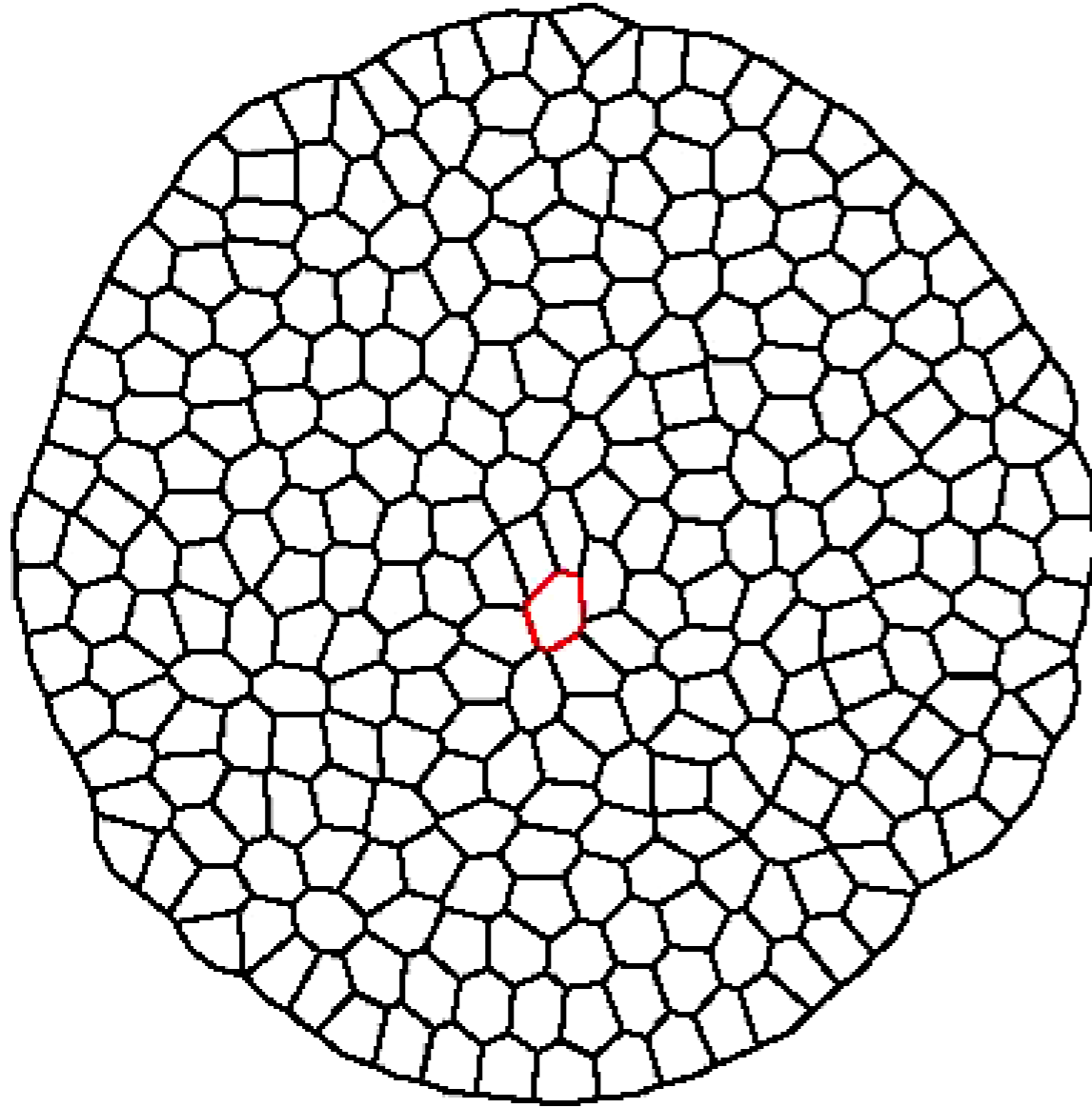
$$\varepsilon = \frac{1}{\beta A_0} \sum_i^N E_i = \sum_i \left[(a_i - 1)^2 + \frac{(p_i - p_0)^2}{r} \right]$$

Non-dimensionalized mechanical energy

Our model has two parameters:

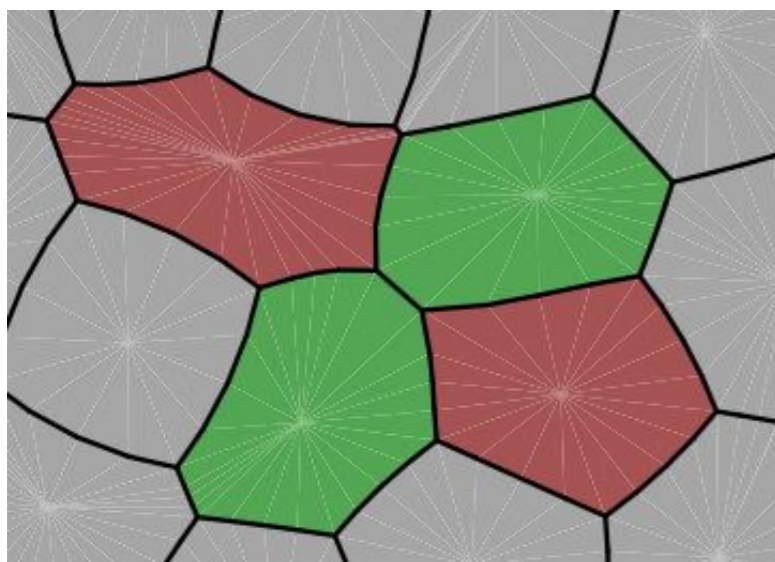
- p_0 = preferred perimeter: interfacial tension generated by adhesion and cortical tension
- r = inverse perimeter modulus: resistance to height fluctuations normalized by perimeter contractility OR ratio between bulk stiffness and interface stiffness

Rearrangements and migration in tissues via T-1 transitions

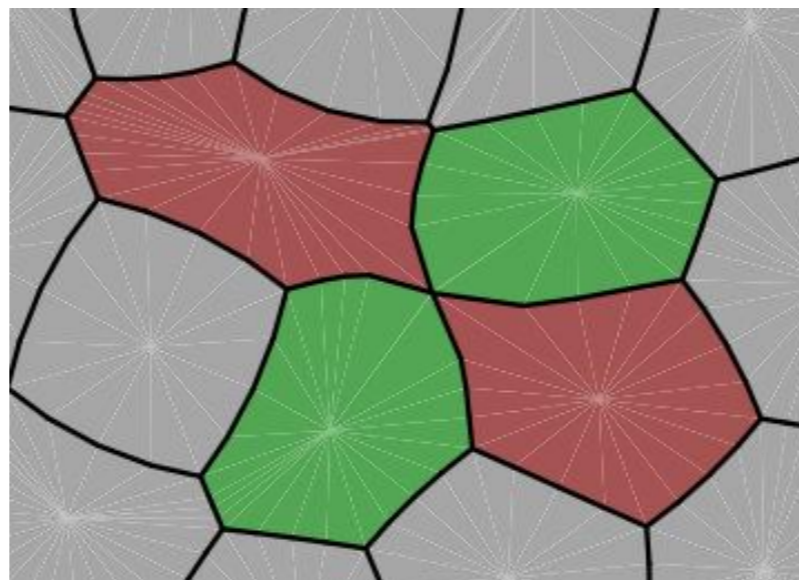


Energy trace for T-1 transitions

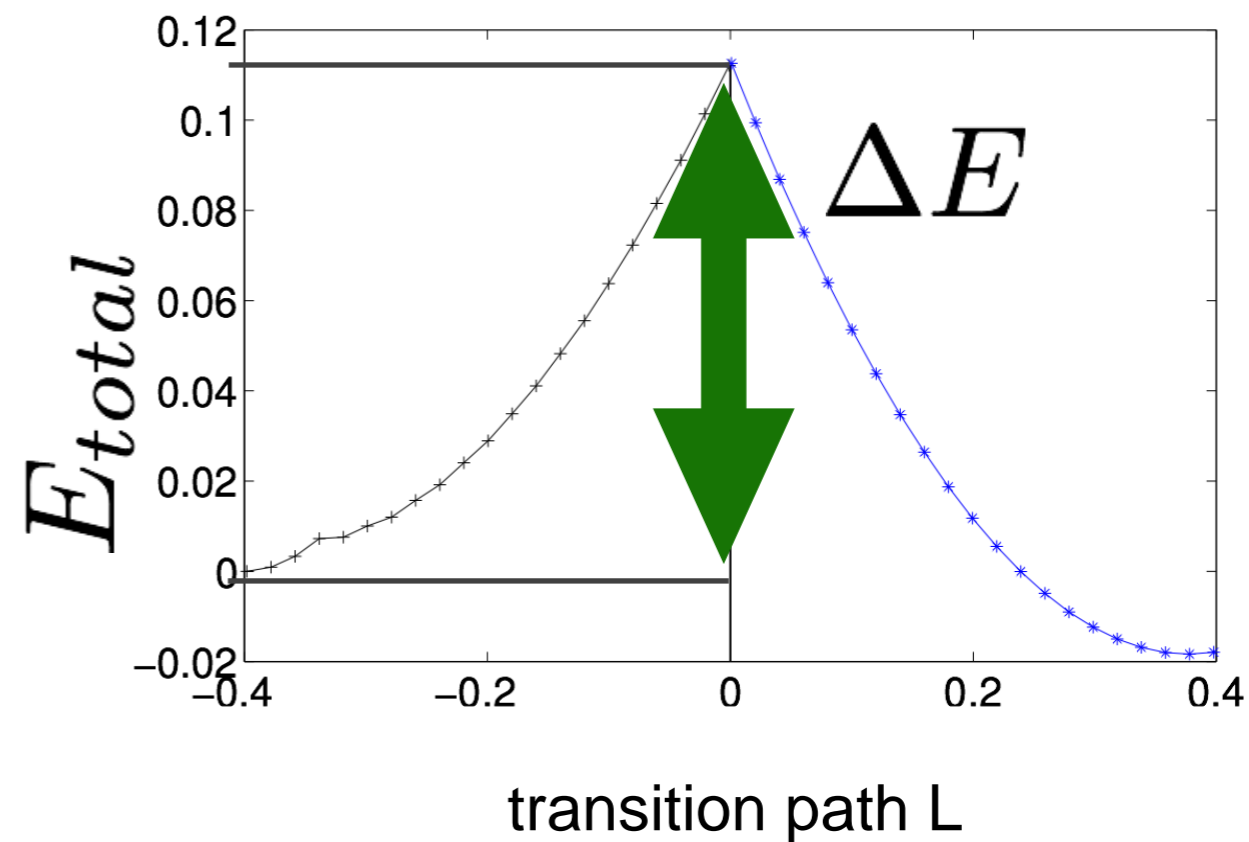
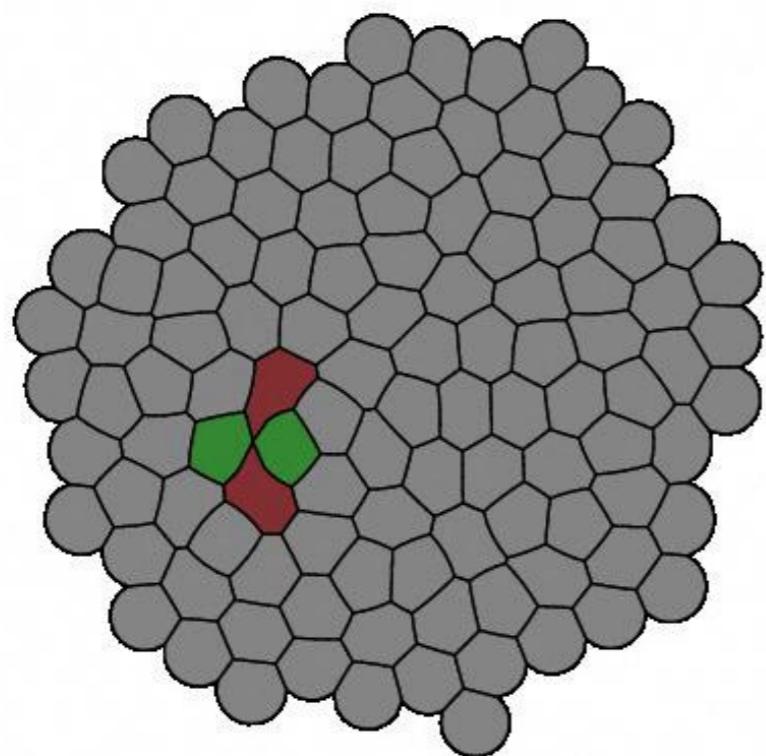
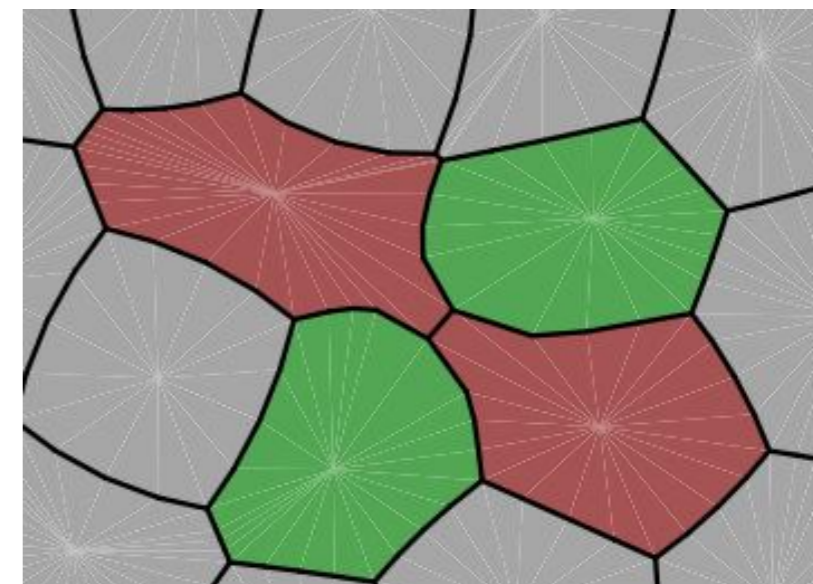
Shrink edge before T-1



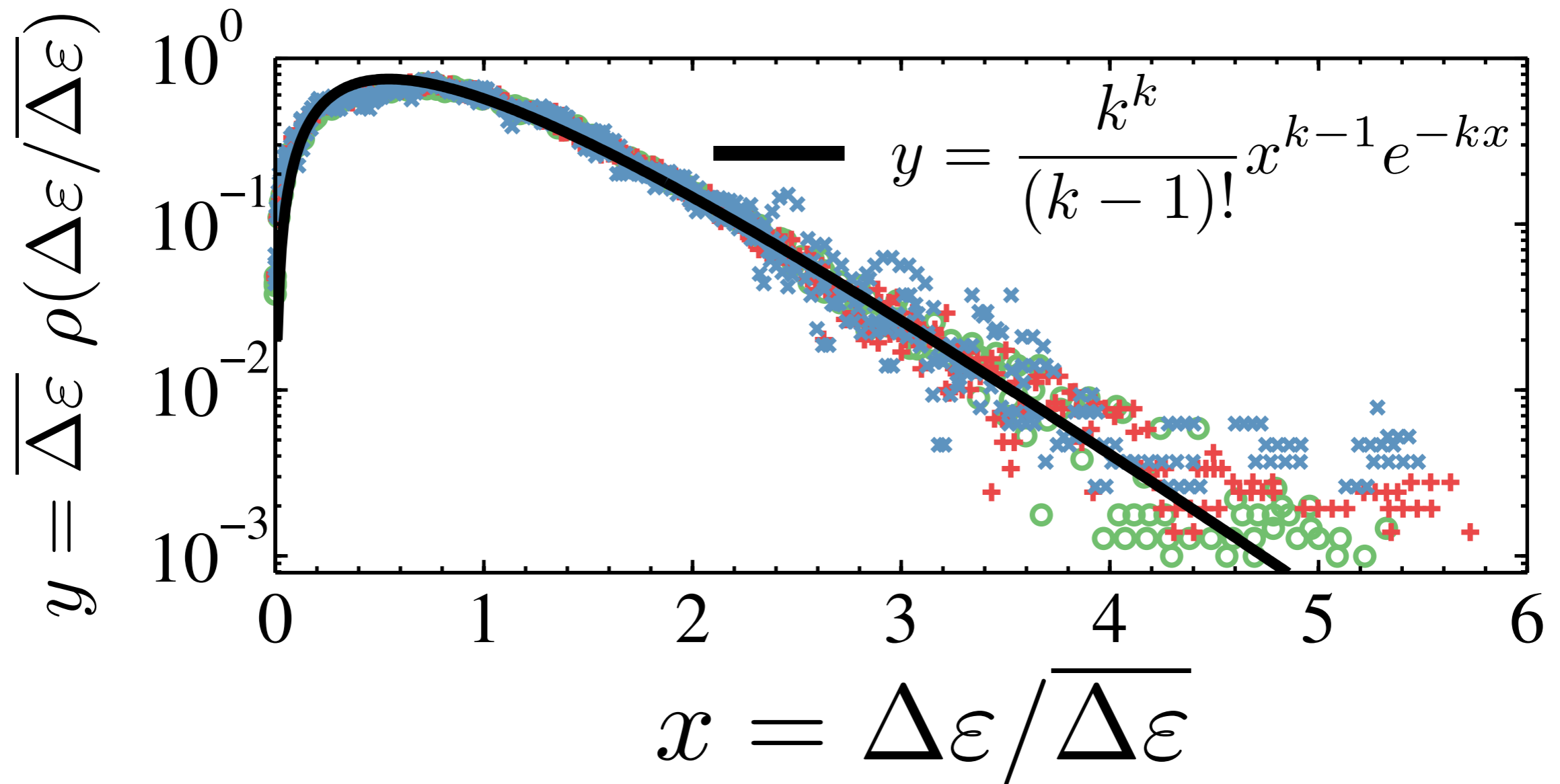
At T-1



Grow edge after T-1

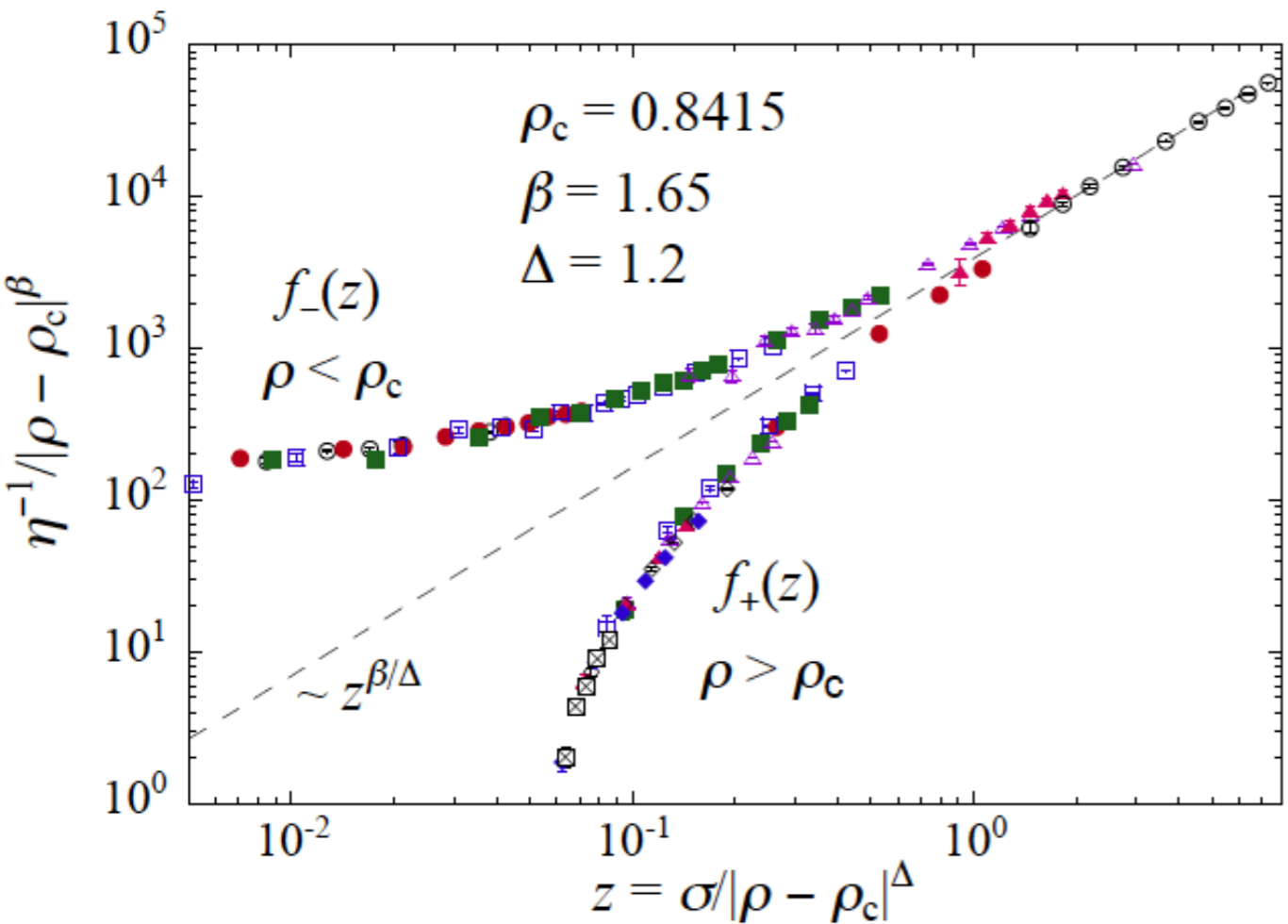


Large tissues: energy barrier statistics determined entirely by the mean energy barrier height

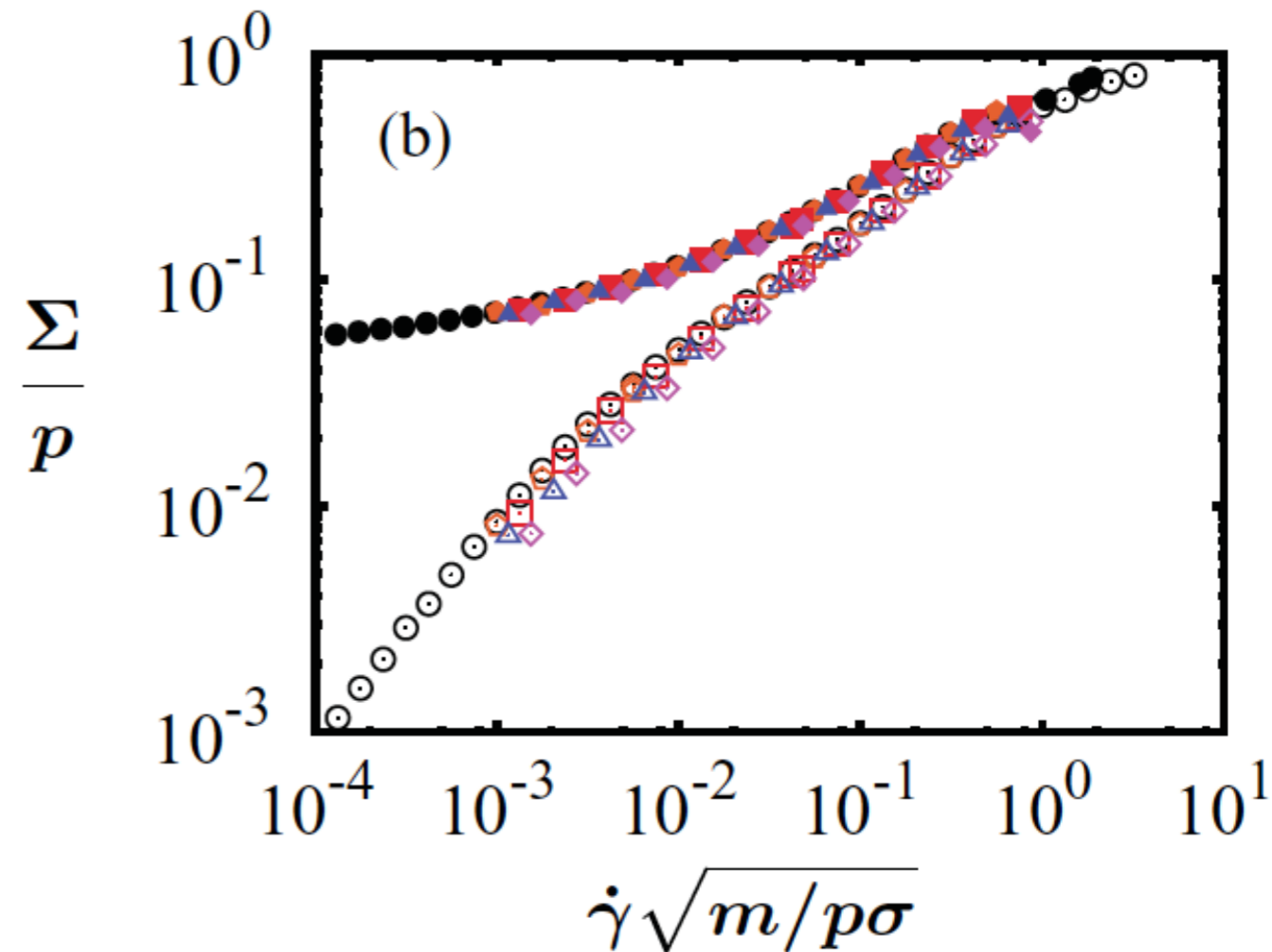


k-gamma distribution with one fit parameter: $k=2.2 \pm 0.2$
(found in many cellular systems)

Proving the existence of a critical “jamming” transition in inert matter:



Olsson and Teitel, PRL 99, 178001 (2007)



Haxton et al PRE 83, 031503 (2011)

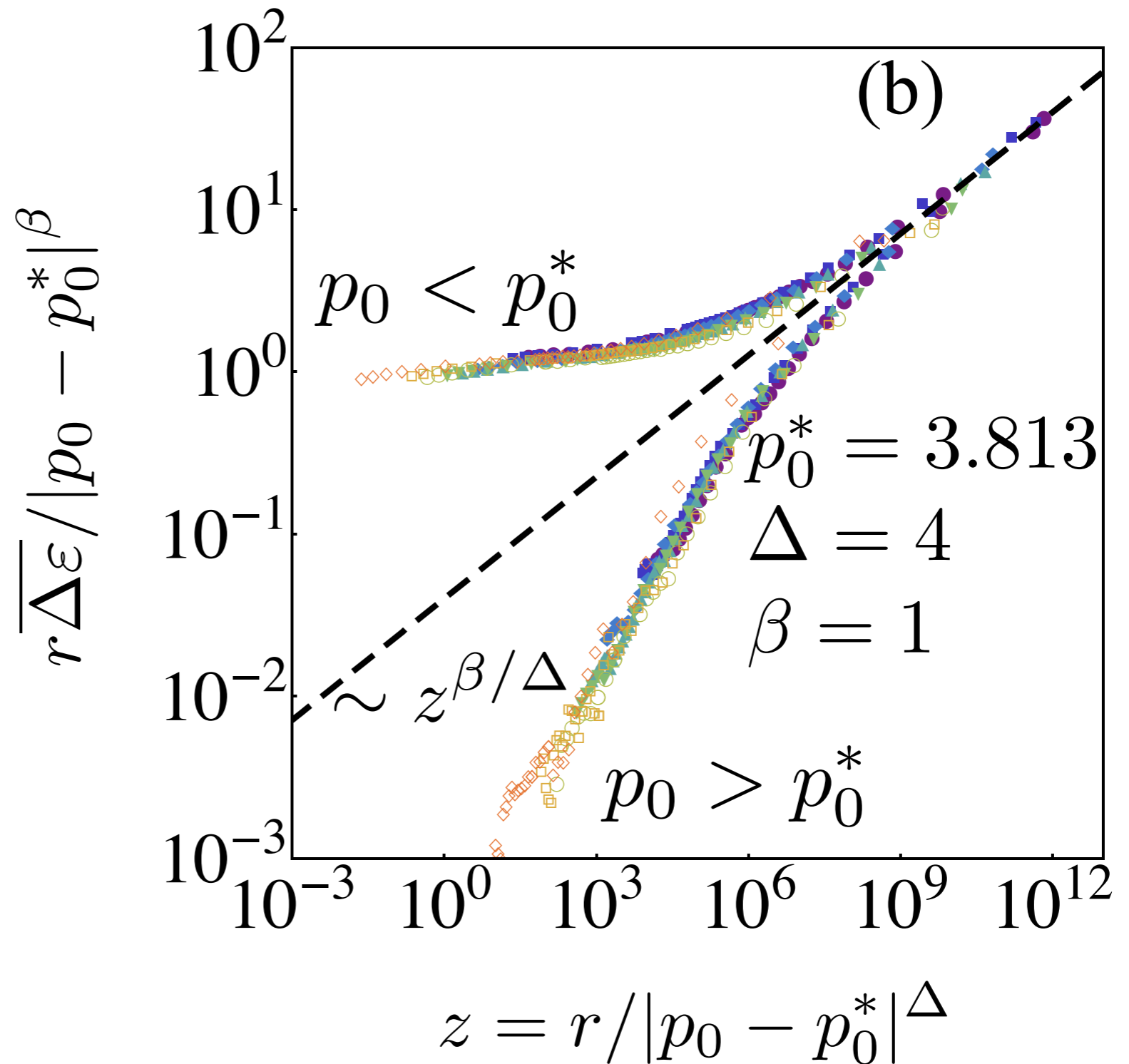
The onset of jamming is controlled by the **density**: $(\rho - \rho_c) \sim \text{pressure}$

A critical rigidity transition controlled by p_0 :

CHEAT SHEET:
Average energy
barrier height \sim
yield stress

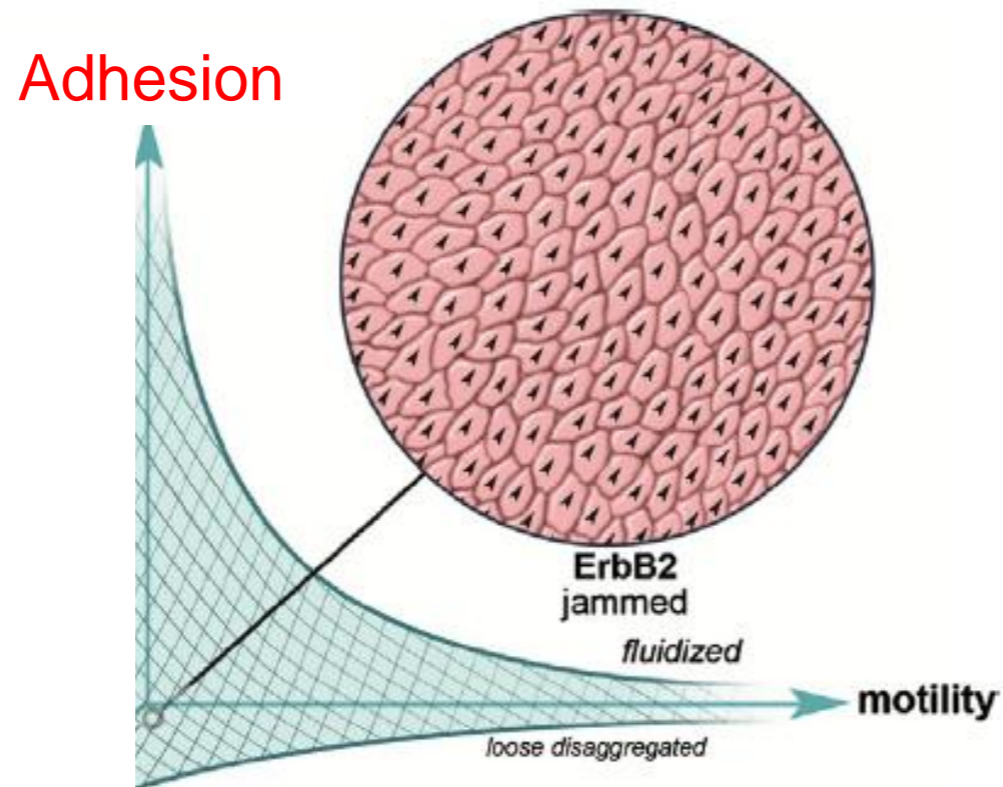
inverse perimeter
modulus $r \sim$ strain
rate

preferred perimeter
 $p_0 \sim$ density



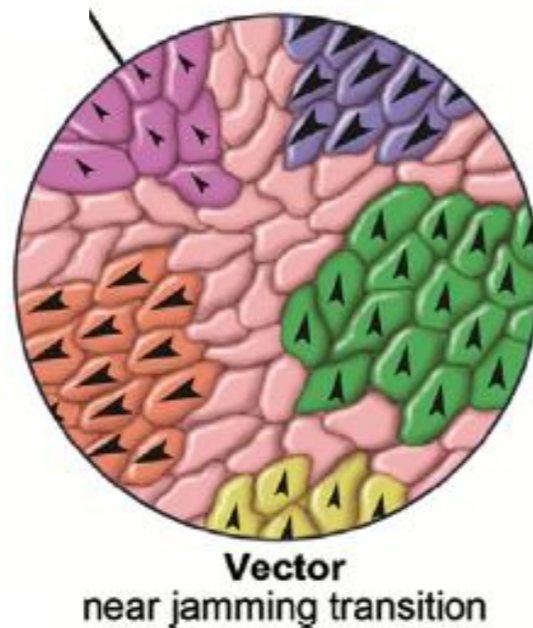
new rigidity phase diagram for biological tissues

Opposite for
confluent
tissues!

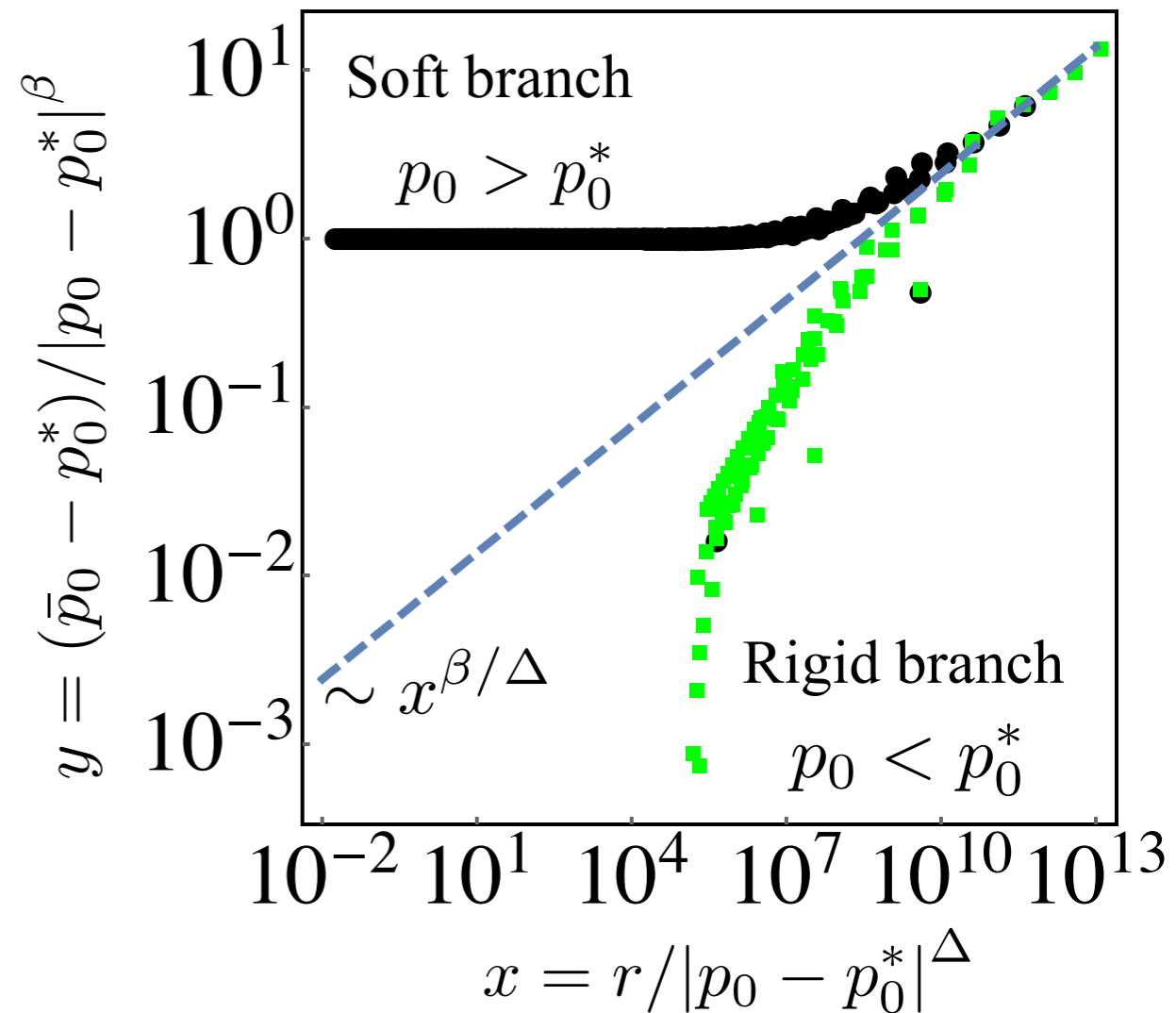
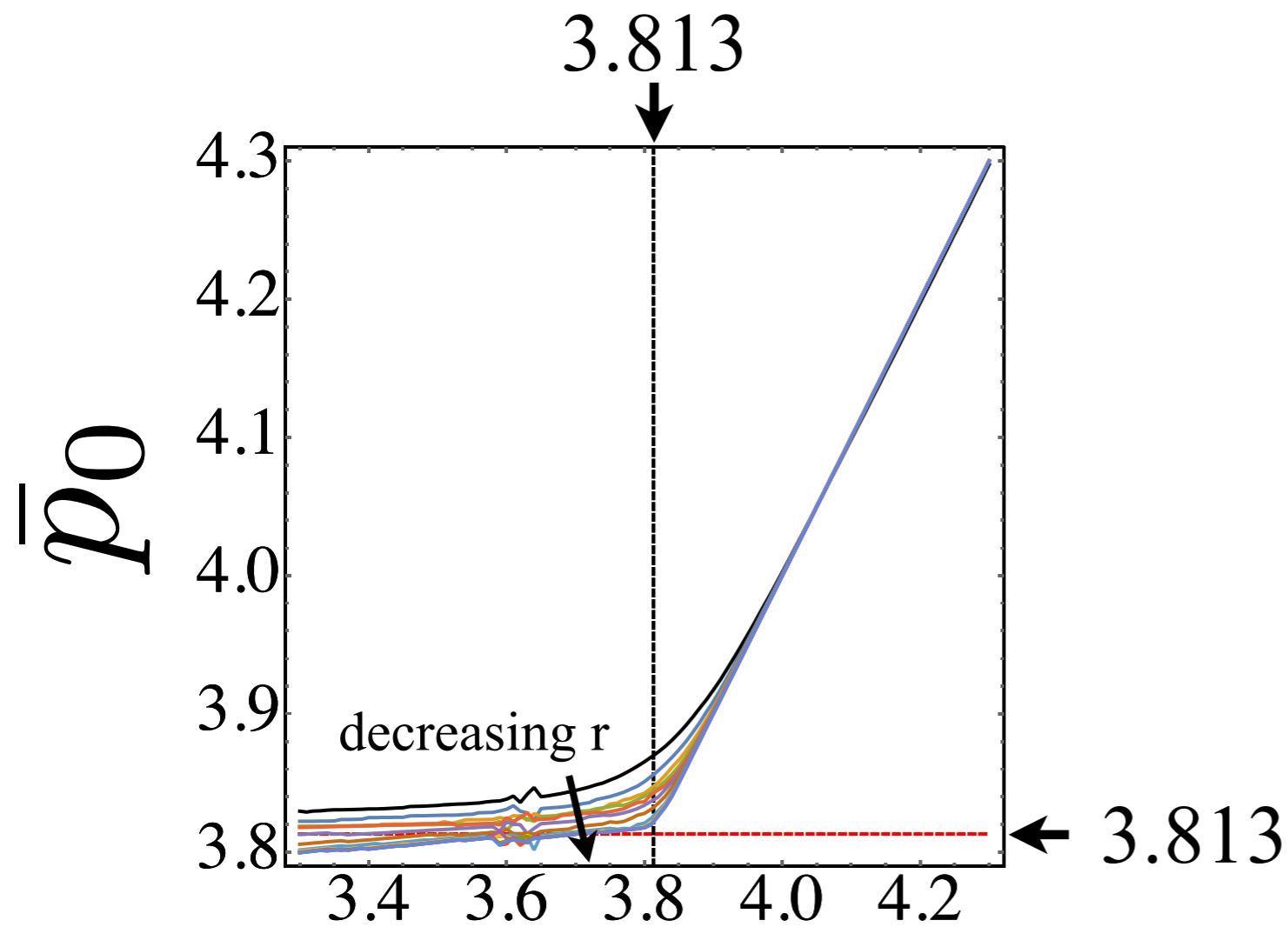


Particulate
matter: axis is
 $1/\text{adhesion}$

more adhesion
means more
gelation means
more solid-like



Experimental prediction

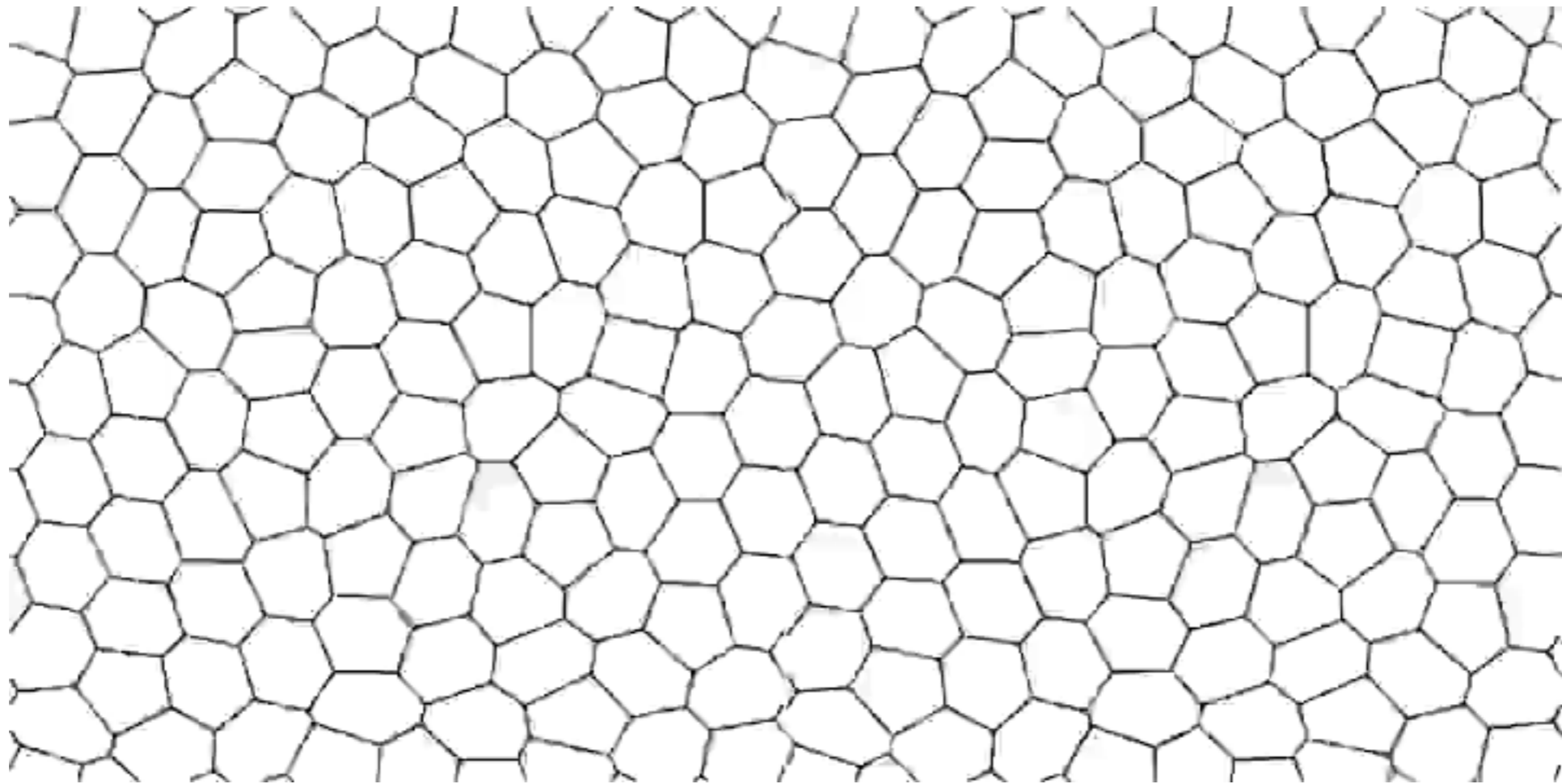


p_0

\bar{p}_0 is the average
OBSERVED perimeter-to-
area ratio:

$$\bar{p}_0 = \langle p \rangle / \sqrt{\langle a \rangle}$$

Effect of finite cell motility?

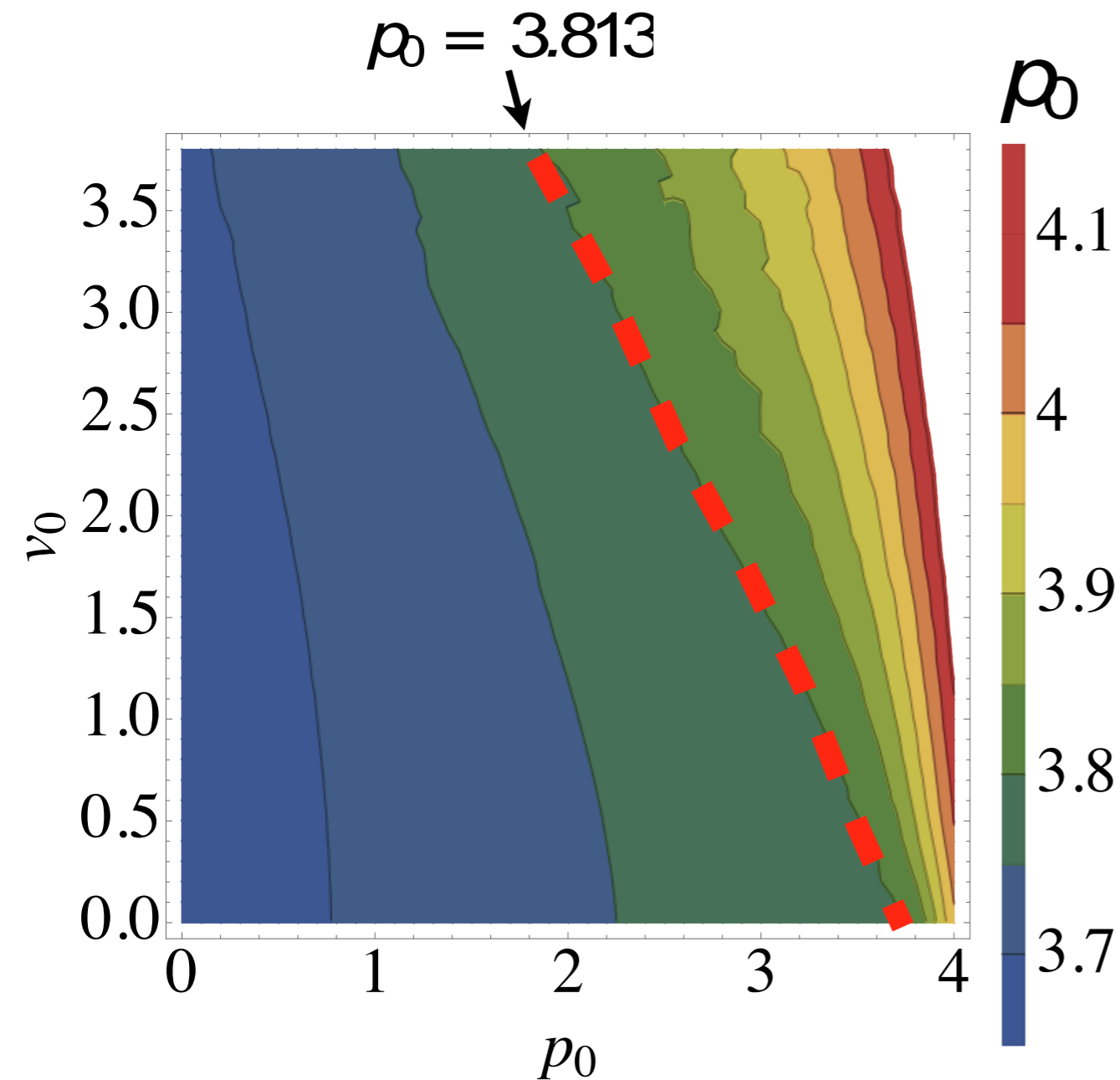


$$p_0 < p_0^*$$

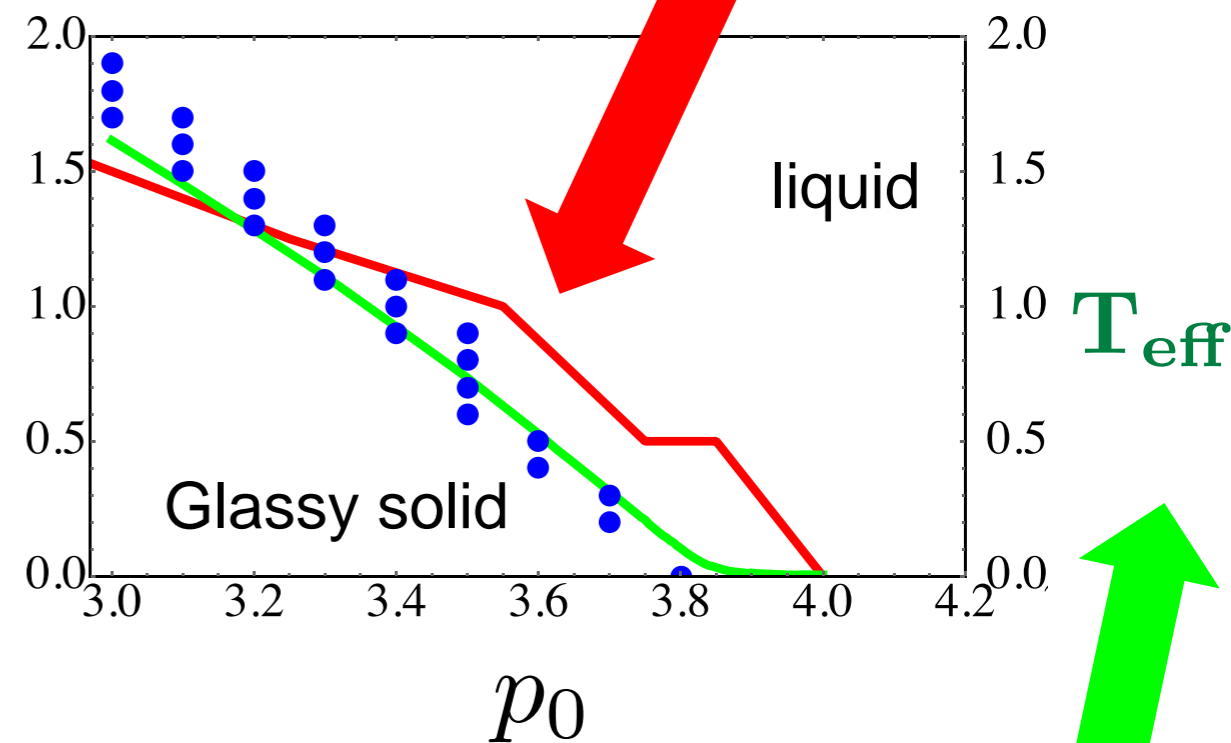
$$p_0 > p_0^*$$

Add an equation for cell polarization (like in self-propelled particle or Vicsek models)

Observed cell perimeter-to-area ratio robust to cell motility



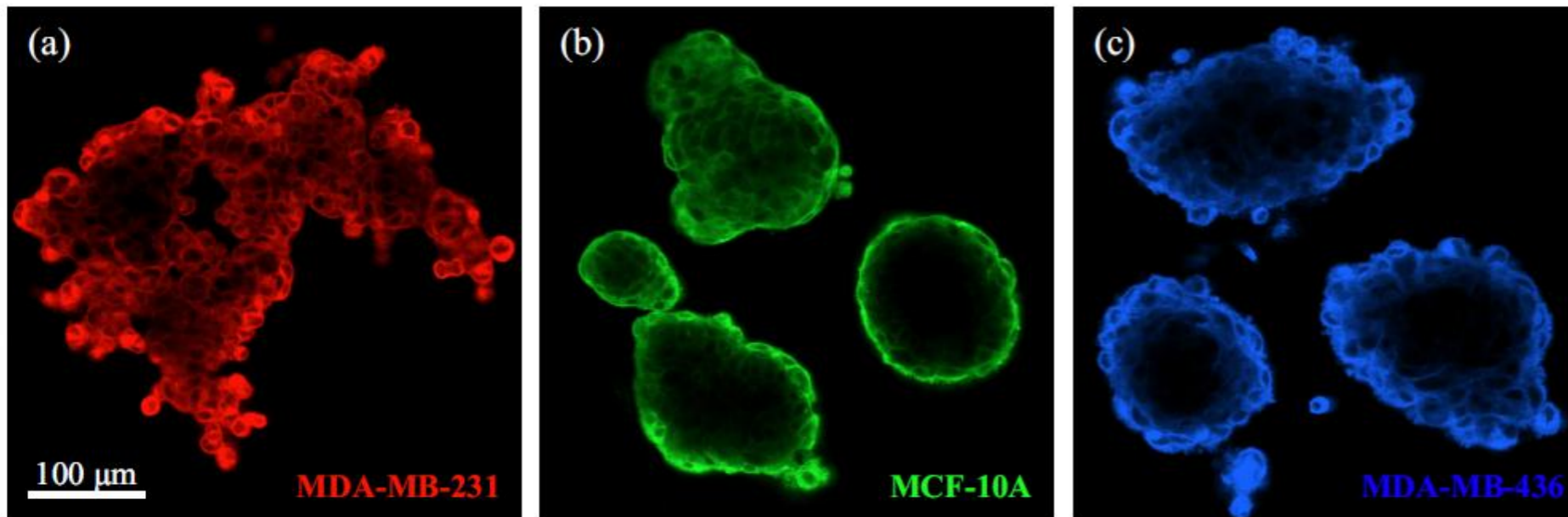
numerical from preliminary MSD data



numerical: from an active vertex model simulation

Analytic: from an extended SGR/trap model

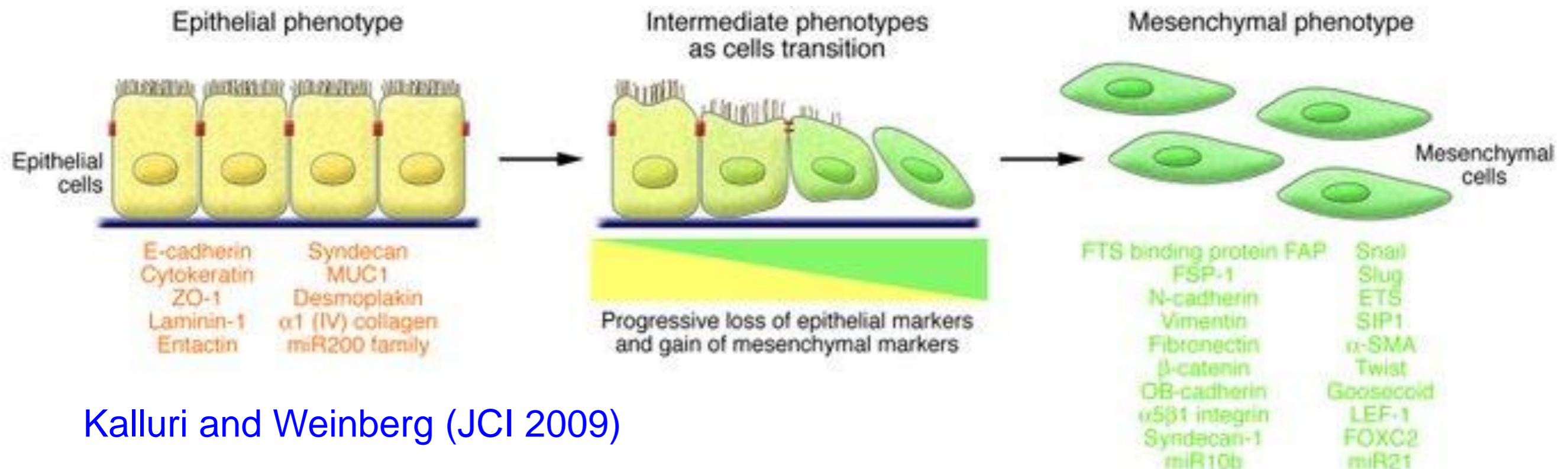
Cancer tumor boundaries and the EMT transition



Cancer cell sorting not driven by differential adhesion/ tissue surface tension!

Pawlizak ... MLM
...Kaes, in preparation (2014)

- Could the EMT transition be thought of as a change in p_0 – the ratio between the cell area and perimeter?
- If so, mesenchymal cells metastasize easily because they are on liquid side of rigidity transition?



Kalluri and Weinberg (JCI 2009)

Conclusions

- Biological tissues are complex materials, with mechanical properties that are important for biological function
- Many tissue types are apparently close to a glass transition
- The vertex model for confluent tissues exhibits a novel type of density-independent rigidity transition
 - excellent scaling collapse
 - control parameter is p_0 , which is proportional to single-cell adhesion or preferred cell perimeter
 - this is opposite of what you'd expect from particulate matter
- This is a rich framework with lots more to do:
 - cancer cell migration
 - EMT transitions
 - collective modes
 - effect of cell motility

Thanks so much for your attention!

Collaborators:

- **Max Dapeng Bi** (SU)
- Jen Schwarz (SU), Jorge Lopez (SU), Eva-Mara Schoetz (UCSD), Marcus Lanio (Princeton), Jared Talbot (Princeton)
- Jeff Amack (Upstate), Guliang Wang (Upstate)

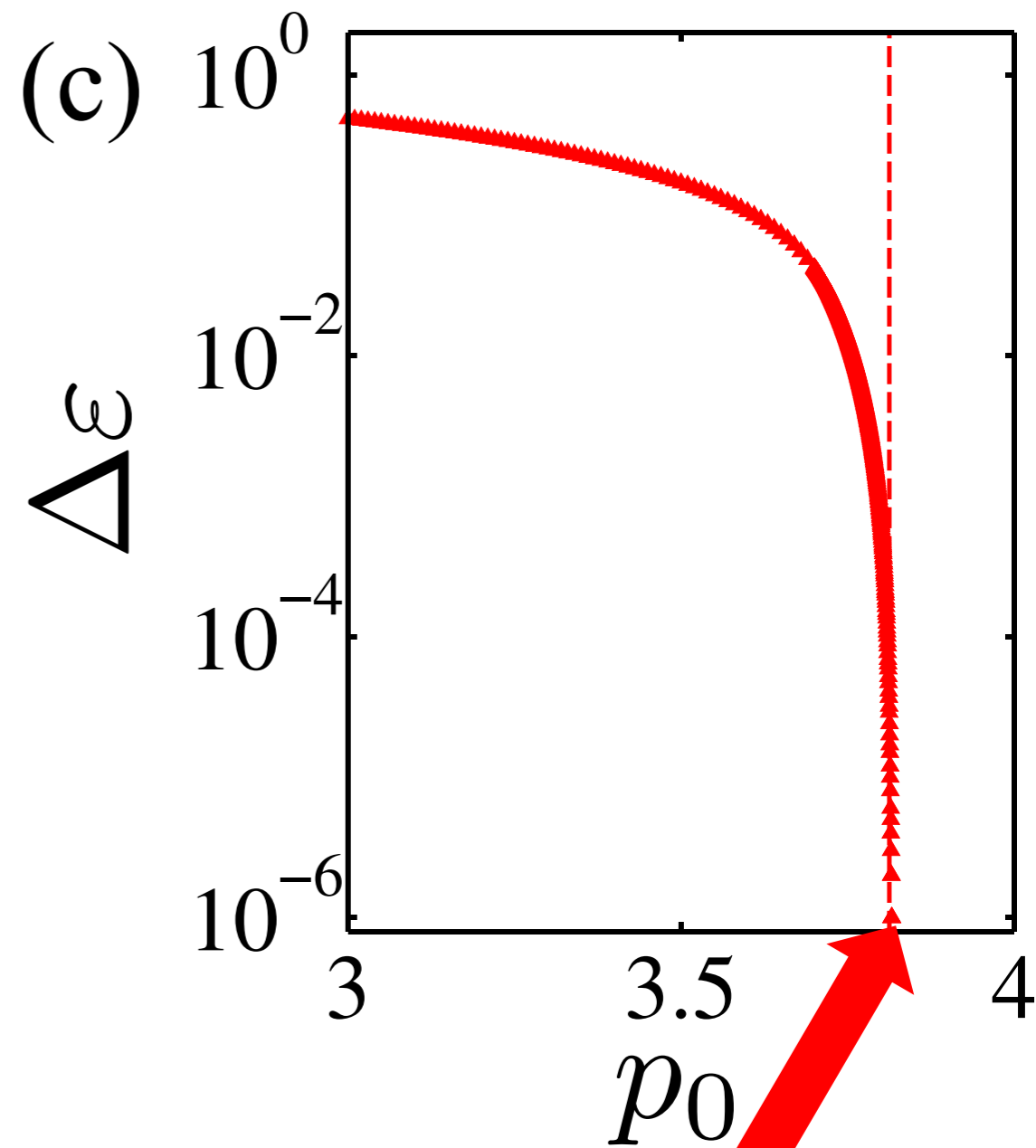
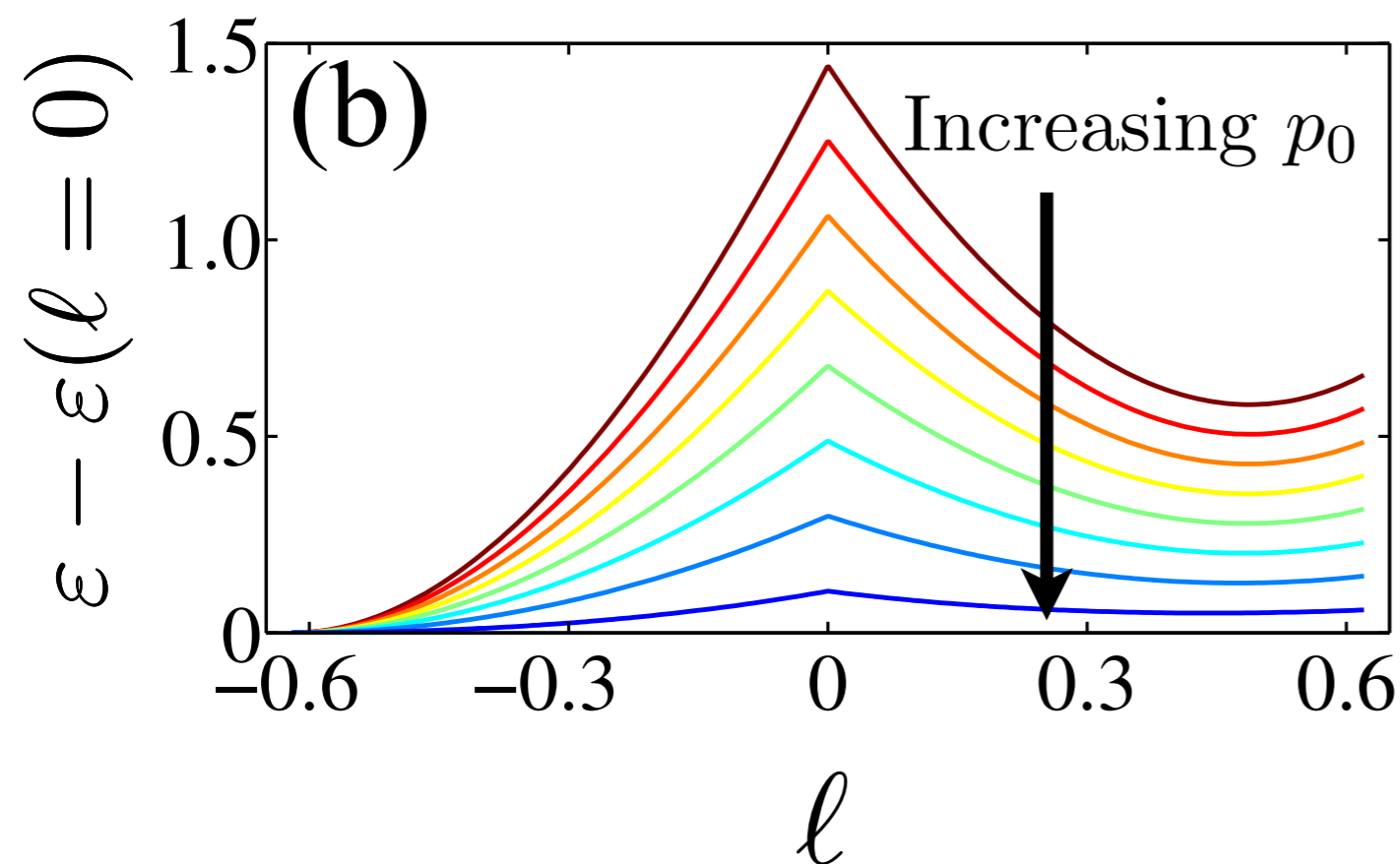
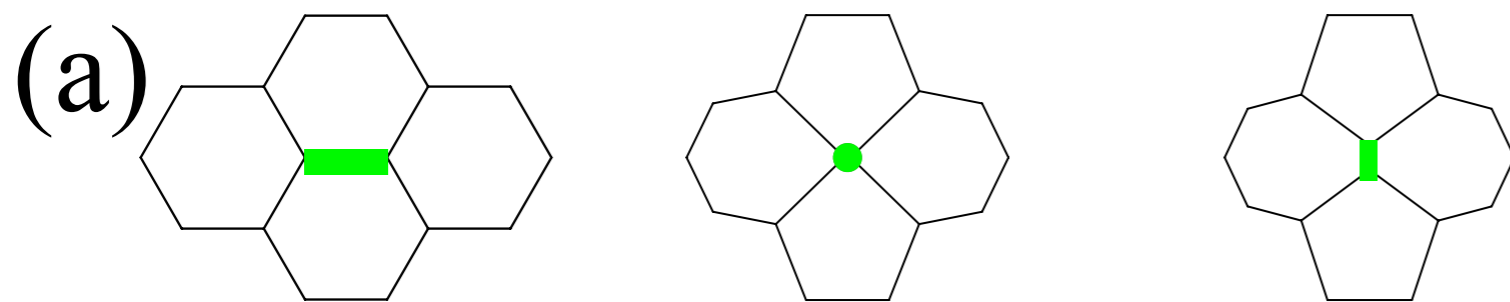
Funding:

- NSF BMMB CMMI-1334611
- NSF DMR CMMT-1352184
- Alfred P. Sloan Foundation
- Soft Interfaces IGERT (DGE-1068780)



Fixed area, four cells

p_0 only control parameter

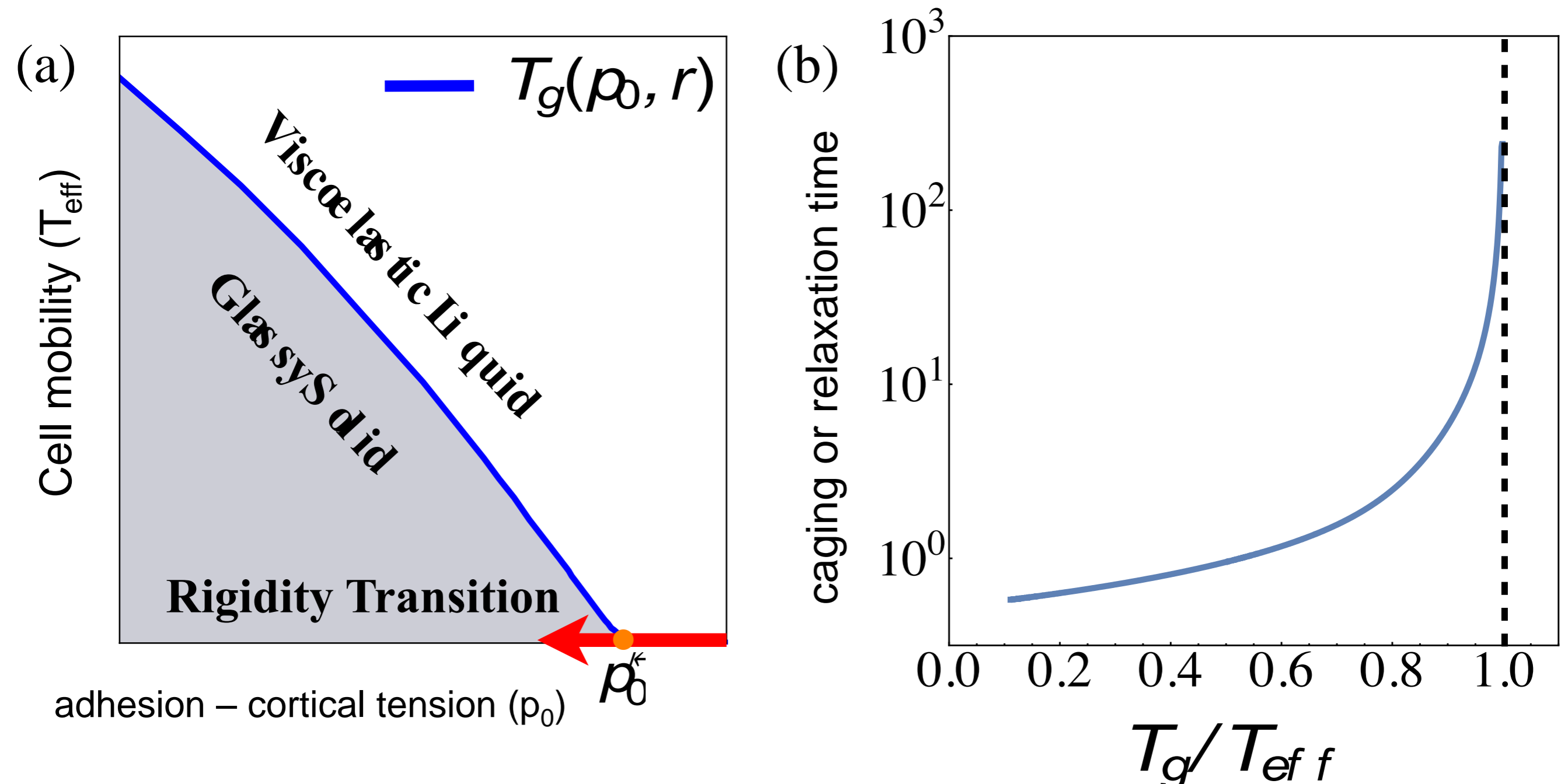


Bi, Lopez, Schwarz, MLM submitted(2014)

Transition point $p_0^* \approx 3.813$
= perimeter of regular
pentagon with unit area

What happens when you
include a finite cell activity
or mobility?

Using a trap or SGR model to go from energy barriers to cell dynamics

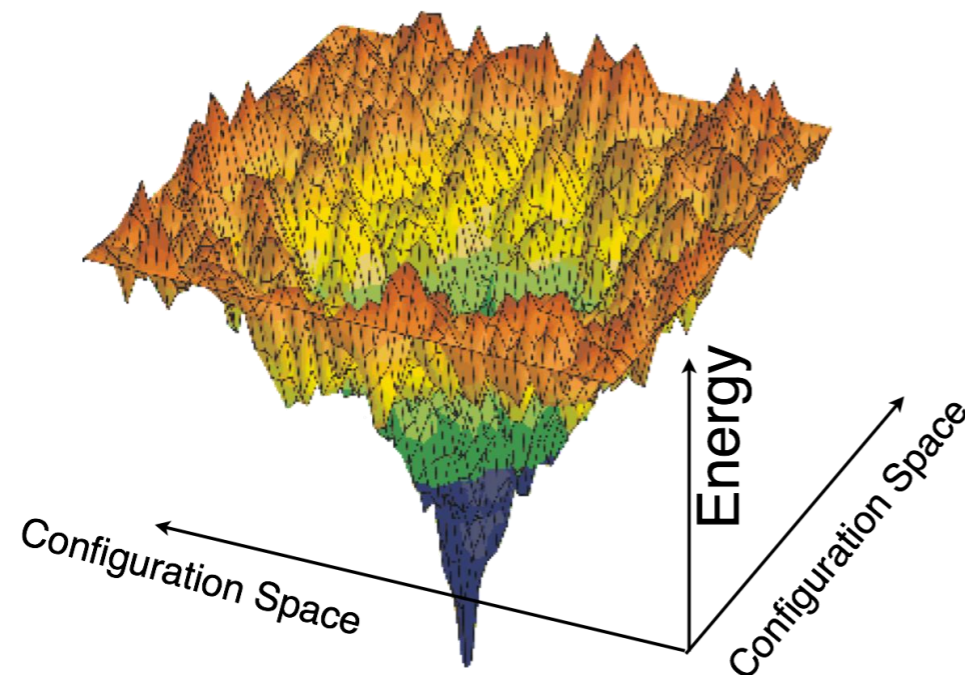


When cells are actively fluctuating and moving, how does this change the transition?

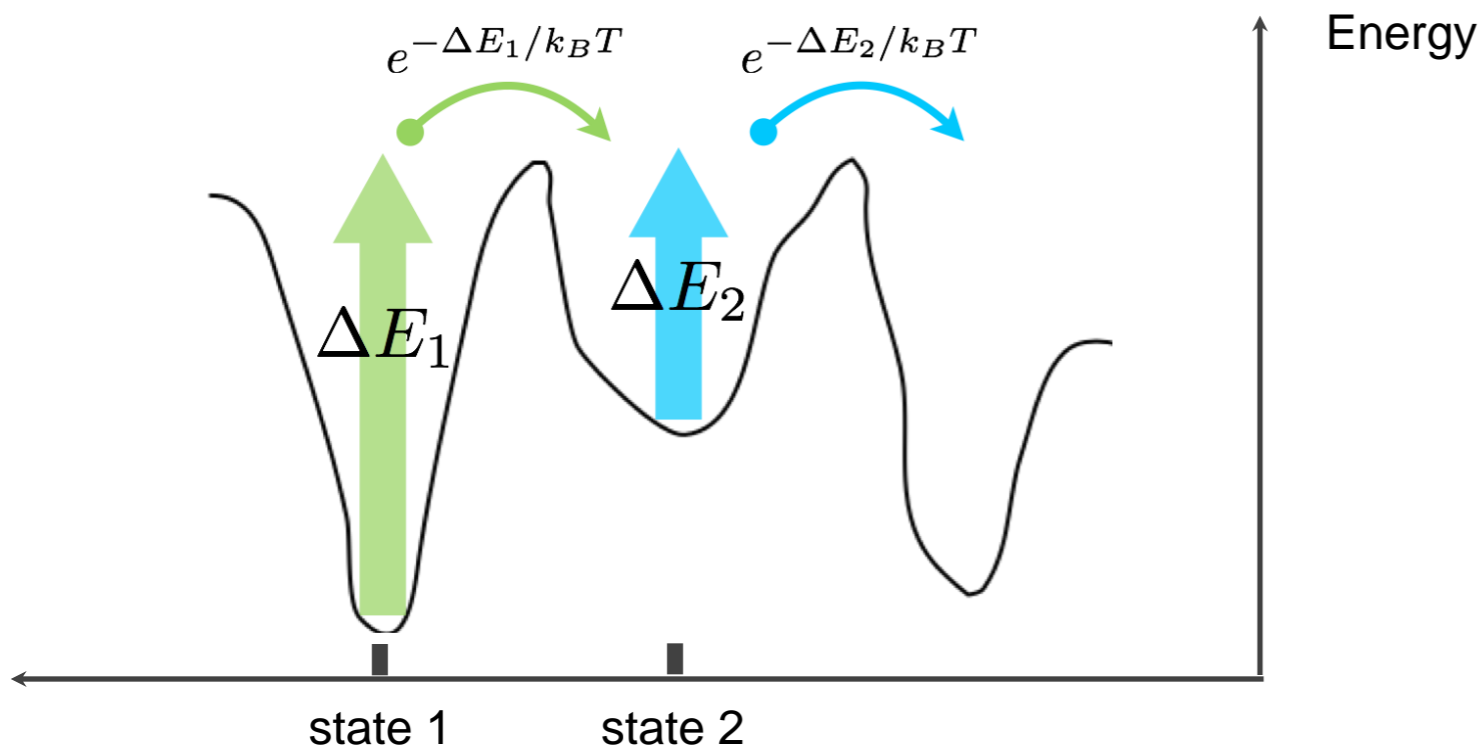
Complex potential
energy landscape

+

system close to energy
landscape surface



System is “trapped” in a
metastable state until a
rare fluctuation allows it to
escape:



trap model **C. Monthus and
J.-P. Bouchaud, J. Phys. A
29, 3847 (1996)**

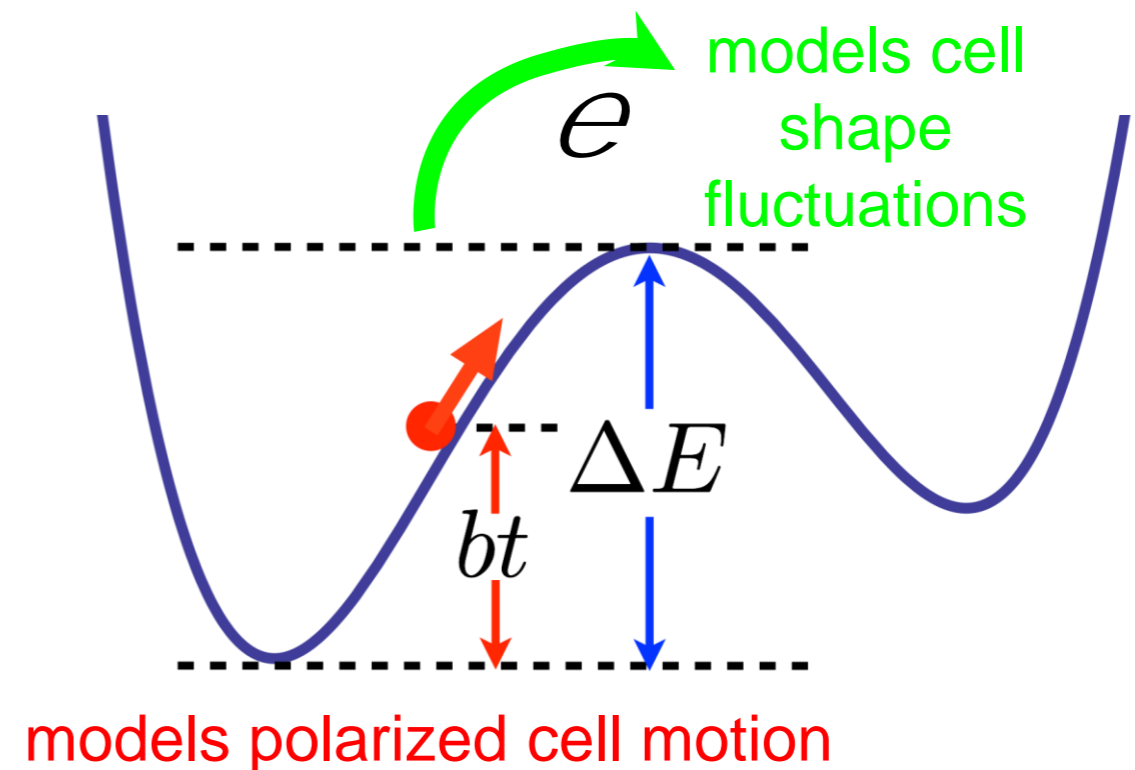
soft glassy rheology
**Sollich et al, PRL 78 2020
(1997)**

From energy barriers to cell migration:

trap model: C. Monthus and J.-P. Bouchaud, J. Phys. A 29, 3847 (1996)

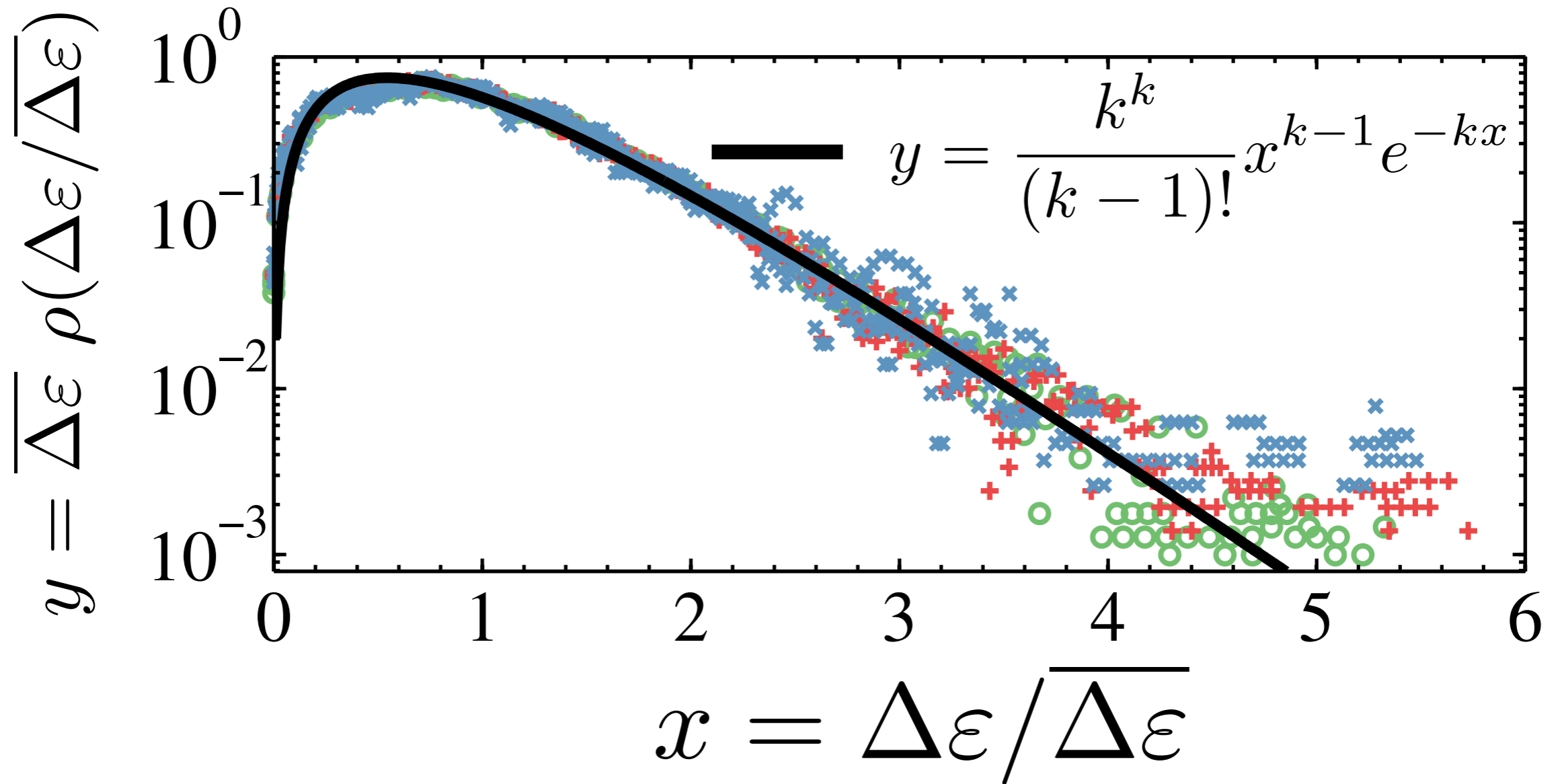
soft glassy rheology: Sollich et al, PRL 78 2020 (1997)

$$R = \omega_0 \exp [-(\Delta E - bt) / \varepsilon]$$

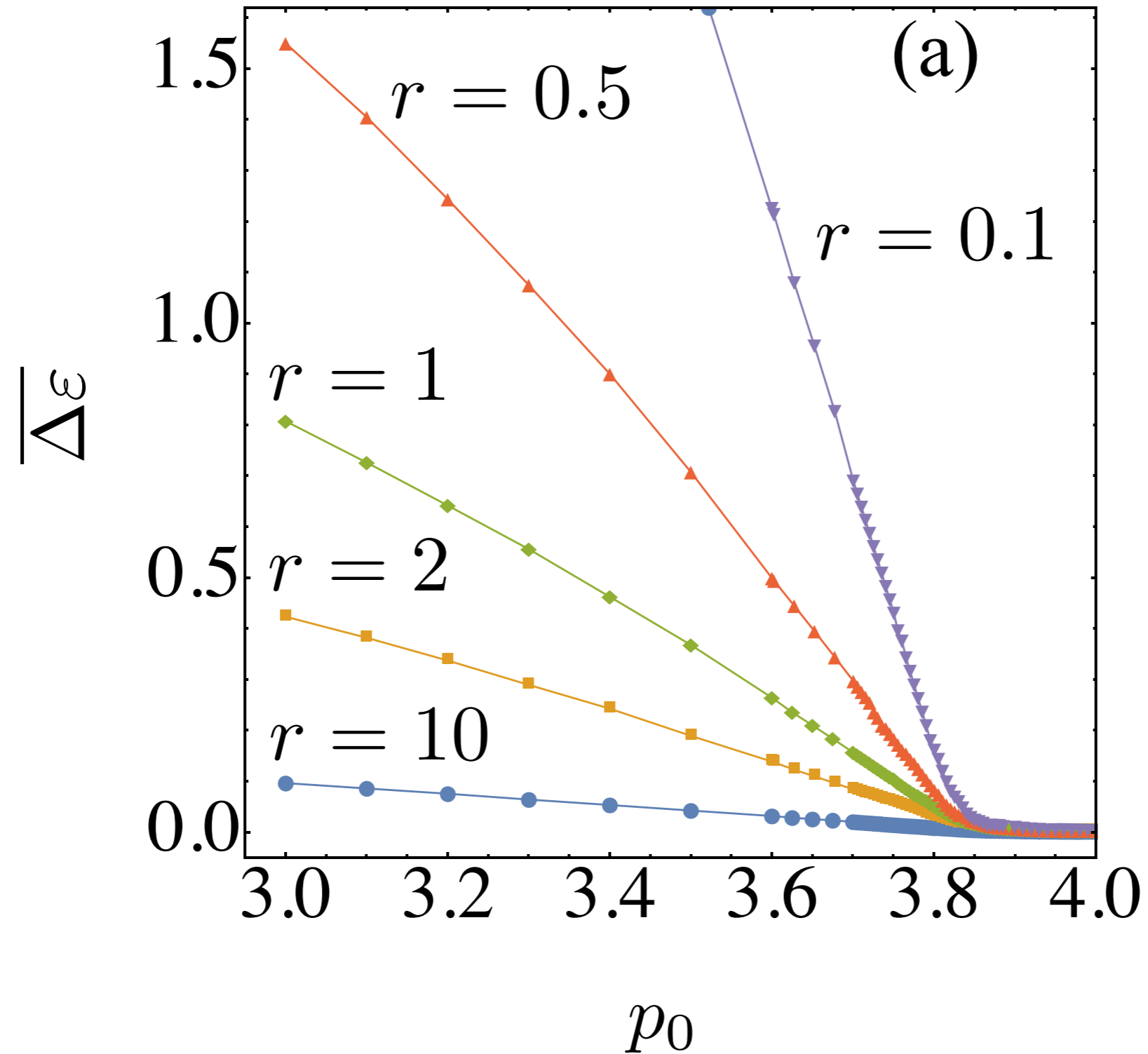


$$\frac{\partial}{\partial t} P(\Delta E, t) = -\omega_0 e^{-[\Delta E - bt] / \varepsilon} P(\Delta E, t) + \rho(\Delta E) \int d\Delta E' \omega_0 e^{-[\Delta E' - bt] / \varepsilon} P(\Delta E', t),$$

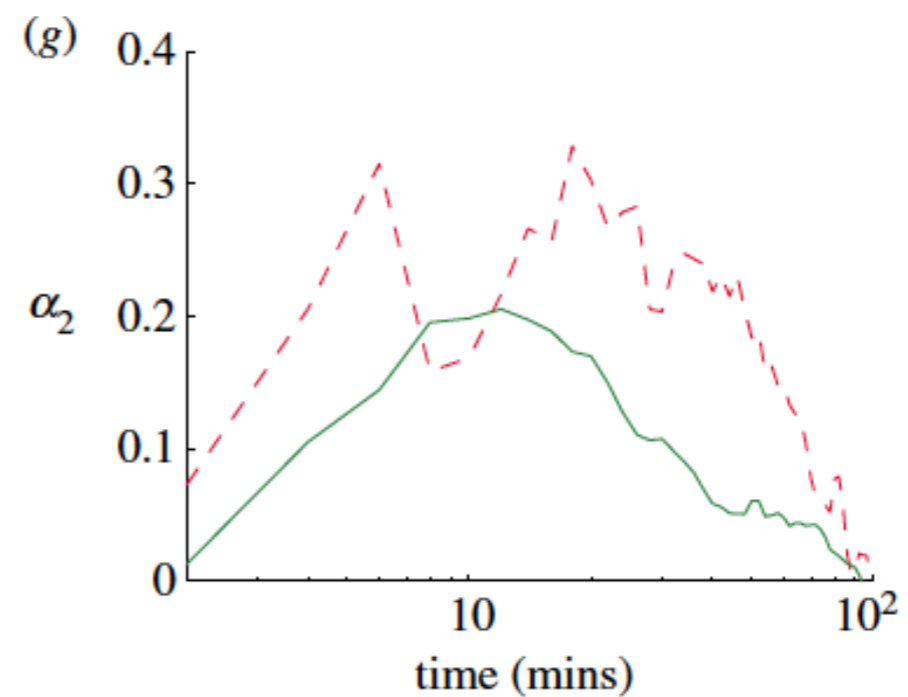
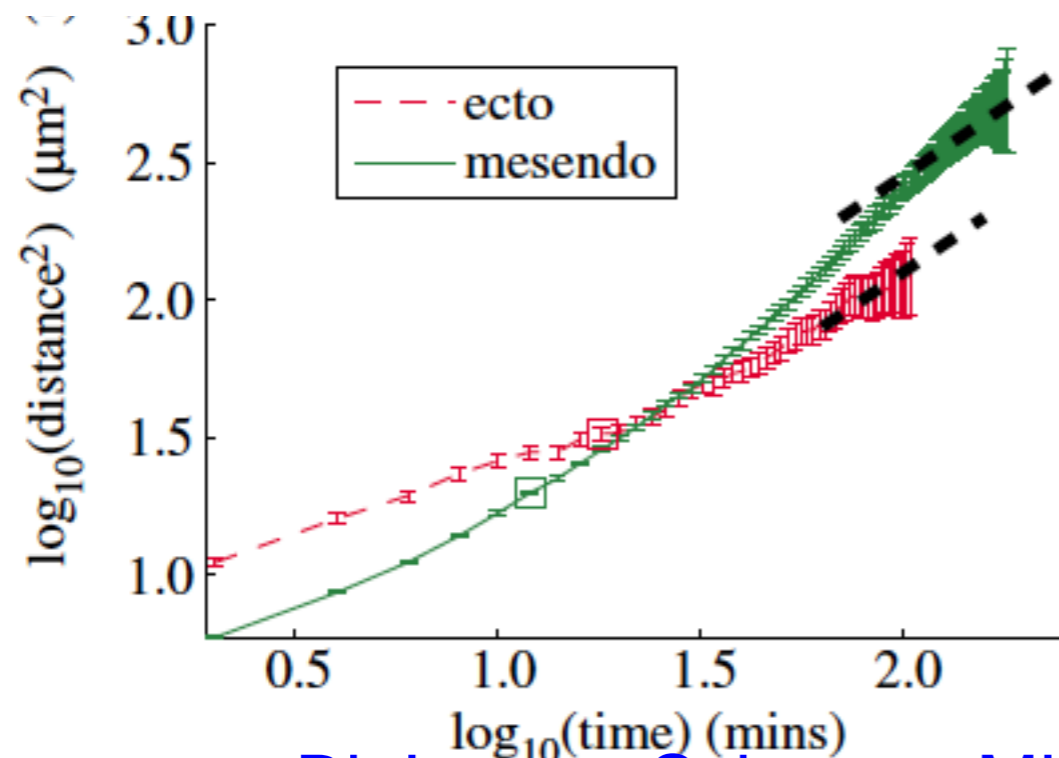
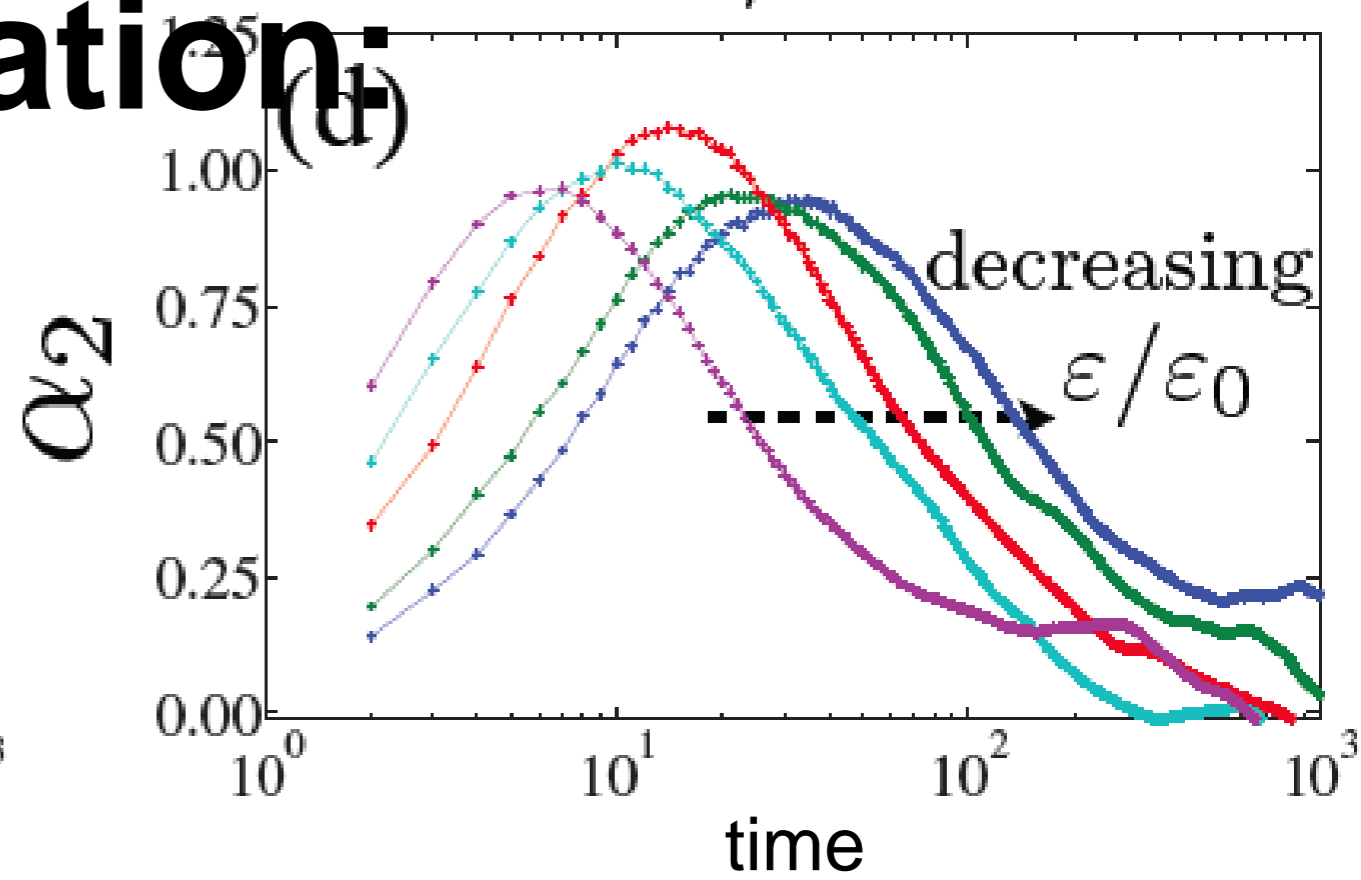
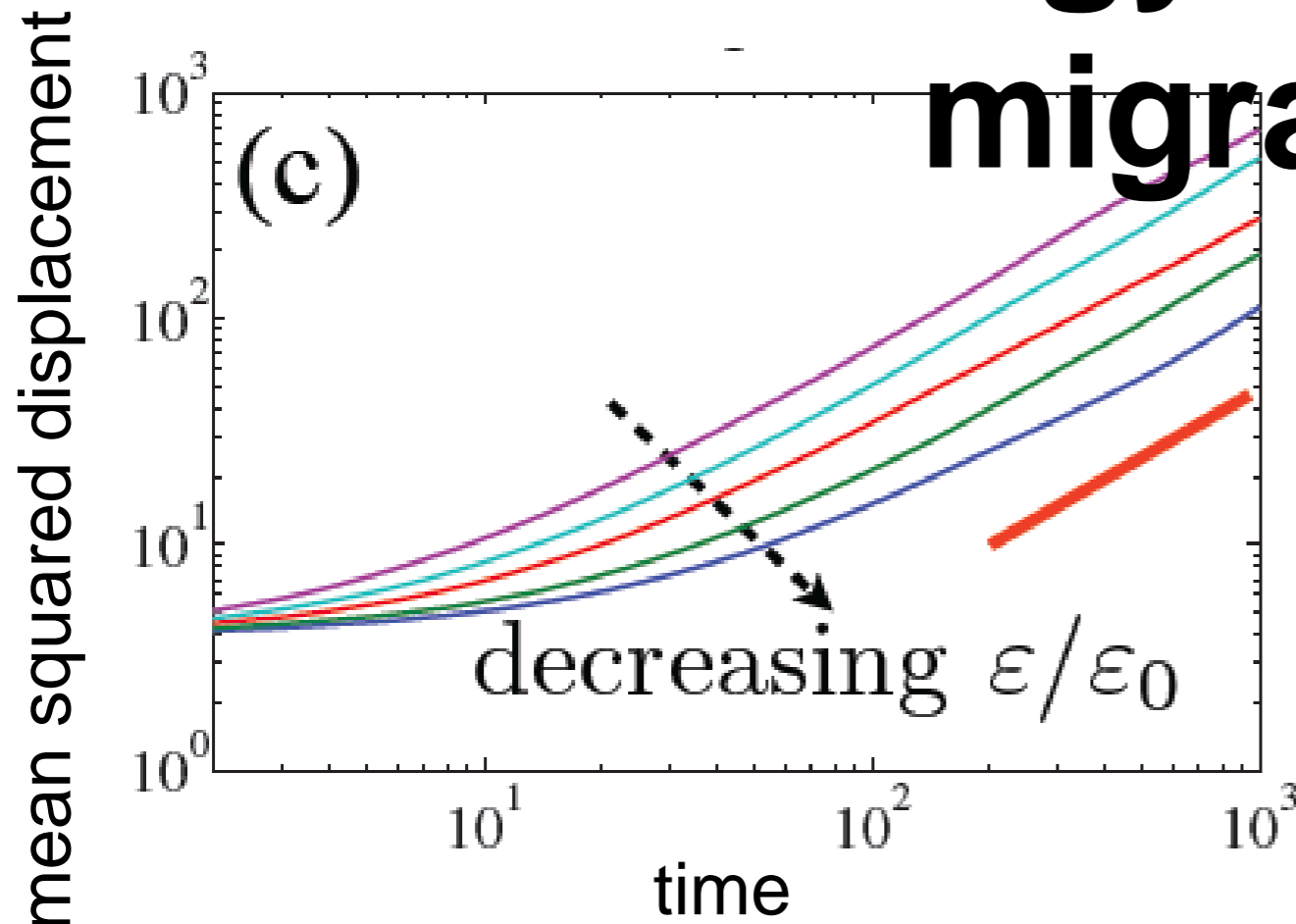
Exponential tail + trap model = glassy dynamics



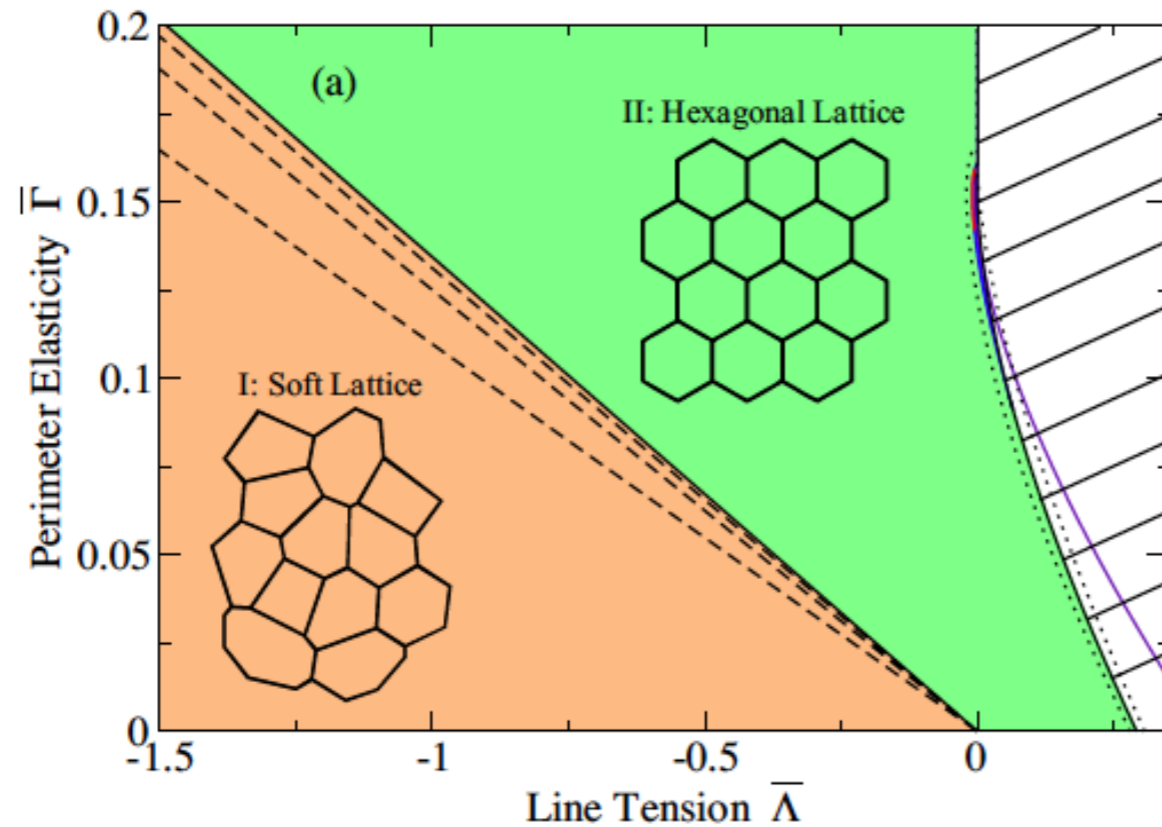
Average energy barrier height vanishes at
 $\rho_0^* \sim 3.813$



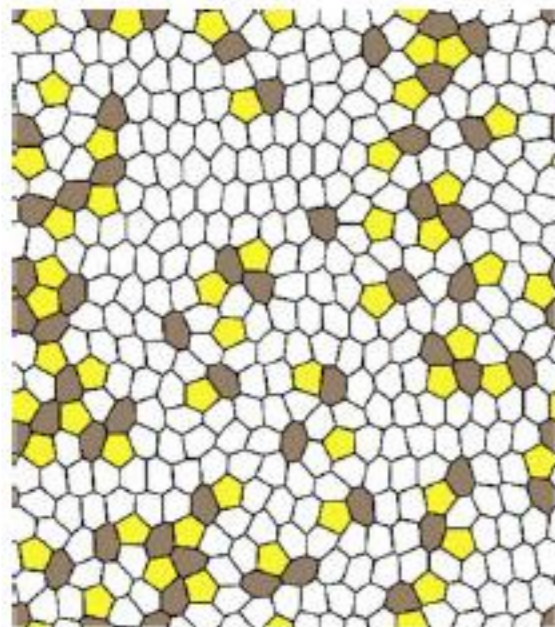
From energy barriers to cell migration:



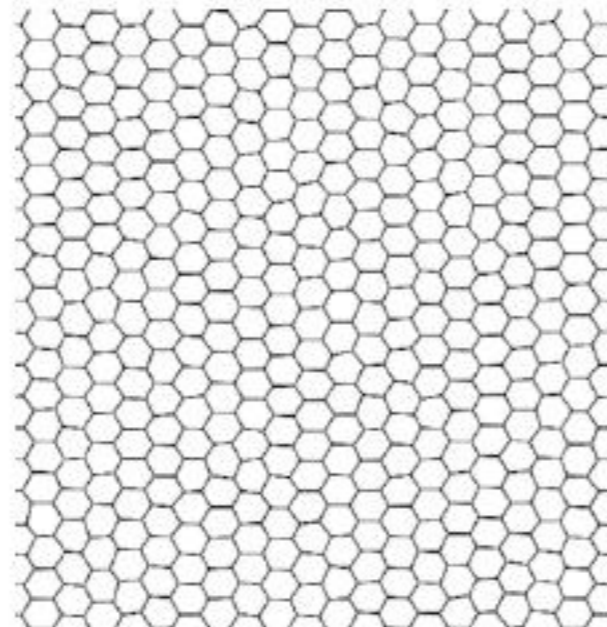
Previous results on ground states:



Staple et al EPJE 33 (2010)



$a = 0.86$

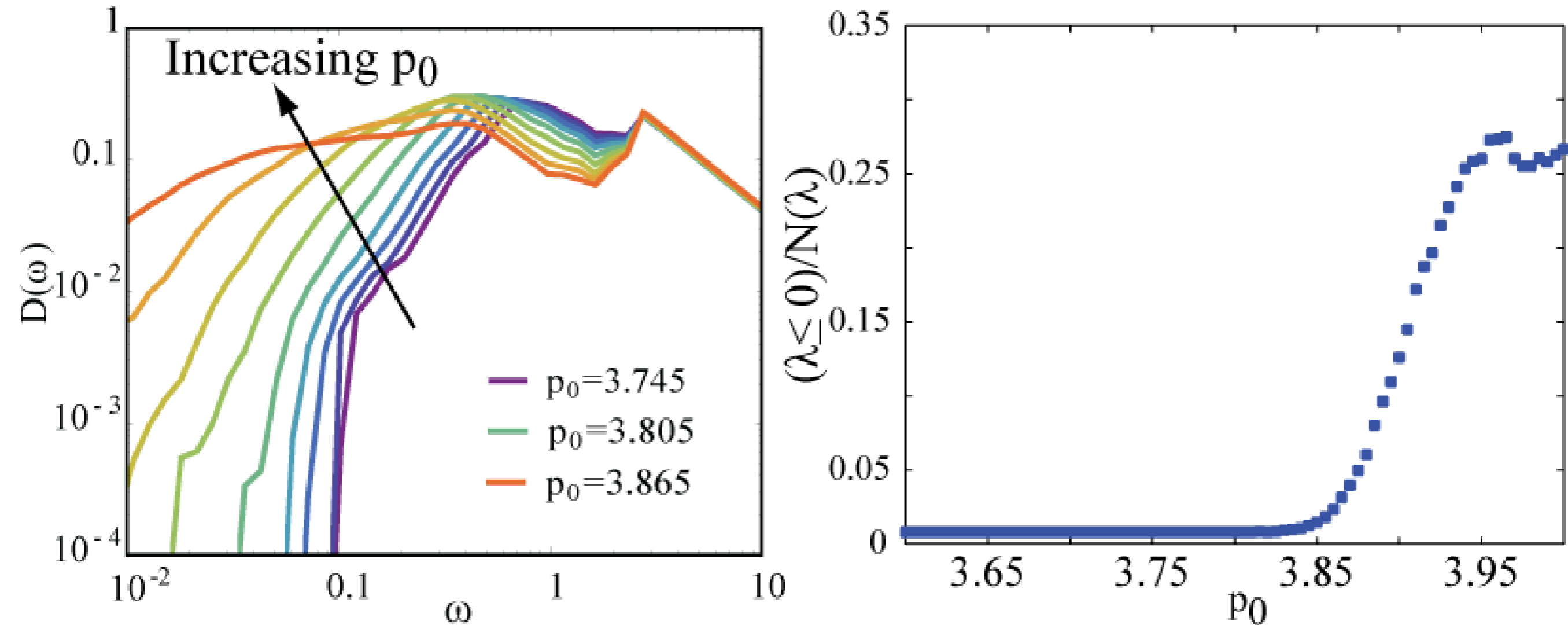


$a = 0.90$

$$a = \frac{4\pi A}{L^2}$$

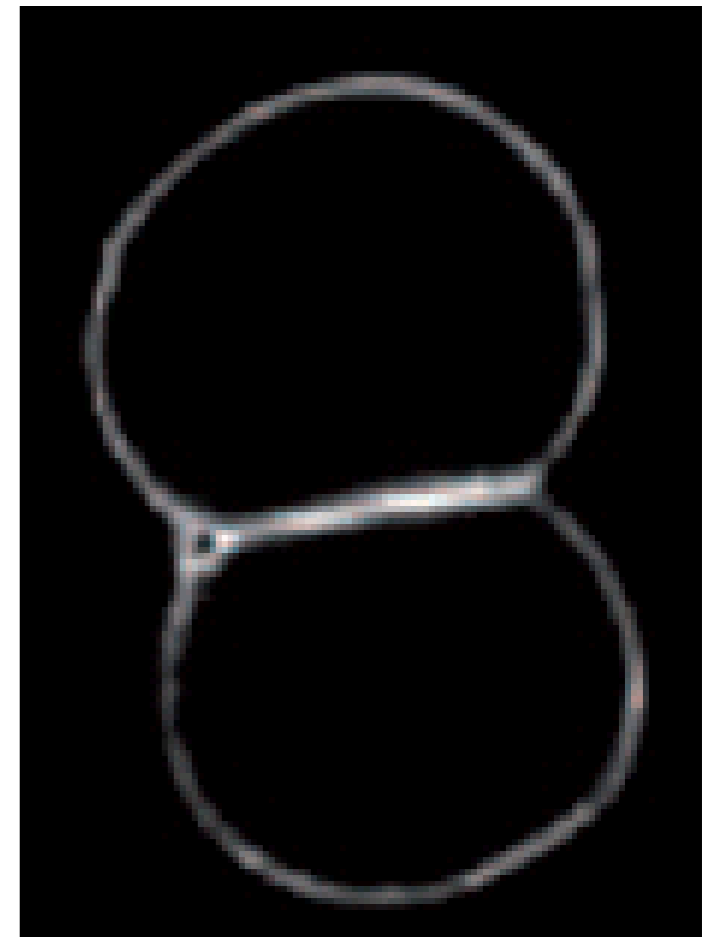
Hocevar and Ziherl PRE 80 11904 (2009)

What about normal modes? Collective behavior? Is it okay to study localized T1s?



“Shape equilibrium” or “vertex” model: what mechanical forces act to generate cell shapes?

1. **Cell-cell adhesion**: cadherins, alpha-catenin, beta catenin, etc.
2. **Active cortical tension**: myosin II, actin (Experiments: Evans, Theory: Joanny, Prost et al)
3. Bulk effects: fluid **resists dialation/compression**, cytoskeleton resists shear
4. Cortical elasticity: cytoskeletal networks



Devries et al,
Development **131**,
4435–4445 (2004)