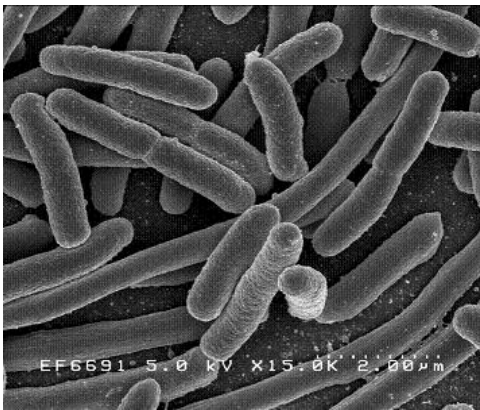


How does a bacterium achieve optimal flux partitioning?

Lei-Han Tang

Department of Physics, Hong Kong Baptist University

Fast growth = efficient usage of resources \oplus speed



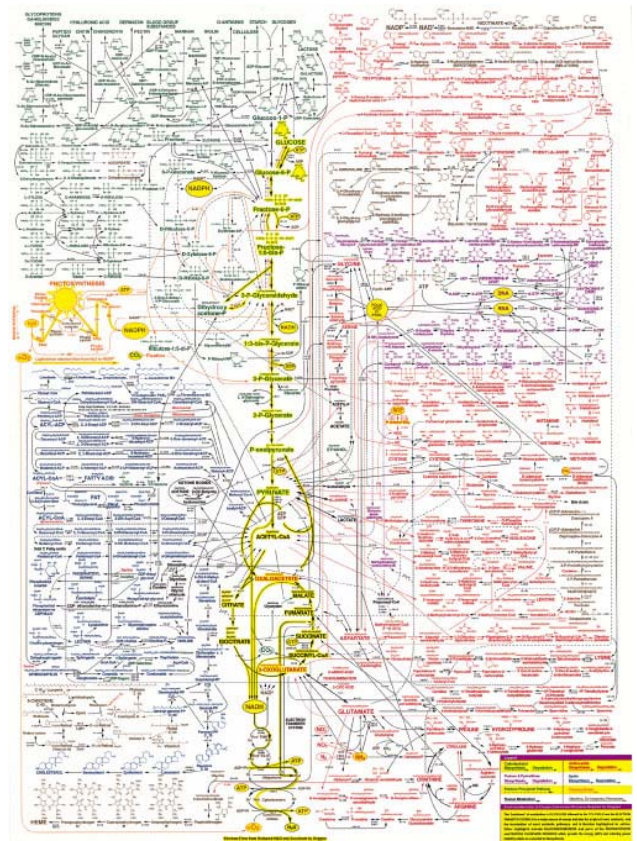
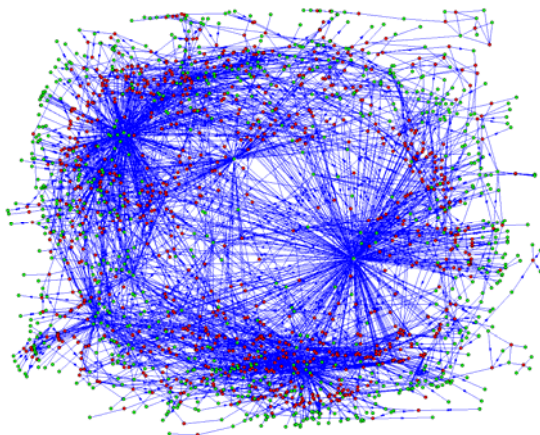
addressed by FBA

regulation at branch points of metabolic flow

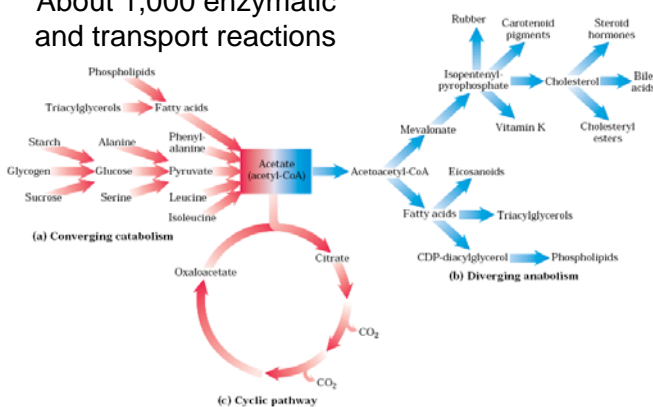
Further coarse-graining (examine global gene expression) to identify nonessential activities and bottlenecks

Makes a run when condition is good
 \Rightarrow Divides in approximately 30 min.

What's underlying cell growth?



About 1,000 enzymatic and transport reactions



The Grand Synthesis

Genomic data



4,200 genes

Gene expression

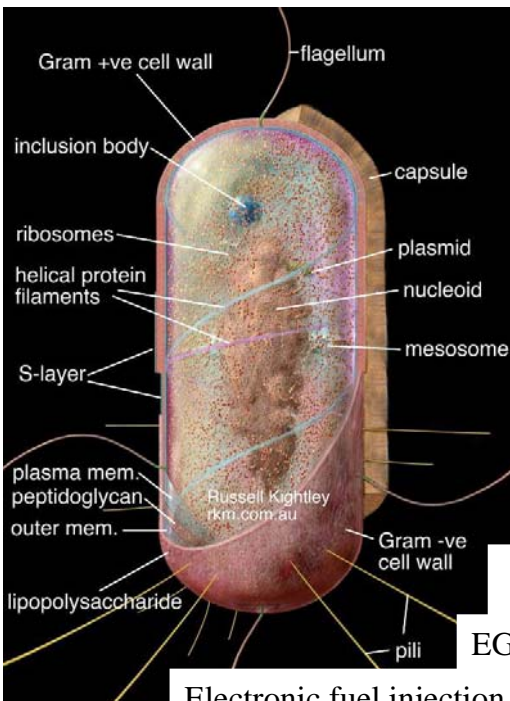


Over 270 transcription factors

Compound concentrations



1,300 interactions by metabolites



John Doyle:

Organized complexity



Electronic fuel injection

Electronic ignition

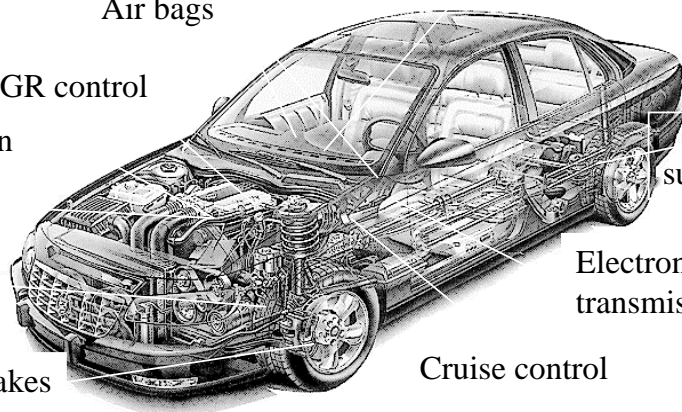
Electric power steering (PAS)

Anti-lock brakes

Air bags

EGR control

Temperature control



Active suspension

Electronic transmission

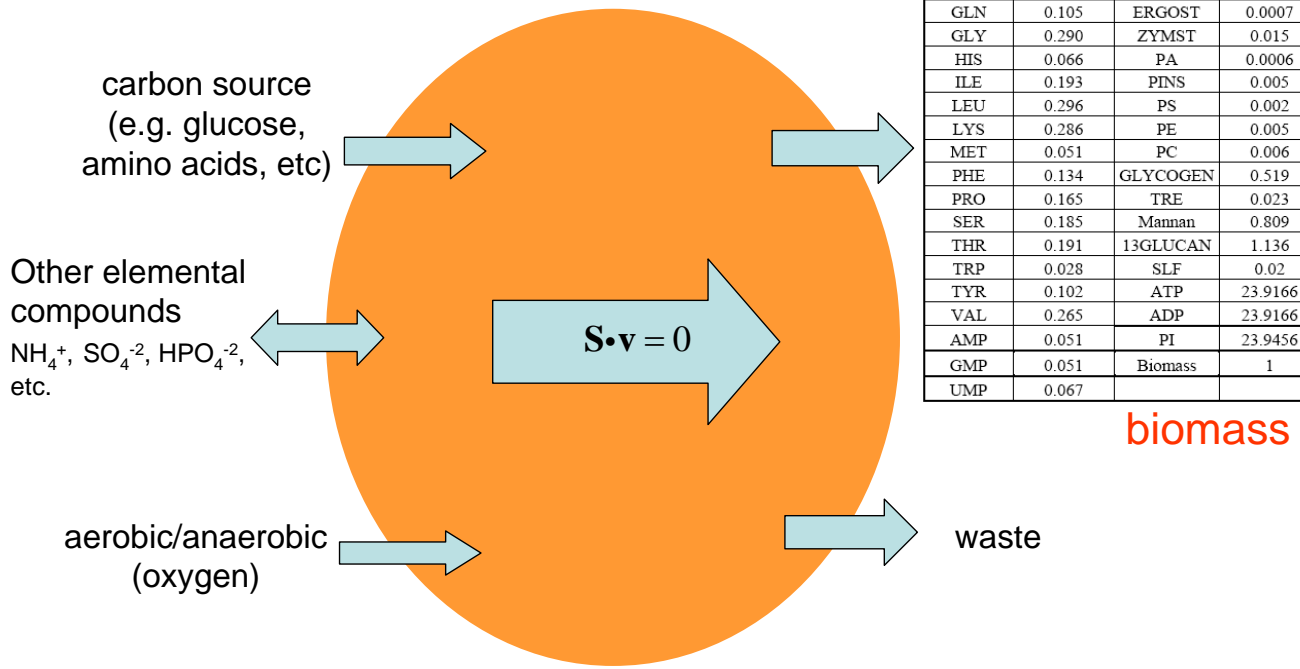
Cruise control

Optimal performance: Flux Balance Analysis (FBA)

N. D. Price, J. L. Reed, B. O. Palsson, *Nature Reviews Microbiology* 2, 886-897 (2004)

Table 9. Cellular components of *S. cerevisiae*

ALA	0.459	CMP	0.05
ARG	0.161	dAMP	0.0036
ASN	0.102	dCMP	0.0024
ASP	0.297	dGMP	0.0024
CYS	0.007	DTMP	0.0036
GLU	0.302	TAGLY	0.007
GLN	0.105	ERGOST	0.0007
GLY	0.290	ZYMST	0.015
HIS	0.066	PA	0.0006
ILE	0.193	PINS	0.005
LEU	0.296	PS	0.002
LYS	0.286	PE	0.005
MET	0.051	PC	0.006
PHE	0.134	GLYCOGEN	0.519
PRO	0.165	TRE	0.023
SER	0.185	Mannan	0.809
THR	0.191	13GLUCAN	1.136
TRP	0.028	SLF	0.02
TYR	0.102	ATP	23.9166
VAL	0.265	ADP	23.9166
AMP	0.051	PI	23.9456
GMP	0.051	Biomass	1
UMP	0.067		



Use linear programming to determine flux pattern for optimal biomass production

⇒ Upper limit on growth speed

All is good, except **cell** does not know linear programming!

And at times, it does not follow the optimal growth predicted by the FBA

What is the basic scheme of metabolic regulation?

- Identify decision points to reach optimal flux pattern

- Regulatory strategies (3-levels)
 - Transcriptional regulation to eliminate futile cycles (as well as setting the enzyme copy number to a suitable level), minutes
 - Small molecule regulation to achieve fast equilibrium, ms to second.
 - Homeostasis of global commodities

- Incorporate proteomic, metabolomic and transcriptomic data to iteratively improve the model parameters

Network simplification to highlight carbon flow

An automated program to reveal carbon flow network

Free-standing

compound	frequency	compound	frequency	formula
h [c]	497	co2 [c]	45	CO2
h2o [c]	293	pyr [c]	16	C3H3O3
pi [c]	149	ac [c]	8	C2H3O2
ppi [c]	75	for [c]	8	CHO2
nh4 [c]	38	succ [c]	7	C4H4O4
o2 [c]	16	fum [c]	3	C4H2O4
		hco3 [c]	2	CHO3

Adenosine deaminase: ~~adn~~ + ~~h~~ + ~~h2o~~ --> ~~ins~~ + ~~nh4~~

Hexokinase: ~~atp~~ + glc-D --> ~~adp~~ + g6p + ~~h~~

Carriers

Cmpd1	Cmpd2	cargo	frequency	Cmpd1	Cmpd2	cargo	frequency
nadh [c]	nad [c]	H	71	glu-L [c]	glu-L [c]	H2NO	13
nadph [c]	nadp [c]	H	49	asn-L [c]	asp-L [c]	H2NO	3
q8h2 [c]	q8 [c]	H2	17	glu-L [c]	akg [c]	H4NO	20
mqn8 [c]	mql8 [c]	H2	16	atp [c]	adp [c]	O3P	136
2dmmql8 [c]	2dmmq8 [c]	H2	10	atp [c]	amp [c]	O6P2	25
trdrd [c]	trdox [c]	H2	10	pep [c]	pyr [c]	HO3P	16
fadh2 [c]	fad [c]	H2	8	gtp [c]	gdp [c]	O3P	6
fum [c]	succ [c]	H2	5	adp [c]	amp [c]	O3P	3

Currency compounds

Nitrogenous: NH4, NO, NO2, NO3, etc.

Phosphates: PO4, diphosphate, etc.

Sulfate/sulfite: SO3, SO4, etc.

Metal ions: Fe, Na, K, etc.

Water, hydrogen, oxygen

Carriers

ATP/ADP/AMP, GTP/GDP,CTP/CMP

nad/nadh, nadp/nadph, fad/fadh

q8/q8h2, mql8/mqn8, 2dmmq8/2dmmql8

akg/glu/gln

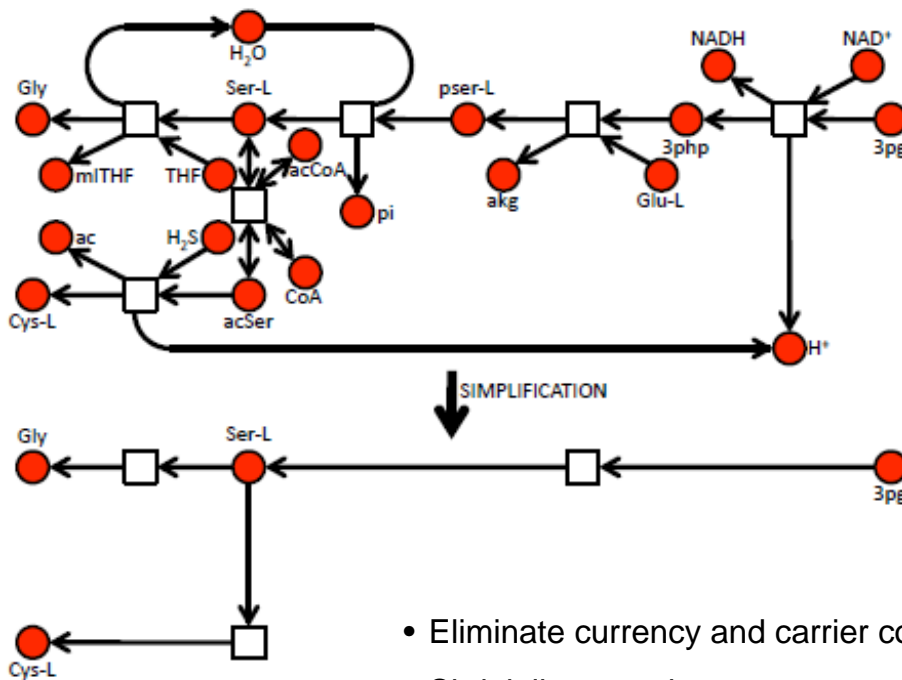
acCoA/CoA, sucCoA/CoA, pep/pyr

Coenzymes/cofactors

ACP, THF, udcpp, etc.

High degree nodes are currency and carrier compounds!

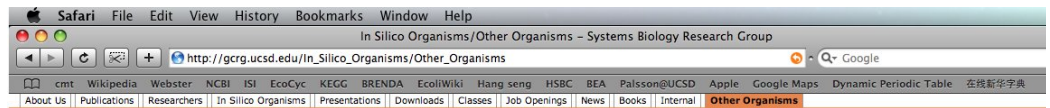
An example of the network simplification scheme



- Eliminate currency and carrier compounds
- Shrink linear pathways

in silico organisms

constructed by Palsson's group at UCSD



Supplementary Table 1 Available predictive genome-scale metabolic network reconstructions

Adam M. Feist, Markus J. Herrgard, Ines Thiele, Jennie L. Reed, Bernhard Ø. Palsson "Reconstruction of Biochemical Networks in Microbial Organisms", Nat. Rev. Micro 2009 Feb;7(2):129-43.

Supplementary Table 2 - Common issues encountered during metabolic network reconstruction

This list includes genome-scale metabolic network reconstructions that have been converted into predictive genome-scale models and whose predictive power has been validated against experimental data.

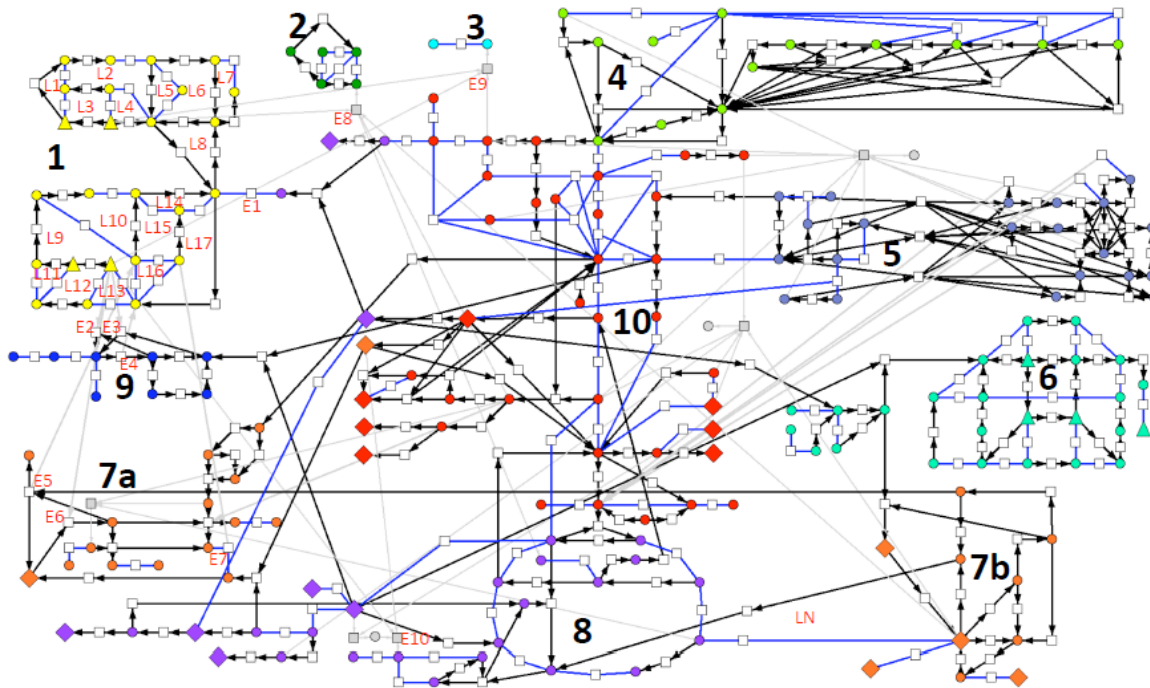
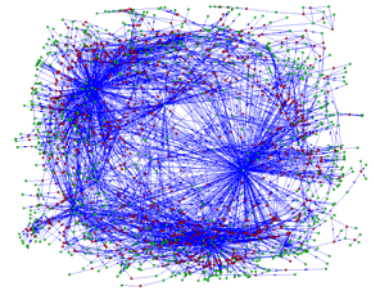
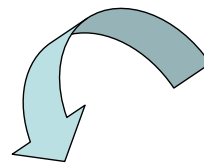
BACTERIA ARCHAEA EUKARYOTES

Organism	Strain	Genes	Status	Version	Genes	Metabolites	Reactions	Compartments	Reference	PMID	Download	Web site
BACTERIA												
<i>Acinetobacter baylyi</i>	ADP1	3,287	F	iAbaylyiV4	774	701	875	3 (c,e,p)	• Durot et al.	18840283	• xls	• Genoscor
<i>Bacillus subtilis</i>		4,225	F	model_v3	844	988	1020	2 (c,e)	• Oh et al.	17573341	• xls	• JBC
<i>Clostridium acetobutylicum</i>	ATCC 824	3,848	F		474	422	552	2 (c,e)	• Senger and Papoutsakis	18767192		
<i>Clostridium acetobutylicum</i>	ATCC 824	3,848	F		432	479	502	2 (c,e)	• Lee et al.	18758767		
<i>Corynebacterium glutamicum</i>	ATCC 1303	3,002	F			411	446	2 (c,e)	• Kjeldsen and Nielsen	18985611	• xls	• B&B
<i>Escherichia coli</i>	K12 MG1655	4,405	F	IJE660	660	438	627	2 (c,e)	• Edwards et al.	10805808		
<i>Escherichia coli</i>	K12 MG1655	4,405	F	IJR904	904	625	931	2 (c,e)	• Reed et al.	12952533	• SBML	• UCSD
<i>Escherichia coli</i>	K12 MG1655	4,405	F	IAF1260	1260	1039	2077	3 (c,e,p)	• Feist et al.	17593909	• SBML	• UCSD
<i>Geobacter sulfurreducens</i>		3,530	F		588	541	523	2 (c,e)	• Mahadevan et al.	16461711	• xls	
<i>Haemophilus influenzae</i>	Rd	1,775	F	IJE296	296	343	488	2 (c,e)	• Edwards et al.	10364169		
<i>Haemophilus influenzae</i>	Rd	1,775	F	ICS400	400	451	461	2 (c,e)	• Schilling et al.	10716908	• pdf	
<i>Helicobacter pylori</i>	26695	1,632	F	IIT341	341	485	476	2 (c,e)	• Thiele et al.	16077130	• SBML	• UCSD
<i>Helicobacter pylori</i>	26695	1,632	F	ICS291	291	340	388	2 (c,e)	• Schilling et al.	12142428	• xls	• UCSD
<i>Lactococcus lactis</i>	ssp. lactis IL1403	2,310	F		358	422	621	2 (c,e)	• Oliveira et al.	15982422	• xls	
<i>Lactobacillus plantarum</i>	WCFS1	3,009	F		721	531	643	2 (c,e)	• Teusink et al.	17062565	• xls	
<i>Mannheimia</i>									• Kim et al.			

- Collection of organism-specific reactions leading to biomass production
- Linear programming to find the optimal state
- Allow for simulation of different growth conditions (nutrient uptake, O₂ availability, etc.)
- **Outcome:** growth rate and **flux pattern**

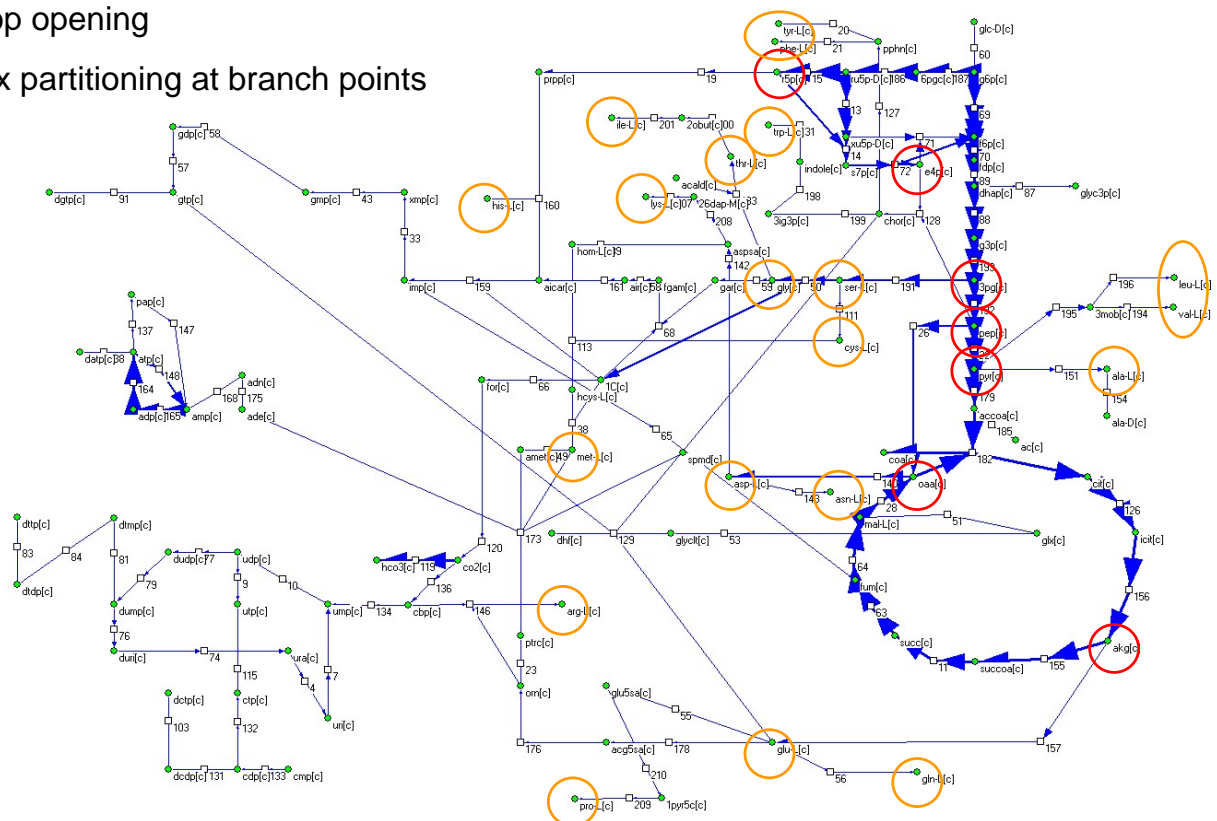
Simplified network based on iJR904

- tree like
- community structure
- loops



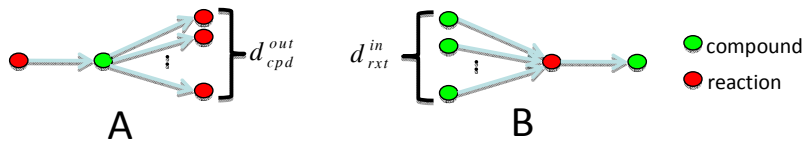
Optimal flow identified by FBA

- Loop opening
- Flux partitioning at branch points



DEGREES OF FREEDOM

Due to flux balance, the actual degree of freedom (312, including 124 transport reactions in iJR904) is much less than the number of reaction flux variables (1075).



Rules to count DoF on a network of compounds and irreversible reactions*:

1. For compounds (figure A):

$$DoF_{cpd} = \sum_{cpd} (d_{cpd}^{out} - 1)$$

Branching increases degrees of freedom

2. For reactions (figure B):

$$DoF_{rxn} = -\sum_{rxn} (d_{rxn}^{in} - 1)$$

Synthesis decreases degrees of freedom

3. Thus

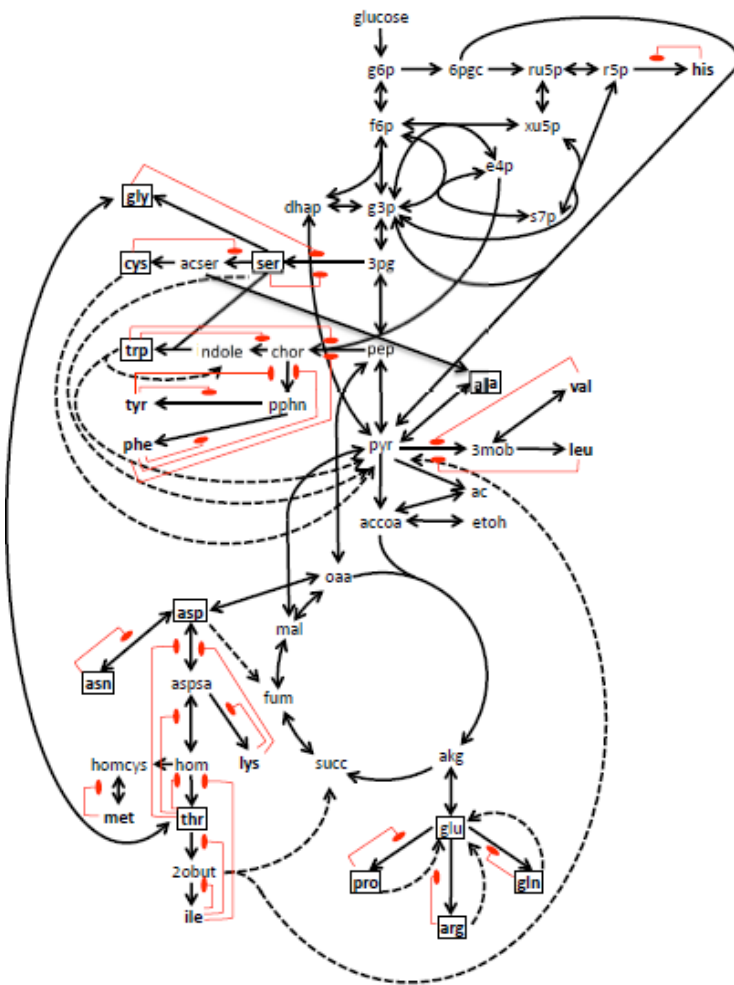
$$DoF_{total} = DoF_{cpd} + DoF_{rxn}$$

How does E. coli do it?

Amino acid biosynthesis subnetwork

Carbon flow network for amino acid metabolism

Amino acids can be both synthesized from sugar or used as carbon source



transcriptional regulation to eliminate futile cycles in carbon flow

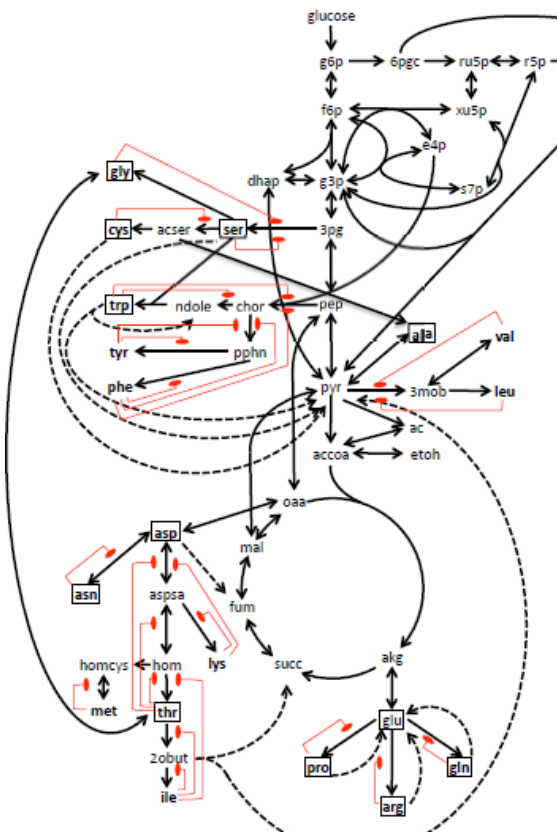
Allen TE, et al. (2003) Genome-scale analysis of the uses of the Escherichia coli genome: model-driven analysis of heterogeneous data sets. J Bacteriol 185(21):6392-6399

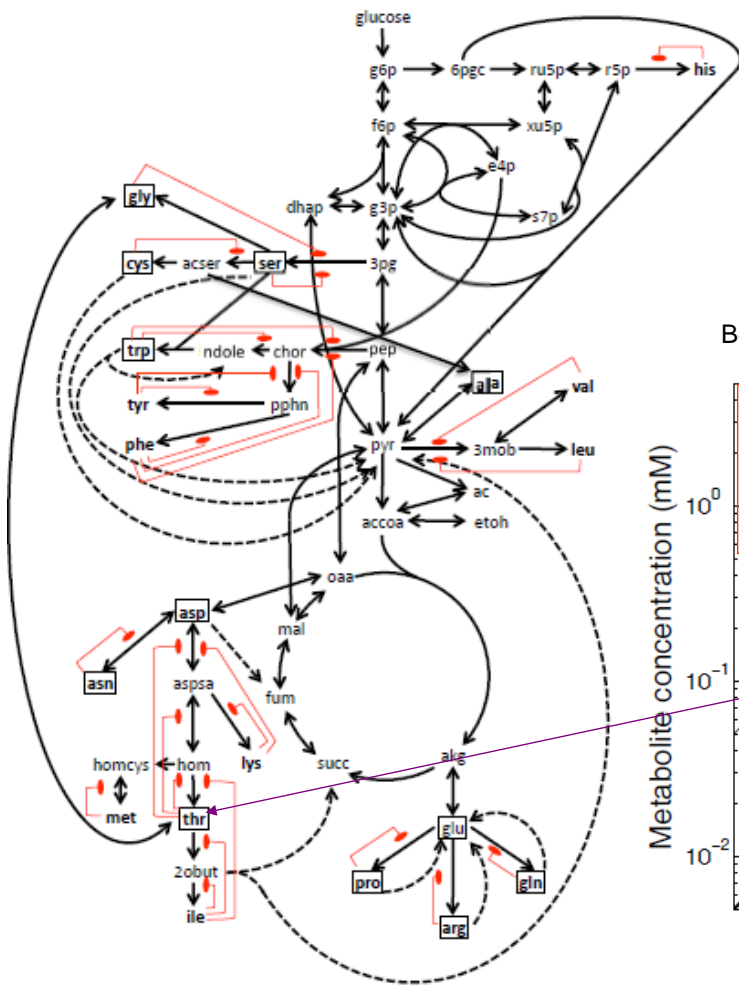
$$\text{Fold} = \log_2(\text{LB}/\text{glucose})$$

Table 4. The TUs and genes for the loop backward pathways of amino acids.

AA	TU	Gene	Reaction	Fold
Trp	tnaCAB	tnaB	$h[e] + trp[e] \rightleftharpoons h + trp$	2.230
		tnaA	$h_2o + trp \rightleftharpoons indole + nh_4 + pyr$	6.296
Ser	sdaCB	sdaC	$ser[e] + h[e] \rightarrow ser + h$	2.614
		sdaB	$ser \rightarrow nh_4 + pyr$	1.014
Asp	aspA-dcuA	dcuA	$asp[e] + succ \rightarrow asp + succ[e]$	-0.281
		aspA	$asp \rightarrow fum + nh_4$	2.505
Gln	yneHG	yneH	$gln + h_2o \rightarrow glu + nh_4$	2.343
Pro	putA*	putA	$fad + pro \rightarrow 1pyr5c + fadh_2 + h$	3.448
			$1pyr5c + (2) h_2o + nad \rightarrow glu + h + nadh$	
Cys	malXY	malY	$cys + h_2o \rightarrow h_2s + pyr + nh_4$	0.815
		metC		-0.513
Arg	astCADBE	astA	$arg + succoa \rightarrow coa + h + sucarg$	-0.368
		astB	$(2) h + (2) h_2o + sucarg \rightarrow co_2 + (2) nh_4 + sucorn$	-0.524
		astC	$akg + sucorn \rightarrow glu + sucgsa$	-0.332
		astD	$h_2o + nad + sucgsa \rightarrow (2) h + nadh + sucglu$	-0.442
		astE	$h_2o + sucglu \rightarrow glu + succ$	0.829
Thr	tdcABCDEF	tdcC	$thr[e] + h[e] \rightarrow thr + h$	-0.029
		tdcB	$thr \rightarrow 2obut + nh_4$	0.222
		tdcE	$2obut + coa \rightarrow for + ppcoa$	-0.795
		prpBCDE	$h_2o + oaa + ppcoa \rightarrow 2mcit + coa + h$	-0.225
	prpBCDE	prpC	$2mcit \rightarrow 2mcacn + h_2o$	-0.121
		prpB	$micit \rightleftharpoons pyr + succ$	-0.678
		acnB	$2mcacn + h_2o \rightarrow micit$	-0.620

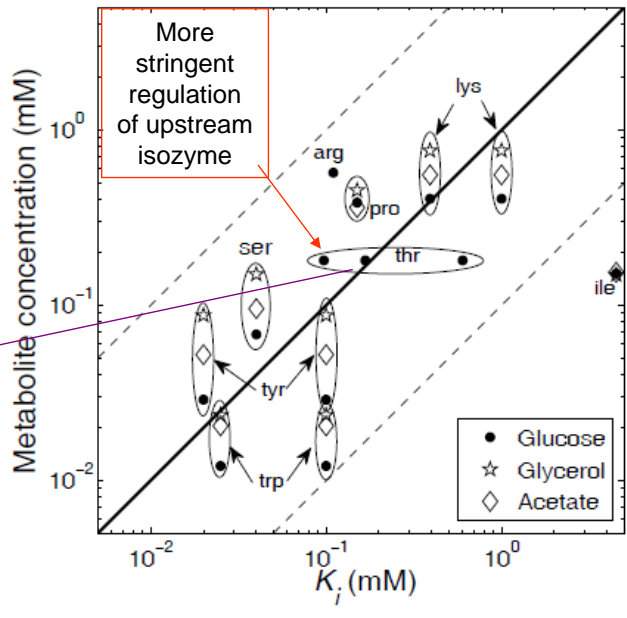
* There is no data of gene putA in measurement of LB/glucose minimal media comparison. We compare change between L-proline minimal media and glucose minimal media using data from same literature. The column Fold is ln-2 fold change of gene expression between growth in LB/L-proline minimal media and growth in glucose minimal media [40]. Significant changes of gene expression are highlighted in bold.





End-product inhibition
via allosteric regulation
ensures appropriate flux
ratio at branching points of
the carbon flow

Bennett et al. (2009) Nat. Chem. Biol. 5, 593-9



Regulation of global commodities

Table 3. Evidences for homeostasis of currencies and carriers.

Compounds	Category	References
NAD ⁺ /NADH, NADP ⁺ /NADPH	redox	[20–22]
SO ₃ ²⁻ , SO ₄ ²⁻ , HS ⁻ , sulfonate	sulfur	[23–25]
NH ₄ ⁺ , glutamine, glutamate	nitrogen	[26, 27]
Fe ²⁺ , Fe ³⁺	ions	[28–32]
Pyruvate	small carbon unit	[33, 34]
ATP/ADP/AMP	energy	[35–37]
PO ₄ ³⁻ , pyrophosphate, metaphosphate	phosphorus	[38]

➤ Regulatory strategies (3-levels)

- Transcriptional regulation to eliminate futile cycles (as well as setting the enzyme copy number to a suitable level), minutes

excess enzymes are kept within 10% of respective population

- Small molecule regulation to achieve fast equilibrium, ms to second.

metabolite concentration measurements show that they are used not only during transition periods to a new mode of growth, but even in steady-state.

- Homeostasis of global commodities

growth rate dependent?

Beyond FBA: **growth physiology**

Is **30 minutes** the physical limit for doubling of any cell?

Where does the bottleneck lie?

Why **doesn't** the bug use multiple carbon source to speed up growth?

What determines the **order of preference** for the sugars?

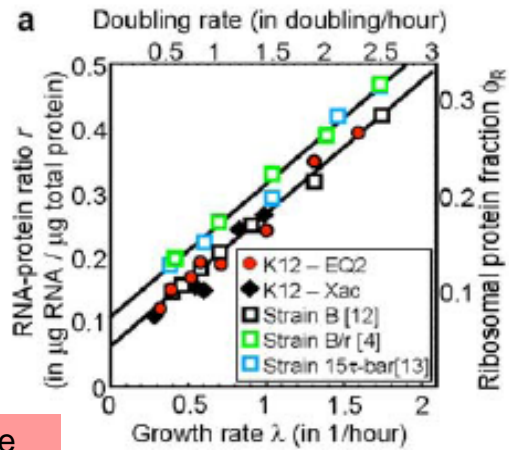
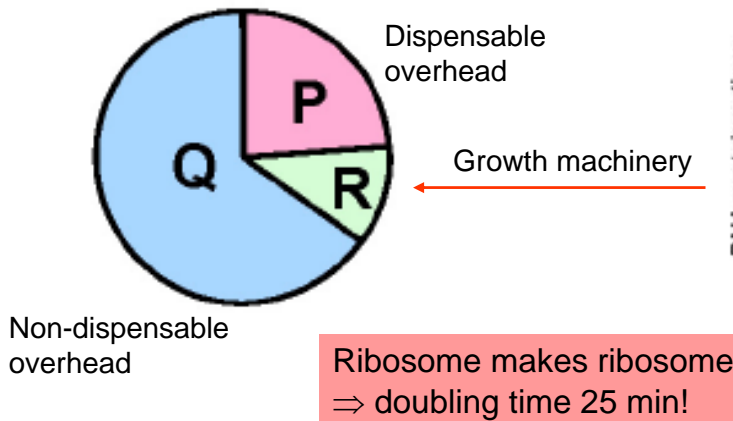
How should one attempt to answer such questions?

Interdependence of cell growth and gene expression: origins and consequences

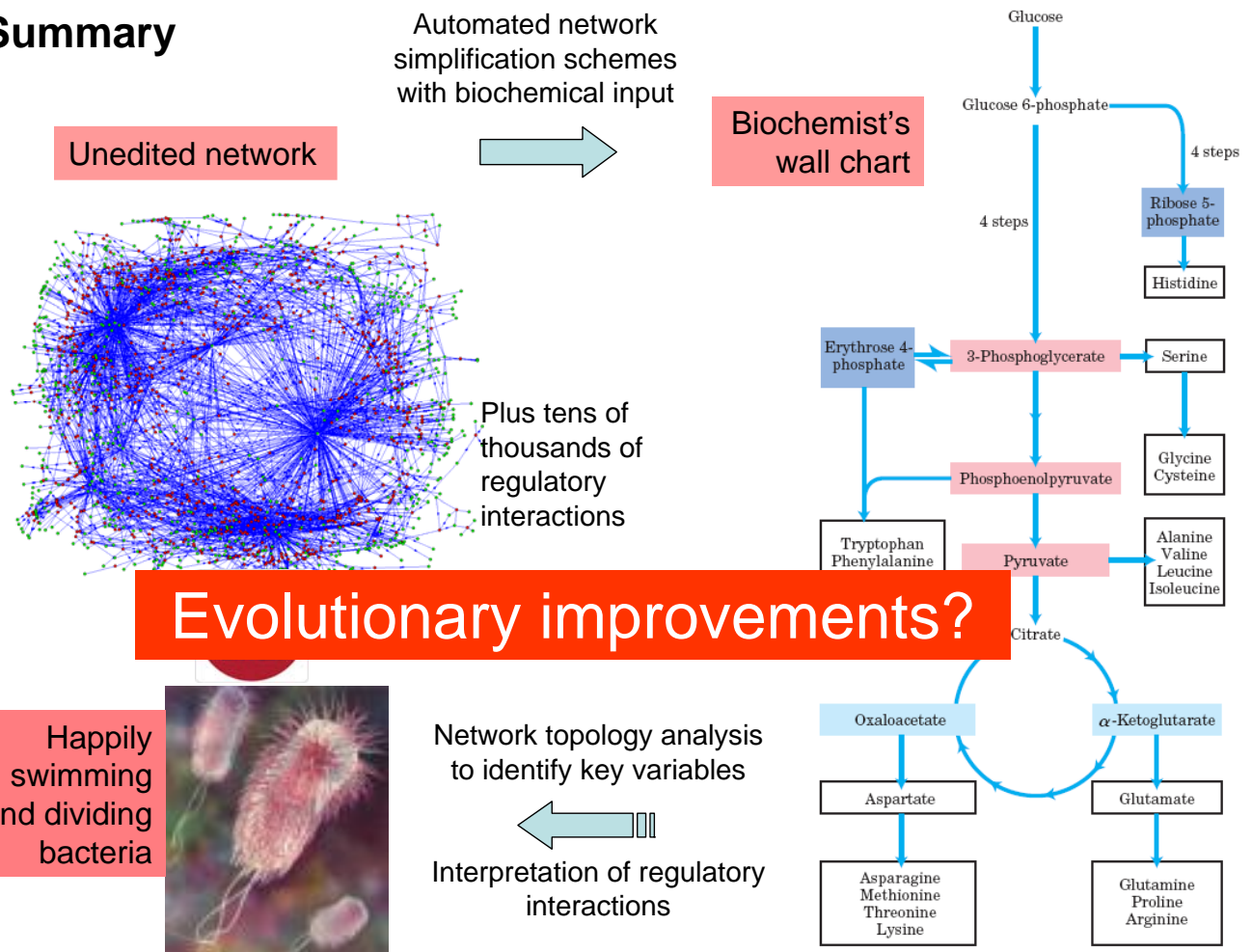
M Scott, C W Gunderson, E M Mateescu, Z Zhang & Terence Hwa, *Science* **330**:1099-102 (2010)

Protein synthesis = major consumer/expenditure of metabolism

Partitioning of protein content in a bacterial cell



Summary



The Team

Prof. Terry Hwa, UCSD

Dr. Pan-Jun Kim, KAIST
and UIUC



Supported by the RGC
of the HKSAR