

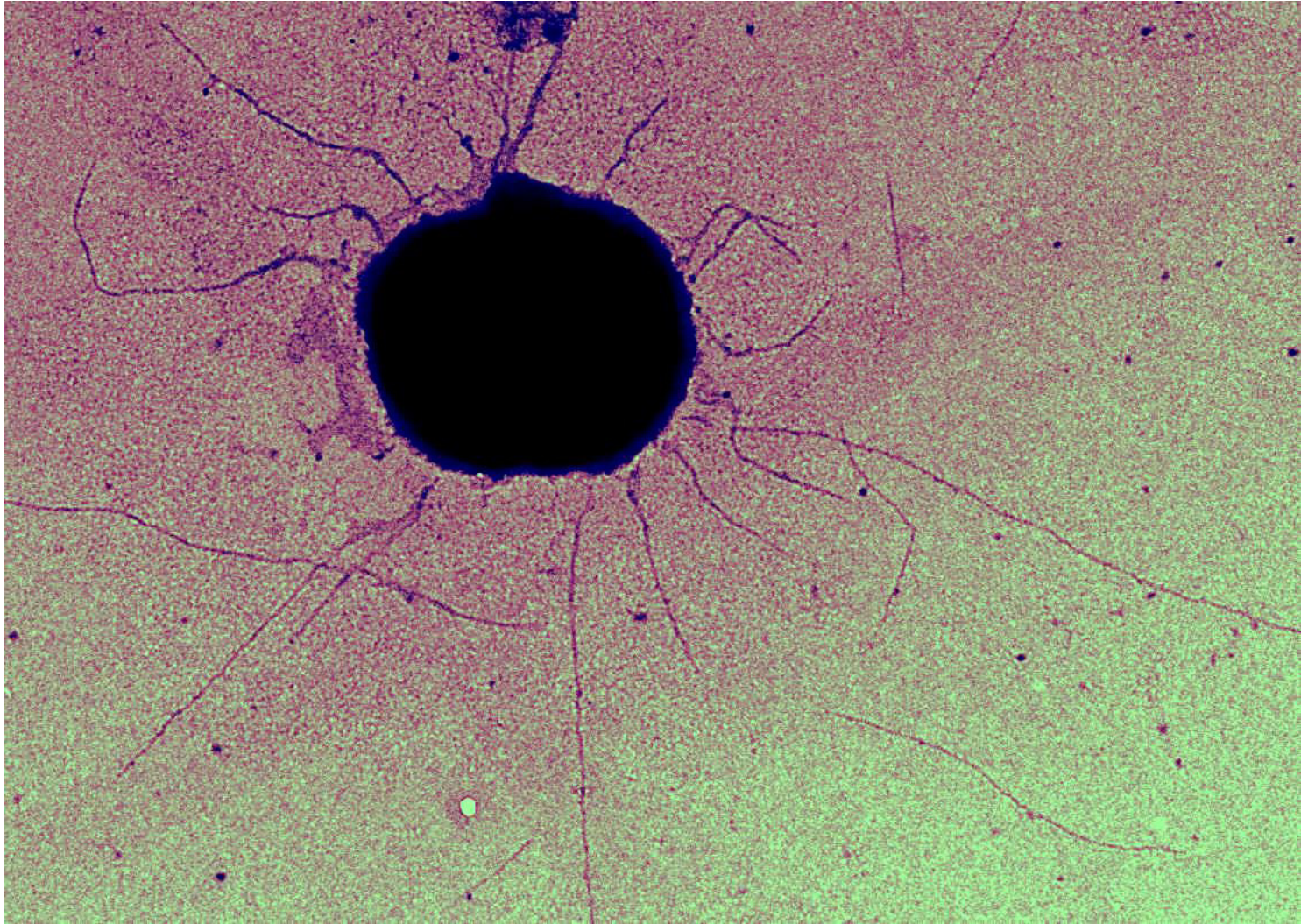
The type IV pilus - a multifunctional bacterial motor

Berenike Maier

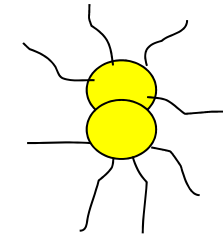
WWU Münster

Kalvi Institute, 1.2.2011

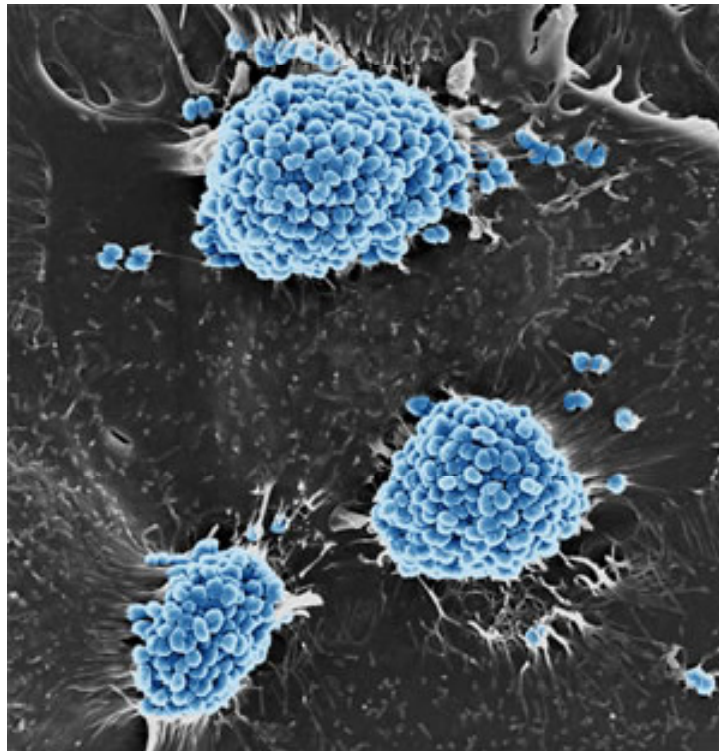
Type IV pili are ubiquitous multifunctional polymers



Type IV pili are ubiquitous multifunctional polymers

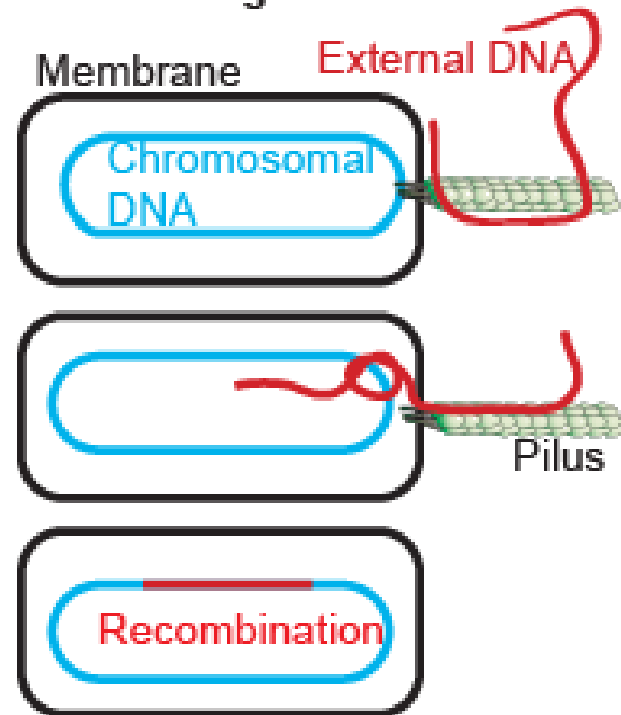


Neisseria gonorrhoeae:
genome size 2Mb



Higashi et al, 2007

Horizontal gene transfer

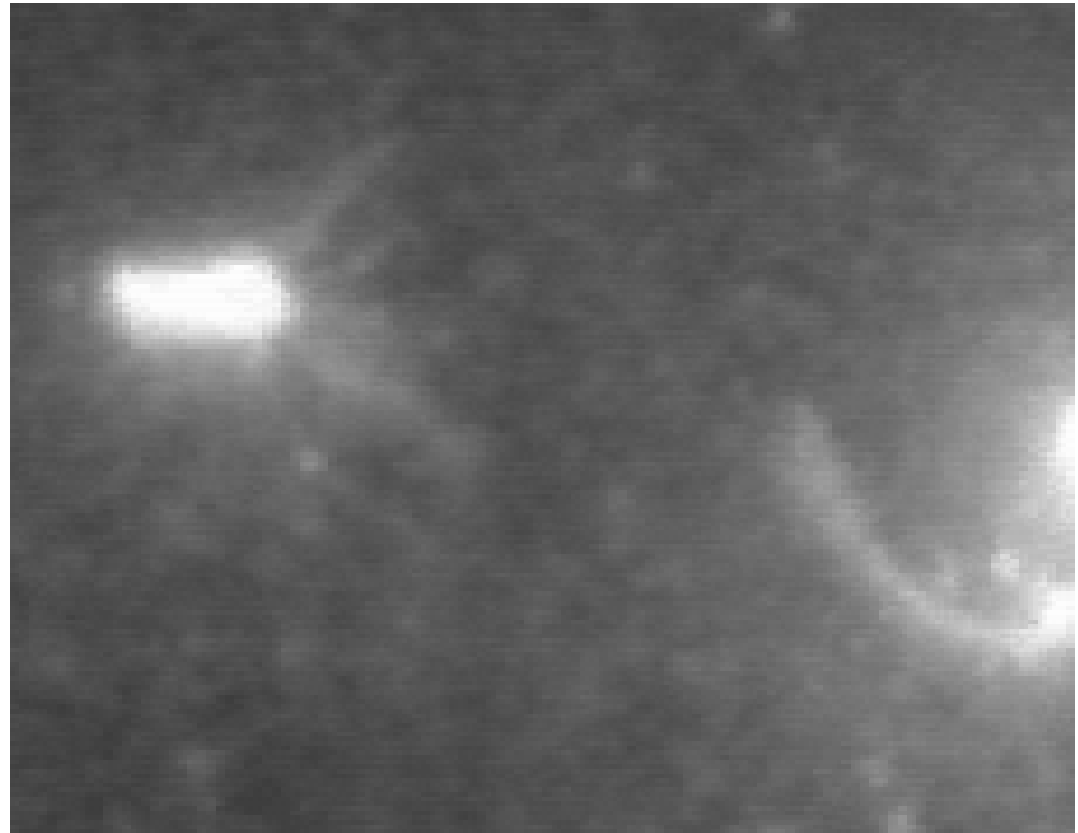


Functions include:

virulence, adhesion, microcolony formation, biofilm formation, motility, gene transfer

Type IV pili are ubiquitous multifunctional polymers

Pseudomonas aeruginosa:



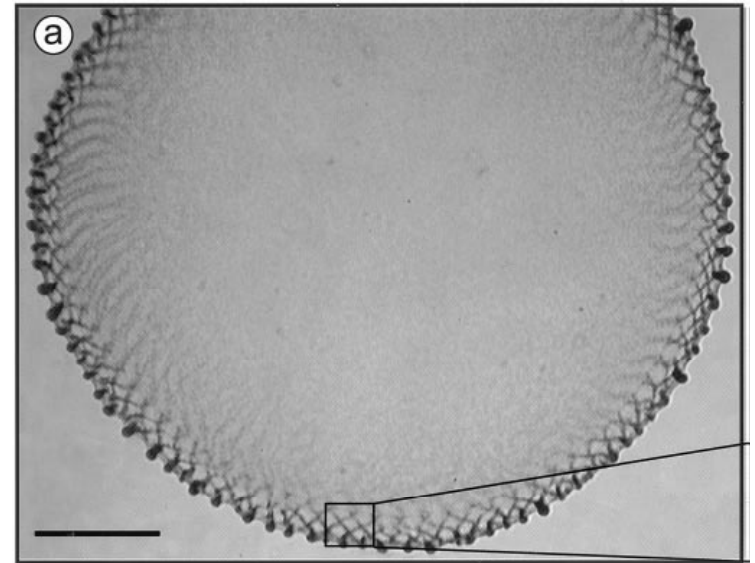
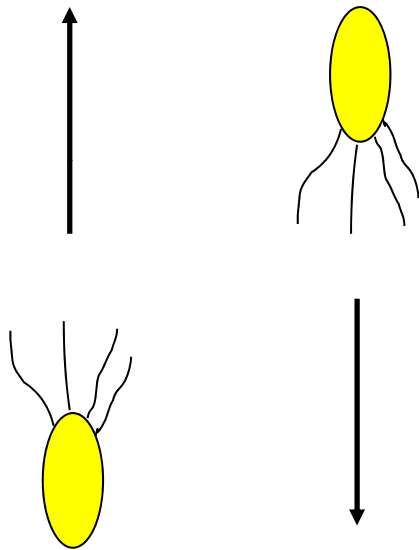
Functions include:
adhesion, motility, biofilm formation, lipid-based chemotaxis

Skerker, Turner, Berg, PNAS 2001

Type IV pili are ubiquitous multifunctional polymers

Myxococcus xanthus:
genome size 9Mb

8 min reversal time

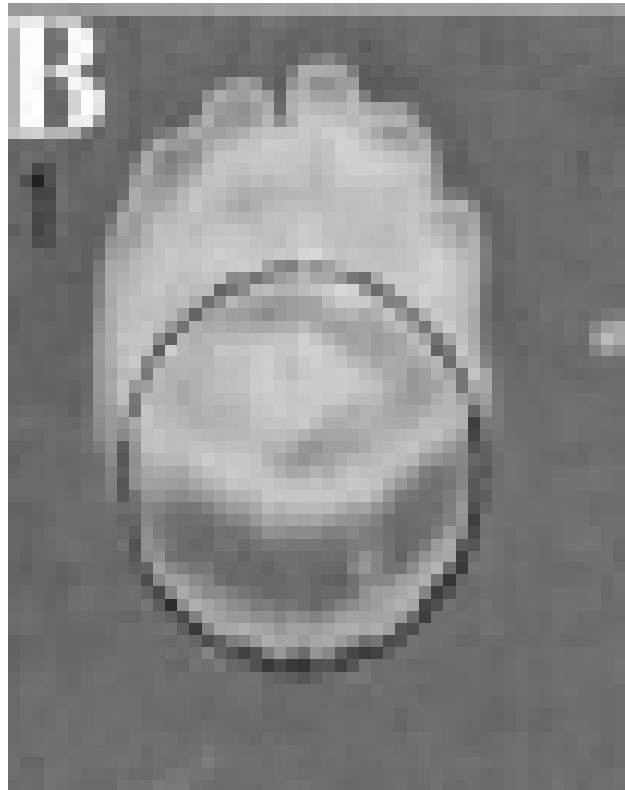


Welch & Kaiser, PNAS 2001

Functions include:
adhesion, motility, fruiting body formation, lipid-based chemotaxis

Type IV pili are ubiquitous multifunctional polymers

Synechocystis PCC6803:

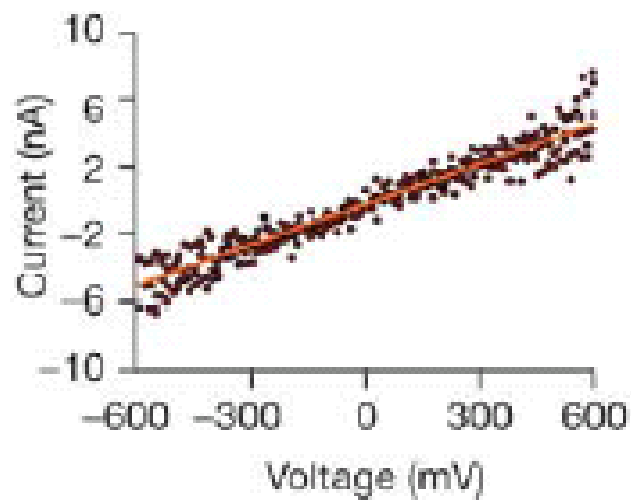
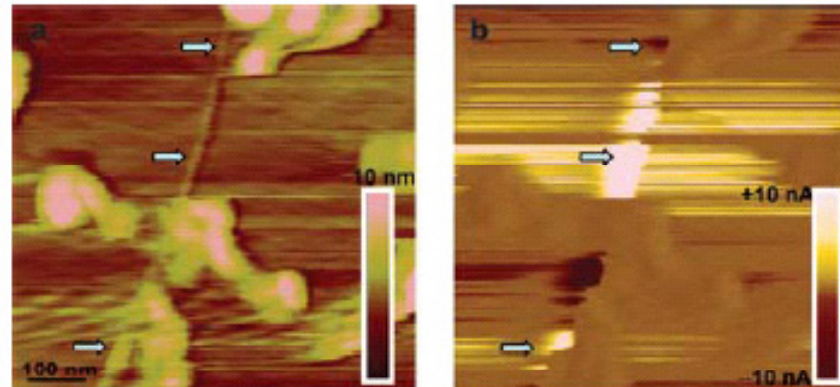


Bhaya et al PNAS 2002

Functions include:
motility, phototaxis

Type IV pili are ubiquitous multifunctional polymers

Geobacter sulfurreductens:



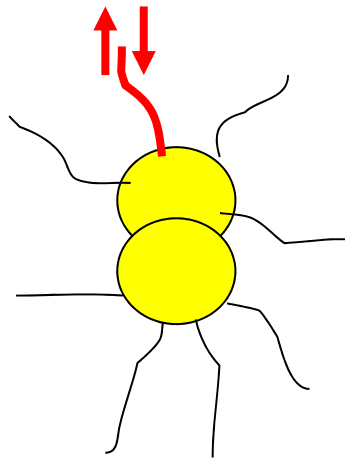
Functions include:
electron transfer

Reguera et al. Nature 2005

The type IV pilus is ubiquitous amongst phylogenetically diverse bacterial species and support different functions

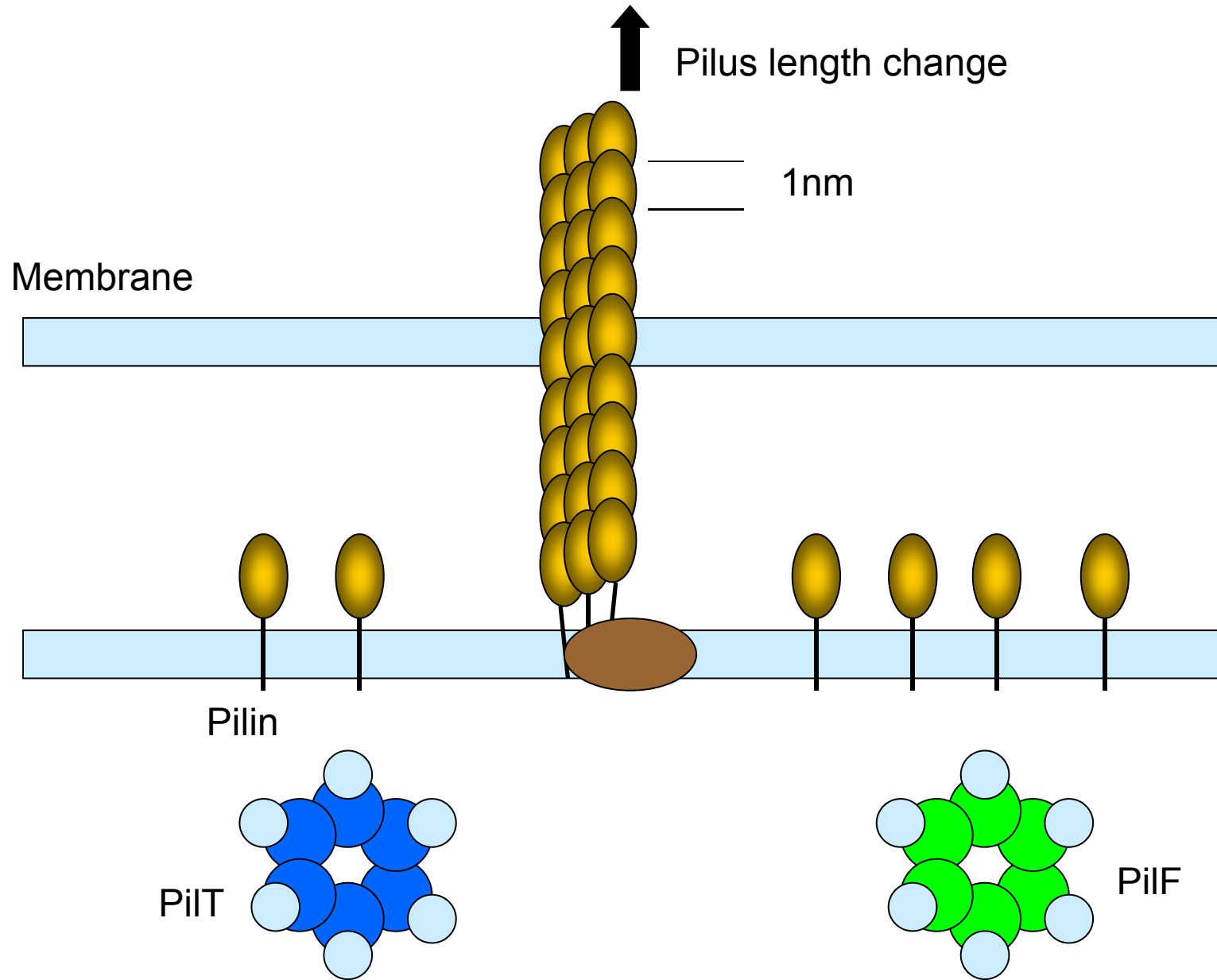
Question: Have the biophysical properties of pili adapted differently to support different phenotypes?

Biophysics of individual pili

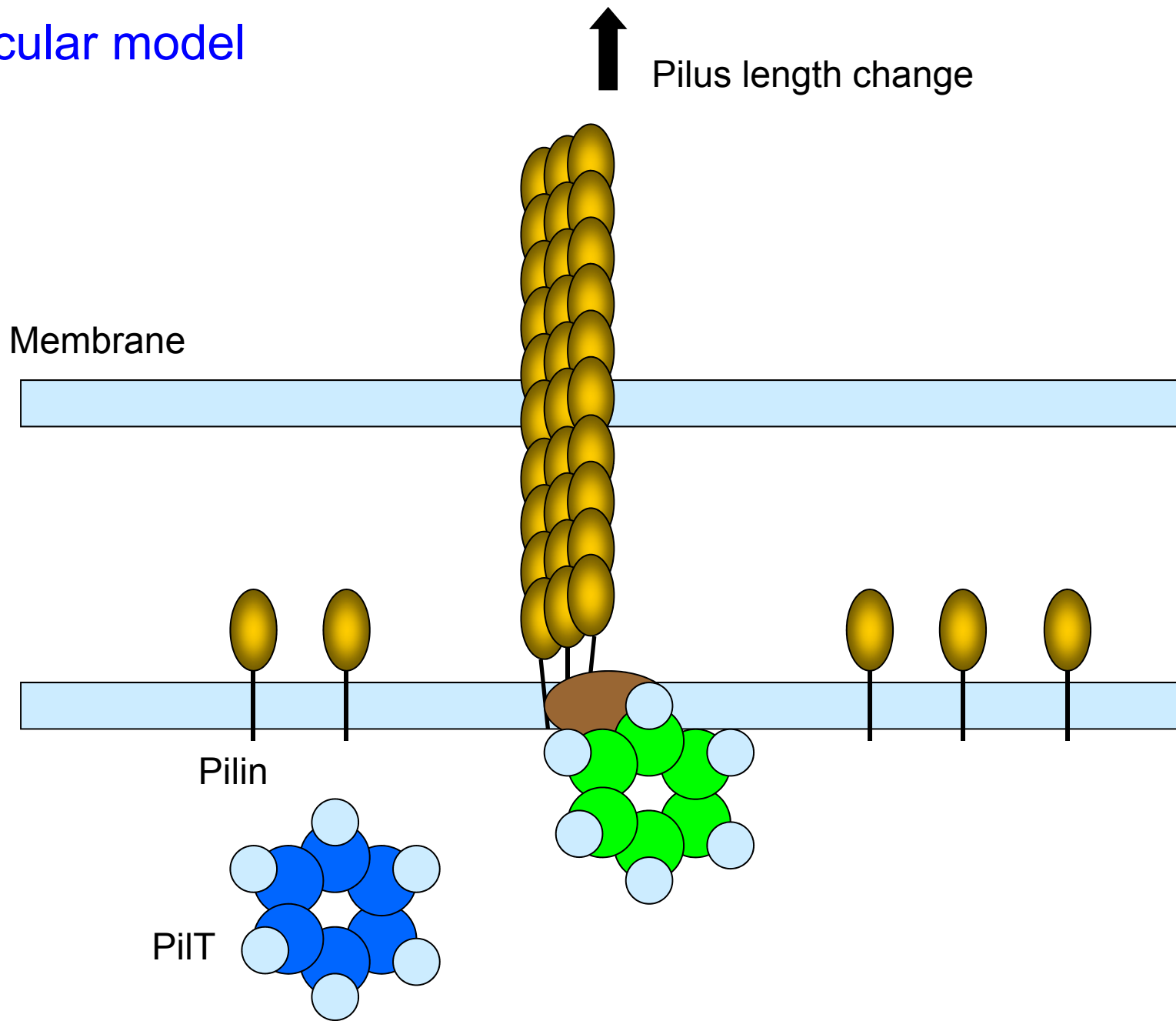


Martin Clausen, Rainer Kurre
Michael Koomey, Oslo University

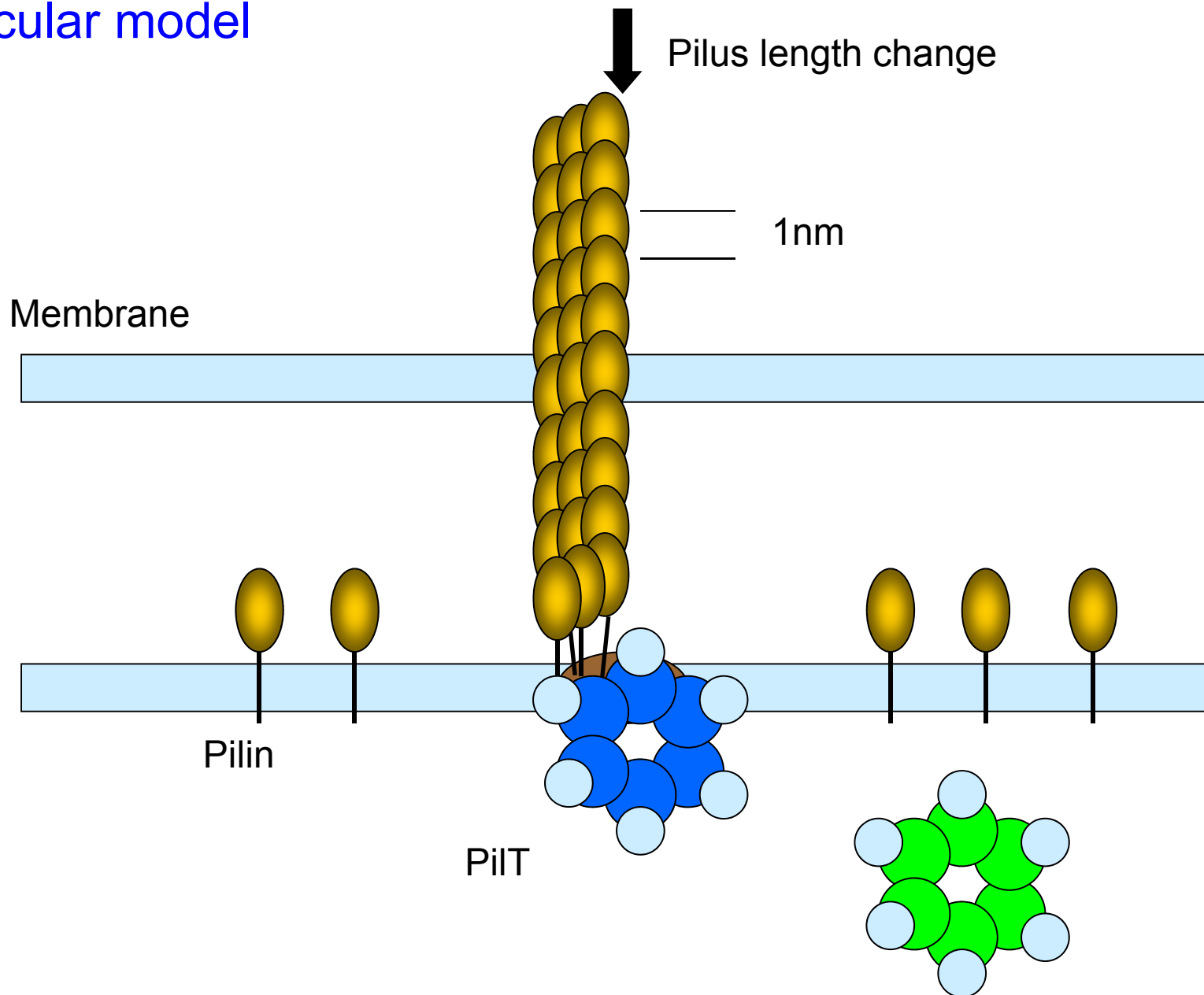
Molecular model of pilus dynamics



Molecular model



Molecular model



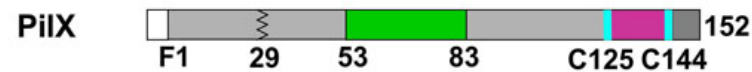
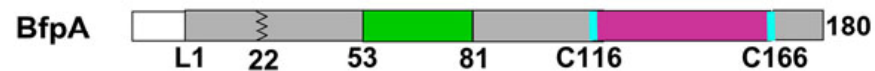
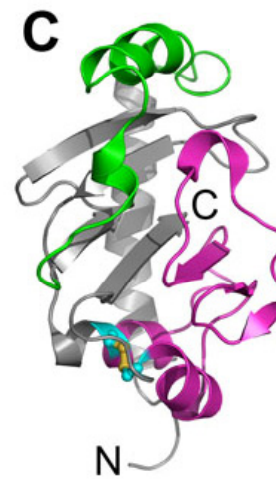
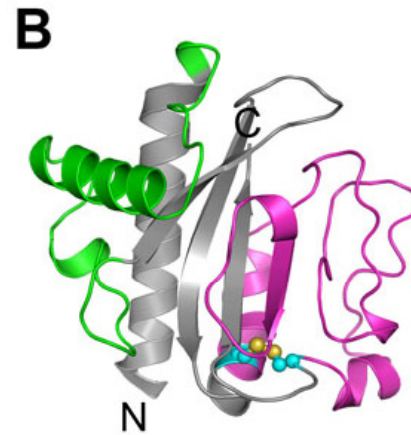
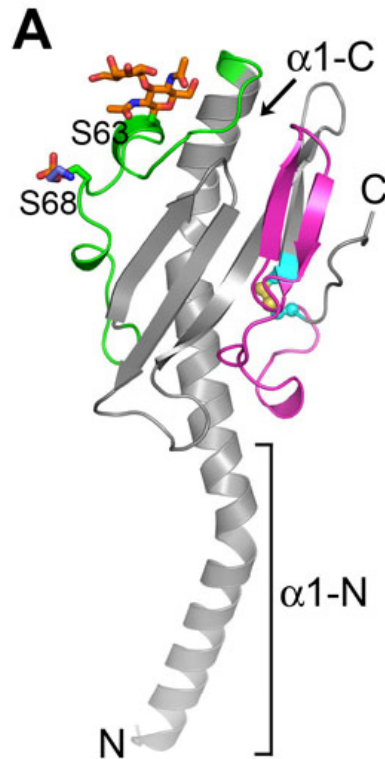
Pilin structure of different species

Neisseria gonorrhoeae

Vibrio cholerae

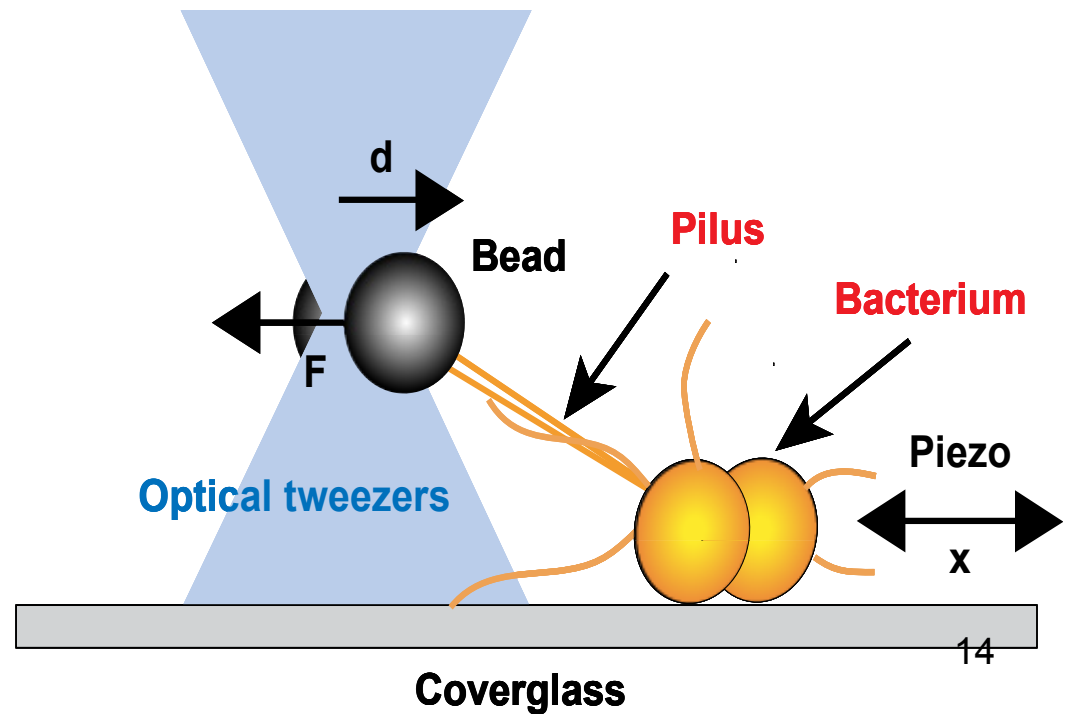
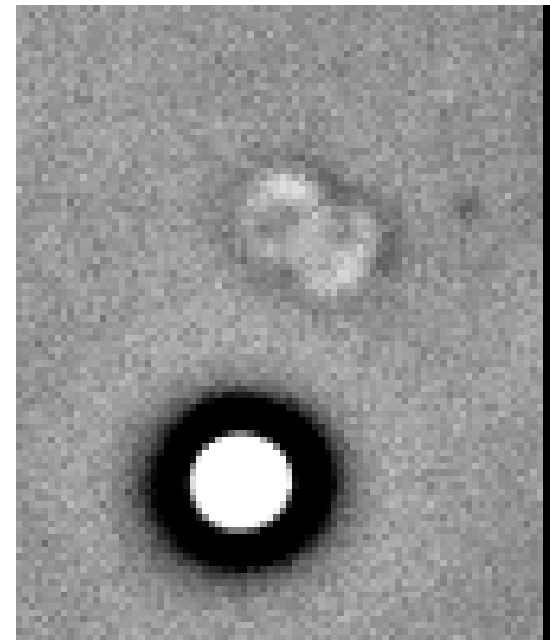
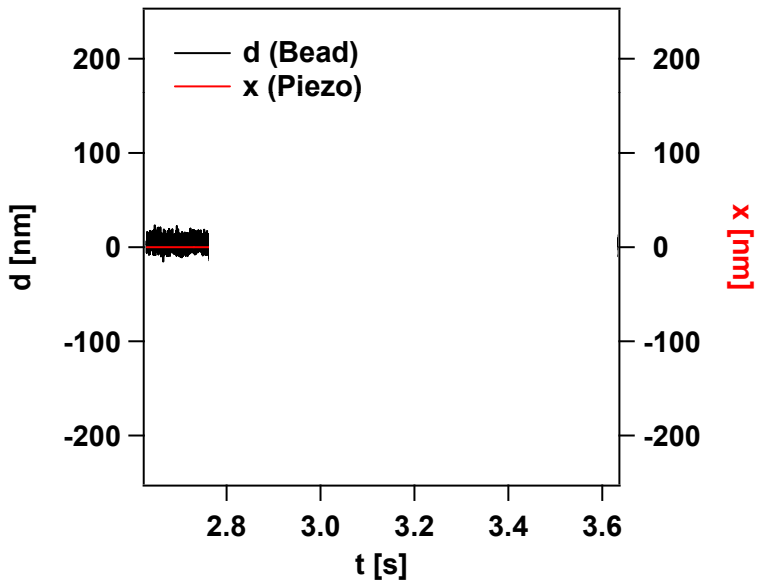
enteropathogenic
Escherichia coli

minor pilin
Neisseria meningitidis



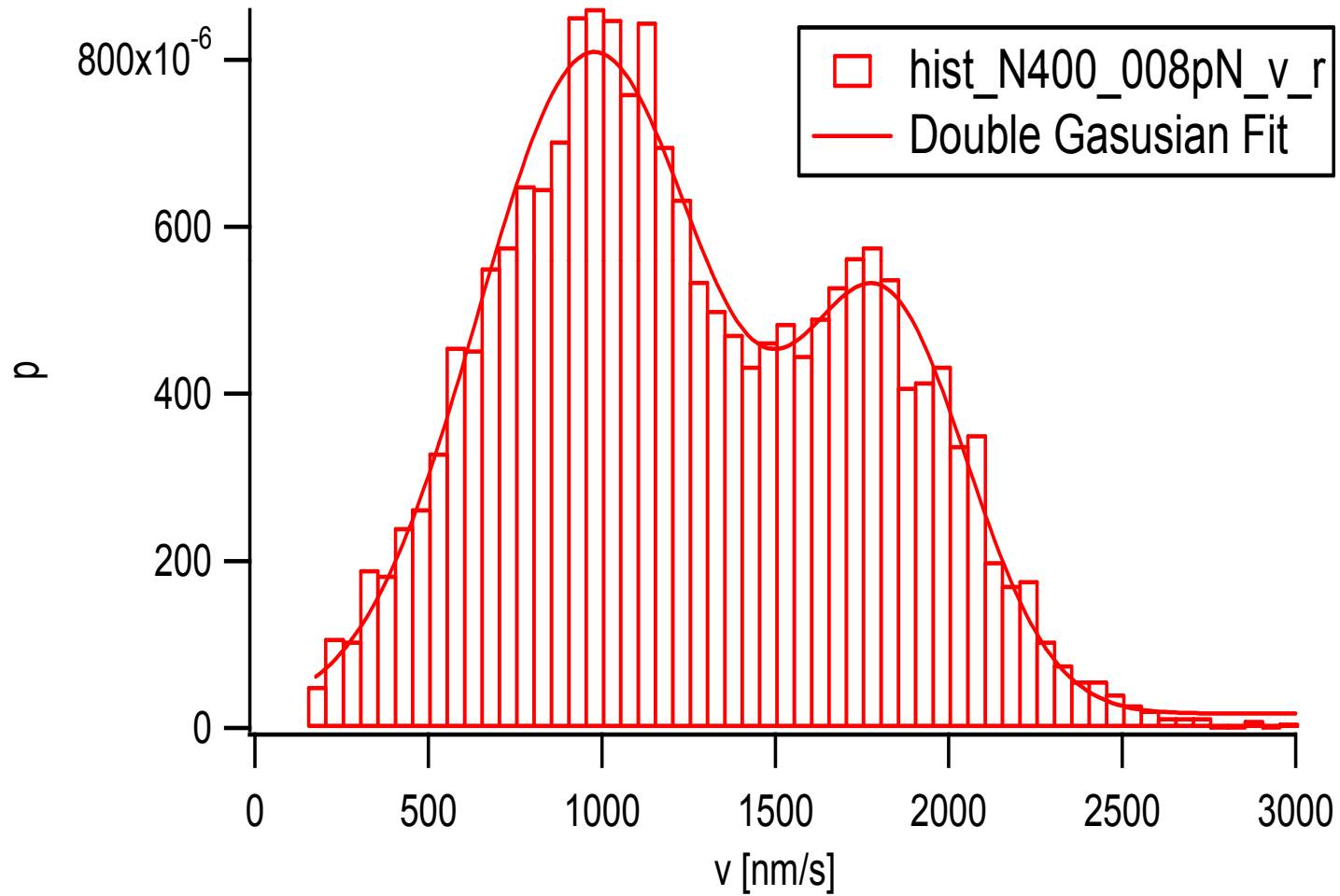
Single pilus dynamics measured with clamped force

Bacterial attachment to a surface via force bead



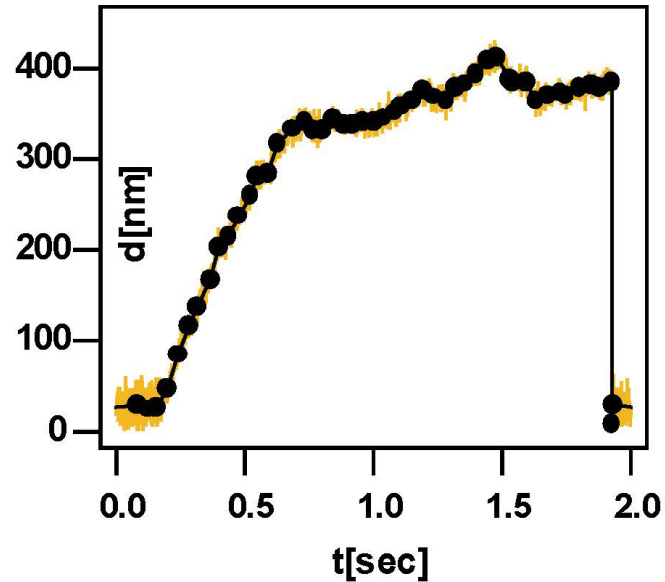
Velocity of type IV pilus retraction is bimodal

N. gonorrhoeae

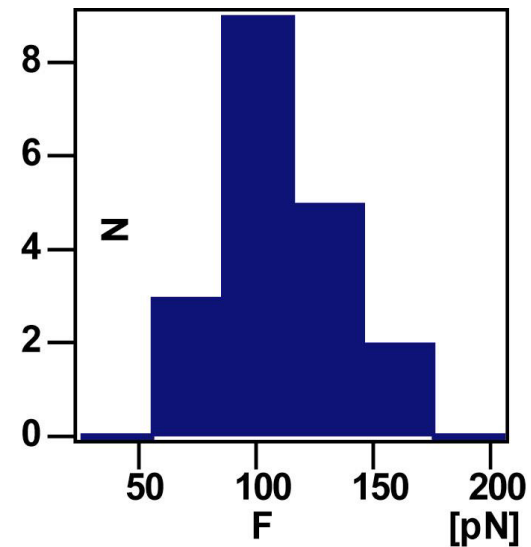


Type IV pili generate force $> 100\text{pN}$ by retraction

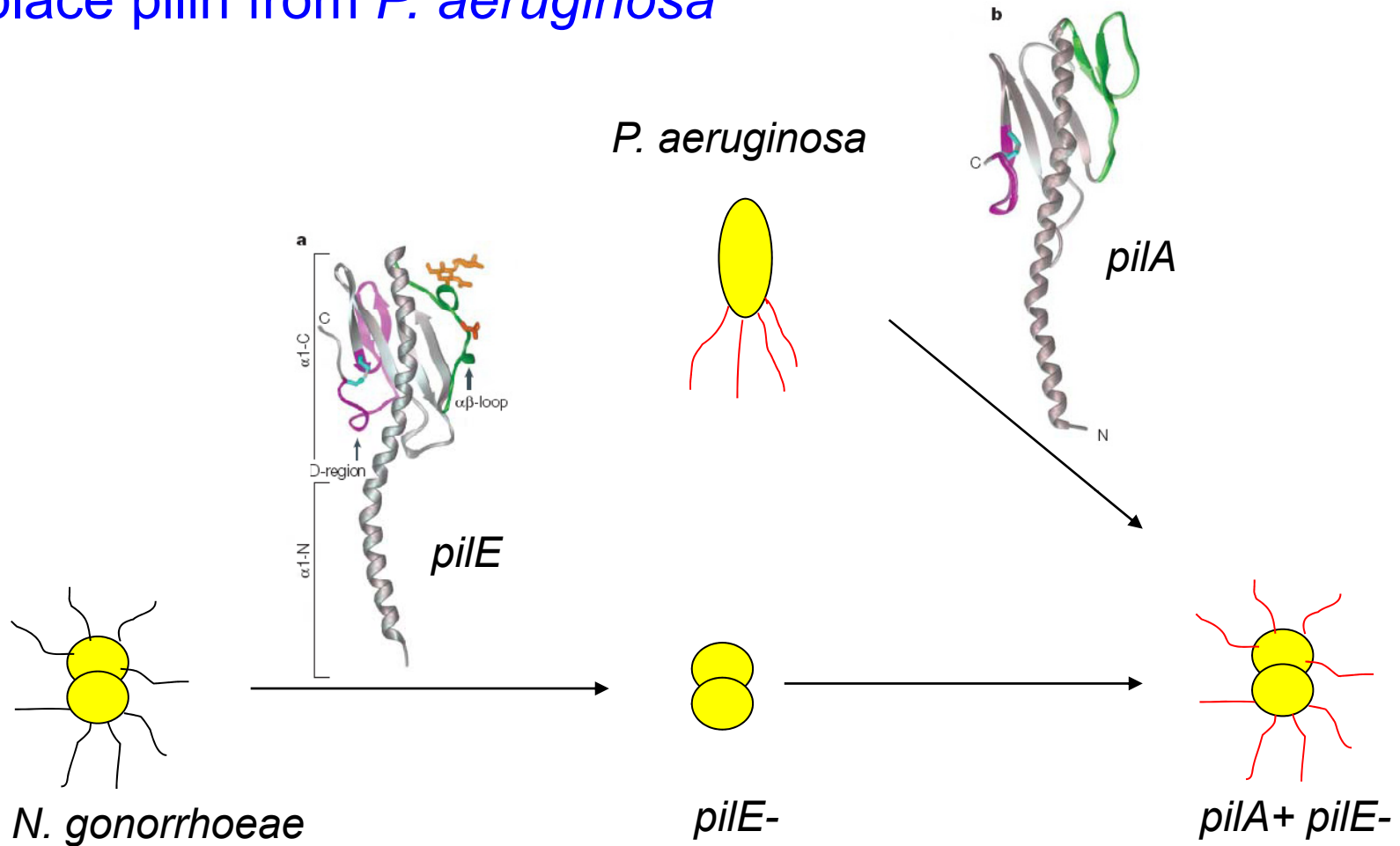
Displacement of the bead from the center of the laser trap:



Distribution of stalling forces:



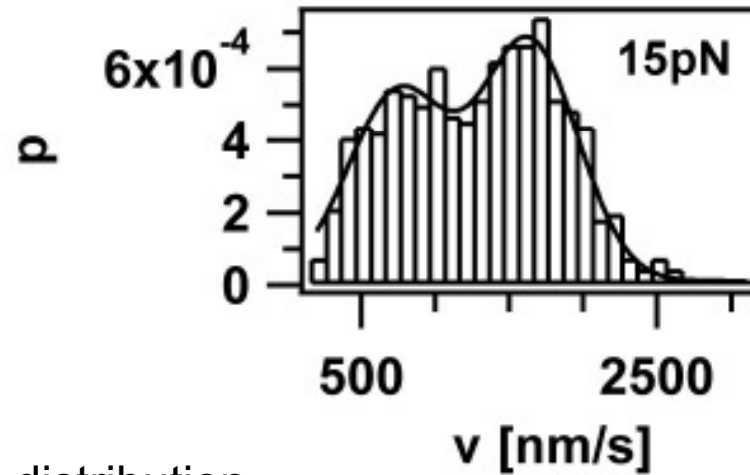
Replace pilin from *P. aeruginosa*



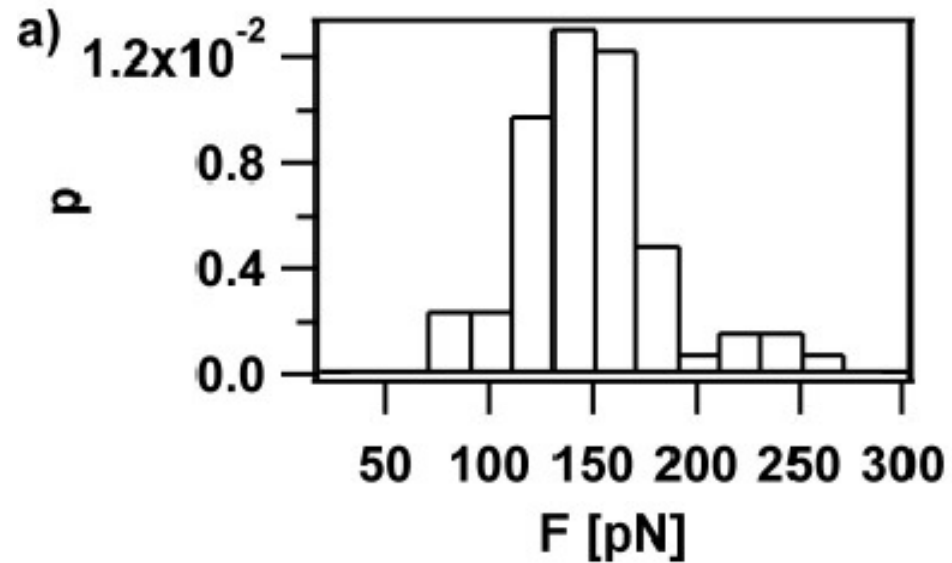
Result: comparable velocity and stalling force

Velocity and force generation in *M. xanthus*

Velocity distribution



Force distribution



Conclusion

The biophysical properties of T4 pili are well conserved at the level of a single pilus.

What about velocity regulation?

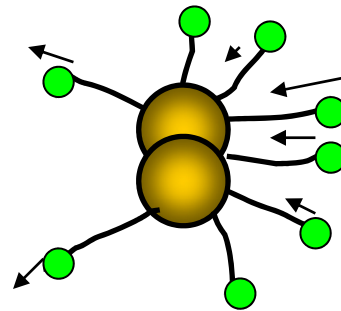
Maier et al, PNAS 2002

Winther-Larsen et al J. Bacteriol. 2007

Clausen et al, J. Bacteriol 2009

Clausen et al, Biophys. J. 2009 19

How is pilus dynamics translated into movement?

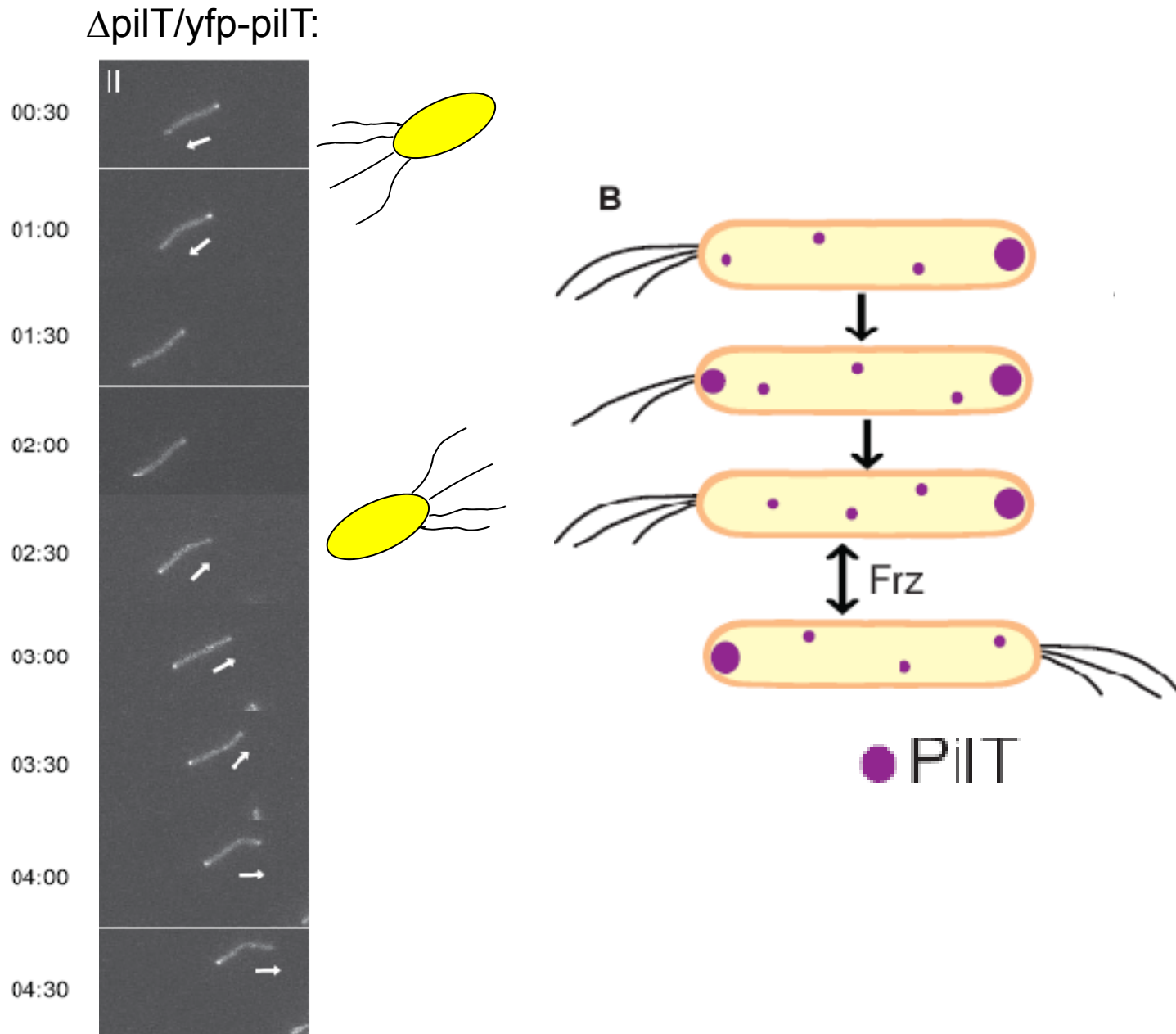


Claudia Holz, Dirk Opitz, Rainer Kurre, Andrea Höne
Lilo Greune, Alexander Schmidt, ZMBE Münster
Iryna Bulhya, Lotte Sogaard-Andersen, MPI Marburg

M. xanthus: Coordinated movement

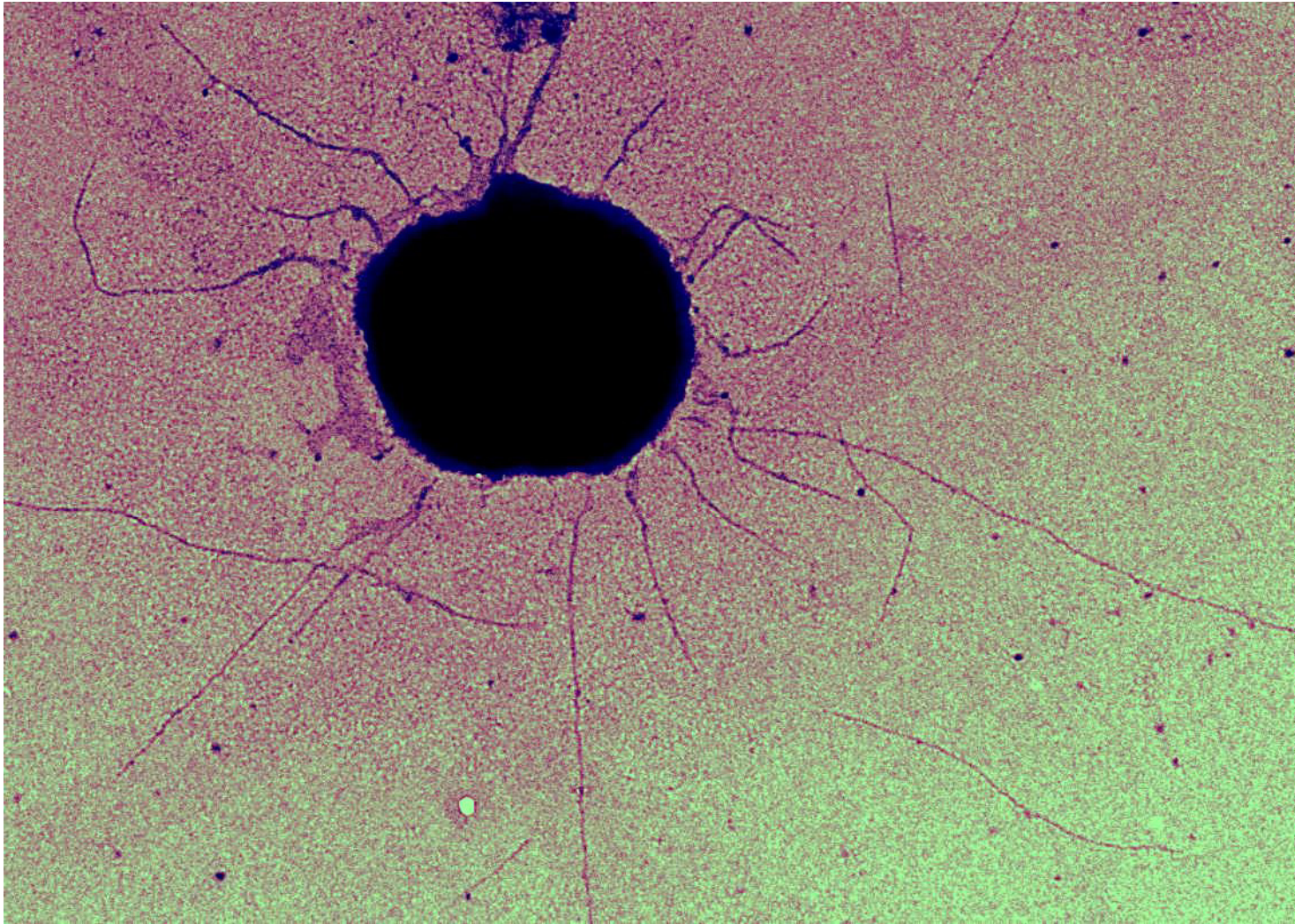


M. xanthus: PilT foci are well coordinated during directional reversal



Pili are randomly distributed around *N. gonorrhoeae*

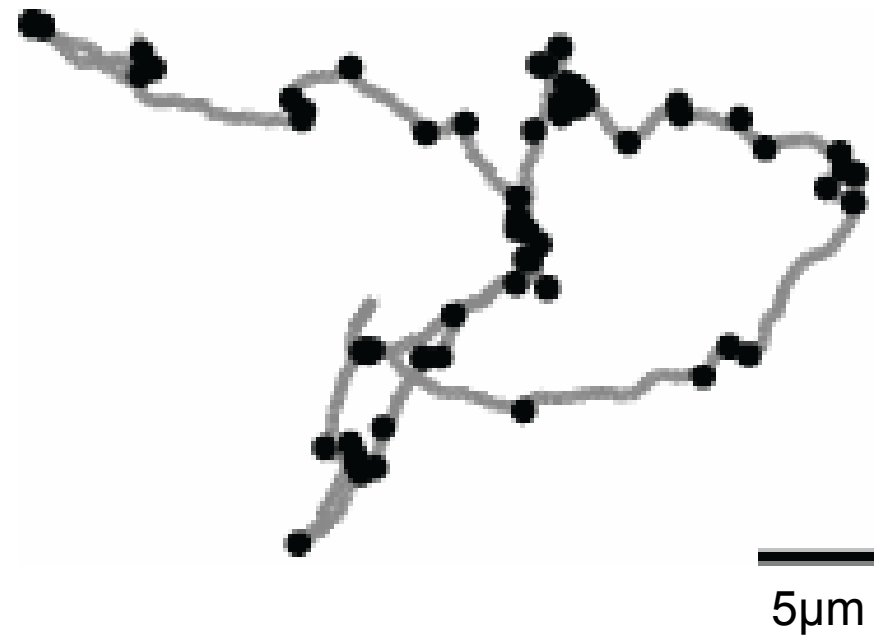
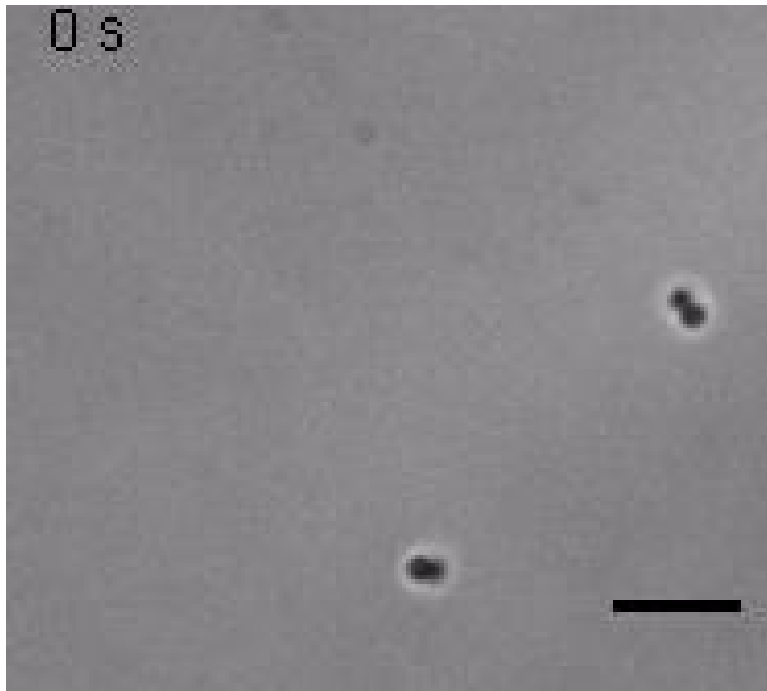
TEM of motile bacterium:



Twitching observed with 3-27 pili

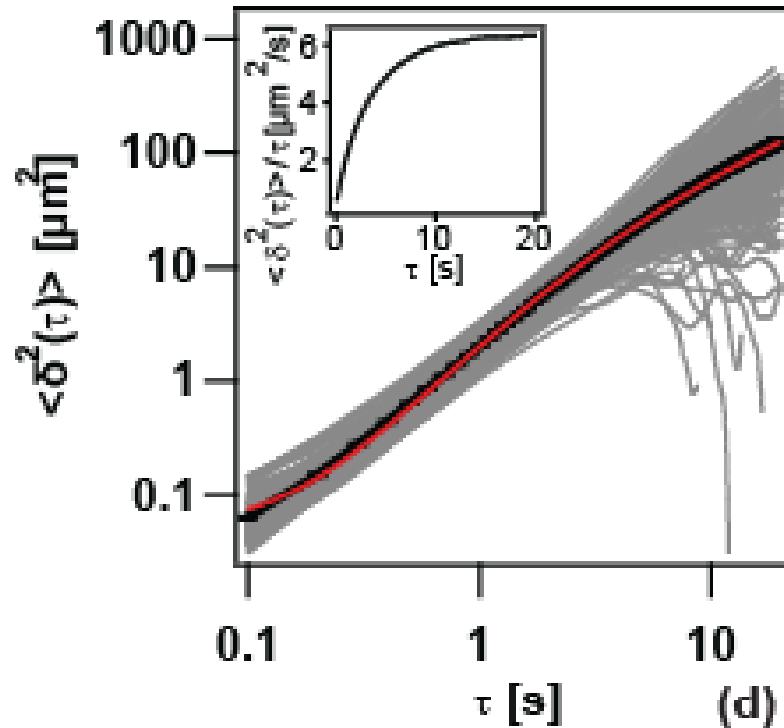
Bacterial movement is persistent

Typical movement of *N. gonorrhoeae* on BSA-treated glass



Bacterial movement is persistent

Mean squared displacement $\langle \delta^2(\tau) \rangle$
of bacteria:



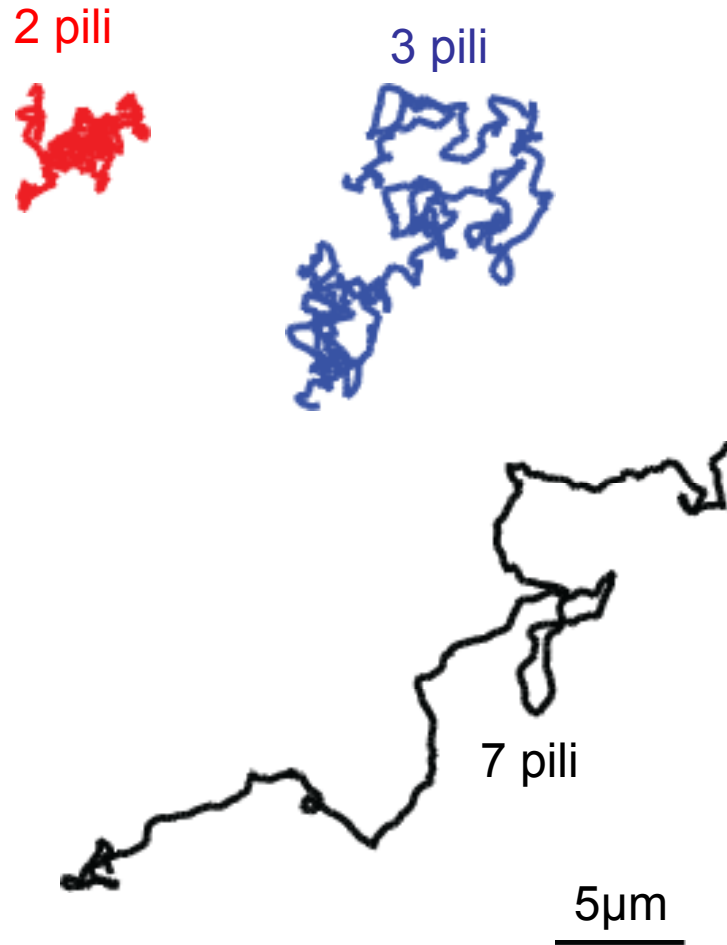
$$\langle \delta^2(\tau) \rangle = 2\tau_c v^2 \left(\tau - \tau_c \left(1 - e^{-\frac{\tau}{\tau_c}} \right) \right) + A$$

$$\tau_c = (1.4 \pm 0.2) \text{ s}$$

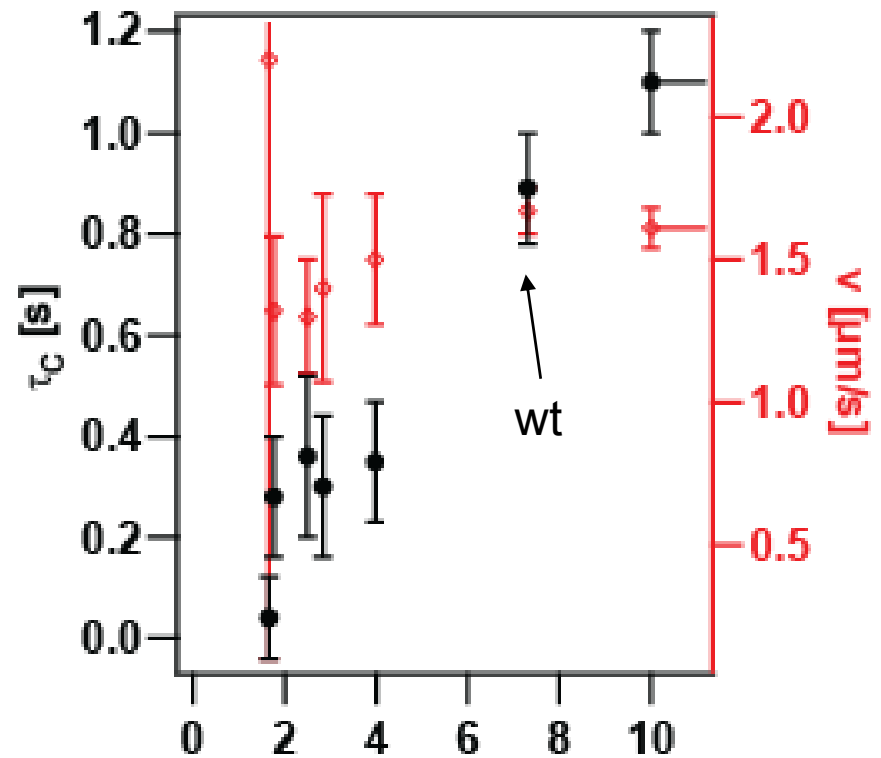
$$v = (1.6 \pm 0.1) \mu\text{ms}^{-1}$$

=> correlation time τ_c longer than pilus retraction time

Multiple pili are required for persistent movement

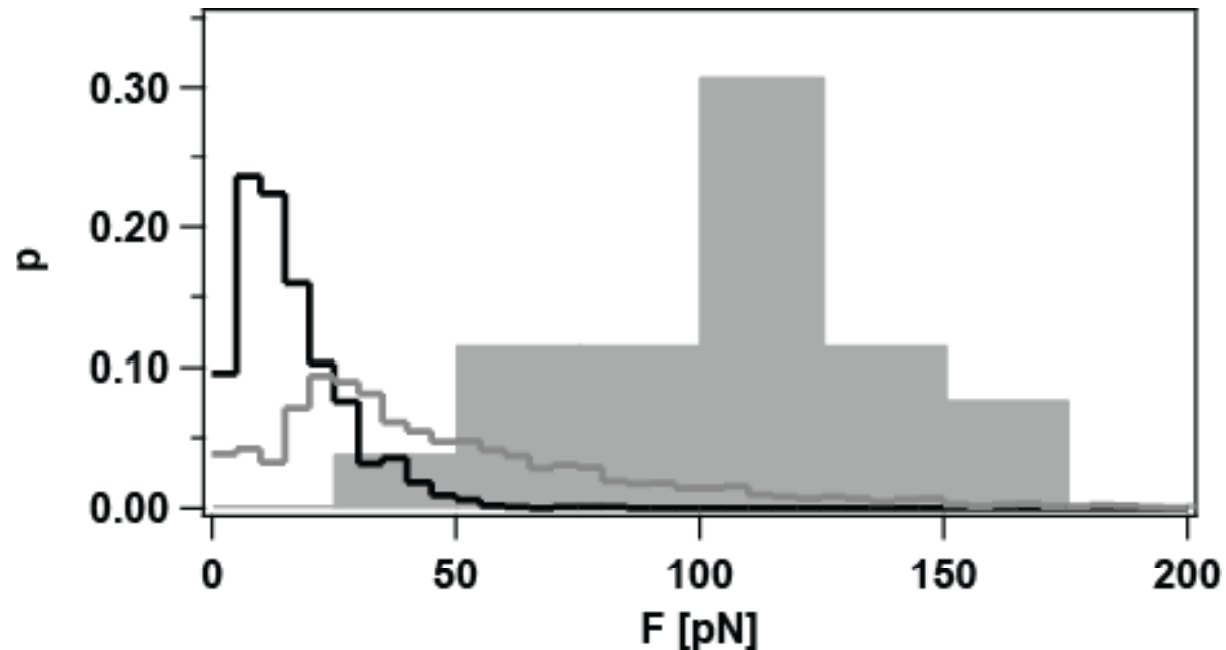


Correlation time τ_c :
Velocity v :



The unbinding force is significantly lower than the motor force

Distribution of forces F :



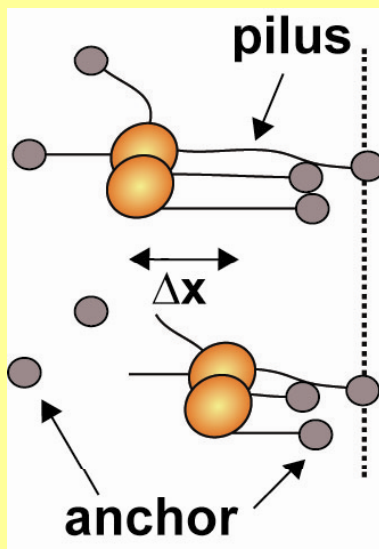
- Unbinding force from BSA-treated glass
- Unbinding force from BSA-treated polystyrol
- Stalling force of individual pilus motors

=> Unbinding force depends on surface and is significantly lower than stalling force

Conclusion

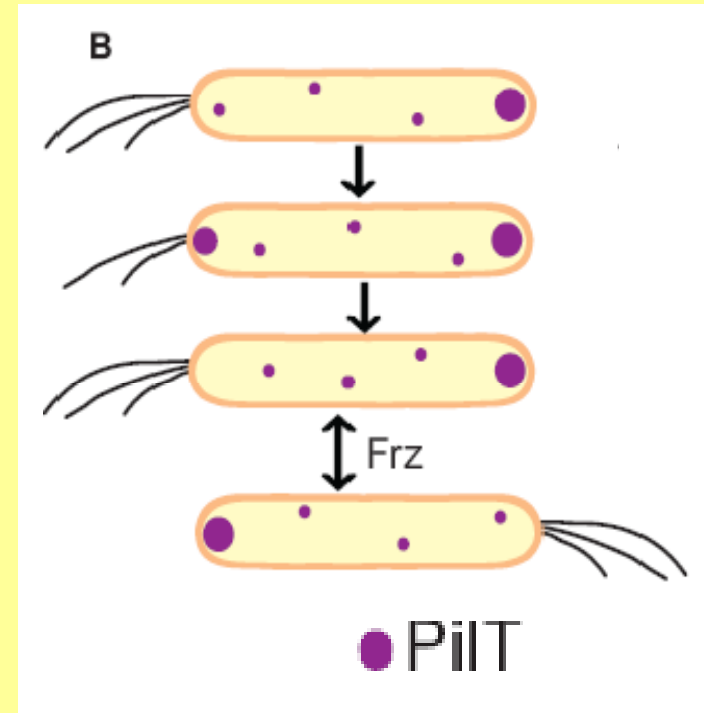
N. gonorrhoeae

- Multiple pili are required for persistent movement
- Unbinding force of pili from surface is significantly lower than motor force



=> consistent with tug-of-war

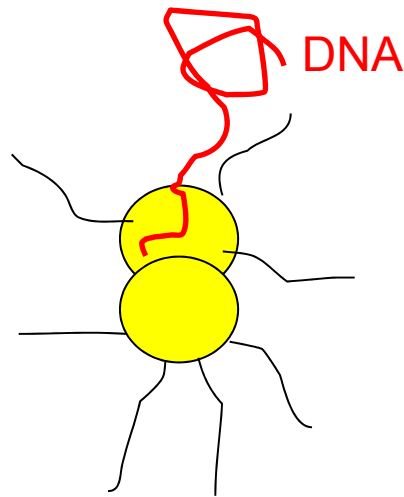
M. xanthus



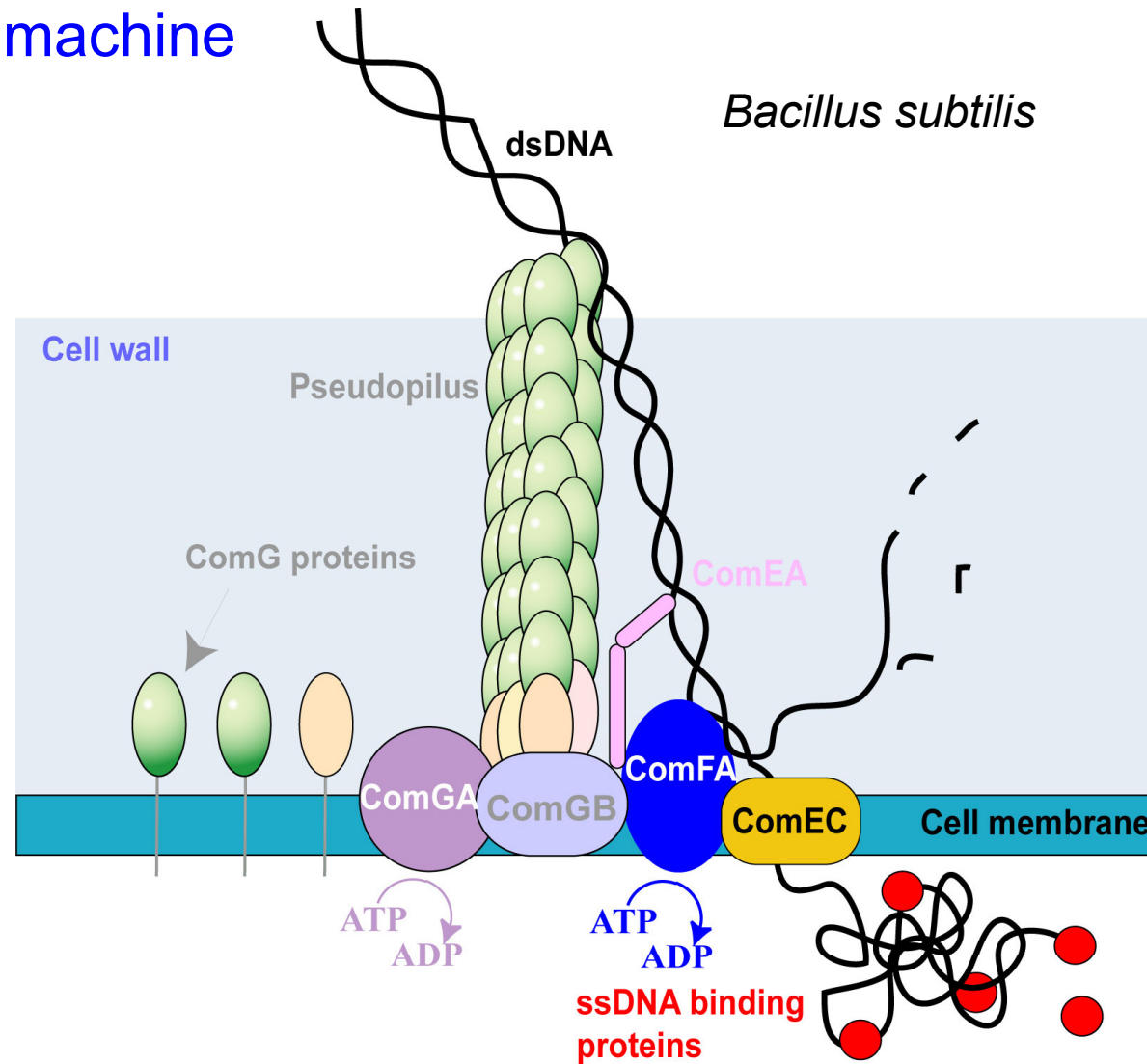
=> highly coordinated motor dynamics

Different machines have evolved for DNA import

Do biophysical properties converge?



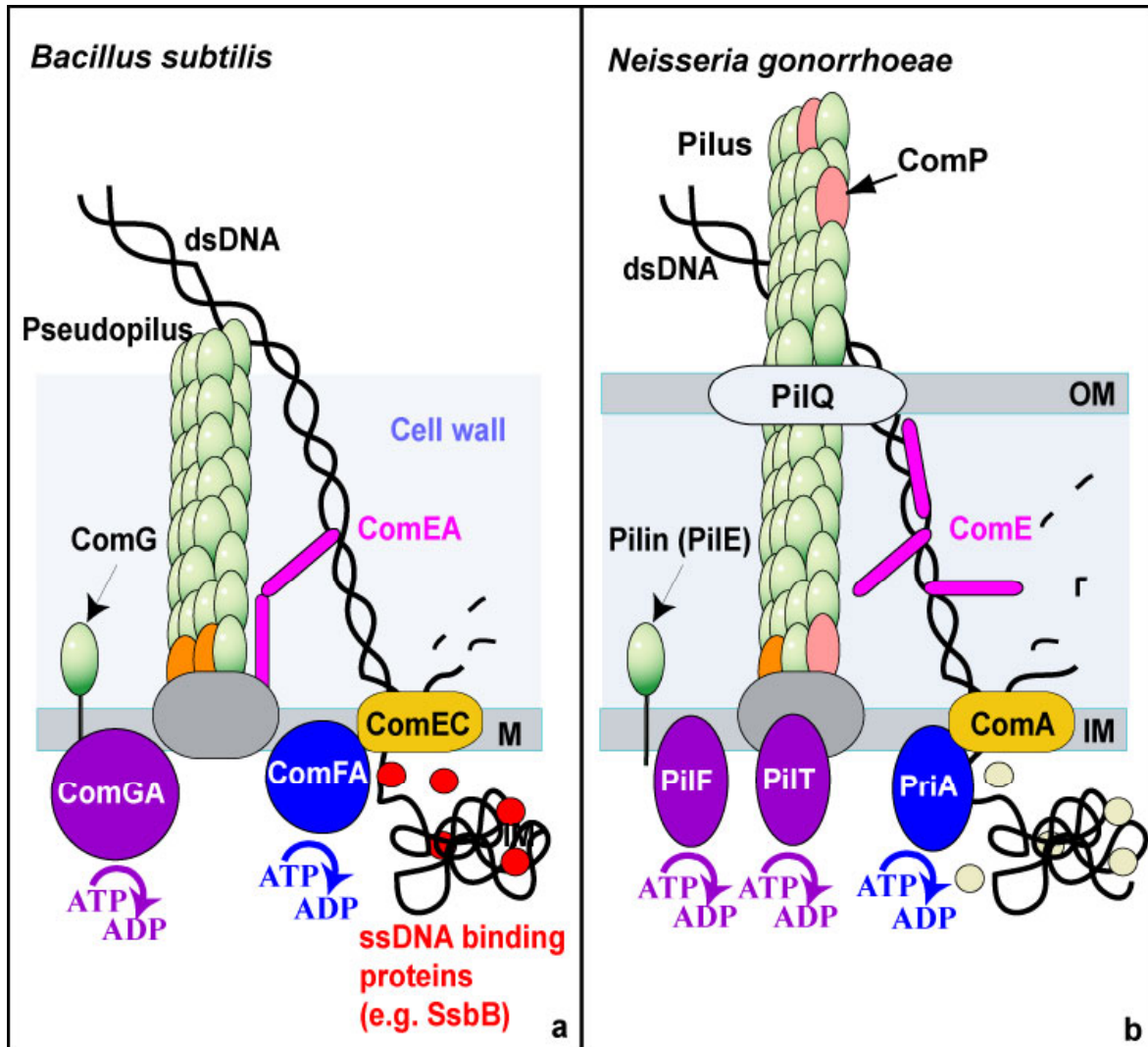
DNA import machine



adapted from Chen & Dubnau 2005

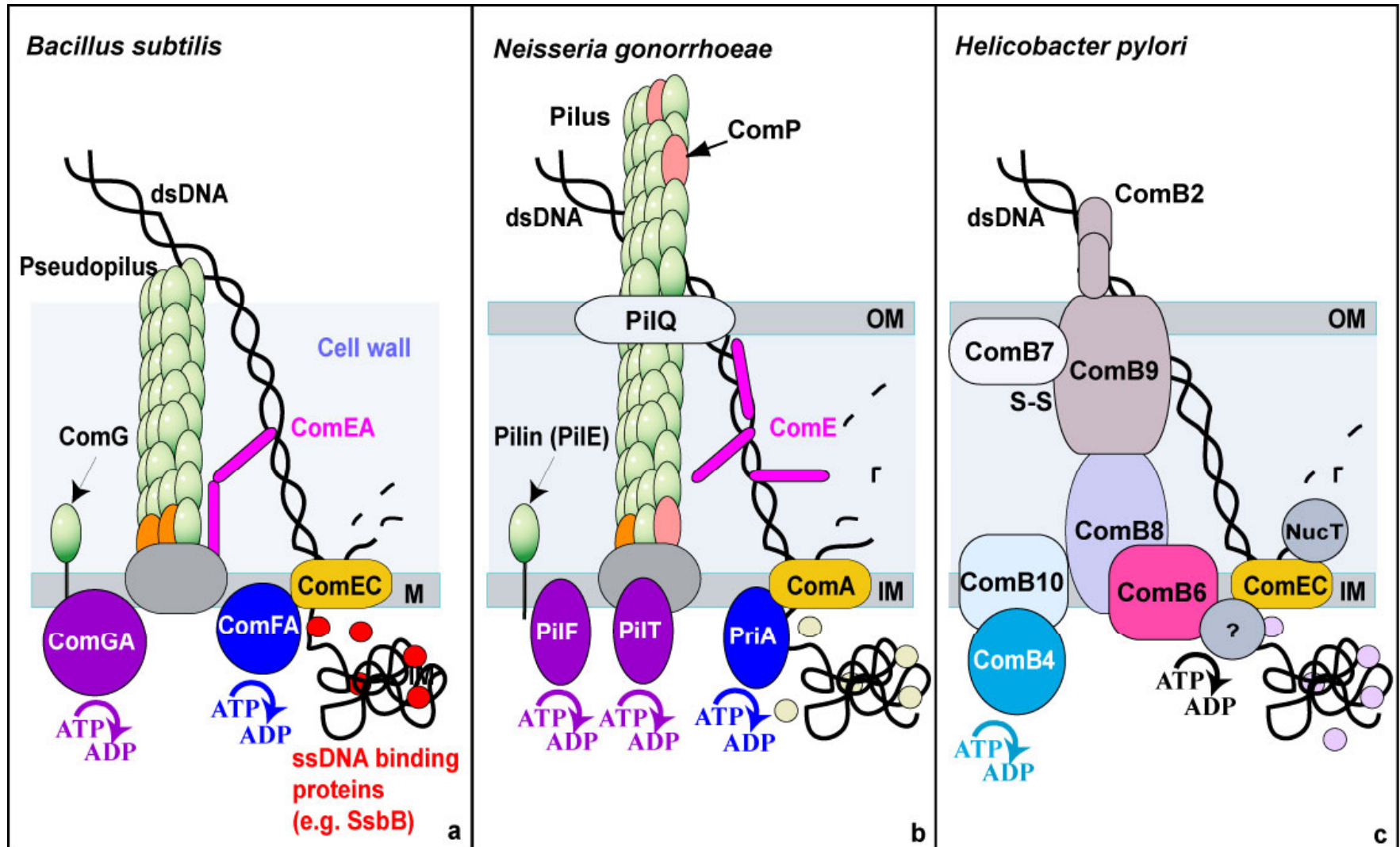
Different machines for DNA import

All naturally competent species use the T4 pilus for DNA import

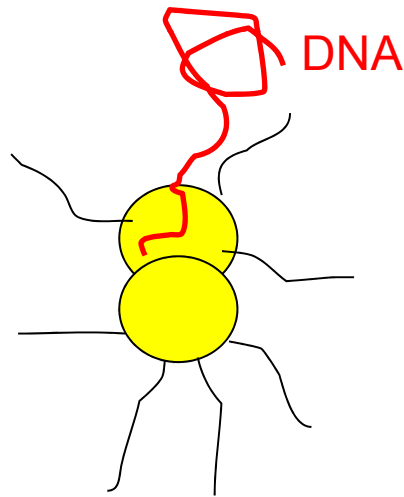


Different machines for DNA import

... with the exception of *Helicobacter pylori*



Molecular mechanism of the DNA import machine

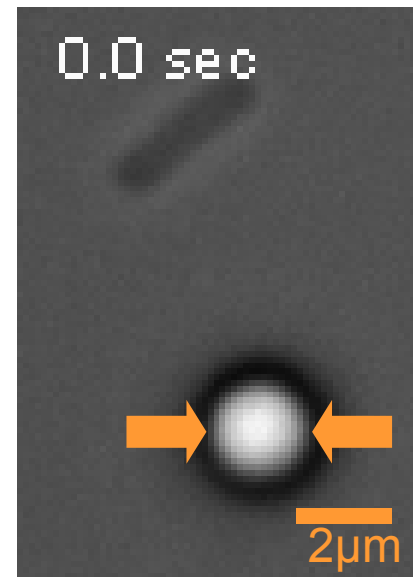
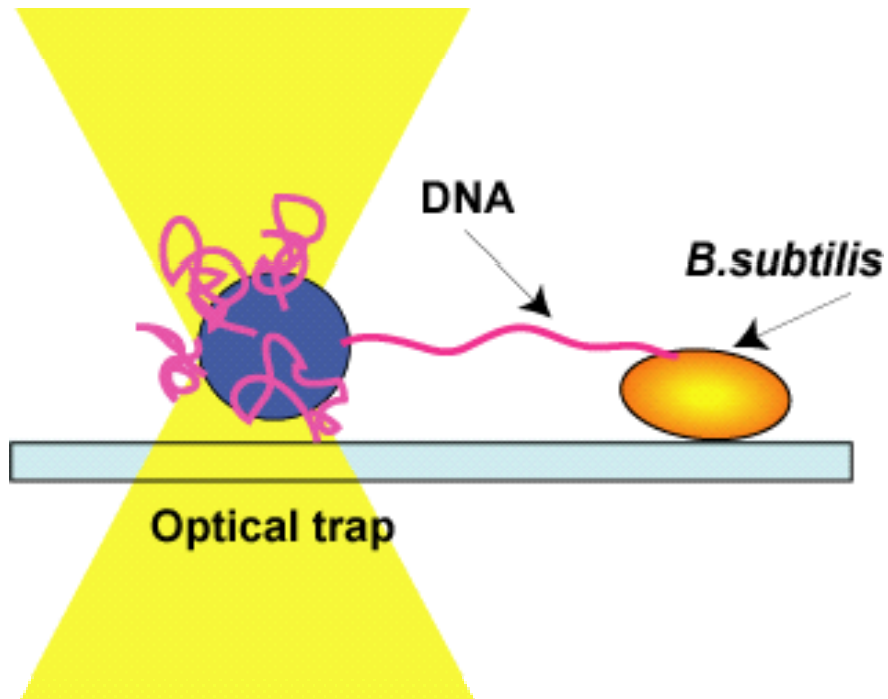


Kerstin Stingl, Stephanie Müller, Martin Clausen

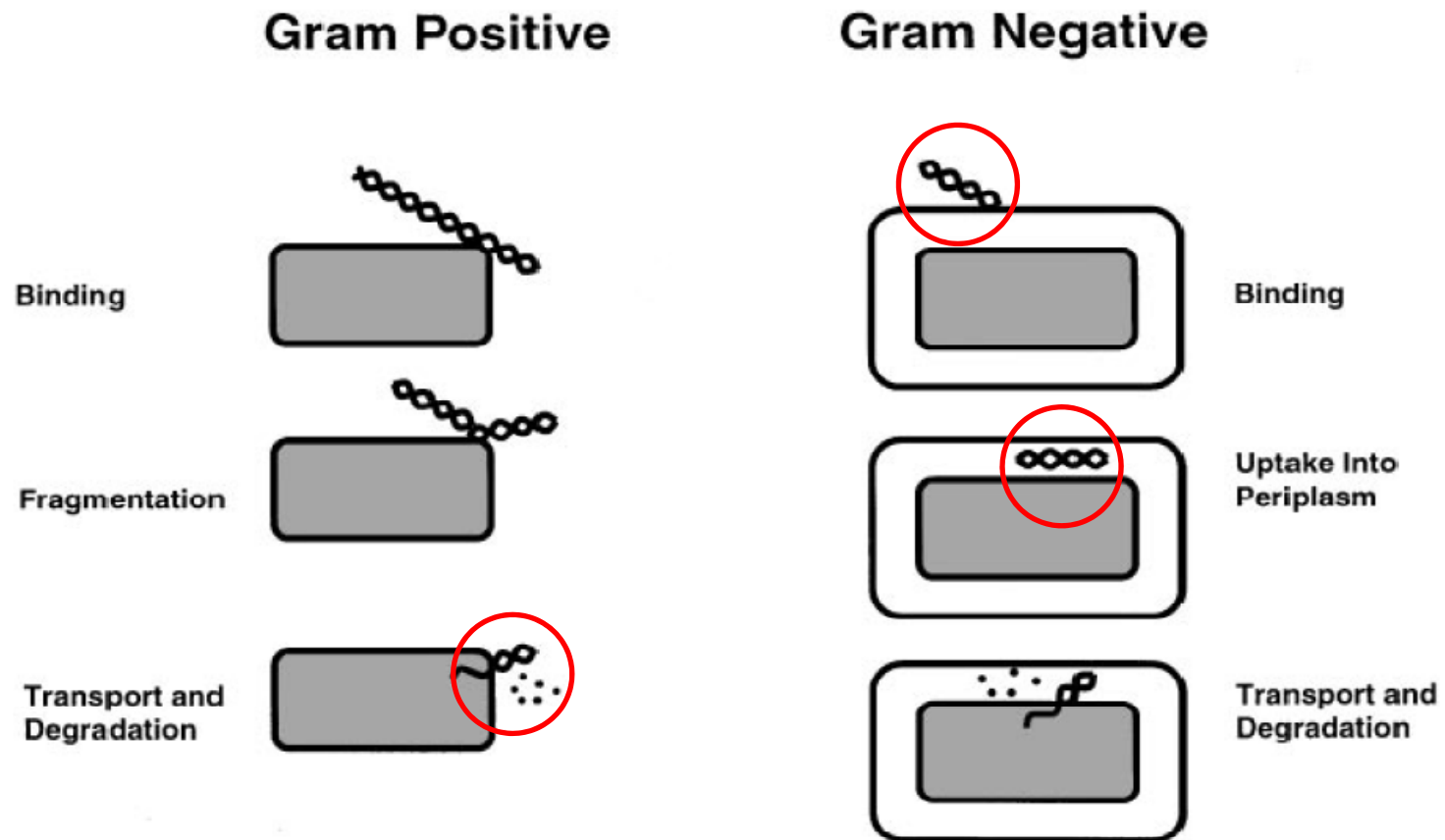
David Dubnau, PHRI Newark

Michael Koomey, Oslo University

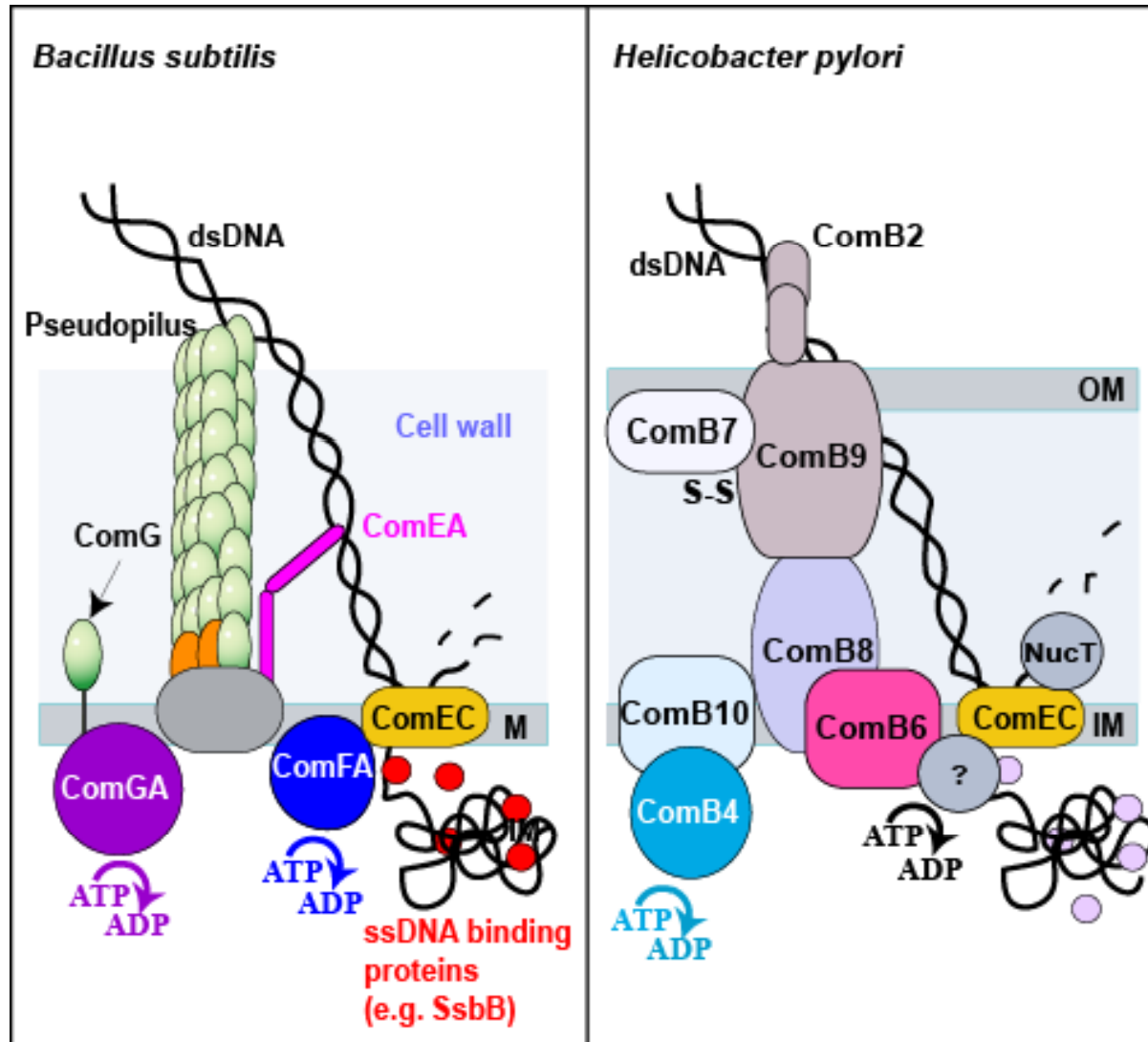
Import of individual DNA by *Bacillus subtilis*



DNA import through two membranes



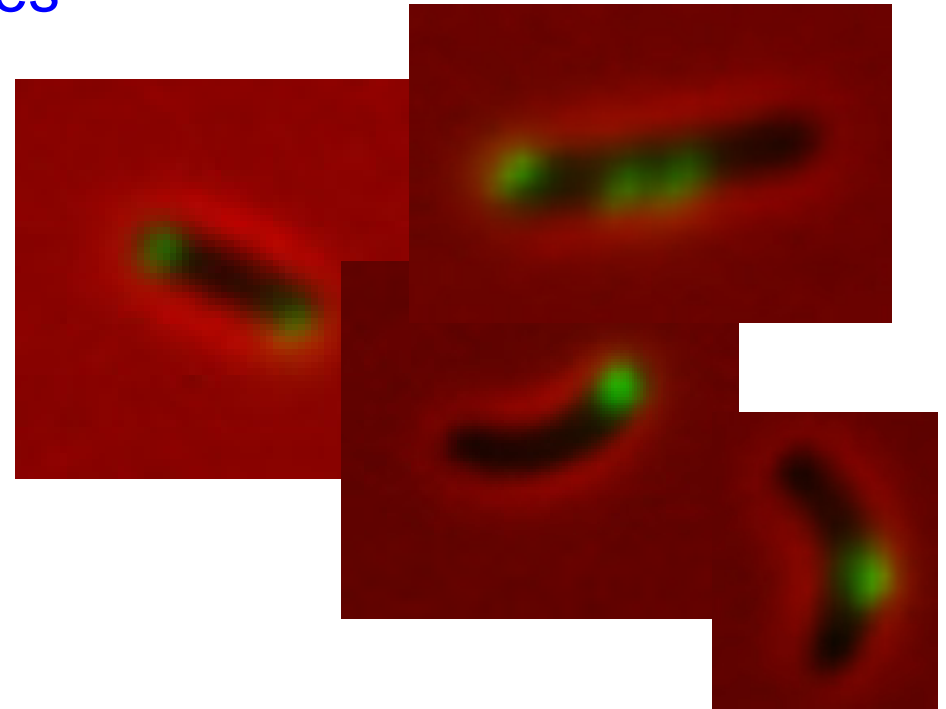
DNA import in *Helicobacter pylori*



DNA-uptake occurs from cell poles

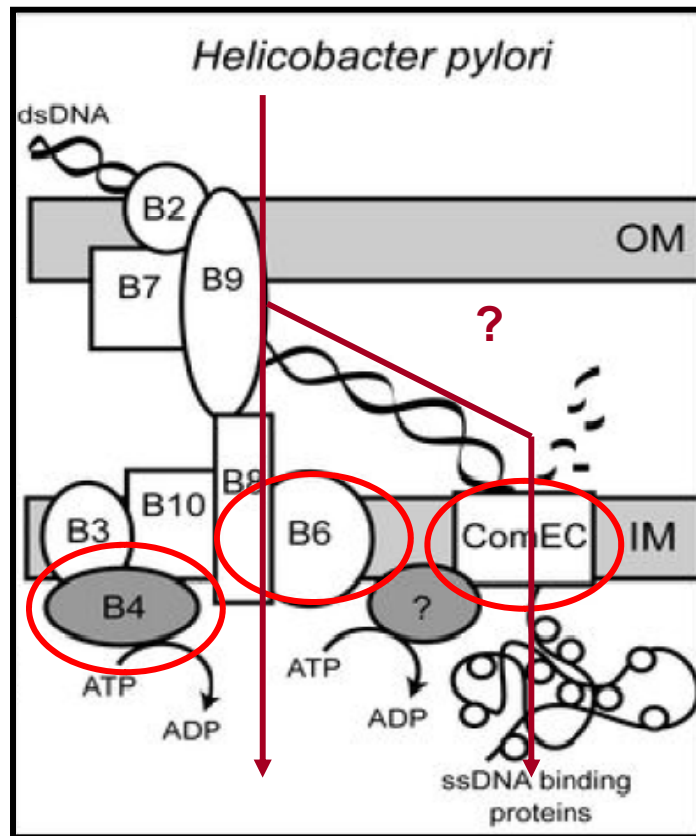
Cy3-DNA-foci in DNase treated

H. pylori:



- Polar localisation
- multiple foci possible
- parental strain: 74 ± 9 % of cells show foci
(one focus: 32 ± 3 %, two foci: 26 ± 4 %, three or more foci: 16 ± 6 %)
- no distinct competence phase (stationary phase: 58 ± 2 %)

DNA-uptake depends on ComB system

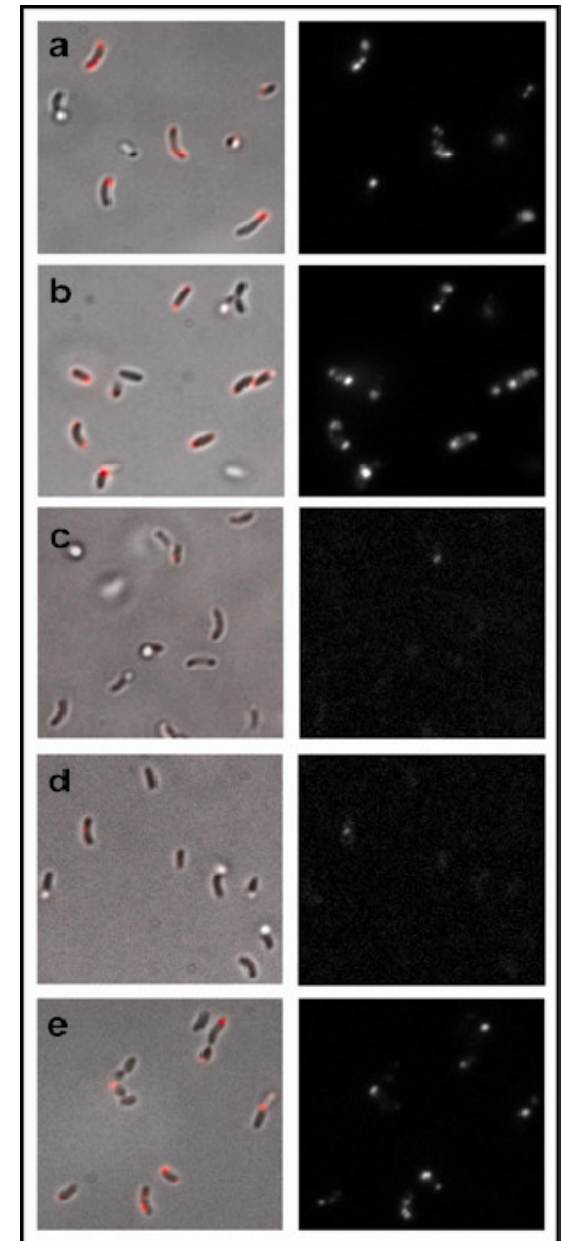


Parental strain

74% ± 9%

Parental strain
with 1kb λ -DNA

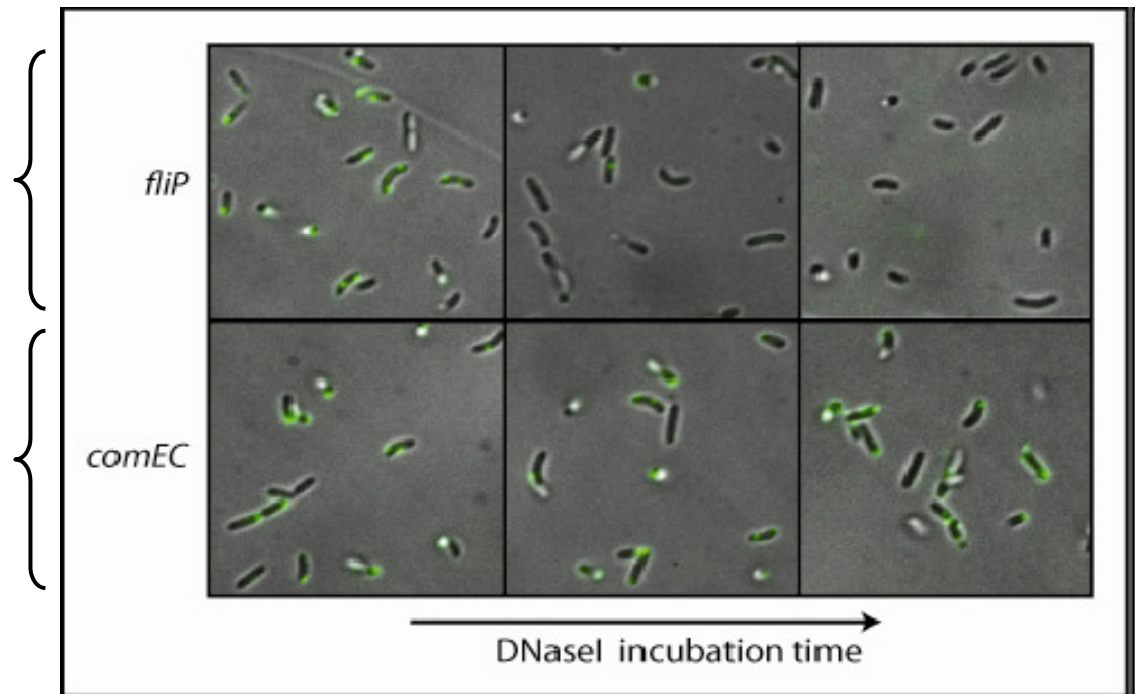
79% ± 7%



Temporal coupling of transport between outer and inner membrane

Foci disappear, but time until foci vanish varies (immediate or up to 60min)

Foci never disappeared (not even after 90min)

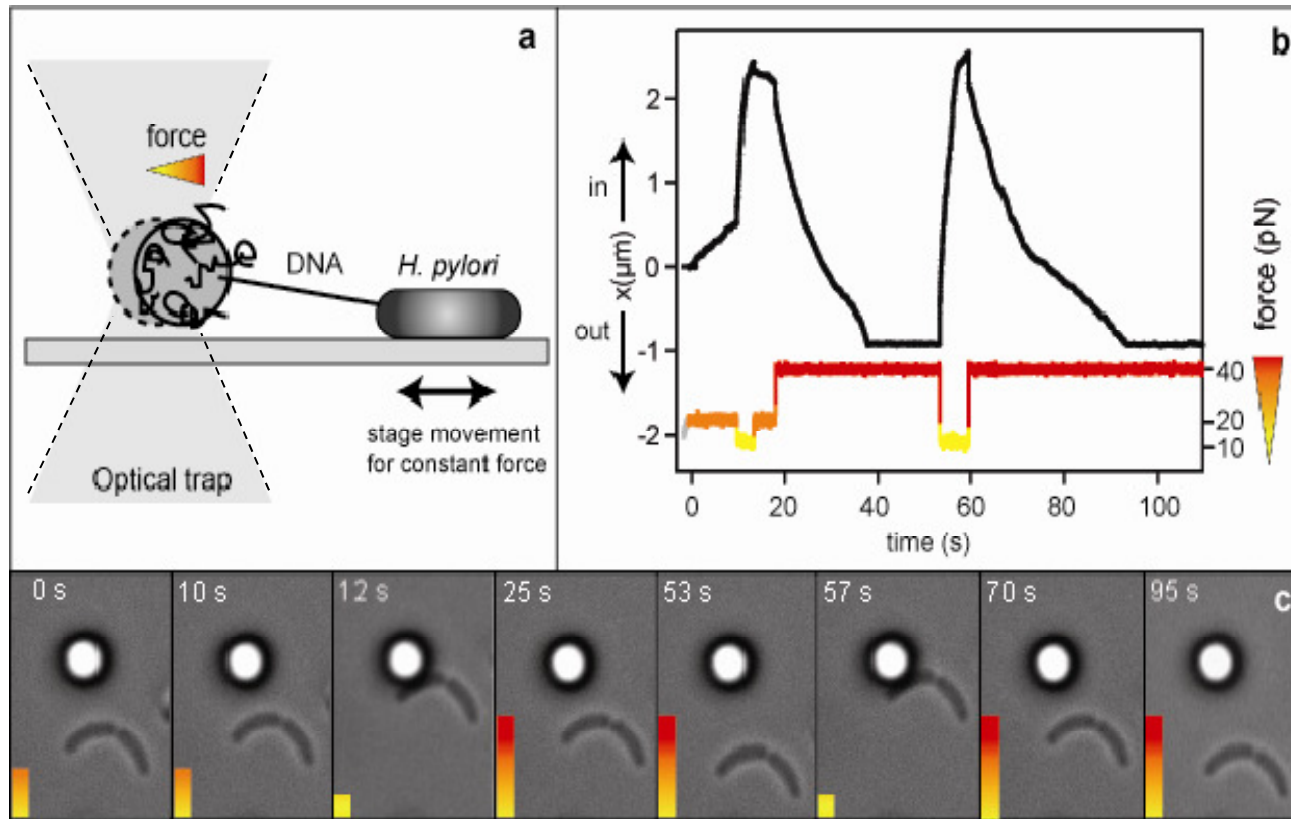


Green foci: imported fluorescently labeled DNA

=> DNA enters periplasm as dsDNA

=> inner and outer membrane transport are temporally uncoupled

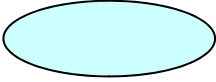
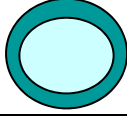
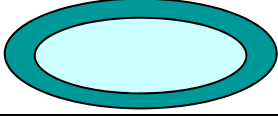
Kinetics and force generation during DNA-uptake in *Helicobacter pylori*



=> Import is reversible at 23 ± 3 pN

=> Average velocity @ 10pN: wt (*flIP*): $v = 1.26 \pm 0.46$ kbp/s

Conclusion

| | <i>B. subtilis</i>  | <i>N. gonorrhoeae</i> *  | <i>H. pylori</i>  |
|--------------------------|---|--|---|
| Velocity (10pN) | 80bps/sec | 80bps/sec | 1.2kbps/sec |
| Maximum force | 80pN | 40pN | 23pN |
| Reversibility | no | yes | yes |
| DNA specificity | high | | weak for om, strong for im |
| Polar foci | yes | no | yes |
| Regulation of competence | quorum sensing nutrient limitation | DNA uptake sequence | none known |

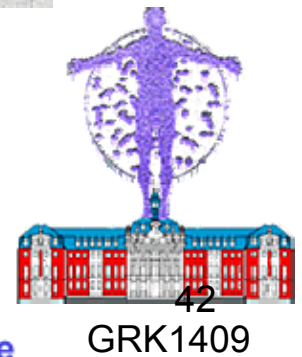
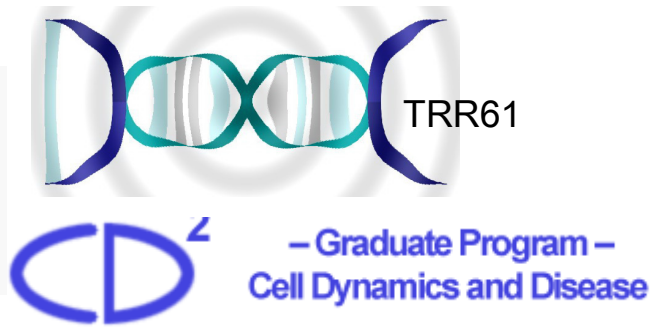
=> inner membrane transport conserved, outer membrane transport species-dependent

Maier, B., Chen, I, Dubnau, D., Sheetz, M. Nat.Struct.Mol.Biol., 2004

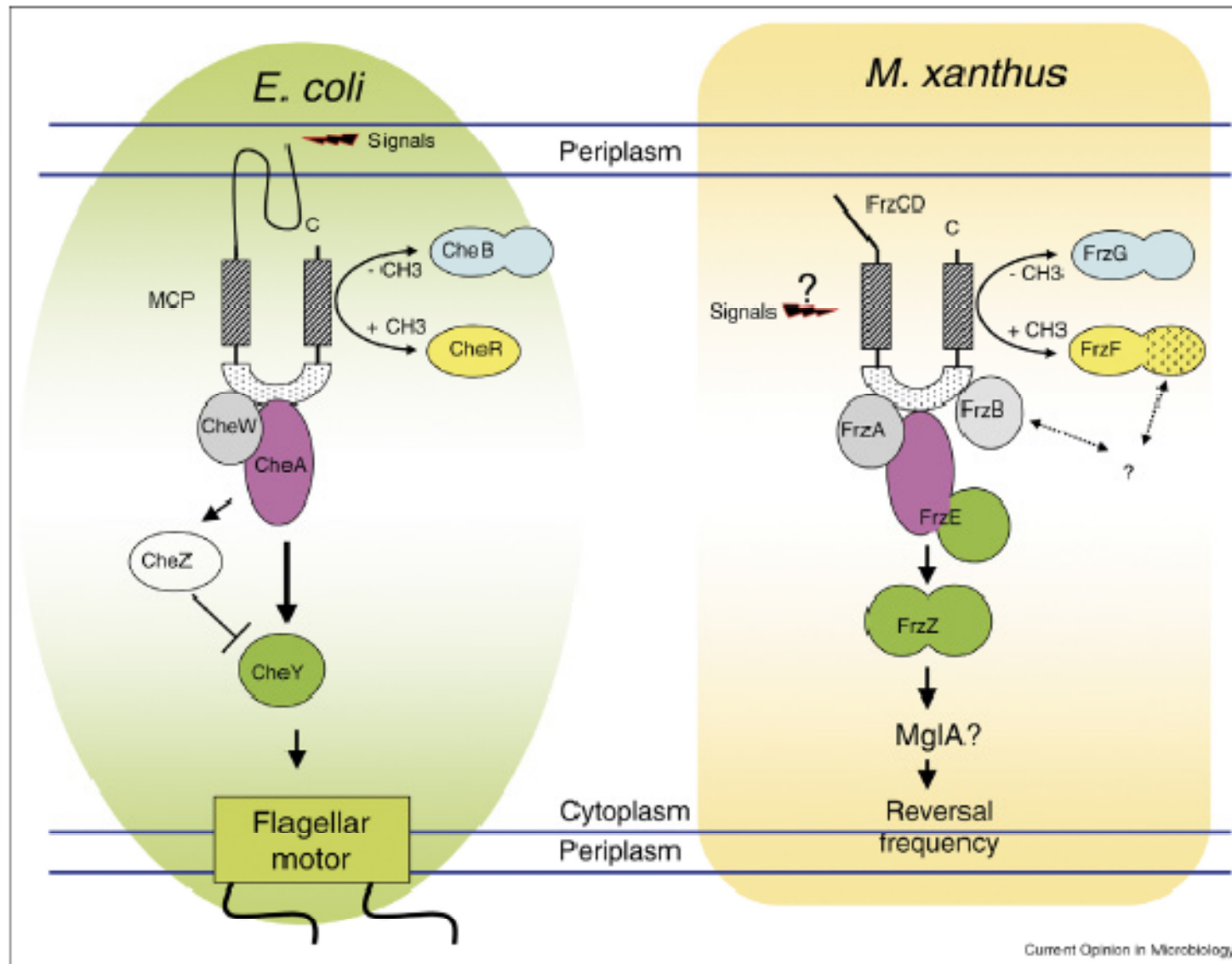
Hahn, Maier, Hajjema, Sheetz, Dubnau, Cell 2005

Stingl, K., Müller, S., Scheidgen-Kleyboldt, G., Clausen, M., Maier, B., PNAS 2010

Stingl, K., Clausen, M., Maier, B. unpublished



The Frz system



The Frizilator

